REVIEW

Emerging Applications of Pickering Emulsions in Pharmaceutical Formulations: A Comprehensive Review

Xingyue Wang, Na Tian, Lili He, Zhixiang Yuan 🕞, Lu Han

College of Pharmacy, Southwest Minzu University, Chengdu, 610041, People's Republic of China

Correspondence: Lu Han, Email lunahan@swun.edu.cn

Abstract: Over the past two decades, particle-stabilized Pickering emulsions (PEs) have emerged as a versatile platform in pharmaceutical formulations, demonstrating distinct advantages over surfactant-based systems through enhanced stability, reduced toxicity, and tunable interfacial properties. These systems exhibit unique drug delivery potential through their precisely controllable architecture, particularly in achieving spatiotemporal drug release patterns, tissue-specific targeting, and enhanced therapeutic payload encapsulation. In this review, the characteristics of PEs are first detailed, followed by an introduction to the main preparation methods and the key parameters for controlling the type, droplet size, and stability of PEs. The third section categorizes and discusses the advantages and disadvantages of various solid particles as emulsifiers. Lastly, emphasis is placed on the application of PEs in the pharmaceutical field, including functionalized designs and various administration routes to enlighten the rational design of PEs for effective drug delivery.

Keywords: Pickering emulsions, drug delivery, solid particle emulsifiers, controlled release

Introduction

Emulsions play a vital role in the pharmaceutical industry. They are widely used in drug delivery systems because they enhance the solubility of hydrophobic drugs, improve pharmacokinetics, and help reduce adverse effects. These formulations are commonly applied in oral, injectable, and topical therapies.¹ However, being thermodynamically unstable systems that disperse immiscible liquids with synthetic surfactants, emulsions have drawbacks including loss of moisture in the epidermis, skin irritation, and challenges in drug distribution and elimination.^{2,3} To address these challenges, alternative stabilization approaches have been explored. Among them, Pickering emulsions (PEs), introduced by Pickering in 1907, utilize solid particles as emulsifiers, offering superior stability against droplet coalescence compared to traditional emulsions. The strategic selection of particles is essential for preparing stable PEs, which have shown promise in drug stability, encapsulation of bioactive ingredients,⁴ slow-release, and targeted drug delivery. Compared with surfactants, solid particles are able to adsorb and be permanently anchored at the oil-water interface.⁵ Notably, recent research has demonstrated the application of pH-responsive PEs in co-encapsulating immune checkpoint inhibitors and chemotherapeutic drugs, presenting a viable approach for oncology treatment.⁶

PEs have found diverse applications in various fields of research and industry such as pharmaceuticals, food production, cosmetics, and agrochemicals. Among these applications, their role in pharmaceutical formulations has garnered particular attention, as they offer enhanced biocompatibility, reduced toxicity, and improved control over drug release.⁷ Unlike conventional surfactants, most solid particles do not typically result in significant in vivo toxicity in formulations and are generally well-tolerated by the body, making them highly suitable for various drug delivery applications. In pharmaceutical formulations, PEs not only enhance drug solubility and bioavailability but also provide sustained release and enzymatic protection, making them valuable for oral and transdermal drug delivery. Their rigid particle-stabilized interface improves skin adhesion, potentially enhancing drug penetration.⁸ Additionally, in injectable

formulations, PEs contribute to controlled drug release, prolonged circulation time, and targeted accumulation in tumor therapy.⁹ These advantages underscore their potential in modern drug delivery systems, warranting further research into their functionalization and in vivo performance.^{10–12}

This review will first summarize the properties and preparation techniques of PEs. Subsequently, the benefits and limitations of current particulate stabilizers in pharmaceutical systems will be discussed. Special attention will be given to the functional designs that endow specific properties to PEs, such as controlled drug release, targeted drug release, stimuli-responsive drug release, etc. In the end, this review will also put specific focus on the use of PEs in various administration routes, helping to address critical formulation challenges in oral, transdermal, subcutaneous, intravascular and intratumoral delivery systems.

