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ORIGINAL RESEARCH

Development and Validation of a Risk Prediction Model Based on Inflammatory and Nutritional Composite Indicators for Posthepatectomy Liver Failure Following Radical Resection of Hepatocellular Carcinoma

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Purpose: A plethora of studies have demonstrated an association between preoperative inflammatory immunonutritional status and the prognosis of patients with hepatocellular carcinoma. Nonetheless, there is a paucity of research examining the predictive value of inflammatory immunonutritional indicators for postoperative liver failure in this patient population. This study seeks to identify independent predictors of post hepatectomy liver failure (PHLF) in patients with hepatocellular carcinoma and to develop a nomogram model.

Patients and Methods: Clinical data were collected from 760 patients diagnosed with hepatocellular carcinoma who underwent surgical treatment at a hospital in China between January 2020 and January 2024. The dataset was randomly divided into a training set (n=570, 75%) and a validation set (n=190, 25%). To identify independent predictors of PHLF in these patients, univariate analysis and least absolute shrinkage and selection operator (LASSO) regression were employed. Subsequently, a multivariate logistic regression model was developed to construct a predictive model. The predictive performance of the nomogram was evaluated using receiver operating characteristic (ROC) curve analysis, calibration curve assessment, and decision curve analysis (DCA).

Results: AAPR, ALBI, GAR, LMR, PNI, INR, APTT, and TT are independent factors associated with PHLF in patients with hepatocellular carcinoma. The C indices for the training and validation datasets were 0.691 (95% CI: 0.634–0.747) and 0.680 (95% CI: 0.556–0.804), respectively. The area under the curve (AUC) and calibration curve analyses demonstrated the nomogram's accuracy in predicting PHLF in this patient population. Furthermore, DCA indicated that the model provides a significant clinical net benefit. A comparison was made of the predictive efficacy of the nomogram prediction model and the associated composite liver function score. ROC curves were plotted for the nomogram prediction model, Child-Pugh score and ALBI score, and AUC values were calculated, which were 0.686 (95% CI 0.635–0.737) for the prediction model, 0.558(95% CI 0.512–0.603) for the Child-Pugh score. The AUC for ALBI score was 0.577 (95% CI 0.530–0.624), indicating that this nomogram prediction model was more effective than other scoring systems in predicting the study population in our center. In this study population, the nomogram model demonstrated an AUC of 0.707 (95% CI 0.620–0.794) for Child-Pugh score grade A and 0.572 (95% CI 0.501–0.643) for Child-Pugh score grade B. For tumors with a diameter of less than 5 cm, the AUC was 0.679 (95% CI 0.608–0.749), and for patients with tumors with a diameter of at least 5 cm, the AUC was 0.715 (95% CI 0.643–0.787).

Conclusion: We have developed an innovative nomogram model designed to predict the incidence of PHLF in patients diagnosed with hepatocellular carcinoma. This nomogram has a good predictive value for PHLF in HCC patients and is important for clinicians to manage patients after hepatectomy.

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Keywords: posthepatectomy liver failure, radical resection of hepatocellular carcinoma, inflammatory and nutritional composite indicators, nomogram

Introduction

Hepatocellular carcinoma (HCC) represents the most prevalent primary solid neoplasm of the liver, with its incidence continuing to escalate over recent decades. Between 2010 and 2021, the global incidence and mortality rates of HCC increased by 26% and 25%, respectively.¹ Currently, a range of therapeutic modalities are available for the management of HCC, encompassing surgical intervention, chemotherapy, targeted therapy, and immunotherapy. However, surgical resection remains the most widely acknowledged treatment associated with favorable prognostic outcomes.² Hepatectomy is frequently linked to various postoperative risks and complications, such as infection, hemorrhage, bile leakage, and PHLF. Despite ongoing advancements in surgical techniques, perioperative management, and surgeon expertise, the incidence of PHLF following hepatectomy has diminished, however, it continues to be a predominant cause of postoperative mortality in patients with HCC.³ In 2011, the International Liver Surgery Study Group (ISGLS) introduced diagnostic criteria for PHLF, significantly enhancing clinicians' ability to diagnose and manage this condition. Nevertheless, the diagnosis of PHLF often remains uncertain until the fifth postoperative day, at which point the efficacy of therapeutic interventions is significantly diminished due to the disease's progression. Considering the elevated mortality risk associated with PHLF and the paucity of effective treatment modalities, the prevention of PHLF continues to be a primary objective in contemporary therapeutic approaches.⁴

In the mid-19th century, Rudolf Virchow initially hypothesized the intricate interconnections between inflammation and cancer. This hypothesis was based on his observations of cancer arising from sites of chronic inflammation and the prevalence of inflammatory cells in tumor biopsies.⁵ The association between chronic inflammation and tumorigenesis is now well-established, with cancer-related inflammation acknowledged as a fundamental characteristic of cancer.⁶ Emerging evidence suggests that inflammation is a critical factor in the pathogenesis of cirrhosis and hepatocellular carcinoma, and it is strongly linked to adverse outcomes following hepatectomy.^{7,8} Hepatic TIMP3 deficiency has been shown to trigger lymphocyte infiltration and hepatocyte death by enhancing TNF-α converting enzyme activity, leading to sustained activation of the TNF signalling pathway. This persistent inflammatory microenvironment has been demonstrated to inhibit liver regeneration and accelerate functional failure of residual liver tissue.⁹ Furthermore, preoperative nutritional status has been identified as a significant prognostic indicator for patients with advanced liver disease.¹⁰ There appears to be a specific correlation between the inflammatory state and nutritional status of patients with hepatocellular carcinoma prior to hepatectomy and the incidence of hepatic failure following surgery. Building upon the aforementioned evidence, certain researchers have initiated investigations into the relationship between inflammatory and nutritional indicators and PHLF.¹¹ However, the prognostic significance of these indicators in predicting liver failure following hepatectomy remains inadequately elucidated. Consequently, the potential of inflammatory and nutritional indices as predictive tools for PHLF warrants further investigation.

Given that a single blood marker is insufficient to comprehensively represent the inflammatory and nutritional status of patients, there has been an increasing emphasis on complex indicators in clinical practice due to their comprehensive nature, predictive efficiency, and stability. Consequently, this study aims to predict the likelihood of PHLF in patients with hepatocellular carcinoma following partial hepatectomy by utilizing complex indicators of inflammation and nutrition.¹² Furthermore, a nomogram was developed based on these complex indicators to enhance predictive accuracy.

Materials and Methods

Data Source

Clinical data were collected from 760 patients diagnosed with hepatocellular carcinoma who underwent partial liver resection at the First Affiliated Hospital of the University of Science and Technology of China between January 2020 and January 2024. Prior to surgery, all patients received comprehensive communication and provided informed consent. The

collection of clinical data for this study received approval from the Medical Research Ethics Committee of the First Affiliated Hospital of the University of Science and Technology of China (approval number: 2024-RE-279).

Patient Admission Standard

The inclusion criteria for this study are: (1) individuals aged over 18 years; (2) a confirmed diagnosis of HCC through pathological examination; (3) the surgical intervention was a partial hepatectomy. The exclusion criteria are: (1) a history of concurrent malignant tumors; (2) the presence of extrahepatic metastasis; (3) Serious extrahepatic diseases (eg, severe cardiovascular disease, active autoimmune disease); (4) incomplete clinical data.

Data Collection and Variable Definition

The collected data encompassed three primary categories: (1) demographic characteristics, including age, gender, body mass index (BMI), and underlying conditions such as diabetes, hypertension, and coronary heart disease; liver function-related data, comprising hepatitis B status, anti-hepatitis B virus therapy, presence of ascites, Child-Pugh score, and cirrhosis; and perioperative laboratory results, which included measurements of red blood cells (RBC), white blood cells(WBC), platelets(PLT), absolute neutrophil count(ANC), hemoglobin(HGB), lymphocytes, monocytes, creatinine(Cr), alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase, gamma-glutamyl transferase(GGT), total bilirubin(TBil), direct bilirubin, indirect bilirubin, total protein(TP), albumin, globulin, and blood glucose(BG), activated partial thromboplastin time (APTT), international normalized ratio (INR), prothrombin time (PT), fibrinogen (FIB), and thrombin time (TT). (2) Intraoperative variables, such as surgery time, hepatic portal blockade time(HPT), intraoperative blood loss, and the volume of intraoperative blood transfusion. (3) Tumor-related characteristics, including tumor size (TS), microvascular Invasion (MVI), and alpha-fetoprotein (AFP) levels. (4) The study also focused on 11 biomarkers associated with immunity, inflammation, and nutrition: ALT (7-56 U/L), AST (10-40 U/L), albumin-alkaline phosphatase ratio (AAPR)(0.4–0.8), albumin-globulin ratio (A/G)(1.1–2.50), albumin-bilirubin (ALBI) grade, and gamma-glutamyl transferase-albumin ratio (GAR), neutrophil to lymphocyte ratio (NLR)(1-3), platelet to lymphocyte ratio (PLR)(50–150), prognostic nutritional index (PNI)(≥45), systemic immunoinflammatory index (SII) (300-900), lymphomononuclear ratio (LMR)(2.2-3.5). Biomarkers are calculated as follows: AAPR = albumin (g/ L)/ALP (IU/L), AG= albumin (g/L)/ globulin (g/L), ALBI= log10 bilirubin (mol/L) \times 0.66 - albumin (g/L) ×0.085, GAR= GGT (U/L)/ albumin (g/ l), NLR = ANC (×10⁹/L)/ absolute value of lymphocytes (×10⁹/L), PLR = peripheral blood plate count ($\times 10^{9}/L$)/ lymphocyte count ($\times 10^{9}/L$), PNI = 5× (serum albumin (g/dL)/ lymphocyte count $(\times 10^9/L)$ +100, SII = peripheral blood plate count $(\times 109/L)$ ×ANC $(\times 10^9/L)$ /lymphocyte absolute value ($\times 10^{9}/L$), LMR= lymphocyte count ($\times 10^{9}/L$)/absolute value of monocytes ($\times 10^{9}/L$). ALBI grades are divided according to cutoff values, as previously mentioned: <-2.60 (ALBI Grade 1), >-2.60 to <-1.39 (ALBI Grade 2), and \geq -1.39 (ALBI Grade 3).

PHLF Definition

Due to challenges associated with data retrieval and clinical practice, certain patients do not undergo concurrent assessments of TBil and INR on or after the fifth postoperative day. To address potential analytical biases, The incidence of PHLF is characterised on the basis of the ISGLS and in the light of the relevant literature.^{13,14}

The specific diagnostic criteria for PHLF are the fifth day after surgery and include the following: TBil value > 24 μ mol/L and INR > 1.2, or TBil > 70.1 μ mol/L (4.1 mg/dL), or INR > 2.5, or ascites drainage flow > 500 mL/ day. In addition, patients who were discharged within 5 days of surgery were classified as not having liver failure, except for patients who were discharged or died due to bleeding.

Hepatectomy Definition

Hepatic resection is a surgical intervention that involves the partial or total removal of liver tissue. It is primarily utilised in the management of primary liver tumours, such as hepatocellular carcinoma, as well as metastatic liver cancer, including cases of liver metastasis from colorectal cancer. The procedure is also employed in the treatment of benign liver tumours, liver injury, and other hepatic diseases. Depending on the extent of the procedure, it can be classified into two categories: partial hepatectomy (eg segmental hepatectomy, wedge resection) and extensive hepatectomy (eg hemihepatectomy, extended hemihepatectomy). The core objective is to achieve radical resection of the diseased tissue while ensuring that the remaining liver function is sufficient to maintain metabolic requirements.¹⁵ In this study, the tumours of patients with HCC who underwent hepatectomy were found to meet the Milan criteria (ie a solitary tumour \leq 5 cm in diameter or a maximum of three nodules, each \leq 3 cm).¹⁶

Statistical Analysis

The training group (75% of the sample size) was used to construct the prediction model, and the validation group (25% of the sample size) was used to verify the accuracy of the model. Continuous variables were expressed as median with quartile, while categorical variables are shown as percentage. Continuous variables were compared using Student's t tests or the Mann–Whitney *U*-tests as appropriate, and Pearson's Chi-squared or Fisher's exact tests were used to compare categorical variables. Univariate logistic regression models and LASSO regression models were used to identify independent predictors of PHLF in the training cohort. The AUC of the ROC curve was calculated to assess the discrimination of the nomogram in the training group and the validation group.

Statistical analysis was performed using R version 4.1.3 software equipped with the "glmnet", "rms", and "rmda" packages and SPSS version 26.0. A P-value < 0.05 was statistically significant.

Results

Patient Baseline Data

As illustrated in Figure 1, the study comprised a total of 760 patients diagnosed with hepatocellular carcinoma who underwent hepatectomy, among whom 134 individuals (17.6%) developed PHLF. The cohort was randomly divided into a training set, consisting of 75% of the cases (n=570), and a validation set, comprising 25% of the cases (n=190). The mean age of patients in the training set was 58.91 ± 10.85 years, with a gender distribution of 486 males (85.3%) and 84 females (14.7%). Similarly, the mean age of the validation set was 58.95 ± 10.09 years, with 153 males (80.5%) and 37 females (19.5%). In the training and validation cohorts, the incidence rates of liver failure among patients with hepatocellular carcinoma who underwent partial hepatectomy were 18.9% and 13.7%, respectively. Statistical analysis revealed no significant differences between the training and validation cohorts (p > 0.05), suggesting that the baseline characteristics of the two groups were comparable (Table 1).



Figure I The flowchart of patient selection.