Characteristics and Preparation of PEs

The discovery of PEs is credited to S.U. Pickering, whose publication was believed to be the first to report the stabilization of O/W emulsions by solid particles adsorbed on the surface of oil droplets. However, Pickering's findings did not gain significant attention until recently. PEs are composed of two immiscible liquids and are stabilized by either organic or inorganic solid particles. Similar to conventional emulsions, PEs can be categorized as O/W, W/O, W/W, O/O, or multi-type (such as W/O/W or O/W/O).^{13–15} In PEs, the active pharmaceutical ingredient (API) can be encapsulated not only within the inner phase but also within the solid particles or attached to the particle surface or the water-oil interface. For instance, Candiani et al successfully encapsulated vitamin D3 within droplets stabilized by flaxseed meal particles.¹⁶ It is worth noting that while most medicinal PEs are predominantly O/W, there is a diverse range of W/O/W and O/W/O PEs that have been developed. Multiple emulsions are particularly interesting for pharmaceutical applications as they allow for the co-encapsulation of three APIs: one within the internal droplets, the second within larger droplets, and the last within the particles.¹⁷

The robust stability of PEs has garnered significant research attention, with its stabilization mechanism primarily revolving around two main types: the interfacial film theory and the three-dimensional network mechanism. The interfacial film theory involves solid particles, such as spherical and rod-shaped nanoparticles and nanofibers, creating a dense film encasing dispersed droplets, preventing coalescence when droplets come into coalescence.¹⁸ Furthermore, oil droplets can become ensnared within particle arrays during particle aggregation in the continuous phase, forming a three-dimensional network that restricts particle mobility and bolsters emulsion stability, exemplified by fat crystals.¹⁹ The stability of PEs is influenced by various factors, with particle wettability being a crucial parameter for solid particles to stabilize these emulsions. For particles to be adsorbed at the interface, they need to be wetted by both liquids. The empirical Finkle's rule states that the type of emulsion depends on the relative wettability of the particles in the two liquids.²⁰ Hence, the wettability of particles at the oil-water interface, quantified by the contact angle, is of great importance. Hydrophilic particles with a contact angle less than 90° tend to favor the formation of oil-in-water emulsions, while hydrophobic particles with a contact angle greater than 90° prefer water-in-oil emulsions (Figure 1). Additionally, factors such as the particle size and shape, pH, external magnetic field, temperature, ionic strength, the crystallinity of the particles, and oil-water ratio can also impact the stability of PEs.^{21,22} Similarly, the coverage of the surface particles, along with their wettability and concentration, also plays a critical role, as higher particle concentration and surface coverage can provide stronger interfacial resistance, thus improving stability. All these key factors that stabilize PEs are interlinked and may influence the nanoparticle wetting properties and, consequently, the obtained emulsion and its stability.23-26

Furthermore, the use of nanoparticles allows for the formation of PEs at the micron or even nanometer scale. For example, Chen et al successfully created nanoscale superstructures by compressing nanogels at the water-oil interface.²⁷ These superstructures were then used to stabilize nanodroplets loaded with paclitaxel (PTX), resulting in a slow drug release profile, enhanced cytotoxicity, and prolonged in vivo blood circulation. This innovative approach offers a versatile strategy for fabricating nanoscale superstructures with deformable nanoparticles, demonstrating the capacity of PEs in the field of nanomedicine.

PEs represent a highly stable form of emulsion, allowing for the utilization of existing surfactant-based emulsion preparation techniques. Currently, PEs are created through rotor-stator homogenization, ultrasonic emulsification, high-

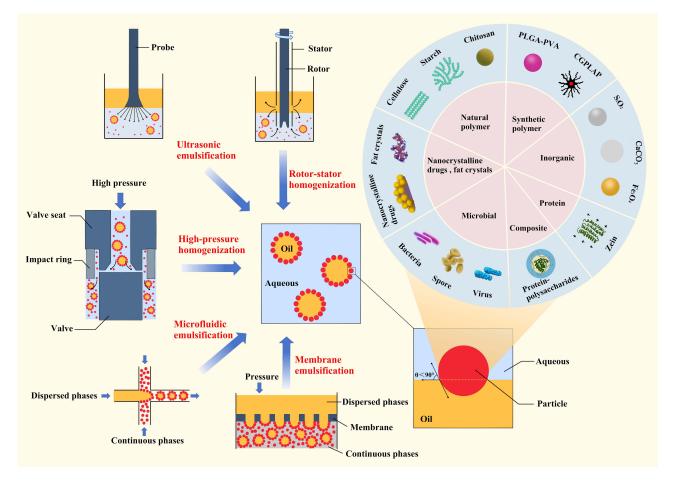


Figure I Schematic representation of the main preparation methods and on the micro and nano scales for O/W PEs. The figure on the right shows a fan schematic of solid particle classification. Abbreviations: PLGA-PVA, poly(lactic-co-glycolic acid)- poly(vinyl alcohol); CGPLAP, cashew tree gum grafted with polylactide; SiO₂, silicon dioxide; CaCO₃, talcum