| Variable | Total (n=760) | Training (n=570) | Validation (n=190) | P value |
|--------------------------------------|------------------------|------------------------|------------------------|---------|
| Failure, n (%) | | | | 0.060 |
| No | 626(82.4) | 462(81.1) | 164(86.3) | |
| Yes | 134(17.6) | 108(18.9) | 26(13.7) | |
| Gender, n (%) | | | | 0.078 |
| Female | 121(15.9) | 84(14.7) | 37(19.5) | |
| Male | 639(84.1) | 486(85.3) | 153(80.5) | |
| Age (years) | 58.92±10.66 | 58.91±10.85 | 58.95±10.09 | 0.970 |
| BMI, n (%) | | | | 0.510 |
| 1 | 33 (4.3) | 25 (4.4) | 8(4.2) | |
| 2 | 390 (51.3) | 287 (50.4) | 103(54.2) | |
| 3 | 337 (44.4) | 258 (4.3) | 79(41.6) | |
| Hepatitis B | | | | 0.801 |
| No | 339(44.6) | 256(44.9) | 83(43.7) | |
| Yes | 421(55.4) | 314(55.1) | 107(56.3) | |
| ART*, n (%) | () | | | 0.669 |
| No | 559(73.6) | 417(73.2) | 142(74.7) | |
| Yes | 201(26.4) | 153(26.8) | 48(25.3) | |
| HBP*. n (%) | | () | | 0.701 |
| No. | 565(743) | 426(74 7) | 139(73.2) | |
| Yes | 195(25.7) | 144(25.3) | 51(26.8) | |
| Diabetes (n%) | 175(25.7) | 111(25.5) | 51(20.0) | |
| No | 674(88.7) | 503(88.2) | 171(90.0) | |
| Yes | 86(11.3) | 67(11.8) | 19(10.0) | |
| CHD* n (%) | 00(11.5) | 07(11.0) | 17(10.0) | 0 772 |
| No | 745(98.0) | 558(97.9) | 187(98.4) | 0.772 |
| Yos | 15(2.0) | 12(21) | 3(1.6) | |
| $\Delta A B C (\times 10^{9} / 1)$ | F 27+2 07 | F 41+2 09 | 5(1.0) E 22+2.02 | 0.249 |
| | 138 00 (134 00 149 00) | 129 00 (126 00 149 00) | J.2212.02 | 0.200 |
| | 130.00 (120.00,147.00) | 130.00 (120.00,140.00) | 136.00 (120.00,147.00) | 0.737 |
| $PPC(w10^{9}/L)$ | 4 40(4 02 4 77) | 4 20(4 02 4 77) | 130.30 (107.00,103.73) | 0.235 |
| $C_{\rm r} ({\rm wms}/{\rm L})$ | 4.40(4.02,4.77) | 4.39(4.03,4.77) | 4.42(4.01,4.80) | 0.895 |
| | 65./U±15.90 | 66.22±16.51 | 64.15±13.85 | 0.121 |
| | 28.00 (19.50, 44.73) | 28.85 (19.02, 46.00) | 26.10 (20.00, 36.00) | 0.210 |
| AST (IU/L) | 32.00 (24.00, 46.00) | 32.00 (23.45,47.22) | 31.00 (24.40,41.92) | 0.475 |
| 1 P*(g/di) | 68.86±6.84 | 69.00±6.58 | 68.41±7.54 | 0.298 |
| A/G | 1.41(1.26,1.64) | 1.46(1.26,1.65) | 1.43(1.28,1.61) | 0.317 |
| BG(mmol/L) | 5.48±1.65 | 5.51±1./3 | 5.39±1.35 | 0.414 |
| | 1.11(0.90,1.42) | 1.11(0.89,1.440 | 1.11(0.92,1.38) | 0.704 |
| PI(s) | 12./6±3.64 | 12.80±4.09 | 12.62±1.69 | 0.538 |
| | 1.05±0.13 | 1.05±0.12 | 1.05±0.13 | 0.621 |
| APTT(s) | 34.31±5.92 | 34.18±5.95 | 34.73±5.81 | 0.269 |
| Fib (g/L) | 2.76(2.31,3.37) | 2.77(2.33,3.48) | 2.73 (2.28,3.23) | 0.370 |
| TT(s) | 17.20(15.40,18.50) | 17.20(15.50,18.40) | 17.10(15.00,18.60) | 0.641 |
| HYD*, n (%) | | | | 0.567 |
| No | 739(97.2) | 554(97.2) | 185(97.4) | |
| Yes | 21(2.8) | 16(2.8) | 5(2.6%) | |
| Child-Pugh | | | | 0.358 |
| | 323(42.5) | 236(41.4) | 87(45.8) | |
| 2 | 434(57.1) | 332(58.2) | 102(53.7) | |
| 3 | 3(0.4) | 2(0.4) | I (0.5) | |

Table I Baseline Characteristics of Patients with Hepatocellular Carcinoma Treated Surgically

(Continued)

| Variable | Total (n=760) | Training (n=570) | Validation (n=190) | P value |
|---------------------|------------------------|-------------------------|-------------------------|---------|
| AAPR | 0.44(0.33,0.56) | 0.44(0.33,0.56) | 0.43(0.33, 0.56) | 0.851 |
| ALBI | | | | 0.967 |
| I | 439(57.8) | 329(57.7) | 110(57.9) | |
| 2 | 315(41.4) | 237(41.6) | 78(41.1) | |
| 3 | 6(0.8) | 4(0.7) | 2(1.1) | |
| GAR | 1.19(0.65,2.30) | 1.23(0.65, 2.41) | 1.08(0.65, 2.17) | 0.320 |
| NLR | 2.10(1.55,2.95) | 2.09(1.57,2.95) | 2.16(1.48,2.99) | 0.792 |
| PLR | 103.00 (76.52,140.95) | 102.88 (77.46,143.06) | 103.50 (71.53,136.14) | 0.296 |
| PNI | 116.32±7.55 | 116.29±7.56 | 116.40±7.53 | 0.865 |
| SII | 295.7 (192.63, 509.93) | 298.62 (196.99, 513.56) | 292.23 (179.00, 470.88) | 0.338 |
| LMR | 3.35(2.55,4.29) | 3.38(2.55,4.30) | 3.23(2.53,4.27) | 0.967 |
| Surgery time (/min) | 185.00 (139.00,240.00) | 185.00 (140.00, 241.75) | 178.50 (130.00, 238.75) | 0.169 |
| HPT*(/min) | 12.00(0.00,30.00) | 12.00(0.00,30.00) | 9.00(0.00,28.75) | 0.400 |
| Cirrhosis, n(%) | | | | 0.204 |
| No | 233(30.7) | 182(31.9) | 51(26.8) | |
| Yes | 527(69.3) | 388(68.1) | 139(73.2) | |
| IBL*(/mL) | 150.00 (100.00,300.00) | 150.00 (100.00,300.00) | 100.00 (50.00,300.00) | 0.122 |
| IBT*(/mL) | 0.00(0.00,300.00) | 0.00(0.00,300) | 0.00(0.00,275.00) | 0.842 |
| TS*(/cm) | 4.50(3.00,7.50) | 4.50(3.00,8.00) | 4.00(3.00,7.00) | 0.102 |
| AFP | | | | 0.241 |
| No | 361(47.5%) | 278(48.8%) | 83(43.7%) | |
| Yes | 399(52.5%) | 292(51.2%) | 107(56.3%) | |
| MVI | | | | 0.960 |
| 0 | 336(44.2) | 259(45.4) | 77(40.5) | |
| I | 286(37.6) | 205(36.0) | 81 (42.7) | |
| 2 | 138(18.2) | 106(18.6) | 32(16.8) | |

Table I (Continued).

Abbreviations: ART, antiretroviral therapy; HBP, high blood pressure; CHD, coronary heart disease; TP, total protein; HYD, hydroperitoneum; HPT, hepatic portal blockade time; IBL, Intraoperative blood loss; IBT, Intraoperative blood transfusion; TS, Tumor size.

Single Factor Logistic Regression Analysis

Using univariate logistic regression analysis, several factors were identified as significant risk factors for PHLF. These factors include AST (P = 0.006, OR = 1.008, 95% CI: 1.002–1.013), INR (P = 0.001, OR = 13.359, 95% CI: 2.718–65.657), APTT (P = 0.001, OR = 1.063, 95% CI: 1.025–1.103), TT (P = 0.028, OR = 1.119, 95% CI: 1.012–1.237), and the Child-Pugh score (P = 0.047, OR = 1.553, 95% CI: 1.007–2.397). Additionally, AAPR (P < 0.001, OR = 0.048, 95% CI: 0.013–0.180), ALBI (P = 0.001, OR = 2.041, 95% CI: 1.358–3.066), GAR (P = 0.005, OR = 1.073, 95% CI: 1.022–1.127), PLR (P = 0.018, OR = 1.003, 95% CI: 1.001–1.006), PNI (P = 0.015, OR = 1.031, 95% CI: 1.006–1.058), and LMR (P < 0.001, OR = 0.717, 95% CI: 0.603–0.852) were also identified as significant risk factors. For detailed results, refer to Table 2. A 10-fold cross-validation of the initial input LASSO regression method was conducted to address collinearity among the relevant indicators, thereby identifying the predictors of postoperative liver failure in patients with hepatocellular carcinoma who underwent partial hepatectomy. Ultimately, eight variables were selected as optimal based on the best lambda value: AABR, ALBI, GAR, LMR, PNI, INR, APTT, and TT (Figure 2).