Abbreviations: PLGA-PVA, poly(lactic-co-glycolic acid)- poly(vinyl alcohol); CGPLAP, cashew tree gum grafted with polylactide; SiO₂, silicon dioxide; CaCO₃, talcum carbonate; Fe₃O₄, ferric oxide.

pressure homogenization, microfluidic emulsification, and membrane emulsification (Figure 1). Among these methods, rotor-stator homogenization, ultrasonic emulsification, and high-pressure homogenization are the most commonly employed for PEs preparation. For instance, during rotor-stator homogenization, a powerful shear field is rapidly generated between the rotor and stator, resulting in the uniform dispersion of the oil and water phases along with the added particles. The droplets interact with solid particles dispersed in the system, promoting particle adsorption at the oil-water interface and contributing to the stabilization of the PEs. It should be noted that while rotor-stator homogenization is frequently used, the shear forces involved can present challenges such as particle aggregation, particle damage or excessive heating. Therefore, precise control of process parameters such as rotational speed, gap size, processing time, and particle concentration, as well as optimization of these parameters, is crucial for obtaining stable PEs with desired properties.¹⁷ The size of PEs' droplets achieved through the mentioned techniques exhibits some degree of variation. The high pressure homogenization method enables the production of nanoscale droplets through the repetition of multiple cycles. Alternatively, the rotor-stator homogenization method achieves similar results by utilizing high-speed stirring.²⁸

Solid Particle Emulsifiers

Solid particles play a vital role in PEs by decreasing oil-water interfacial tension and thus preventing droplet coalescence. Many natural organic particles, like polysaccharides, proteins, and solid lipid particles, are considered ideal materials for drug delivery using PEs. Several studies have confirmed the safety of these organic particles and their versatility for various applications, including pharmaceuticals and the food industries. Notably, starch, cellulose, alginate, albumin, and

poly(lactic-co-glycolic acid) (PLGA) are examples of organic particles that are approved for clinical use in drug delivery systems.^{4,29-32} In addition, inorganic particles, fat crystals, microbial particles, and other particles shown in Solid Particle Emulsifiers are suitable for stabilizing PEs. Different types of solid particles used to stabilize PEs not only exhibit varying toxicity profiles but also significantly influence the emulsions' physical stability and drug release behavior. For instance, synthetic particles such as strongly cationic silica nanoparticles or non-degradable iron oxide particles often exhibit systemic toxicity due to prolonged tissue retention in vivo. In contrast, natural macromolecules such as proteins and bacterial derivatives tend to be more biocompatible, though they have the potential risk of eliciting immunogenicity risks.^{33,34} Furthermore, the physicochemical characteristics of these particles, such as wettability, size, concentration, storage stability, and responsiveness to environmental stimuli, play a critical role in determining the overall stability of PEs. A rational selection of particle materials, properties, and concentrations is imperative when designing PEs. In fact, some formulations are engineered so that surface-bound particles can degrade, aggregate, or detach from the oil–water interface in response to internal stimuli (eg, changes in pH, redox conditions, or enzymatic activity) or external triggers (eg, light or temperature).^{35–37} This controlled destabilization mechanism is harnessed to achieve stimulus-responsive drug release. Collectively, these factors underscore that the selection and design of the stabilizing particles are pivotal not only for optimizing the emulsions' stability and in vivo toxicity but also for their efficacy in targeted drug delivery.

Natural Polymer Nanoparticles

In emulsions, it is important to prevent droplets from coalescence to maintain stability. Some polysaccharides, which are natural polymers, are insoluble in both water and oil. For instance, the most representative polysaccharides such as gum Arabic, modified cellulose, and starch can be utilized to stabilize emulsions.³⁸ In recent years, polysaccharide-based nanoparticles have shown tremendous capacity for applications in the fields of biotechnology, pharmaceuticals, and food industries. The information detailed is succinctly presented in Table 1.

Cellulose, a widely found biopolymer, is an ideal material option due to its sustainability, biodegradability, and non-toxic nature. Cellulose nanocrystals (CNCs) and cellulose nanofibrils are the primary forms of cellulose nanoparticles.³⁹ CNCs, prepared by acid hydrolysis, possess a highly crystalline structure and have rod-like nanosizes. They can form a stable interfacial film at the oil-water interface, thereby maintaining the stability of the emulsion.⁶³ Dong et al successfully synthesized spherical cellulose nanocrystals (S-CNCs) with diameters ranging from 30 nm to 60 nm using mixed acid hydrolysis of mercerized microcrystalline cellulose combined with ultrasonic treatment.³⁹ The PEs' droplets stabilized by S-CNCs showed remarkable stability across a wide range of pH, ionic strength, and temperature variations. Consequently, the utilization of S-CNCs as novel stabilizers for PEs presents promising opportunities in the fields of biomedical, cosmetic, and food applications. Furthermore, bacterial cellulose nanocrystals (BCNs) proved to be another promising emulsifier for the preparation of stable oil-in -water PEs.⁴⁰ Bacterial cellulose (BC) can be further purified with sodium hydroxide to yield a form of BC with an endotoxin value below 20 endotoxin units, meeting the acceptable range for implant applications as defined by the US Food and Drug Administration. This suggests that BCNs may be suitable for intravenous applications.⁶⁴ However, the wider application of nanocellulose-based PEs is hindered by safety concerns, and there is a lack of established protocols for evaluating the immunotoxicity, genotoxicity, and nutrient absorption associated with these emulsions. Additionally, the lack of standardized nanocellulose products, the limited number of manufacturers, and high production costs pose significant challenges.⁶⁵

Starch, the second most abundant natural polymer after cellulose, finds extensive usage in the food and pharmaceutical industries.⁶⁶ The inherent hydrophilic structure of natural starch granules restricts their ability to effectively stabilize the oil-water interface. Consequently, chemical modifications were performed to enhance their hydrophobic properties, thereby facilitating improved adsorption at the oil-water interface.⁶⁷ One commonly used modification method is the application of octenyl succinic anhydride (OSA). For instance, Wang et al conducted a study in which enzymatically branched glutinous maize and potato starch were recrystallized to form starch globule crystals.⁴² These crystals were then modified with OSA to increase their hydrophobicity, ultimately serving as emulsifiers for the preparation of PEs. The researchers discovered that the resulting octenyl succinate starch spherocrystals played a role in protecting the stability of encapsulants and slowing down lipid oxidation. This

Table I Examples of PEs Stabilized by Solid Particles

Particles					References	
Composition	Shape	Types	Droplet size	Emulsification Method		
Natural polymer	Cellulose nanocrystals	Spherical	o /w	≈32.17–32.83 µm	Ultrasonic pulverization (30%, 3min)	Dong et al, 2021 ³⁹
nanoparticles	Bacterial cellulose nanocrystals	Needle-like	0 /W	≈4 μm	Ultrasonic device (2 W/mL, 20s)	Kalashnikova et al, 2011 ⁴⁰
	Microfibrillated cellulose	Fiber	0 /W	≈28.44–29.10 μm	High-pressure homogenizer (pre-emulsion high- speed homogenizer)	Bouhoute et al, 2021 ⁴¹
	Starch globule crystals modified with OSA	Spherical	0 /W	≈I-50 μm	High-pressure homogenizer (300 bar) (pre-emulsion high-speed homogenizer)	Wang et al, 2017 ⁴²
	Butyric acid -modified porous starch	Spherical	O/W	≈15-30 μm	Homogenizer (18000 r/min, 120 s)	Ma et al, 2023 ⁴³
	Chitosan nanoparticles	-	O/W	-	Sonicated (0.2 kW, 3min)	Alehosseini et al, 2021 ⁴⁴
	Lignosulfonate nanoparticles	Spherical	O/W	≈2-18 μm	Ultrasonic Homogenizer (160W, 1min)	Shomali et al, 2022 ⁴⁵
	Alkaline lignin-graft-PNIPAM copolymer nanoparticles	Spherical	O/W	≈180-200 μm	Sonicator (50%, 30s)	Dai et al, 2019 ⁴⁶
Synthetic polymer nanoparticles	Cashew tree gum grafted with polylactide	-	O/W	≈I-3.5 μm	Spontaneous emulsification	Richter et al, 2018 ⁴⁷
	Plga	Spherical	W/O	≈25 μm	Three-way stop cock	Deschamps et al, 2019 ⁴⁸
Inorganic nanoparticles	SiO ₂ nanoparticles	Spherical	O/W	≈60 μm	Rotor-stator homogenizer (14,000 rpm, 6 min)	Heidari et al, 2022 ⁴⁹
	Phosphatidylcholine-kaolinites	Octahedral	O/W	-	High-shear homogenizer (8000rpm, 30s)	Tang et al, 2019 ⁵⁰
	Pbs quantum dots	Spherical	O/W	≈0.5–10 μm	Homogenizer	Huang et al, 2022 ⁵¹
	Titania (tio ₂) and iron oxide (Fe ₃ O ₄) nanoparticles	Spherical	W/O	≈100 μm	Ultrasonic Vibration Processor (200 W, 3 min)	Xie et al, 2017 ⁵²
Protein nanoparticles	Recombinant oil bodies protein nanoparticles	Spherical	O/W	≈10 μm	High-pressure homogenizer (100bar,5 cycles) (pre- emulsion high-speed blender)	Sun et al, 2023 ⁵³
	Groel-derived protein nano-ring	Ring-like	O/W	≈200-400 nm	Ultrasound equipment (40%, 2min)	Xu et al, 2020 ⁵⁴
	Sodium caseinate-modified zein nanoparticles	Spherical	O/W	≈10-50 μm	A high-speed homogenizer (12000rpm, 2min)	Feng et al, 2016 ⁵⁵