Development of Nomogram in Training Set

Eight factors, including AABR, ALBI, GAR, PNI, LMR, INR, APTT, and TT, were selected as predictors to develop a forecasting model for PHLF in patients undergoing partial liver resection for HCC. The resulting nomogram, depicted in Figure 3, indicates that AAPR and LMR significantly impact patient outcomes. By summing the points assigned to

| Variables | OR | Lower | Upper | Ρ |
|---------------------------|----------------|----------------|----------------|----------------|
| Genders (n%) | 0.754 | 0.400 | 1.418 | 0.381 |
| Age (years) | 1.009 | 0.990 | 1.029 | 0.353 |
| BMI | 1.105 | 0.765 | 1.595 | 0.595 |
| Hepatitis B (n%) | 0.933 | 0.613 | 1.421 | 0.748 |
| ART* (n%) | 0.836 | 0.514 | 1.360 | 0.471 |
| HBP* (n%) | 1.044 | 0.647 | 1.685 | 0.860 |
| Diabetes* (n%) | 0.725 | 0.357 | 1.471 | 0.373 |
| CHD* (n%) | 0.853 | 0.184 | 3.950 | 0.839 |
| WBC (x10 ⁹ /L) | 1.080 | 0.985 | 1.185 | 0.103 |
| HGB (g/l) | 1.002 | 0.999 | 1.006 | 0.226 |
| PLT (x10 ⁹ /L) | 1.000 | 0.997 | 1.002 | 0.813 |
| RBC (x10 ⁹ /L) | 0.824 | 0.635 | 1.069 | 0.144 |
| Cr (umol/L) | 1.001 | 0.989 | 1.014 | 0.851 |
| ALT (IU/L) | 1.004 | 0.998 | 1.009 | 0.195 |
| AST (IU/L) | 1.008 | 1.002 | 1.013 | 0.006 |
| TP*(g/dl) | 0.993 | 0.962 | 1.025 | 0.655 |
| A/G | 0.238 | 0.111 | 0.508 | 0.000 |
| BG*(mmol/L) | 1.030 | 0.918 | 1.155 | 0.617 |
| ALT/AST | 1.117 | 0.857 | 1.455 | 0.414 |
| PT(s) | 1.014 | 0.972 | 1.059 | 0.513 |
| INR | 13.359 | 2.718 | 65.657 | 0.001 |
| APTT(s) | 1.063 | 1.025 | 1.103 | 0.001 |
| FIB (g/L) | 1.083 | 0.894 | 1.312 | 0.416 |
| TT(s) | 1.119 | 1.012 | 1.237 | 0.028 |
| HYD* | 1.442 | 0.456 | 4.562 | 0.533 |
| Surgery time(min) | 1.000 | 0.998 | 1.003 | 0.855 |
| HBT(min) | 1.004 | 0.995 | 1.013 | 0.372 |
| Cirrhosis | 1.142 | 0.723 | 1.803 | 0.569 |
| IBL(mL) | 1.000 | 1.000 | 1.001 | 0.352 |
| IBT(mL) | 1.000 | 1.000 | 1.000 | 0.633 |
| Tumor size(cm) | 0.968 | 0.915 | 1.024 | 0.256 |
| MVI | 1.015 | 0.792 | 1.301 | 0.906 |
| AFP | 0.682 | 0.447 | 1.041 | 0.076 |
| Child Pugh | 1.553 | 1.007 | 2.397 | 0.047 |
| AAPR | 0.048 | 0.013 | 0.180 | 0.000 |
| ALBI | 2.041 | 1.358 | 3.066 | 0.001 |
| GAR | 1.073 | 1.022 | 1.127 | 0.005 |
| NLR | 1.035 | 0.985 | 1.088 | 0.169 |
| | 1.003 | 1.001 | 1.006 | 0.018 |
| PLR | 1.005 | | | |
| PLR PNI | 1.031 | 1.006 | 1.058 | 0.015 |
| PLR PNI SII | 1.031 1.000 | 1.006 1.000 | 1.058 1.001 | 0.015 0.069 |

Table 2UnivariateAnalysisofPatientswithHepatocellularCarcinomaTreatedSurgically in theTrainingCohort

Abbreviations: ART, antiretroviral therapy; HBP, high blood pressure; CHD, coronary heart disease; TP, total protein; HYD, hydroperitoneum; HPT, hepatic portal blockade time; IBL, Intraoperative blood loss; IBT, Intraoperative blood transfusion; TS, Tumor size.



Figure 2 Clinical feature selection by LASSO. (A) Plot of LASSO coefficient profiles of the 12 features. The log (lambda) sequence was plotted against a coefficient profile plot. There were 8 features with non-zero coefficients generated by the ideal lambda (λ =2.539628921009624); (B) 10-fold cross-validation for LASSO model parameter adjustment. The binomial deviation curve was displayed with log (lambda). The minimum criteria and its one standard error were used to construct dotted vertical lines at the optimal values (the 1-SE criteria).

each variable along the vertical line and referencing the point axis, the likelihood of a patient experiencing PHLF can be estimated.

Model Verification

The concordance index (C-index) for the training and validation datasets was 0.691 (95% CI: 0.634–0.747) and 0.680 (95% CI: 0.556–0.804), respectively, aligning with the results of the ROC curve analysis (Figure 4). These findings indicate that the nomogram model serves as an effective predictor of PHLF) incidence in patients with HCC. Furthermore, calibration curves for both the training and validation cohorts demonstrate that the predicted probabilities closely match the observed outcomes, signifying successful model calibration (Figure 5). As illustrated in Figure 6, DCA demonstrates that the nomogram exhibits an exceptional overall net benefit across a wide and practical range of threshold probabilities, suggesting a significant potential for clinical utility.



Figure 3 Nomogram model for predicting PHLF in patients with HCC.

Prob of S-AKI ?

Comparison of the Predictive Efficacy of Column-Line Graphical Prediction Models and Associated Liver Function Composite Scores

0.1

0.3

0.5

0.7

In this study, the predictive efficacy of the nomogram model was compared with that of the associated liver function composite score. The ROC curves for the nomogram prediction model, Child-Pugh score and ALBI score were plotted, and the AUC values were calculated. The AUC of the nomogram model was 0.6863 (95% CI 0.63536–0.73715), the AUC of Child-Pugh grading was 0.5578 (95% CI 0.51249–0.60302), and the AUC of ALBI scoring was 0.5774 (95% CI 0.530 37–0.62449). The results illustrated that both the aforementioned column line drawing prediction model and the results indicated that the above nomogram prediction model and scoring system had predictive value for PHLF, and the AUC value of this nomogram prediction model was larger compared with Child-Pugh score (NRI=0.4474, P<0.001) and ALBI score (NRI=0.5061, P<0.001). The comparison of the ROC curves provides a more intuitive illustration of the differences between the scoring models, thereby suggesting that this nomogram model is more effective than other scoring systems in predicting the study population in our center (see Table 3 and Figure 7).