(Continued)

Wang et al

Table I (Continued).

Particles					References		
Composition		Shape	Types	Droplet size	Emulsification Method	-	
Nanocrystalline drugs, fat crystals	Curcumin nanocrystalline	Spherical	O/W	≈I56.88–I70.44 nm	A high-pressure homogenizer (800 bar, 50 cycles)	Wang et al, 2023 ⁵⁶	
	Diacylglycerol-based solid lipid nanoparticles	Spherical	W/O	≈20 μm	A rotor-stator homogenizer (10000 rpm, 2min)	Li et al, 2021 ⁵⁷	
Microbial nanoparticles	Tobacco mosaic virus-like nanorods	Rod-like	O/W	≈2-8 μm	Shaken by hand (2 min)	Wang et al, 2017 ⁵⁸	
	Ganoderma lucidum spores particles	Oval	O/W	≈60-70 μm	A Branson digital 450 W Sonifier (50%, 20min)	Zhu et al, 2019 ⁵⁹	
	Baker's yeast and lactic acid bacteria	Spherical and rod-like	O/W	≈30-100 μm	Vortexing or impeller-type mixing	Firoozmand et al, 2016 ⁶⁰	
Composite particles	Cellulose nanofibrils modified by soy protein isolate nanoparticles	Spherical and fiber	O/W	≈I0-40 μm	Homogenized (16000rpm, 2min)	Zhang et al, 2019 ⁶¹	
	Fe ₃ O ₄ cellulose nanocrystal	Rod-like	0 /W	≈7.5 μm	Ultrasound (60W, 3min) (pre-emulsion high-speed homogenizer)	Low et al, 2019 ⁶²	

finding contributes to our current understanding of emulsifiers for edible particles. To address the inherent limitations of OSA modification, Ma et al successfully synthesized butyric acid (BA)-modified porous starch (PS) with varying degrees of substitution.⁴³ Through an esterification reaction, BA-PS with high degree of substitution can effectively stabilize the encapsulation of PTX in PEs, significantly enhancing emulsification and PTX retardation capabilities. This suggests that BA-PS has the capacity to be utilized as an emulsifier for slow-release drug delivery. Furthermore, there is still ample room for exploration in the application and development of environmentally friendly methods for producing starch granules to stabilize PEs.⁶⁸

Chitosan is derived from chitin through deacetylation in the presence of hot alkali, and it possesses remarkable properties such as biocompatibility, biodegradability, mucous adhesion, and low toxicity.⁶⁹ In comparison to traditional emulsions, PEs incorporating chitosan enhance the bioavailability of various compounds. The stability of the PEs can be achieved through the use of self-aggregated chitosan particles,⁷⁰ chitosan nanocrystals (also known as chitosan whiskers, nanowhiskers, or rod-shaped chitosan), or complexes of chitosan with proteins^{71,72} such as corn protein or gelatin.⁷³ As an example, Alehosseini et al tried to explore the PEs based on chitosan nanoparticles (CSNPs) as food-grade delivery systems for d-limonene, which is a bioactive compound.⁴⁴ If they are able to provide data related to the in vivo safety studies of the obtained emulsion, the stable PEs of CSNPs holds promise for application in the delivery of active drug nanoparticles.