Comparison of Prediction Performance of Nomogram Prediction Models Under Different Populations

The available data population was divided into Child-Pugh grade A (n=323) and Child-Pugh grade B(n=434) for the prediction of the model, respectively, where the AUC for Child-Pugh grade A was 0.7070 ((95% CI 0.62003-0.79400), and the AUC for Child-Pugh grade B was 0.572 (95% CI 0.50193-0.64345). The population was divided into two groups for the purpose of prediction by the model: those with tumor diameters less than 5cm (n=405) and those with tumor diameters greater than or equal to 5cm (n=355). The AUC for the former group was 0.6785 (95% CI 0.60784-0.74914),



Figure 4 The area under the ROC curves AUCs of the nomogram for mortality from PHLF in patients with HCC in training set (A) and validation set (B).



Figure 5 Calibration curves of the predicted nomogram. Evaluation of the predictive performance for mortality from PHLF in patients with HCC of the nomogram in the training set (\mathbf{A}) and validation set. (\mathbf{B}) .



Figure 6 Decision curve analysis of the nomogram. (A) Decision curve analysis in the training set; (B) Decision curve analysis in validation set.

| | C , | | | | |
|---------------------------------|----------------------------|----------------------------|---|------------------|------------------|
| | AUC | Standard Error | 95% CI | NRI | P |
| New model Child-Pugh ALBI | 0.6863 0.5578 0.5774 | 0.0260 0.0231 0.0240 | 0.63536–0.73715 0.51249–0.60302 0.53037–0.62449 | 0.4474 0.5061 | <0.001 <0.001 |

Table 3 PHLF Prediction Model with Associated Scoring System AUC

and that for tumor diameters greater than or equal to 5cm was 0.7153 (95% CI 0.64339–0.78730), see Table 4. It is evident that the model demonstrates a certain degree of prediction performance across all populations, however, the prediction performance is more pronounced in the Child-Pugh grade A and tumor diameter greater than or equal to 5cm population. The predictive performance of the nomogram model can be evaluated more intuitively by the ROC curve (Figure 8).

Discussion

In recent years, notwithstanding the ongoing advancements in medical technology, PHLF remains a significant fatal complication following liver cancer surgery. Current reports indicate that the incidence of postoperative hepatocellular carcinoma complicated by liver failure ranges from 3% to 48.5%, with most studies reporting an incidence of approximately 10% to 20%.^{17,18} The substantial variability in PHLF incidence may be attributed to differences in study design, patient selection, surgical techniques, and varying interpretations of the definition of liver failure. In this study, the incidence of PHLF was 17.6%, aligning with rates reported in the majority of existing literature. Currently, the diagnostic criteria for PHLF predominantly follow the guidelines established by the ISGLS in 2011. According to these criteria, PHLF is diagnosed when, on or after the fifth postoperative day, a patient's INR and TBIL levels exceed the normal range as defined by the local laboratory.¹³ According to the established diagnostic criteria, PHLF is typically diagnosed on or after the limitations of conventional indicators in assessing liver failure following hepatectomy, there is an ongoing pursuit for accessible and reliable predictors associated with PHLF. Recent studies have demonstrated that immune, inflammatory, and nutritional indicators are significant predictors of prognostic risk in solid tumors, such as HCC,¹⁹ nasopharyngeal carcinoma,²⁰ and pancreatic cancer.²¹ This research involved comprehensive collection of biomarkers associated with immunity, inflammation, and nutrition from preoperative peripheral blood tests



Figure 7 Comparison of PHLF prediction model and related scoring system ROC curves.

| n | AUC | Standard Error | 95% CI |
|-----|-------------------------------|---|--|
| 323 | 0.7070 | 0.0444 | 0.62003–0.79400 |
| 434 | 0.5727 | 0.0361 | 0.50193-0.64345 |
| 405 | 0.6785 | 0.0360 | 0.60784-0.74914 |
| 355 | 0.7153 | 0.0367 | 0.64339–0.78730 |
| | n 323 434 405 355 | n AUC 323 0.7070 434 0.5727 405 0.6785 355 0.7153 | n AUC Standard Error 323 0.7070 0.0444 434 0.5727 0.0361 405 0.6785 0.0360 355 0.7153 0.0367 |

Table 4 Comparison of Predictive Performance of PredictiveModels in Different Populations

of HCC patients, and subsequently calculated composite indicators pertaining to these domains. To the best of our knowledge, this study represents the first comprehensive assessment of the prognostic significance and clinical relevance of a combination of immune, inflammatory, and nutritional markers in predicting PHLF following hepatocellular carcinoma resection. In contrast to conventional predictive indicators, the nomogram employed in this study emphasizes the integration of composite biological markers, thereby providing a more holistic evaluation of the preoperative status of patients with liver cancer. Our results showed that AAPR, ALBI, GAR, PNI, LMR, INR, APTT, TT was associated with a significant PHLF occurred postoperatively in patients with HCC.

Among the various clinical scoring systems, the Child-Pugh scoring system is the most frequently utilized. However, it includes two subjective parameters, hepatic encephalopathy and ascites, which present certain limitations.²² In response to these limitations, Johnson²³ introduced the ALBI scoring system, which offers a more objective assessment compared to the Child-Pugh system, with easily obtainable indicators. Numerous studies have validated the ALBI score as a significant prognostic tool for hepatocellular carcinoma surgery.^{24,25} In this study, the truncation values of the ALBI score were determined to be -0.260 and -0.139, indicating that an elevated ALBI score is associated with an increased risk of postoperative PHLF. Qin²⁶ similarly identified that an ALBI score of grades 2 or higher serves as an independent risk factor for PHLF. The AAPR was introduced by Anthony²⁷ in 2015 as a novel inflammatory marker, combining



Figure 8 The area under the ROC curves AUCs of the nomogram for mortality from PHLF in patients with HCC in Child-Pugh grade A (A), Child-Pugh grade B (B), tumor diameters less than 5cm (C), tumor diameters greater than or equal to 5cm (D).

serum albumin and alkaline phosphatase levels. Their research demonstrated that a lower AAPR value correlates with reduced overall survival in patients with hepatocellular carcinoma.

Numerous studies have corroborated that a low AAPR serves as an independent risk factor for HCC patients undergoing hepatectomy, with diminished preoperative AAPR levels often correlating with reduced overall survival and relapse-free survival rates.^{27,28} The present study demonstrated that the incidence of PHLF escalates as AAPR decreases, aligning with findings from existing research. Although the direct association between AAPR and PHLF lacks unequivocal empirical support, the components of AAPR, namely albumin and alkaline phosphatase, are integral to the prognostic evaluation of liver diseases, including possible association with PHLF. The AAPR, serving as a comprehensive marker of both inflammation and nutritional status, may represent a viable preoperative predictor for PHLF. Lymphocytes constitute a fundamental component of the immune system and play a crucial role in the defense mechanisms within the microenvironment where hepatocellular carcinoma develops, particularly in the context of chronic inflammation induced by factors such as hepatitis B or C virus, alcohol consumption, or metabolic syndrome.²⁹ Furthermore, a correlation between the abundance of tumor-infiltrating lymphocytes and the prognosis of hepatocellular carcinoma has been established.³⁰ Furthermore, inflammation is crucial in regulating liver regeneration, a process vital for the restoration of liver function following hepatectomy.³¹ The LMR, serving as an indicator of the balance between inflammatory response and immune status, may be associated with the incidence of PHLF. This study finds that a low LMR is more likely to contribute to the occurrence of PHLF, aligning with existing research findings.³² The mechanism by which preoperative patient inflammatory markers contribute to PHLF remains to be elucidated. However, extant research suggests that numerous inflammatory cytokines play a significant role in the anabolism of liver cells. However, extant research has demonstrated that a multitude of inflammatory cytokines exert a pivotal function in the anabolism of liver cells. For instance, transforming growth factor β (TGF- β) has been observed to inhibit albumin production in normal human hepatocytes and hepatocellular carcinoma HepG2 cells, achieving a decrease in albumin mRNA levels of 2-4-fold. Furthermore, lipopolysaccharide (LPS)-induced signalling activation and an increase in NF-kB activity have been shown to significantly reduce albumin expression.^{33,34} Moreover, it is evident that the process of hepatic fibrosis may be precipitated by chronic inflammation. This is characterised by the activation of hepatic stellate cells (HSCs) by inflammatory cells, such as macrophages and neutrophils. Consequently, these HSCs undergo a transformation into myofibroblasts, which in turn secrete excess extracellular matrix, ultimately resulting in hepatic fibrosis.³⁵ The complex interplay of fibrosis, inflammation and progressive cellular damage that is characteristic of chronic disease fundamentally impairs liver regeneration due to the destruction of tissue structure.³⁶ It is also possible that this is related to the occurrence of PHLF.