Moreover, both lignin and agarose (agarose microgels)⁷⁴ are natural polymers. Lignin, a phenolic polymer, creates a three-dimensional network structure in the cell wall.⁷⁵ Lignin has extensive applications in packaging materials, hydrogels, and even as a carrier for bioactive substances like enzymes and drugs.⁷⁶ Nevertheless, the limited solubility and dispersibility of structurally diverse lignin pose challenges for its advancements in the realm of green energy and materials. Consequently, diverse approaches must be employed to improve its properties. For instance, Shomali et al utilized carboxylated lignosulfonate with different grafted alkyl side chains to synthesize lignosulfonate nanoparticles (LNPs).⁴⁵ By adjusting the length of the alkyl chains, the resulting LNPs exhibited a higher affinity to the oil-water interface. As a result, a green, environmentally friendly, and simple water-based acid precipitation process was employed to obtain emulsions with smaller and more stable droplet sizes. In addition, Zhang et al have reported a green and simple method for the production of LNPs using beet alkaloid and lactic acid deep eutectic solvent through water droplet-induced self-assembly, which effectively addresses the issue of environmentally unfriendly organic solvents.⁷⁶ Moreover, Dai et al have developed a novel multi-functional PEs stabilized by LNPs to enhance the stability and thermal-controlled release of trans-resveratrol.⁴⁶ The research findings suggest that LNPs-based PEs hold great capacity in the storage and thermal-controlled release of light-unstable and poorly water-soluble drugs.

While certain natural substances such as proteins or polysaccharides are utilized in the formulation of these emulsions, the resultant particles typically exhibit polydispersity, instability, or excessive hydrophilicity, rendering them inadequate for emulsion stabilization. Indeed, the challenges associated with quality control of natural materials, influenced by factors like production locale and processing parameters, further complicate matters.⁷⁷ Consequently, research pertaining to PEs in pharmaceutical applications warrants further exploration.

Synthetic Polymer Nanoparticles

Biodegradable polymer particles have garnered attention specially in the field of drug delivery due to their reduced toxicity, ability to control drug release kinetics, and enhanced biocompatibility.⁷⁸ Biodegradable polymers comprise synthetic polymers like polylactic acid (PLA), and polyglycolic acid, as well as their copolymers, such as PLGA, and polycaprolactone.¹⁷ For instance, to enhance Amphotericin B (AmB) oral bioavailability and control drug release, Richter et al first reported the utilization of self-assembled cashew gum grafted with PLA nanoparticles, obtained through nano-precipitation, to obtain stable AmB-loaded PEs through spontaneous emulsification.⁴⁷ Biodegradable particles made of PLGA offer the advantages of biocompatibility and rapid clearance from the body, which can help prevent inflammatory reactions due to accumulation. In a separate study, Deschamps et al employed a simple emulsification technique using two connected syringes to incorporate PLGA nanoparticles into the aqueous phase, resulting in the formation of an oil-in-water lipiodol emulsion with an average droplet size of

approximately 40 µm, which is comparable to the diameter of blood vessels near tumors.⁴⁸ They loaded doxorubicin (DOX) into the internal phase and found that complete release of DOX from the stable lipiodol emulsion required up to 10 days, demonstrating the capacity of these solid particle-stabilized emulsions for efficient encapsulation and sustained release of chemotherapy drugs for the treatment of tumors. In line with prior research, in vitro experiments have revealed that PLGA nanoparticles exhibit no discernible toxicity until reaching relatively high concentrations of 3 mg/mL.⁷⁹ However, the toxicity of PLGA up to 3 mg/mL needs further exploration, because stabilization of PEs requires larger concentrations of solid particles than this. Notably, in a significantly dynamic physiological environment, the interaction between nanoparticles and cellular components would occur at a reduced concentration level necessitating continuous fluid replenishment, thereby mitigating the risk of cytotoxic effects.⁴⁸ As a result, more in-depth studies on toxicity tests including genotoxicity assay are needed, which hold significance for use in the injectable administration of chemotherapeutic drug.⁸⁰