Patients with advanced chronic illnesses frequently experience malnutrition and are unable to achieve sufficient nutritional intake solely through oral consumption. Liver disease is no exception, with nutritional status recognized as a prognostic indicator for individuals with advanced liver disease.^{10,37} Regrettably, the nutritional challenges faced by patients with chronic liver disease are often underestimated, and comprehensive pre-surgical nutritional assessments are frequently neglected. Nutritional therapeutic interventions for patients with chronic liver disease are frequently underutilized. Serum albumin accounts for more than 50% of total serum protein in healthy individuals and is a marker of the liver's ability to synthesise it, as well as a major indicator of human nutrition. It is widely acknowledged that low serum albumin levels are a significant predictor of complications, progression, survival and recurrence in a variety of tumours, including those that develop in the liver.³⁸ Albumin deficiency has been demonstrated to result in an excessive inflammatory response.³⁹ In murine models of acetaminophen-induced hepatitis, albumin fusion has been demonstrated to ameliorate hepatic redox and inflammatory conditions, thereby suggesting that serum albumin possesses antioxidant and anti-inflammatory properties.⁴⁰ Albumin modulates the immune and inflammatory response through binding lipopolysaccharide and bacterial products, reactive oxygen species, nitric oxide, prostaglandins, and endosomal TLR signaling.⁴¹ Consequently, the infusion of serum albumin may represent a novel therapeutic approach to prevent systemic inflammatory response, PHLF and postoperative mortality. Nutritional indicators in this study were also calculated based on albumin. Numerous studies have demonstrated that PNI serves as a potential prognostic indicator in various cancers, including liver, stomach, ovarian, and lung cancers.^{42–44} In the present study, PNI exhibits a degree of predictive value for PHLF. This is very important, and early nutritional therapy for patients with hepatocellular carcinoma may help reduce the occurrence of PHLF, and it is worth further exploring the predictive value of PNI in PHLF. GGT mainly exists on liver cell membrane and microsomes, and is often increased due to liver cell inflammation when liver parenchyma is compressed.⁴⁵ The GGT in serum mainly comes from liver. In recent years, GGT has gradually been recognized as an independent prognostic indicator related to tumors, including urinary system tumors and liver cancer.^{46,47} Recent studies have also shown that the higher the serum GGT level, the worse the prognosis of HCC patients.⁴⁸ Originating in the liver, albumin is the most dominant protein in plasma, accounting for about 50% of total plasma protein, and has been advocated as a marker of the nutritional status of individual patients. The reasons for the low level of albumin before operation may include: less albumin synthesis and secretion due to liver dysfunction; Tumor-related inflammatory responses lead to accelerated protein breakdown.⁴⁹ Patients with higher albumin levels tend to have better postoperative recovery speed and prognosis.⁵⁰ In this study, GAR was the ratio of GGT to albumin, and as the ratio increased, patients' risk of developing PHLF also increased, that is, high GGT and low albumin levels would have a worse prognosis, which is consistent with the current study.

The liver serves as the primary site for the synthesis of most clotting factors, as well as several proteins involved in fibrinolysis and anticoagulation. Chronic liver disease can substantially impact the synthesis of these factors, consequently affecting the systemic levels of pro-coagulant and anticoagulant factors.⁵¹ Current research has demonstrated that, in addition to the liver's influence on clotting factor synthesis, the activity of these factors also diminishes as liver disease progresses.⁵² The APTT serves as a measure of coagulation function, specifically reflecting the intrinsic coagulation pathway and the overall activity of coagulation factors during the initial phase. Prolongation of APTT is observed in patients with impaired hepatic function. The INR is another parameter of coagulation function, primarily utilized to assess the activity of vitamin K-dependent coagulation factors.⁵³ TT predominantly indicates alterations in the quantity or functionality of fibrinogen. The liver serves as the principal site for the synthesis of vitamin K-dependent clotting factors and fibrinogen. Consequently, impaired liver function leads to a reduction in the synthesis of these components, resulting in prolonged INR and TT. This study found that elevated APTT, INR, and TT were associated with an increased likelihood of developing PHLF. This association may be attributed to diminished liver function prior to surgery in patients exhibiting elevated levels of these indicators. The findings align with the established understanding that poorer preoperative liver function increases the risk of PHLF.

In this study, we constructed and validated a simple nomogram model based on patients' more readily available preoperative laboratory indicators, partially compounded and formed by correlation calculations, for predicting the postoperative development of PHLF in patients with HCC. The designed nomogram was validated and performed well in discrimination, calibration and clinical application. In addition, the nomogram provided valuable information for determining the appropriate treatment regimen for high-risk patients who may develop PHLF. Here we give an example of how to use the nomogram model, assuming a patient with hepatocellular carcinoma who is proposed to undergo hepatectomy, with an AAPR of 0.25 g/IU, ALBI of grade 3, GAR of 20 U/g, PNI of 170, LMR of 2, INR of 1.1, APTT of 35s, TT of 20s, according to Figure 3, obtaining a line graph with the score corresponding to each parameter on the "point" axis was obtained according to Figure 3. The total score was calculated as the sum of the scores for all parameters [70 (AAPR) + 10 (ALBI) + 10 (GAR) + 30 (PNI) + 87.5 (LMR) + 12.5 (INR) + 40 (APTT) + 18 (TT) = 268]. This score corresponds to a 65% risk of developing PHLF. In this study, we sought to ascertain the comparative performance of the nomogram with existing liver function scoring systems. The results demonstrated the nomogram to be superior in terms of predictive performance. Furthermore, a subgroup analysis of the existing population was conducted, which revealed that the nomogram exhibited superior predictive capabilities in Child-Pugh score grade A (AUC=0.707, 95CI 0.620–0.794) and tumor diameter size \geq 5cm (AUC=0.715, 95CI 0.643–0.788).

Limitations

The present study is subject to certain limitations. Firstly, although the patient sample size was large, it was not validated using an external dataset. Secondly, a large number of potential factors affecting postoperative complications in hepatic failure in patients with primary hepatocellular carcinoma may have been overlooked, and this Nomogram may have failed to include other important risk variables. Finally, as this was a single-centre study, further validation may be required to ensure the generalisability of the Nomogram.

Conclusion

In conclusion, this study utilized eight preoperative indices—AAPR, ALBI, GAR, PNI, LMR, INR, APTT, TT—related to immunity, inflammation, nutrition, and blood coagulation to develop a nomogram for predicting postoperative PHLF in patients with hepatocellular carcinoma. This nomogram has a good predictive value for PHLF in HCC patients and is important for clinicians to manage patients after hepatectomy.

Data Sharing Statement

The datasets generated and analyzed in this study will be available by the corresponding author upon reasonable request.

Ethics Statement

This study was conducted following the Declaration of Helsinki and was approved by the Ethical Committee of the First Hospital Affiliated to the University of Science and Technology of China. All patient data were analyzed in anonymity. Patient consent was waived by the ethics committee, as no individual data were published, nor was any intervention performed on patients.

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Author Contributions

All authors made significant contributions to the reported work, including conception, study design, execution, data acquisition, analysis, and interpretation. They also participated in drafting, revising, or critically reviewing the article, and gave final approval of the manuscript. Furthermore, they agreed on the journal to which the article would be submitted, and agreed to be accountable for all aspects of the work.

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Disclosure

The authors declare no conflicts of interest in this work.

References

- 1. Tan EY, Danpanichkul P, Yong JN, et al. Liver cancer in 2021: global burden of disease study. J Hepatol. 2024. doi:10.1016/j.jhep.2024.10.031
- Gundavda KK, Patkar S, Varty GP, Shah N, Velmurugan K, Goel M. Liver resection for hepatocellular carcinoma: recent advances. J Clin Exp Hepatol. 2025;15(1):102401. doi:10.1016/j.jceh.2024.102401
- 3. Sparrelid E, Olthof PB, Dasari BVM, et al. Current evidence on posthepatectomy liver failure: comprehensive review. BJS Open. 2022;6(6). doi:10.1093/bjsopen/zrac142
- 4. Rahnemai-Azar AA, Cloyd JM, Weber SM, et al. Update on liver failure following hepatic resection: strategies for prediction and avoidance of post-operative liver insufficiency. *J Clin Transl Hepatol*. 2018;6(1):97–104. doi:10.14218/jcth.2017.00060
- 5. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? Lancet. 2001;357(9255):539-545. doi:10.1016/s0140-6736(00)04046-0

6. Punt S, Dronkers EA, Welters MJ, et al. A beneficial tumor microenvironment in oropharyngeal squamous cell carcinoma is characterized by a high T cell and low IL-17(+) cell frequency. *Cancer Immunol Immunother*. 2016;65(4):393–403. doi:10.1007/s00262-016-1805-x

- Wu TJ, Chang SS, Li CW, et al. Severe hepatitis promotes hepatocellular carcinoma recurrence via NF-κB pathway-mediated epithelialmesenchymal transition after resection. *Clin Cancer Res.* 2016;22(7):1800–1812. doi:10.1158/1078-0432.Ccr-15-0780
- 8. Peng W, Zhang XY, Li C, Wen TF, Yan LN, Yang JY. Spleen stiffness and volume help to predict posthepatectomy liver failure in patients with hepatocellular carcinoma. *Medicine*. 2019;98(18):e15458. doi:10.1097/md.00000000015458
- 9. Kim G, Chen Z, Li J, et al. Gut-liver axis calibrates intestinal stem cell fitness. Cell. 2024;187(4):914-930.e20. doi:10.1016/j.cell.2024.01.001
- 10. Cheung K, Lee SS, Raman M. Prevalence and mechanisms of malnutrition in patients with advanced liver disease, and nutrition management strategies. *Clin Gastroenterol Hepatol.* 2012;10(2):117–125. doi:10.1016/j.cgh.2011.08.016
- 11. Fang T, Long G, Wang D, et al. A nomogram based on preoperative inflammatory indices and ICG-R15 for prediction of liver failure after hepatectomy in HCC patients. *Front Oncol.* 2021;11:667496. doi:10.3389/fonc.2021.667496
- 12. Mangoni AA, Zinellu A. The diagnostic role of the systemic inflammation index in patients with immunological diseases: a systematic review and meta-analysis. *Clin Exp Med*. 2024;24(1):27. doi:10.1007/s10238-024-01294-3