Inorganic Nanoparticles

Inorganic solid particles commonly employed for the preparation of PEs comprise SiO₂, TiO₂, ZnO, CaCO₃, Fe₃O₄⁸¹ carbon nanotubes, kaolin, quantum dots (QD), and various modified inorganic solid particles. Extensive research has demonstrated that modified solid particles can enhance the stability of PEs and regulate their demulsification.⁸² Silica nanoparticles, also known as nano-silica, are extensively utilized owing to their cost-effectiveness, favorable biocompatibility, low in vivo toxicity, and antibacterial properties. Therefore, SiO2 is a frequently utilized material in the synthesis of PEs.⁸³ For example, Heidari et al employed the Taguchi approach to modify SiO₂ nanoparticles using cetyltrimethylammonium bromide to stabilize PEs, making them bioactive compounds delivery system.⁴⁹ In the study by Tang et al, natural kaolinite was modified to prepare phosphatidylcholine-kaolinite and used to stabilize PEs encapsulating curcumin (Cur).⁵⁰ The resulting PEs were effectively absorbed by cells, non-toxic, and biocompatible, and they could completely release Cur in the intestines to improve the bioavailability of Cur. These findings may contribute to the effective encapsulation of lipophilic food or drugs in PEs to enhance their bioavailability. Some inorganic particles also exhibit characteristics of responsiveness to external stimuli. For example, Huang et al developed QD-based nanoreactors by creating amphiphilic PbS QD-stabilised PEs through the formation of Janus ligand shells. Their research offers valuable insights for enhancing the effectiveness of nanoreactors and microemulsions in facilitating multiphase photocatalytic reactions.⁵¹ In addition, Xie et al proposed a photomagnetic dual-responsive PEs micro-reactor, in which photosensitive TiO₂ and Fe₃O₄ nanoparticles are co-adsorbed at the oil-water interface of the emulsion droplets.⁵² By approaching each other under an external magnetic field, the reactants came into contact and triggered a chemical reaction under ultraviolet light irradiation. Their research has opened up broad prospects for the construction of multifunctional PEs. Further research could investigate the utilization of visible light instead of ultraviolet light in photomagnetically driven PEs microreactors, offering significant potential for advancements in the field of life sciences. However, there is relatively limited research on the in vivo safety of these inorganic nanoparticles, which hinders their application as drug carriers.⁸⁴

Protein Nanoparticles

Protein nanoparticles possess several advantages that make them well-suited for the preparation and stabilization of traditional PEs. These advantages include their natural origin, non-toxicity, biocompatibility, biodegradability, potential health benefits, good surface activity, and emulsion stability.⁸⁵ In the context of PEs, protein nanoparticles can take on various shapes, including non-deformable solid spherical nanoparticles,⁵³ nanofibers, nanotubes, nanogels, nanocages,⁸⁶ and plate-like nanoparticles. Protein-based PEs have greater applications in the food and pharmaceutical industries compared to traditional inorganic particle-stabilized PEs. These applications include substituting partially hydrogenated fats in food,⁸⁷ encapsulating, stabilizing, and releasing active ingredients,⁵³ and inhibiting lipid oxidation.⁸⁸ For instance, in the work of Sun et al, the recombinant oil bodies protein (ROP) was induced using guanidine hydrochloride.⁵³ In this approach, the ROP first formed a core-shell structure to encapsulate Cur, and then the formed ROP-Cur colloidal particles were used to stabilize PEs for the preparation of a double-chamber microdroplet system to achieve co-encapsulation of Cur and vitamin D. This study provides an important avenue for developing nutrition delivery systems and improving the stability and bioactivity of hydrophobic nutrients. Furthermore, Xu et al utilized a GroEL-derived

protein nanoring from Escherichia coli to stabilize PEs for encapsulating β -carotene.⁵⁴ In the context of promoting biocompatibility and sustainability, this work offers a more environmentally friendly solution for emulsion-based applications. Moreover, Feng et al successfully prepared a nanocomplex of corn zein and sodium caseinate by surface adsorption, which enhanced the stability and dispersibility of corn zein colloidal particles in the aqueous phase.⁵⁵ This approach was employed to explore a novel method of utilizing food-grade protein surface modification to stabilize PEs. The aforementioned studies indicate that proteins have the ability to stabilize emulsions. However, it is important to note that protein-stabilized PEs are highly susceptible to alterations in environmental factors such as pH, ionic strength, and temperature. If these conditions surpass certain thresholds, the PEs have a tendency to flocculate and expedite the destabilization process. Consequently, further adjustments are required in order to utilize protein particles effectively in the future for the production of PEs.^{89,90}