- 13. Rahbari NN, Garden OJ, Padbury R, et al. Posthepatectomy liver failure: a definition and grading by the international study group of liver surgery (ISGLS). *Surgery*. 2011;149(5):713-724. doi:10.1016/j.surg.2010.10.001
- 14. Wang X, Wang W, Lin X, et al. Inflammatory markers showed significant incremental value for predicting post-hepatectomy liver failure in hepatocellular carcinoma patients. *Life*. 2023;13(10). doi:10.3390/life13101990
- 15. Fong Y. Surgical therapy of hepatic colorectal metastasis. CA Cancer J Clin. 1999;49(4):231-255. doi:10.3322/canjclin.49.4.231
- 16. Fan ST. Hepatocellular carcinoma--resection or transplant? *Nat Rev Gastroenterol Hepatol*. 2012;9(12):732–737. doi:10.1038/nrgastro.2012.158 17. Chen X, Kuang M, Hu ZH, et al. Prediction of post-hepatectomy liver failure and long-term prognosis after curative resection of hepatocellular
- carcinoma using liver stiffness measurement. Arab J Gastroenterol. 2022;23(2):82–88. doi:10.1016/j.ajg.2022.01.001
 8. Marath K, Tiwari A, Court C, et al. Postonerative liver failure: definitions risk factors, prediction models and prevention strategies. I Gastroenterol.
- Merath K, Tiwari A, Court C, et al. Postoperative liver failure: definitions, risk factors, prediction models and prevention strategies. J Gastrointest Surg. 2023;27(11):2640–2649. doi:10.1007/s11605-023-05834-2
- 19. Wang C, He W, Yuan Y, et al. Comparison of the prognostic value of inflammation-based scores in early recurrent hepatocellular carcinoma after hepatectomy. *Liver Int.* 2020;40(1):229–239. doi:10.1111/liv.14281
- 20. Miao J, Xiao W, Wang L, et al. The value of the prognostic nutritional index (PNI) in predicting outcomes and guiding the treatment strategy of nasopharyngeal carcinoma (NPC) patients receiving intensity-modulated radiotherapy (IMRT) with or without chemotherapy. J Cancer Res Clin Oncol. 2017;143(7):1263–1273. doi:10.1007/s00432-017-2360-3
- 21. Fang L, Yan FH, Liu C, et al. Systemic inflammatory biomarkers, especially fibrinogen to albumin ratio, predict prognosis in patients with pancreatic cancer. *Cancer Res Treat*. 2021;53(1):131–139. doi:10.4143/crt.2020.330
- 22. Johnson PJ, Berhane S, Kagebayashi C, et al. Assessment of liver function in patients with hepatocellular carcinoma: a new evidence-based approach-the ALBI grade. J Clin Oncol. 2015;33(6):550-558. doi:10.1200/jco.2014.57.9151
- 23. Guo X, Zou Q, Yan J, Zhen X, Gu H. Prognostic effect of pretreatment albumin-to-alkaline phosphatase ratio in human cancers: a meta-analysis. *PLoS One.* 2020;15(8):e0237793. doi:10.1371/journal.pone.0237793
- 24. Wu B, Hu X, Jin H, et al. Albumin-bilirubin and platelet-albumin-bilirubin grades for hepatitis B-associated hepatocellular carcinoma in Child-Pugh A patients treated with radical surgery: a retrospective observational study. *Medicine*. 2019;98(43):e17394. doi:10.1097/ md.000000000017394
- 25. Geng L, Zong R, Shi Y, Xu K. Prognostic role of preoperative albumin-bilirubin grade on patients with hepatocellular carcinoma after surgical resection: a systematic review and meta-analysis. *Eur J Gastroenterol Hepatol*. 2020;32(7):769–778. doi:10.1097/meg.00000000001618
- 26. Qin FF, Deng FL, Huang CT, et al. Interaction between the albumin-bilirubin score and nutritional risk index in the prediction of post-hepatectomy liver failure. *World J Gastrointest Surg.* 2024;16(7):2127–2134. doi:10.4240/wjgs.v16.i7.2127
- 27. Chan AW, Chan SL, Mo FK, et al. Albumin-to-alkaline phosphatase ratio: a novel prognostic index for hepatocellular carcinoma. *Dis Markers*. 2015;2015;564057. doi:10.1155/2015/564057
- 28. Huang W, Wei S, Dong X, et al. Preoperative albumin-alkaline phosphatase ratio affects the prognosis of patients undergoing hepatocellular carcinoma surgery. *Cancer Biomark*. 2024;39(1):15–26. doi:10.3233/cbm-230108
- 29. Wang Y, Zhang CY. The roles of liver-resident lymphocytes in liver diseases. Front Immunol. 2019;10:1582. doi:10.3389/fimmu.2019.01582
- 30. Yao W, He JC, Yang Y, et al. The prognostic value of tumor-infiltrating lymphocytes in hepatocellular carcinoma: a systematic review and meta-analysis. *Sci Rep.* 2017;7(1):7525. doi:10.1038/s41598-017-08128-1
- 31. Li L, Zinger J, Sassen SDT, Juffermans NP, Koch BCP, Endeman H. The relation between inflammatory biomarkers and drug pharmacokinetics in the critically ill patients: a scoping review. *Crit Care*. 2024;28(1):376. doi:10.1186/s13054-024-05150-4
- 32. Yang YT, Jiang JH, Yang HJ, Wu ZJ, Xiao ZM, Xiang BD. The lymphocyte-to-monocyte ratio is a superior predictor of overall survival compared to established biomarkers in HCC patients undergoing liver resection. *Sci Rep.* 2018;8(1):2535. doi:10.1038/s41598-018-20199-2
- 33. Wang X, Li W, Lu J, Li N, Li J. Lipopolysaccharide suppresses albumin expression by activating NF-kappaB in rat hepatocytes. J Surg Res. 2004;122(2):274–279. doi:10.1016/j.jss.2004.07.008
- 34. Busso N, Chesne C, Delers F, Morel F, Guillouzo A. Transforming growth-factor-beta (TGF-beta) inhibits albumin synthesis in normal human hepatocytes and in hepatoma HepG2 cells. *Biochem Biophys Res Commun.* 1990;171(2):647–654. doi:10.1016/0006-291x(90)91195-x
- 35. Ma X, Huang T, Chen X, et al. Molecular mechanisms in liver repair and regeneration: from physiology to therapeutics. *Signal Transduct Target Ther.* 2025;10(1):63. doi:10.1038/s41392-024-02104-8
- 36. Tincopa MA, Anstee QM, Loomba R. New and emerging treatments for metabolic dysfunction-associated steatohepatitis. *Cell Metab.* 2024;36 (6):1430. doi:10.1016/j.cmet.2024.04.016
- 37. Shah ND, Barritt IV AS. Nutrition as therapy in liver disease. Clin Ther. 2022;44(5):682-696. doi:10.1016/j.clinthera.2022.04.012
- 38. Gupta D, Lis CG. Pretreatment serum albumin as a predictor of cancer survival: a systematic review of the epidemiological literature. *Nutr J*. 2010;9:69. doi:10.1186/1475-2891-9-69
- 39. Musliha A, Dermawan D, Rahayu P, Tjandrawinata RR. Unraveling modulation effects on albumin synthesis and inflammation by Striatin, a bioactive protein fraction isolated from Channa striata. *Silico Proteomics in Vitro Approaches Heliyon*. 2024;10(19):e38386. doi:10.1016/j. heliyon.2024.e38386
- 40. Tanaka R, Ishima Y, Maeda H, et al. Albumin fusion prolongs the antioxidant and anti-inflammatory activities of thioredoxin in mice with Acetaminophen-induced hepatitis. *mol Pharm*. 2014;11(4):1228–1238. doi:10.1021/mp400690v
- 41. Casulleras M, Flores-Costa R, Duran-Güell M, et al. Albumin internalizes and inhibits endosomal TLR signaling in leukocytes from patients with decompensated cirrhosis. *Sci Transl Med.* 2020;12(566). doi:10.1126/scitranslmed.aax5135
- 42. Nogueiro J, Santos-Sousa H, Pereira A, et al. The impact of the prognostic nutritional index (PNI) in gastric cancer. *Langenbecks Arch Surg.* 2022;407(7):2703–2714. doi:10.1007/s00423-022-02627-0
- 43. Tan X, Chen H. The prognostic value of prognostic nutritional index in patients with ovarian cancer: a systematic review and meta-analysis. *Nutr Cancer*. 2023;75(1):73–81. doi:10.1080/01635581.2022.2104879
- 44. Wang Z, Wang Y, Zhang X, Zhang T. Pretreatment prognostic nutritional index as a prognostic factor in lung cancer: review and meta-analysis. *Clin Chim Acta*. 2018;486:303–310. doi:10.1016/j.cca.2018.08.030
- 45. Ahn SB. Noninvasive serum biomarkers for liver steatosis in nonalcoholic fatty liver disease: current and future developments. *Clin mol Hepatol.* 2023;29(Suppl):S150–s156. doi:10.3350/cmh.2022.0362

- 46. Takemura K, Board PG, Koga F. A systematic review of serum γ-glutamyltransferase as a prognostic biomarker in patients with genitourinary cancer. *Antioxidants*. 2021;10(4). doi:10.3390/antiox10040549
- Zhang LX, Lv Y, Xu AM, Wang HZ. The prognostic significance of serum gamma-glutamyltransferase levels and AST/ALT in primary hepatic carcinoma. BMC Cancer. 2019;19(1):841. doi:10.1186/s12885-019-6011-8
- 48. Liu X, Hou Y, Wang X, et al. Machine learning-based development and validation of a scoring system for progression-free survival in liver cancer. *Hepatol Int.* 2020;14(4):567–576. doi:10.1007/s12072-020-10046-w
- Aday U, Tatli F, Akpulat FV, et al. Prognostic significance of pretreatment serum lactate dehydrogenase-to-albumin ratio in gastric cancer. Contemp Oncol. 2020;24(3):145–149. doi:10.5114/wo.2020.10021950
- Wang ZX, Peng W, Zhang XY, Wen TF, Li C. Prognostic significance of postoperative change of PALBI grade for patients with hepatocellular carcinoma after hepatectomy. *Medicine*. 2021;100(11):e24476. doi:10.1097/md.00000000024476
- 51. Lisman T, Caldwell SH, Burroughs AK, et al. Hemostasis and thrombosis in patients with liver disease: the ups and downs. J Hepatol. 2010;53 (2):362–371. doi:10.1016/j.jhep.2010.01.042
- 52. Kopec AK, Luyendyk JP. Coagulation in liver toxicity and disease: role of hepatocyte tissue factor. *Thromb Res.* 2014;133 Suppl 1(01):S57–9. doi:10.1016/j.thromres.2014.03.023
- 53. Khatib R, Ludwikowska M, Witt DM, et al. Vitamin K for reversal of excessive vitamin K antagonist anticoagulation: a systematic review and meta-analysis. *Blood Adv.* 2019;3(5):789–796. doi:10.1182/bloodadvances.2018025163

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