Nanocrystalline Drugs, Fat Crystals

Recently, a new delivery system called drug nanocrystal (NC) self-stabilized PEs (NSSPE) has been developed for poorly water-soluble drugs. This system is stabilized by insoluble drug nanocrystals, eliminating the need for additional stabilizers. NSSPE offers the combined benefits of NC and PEs while addressing the side effects associated with inorganic solid particles like silica. One of the key advantages of NSSPE is its ability to enhance the water solubility of insoluble drugs such as quercetin, geraniol,⁹¹ and silymarin flavonoids,⁹² leading to improved oral bioavailability.⁹³ This delivery system, consisting of just three components (water, oil, and drug), significantly increases the drug loading capacity as the drug no longer needs to be dissolved in the oil phase.⁹⁴ In the work by Sheng et al, Cur was adopted as a typical Biopharmaceutics Classification System (BCS) IV drug, and a stable PEs of Cur-NCs was prepared using either poorly digestible (isopropyl palmitate, IPP) or easily digestible (soybean oil, SO) oils.⁹⁴ After optimization, they found that the concentration of Cur in the formulation far exceeded its solubility in IPP or SO. Their research provides a new strategy for improving the oral bioavailability of Cur and other BCS IV drugs. Additionally, Wang et al proposed the introduction of traditional Chinese medicine volatile oil - turmeric oil as the oil phase in the concept of "drug excipient combination" to prepare a drug called NSSPE, and Cur-NSSPE lyophilized powder was prepared using the freeze-drying method.⁵⁶ This Cur-NSSPE lyophilized powder showed more significant advantages in its anti–inflammatory effects both in vitro and in vivo compared to Cur-API and Cur-NCs.

Solid lipid nanoparticles (SLNs), commonly referred to as fat crystals, have been employed as colloidal nanocarrier systems in drug delivery.⁹⁵ Nevertheless, there is a lack of pharmaceutical applications that utilize fat crystal stabilized PEs. These fat crystals, however, have demonstrated their capability as suitable candidates for this purpose due to their exceptional biocompatibility, minimal toxicity, ability to safeguard thermally unstable molecules against degradation, and their specific targeting capabilities. Lipid crystals, such as triglycerides, contain small amounts of monoglycerides and diglycerides and can serve as solid particles with unique temperature-responsive properties.⁵⁷ They provide stability to W/O emulsions and effectively build the emulsion droplet by forming an interfacial film and a crystalline network, creating the desired texture.^{19,96} In the work of Li et al, they developed SLNs based on medium- and long-chain diglycerides (MLCD) as interface stabilizers for W/O PEs.⁵⁷ By using MLCD as the lipid core and controlling the crystallization curve, crystal form, contact angle, and interface adsorption curve through the selection and concentration of surfactants, the rigidity of SLNs can be adjusted. The high rigidity of SLNs makes water-in-oil PEs a viable option for reducing saturated fatty acid levels and serving as a carrier for water-soluble bioactive compounds. While researchers have demonstrated the safety of the SLNs, the inclusion of surfactants to enhance SLNs properties in the formulation of PEs drug delivery systems necessitates further research to ensure safety.⁹⁷

Microbial Nanoparticles

Various microscopic particles can be found in nature, including viruses, spores, bacteria, and yeast, at both micro- and nano-scale levels. These particles exhibit uniform size distribution and can be tailored for pharmaceutical purposes.¹⁷ For example, virus-like nanoparticles lack genetic material and consist of protein shells. They are robust structures capable of encapsulating drug molecules at the nanoscale. Within nanomedicine, viruses have been engineered as intelligent carriers

for targeted drug delivery.⁹⁸ Nonetheless, concerns regarding the immunogenicity, cytotoxicity, inflammatory response, and other toxicities associated with mammalian viral vectors have prompted the exploration of plant viruses or plant virus nanoparticles as alternative nanocarriers.⁹⁹ Extensive research has revealed the ability of these natural particles to stabilize PEs, enhancing biocompatibility, bioactivity, and bioavailability in body fluids while reducing toxicity. Notably, the Tobacco mosaic virus can self-assemble into nanorods with amphiphilic properties and inherent self-assembly capability, demonstrating significant potential for drug delivery and viral detection.^{58,99} Similarly, Ganoderma lucidum spores have been identified as effective stabilizers of PEs due to their unique organic composition and structure.⁵⁹ Furthermore, in a study by Firoozmand et al, yeast and lactic acid bacteria were employed as micrometersized colloidal particles to stabilize oil-in-water emulsions, resulting in long-term stability to produce a monophasic emulsion with high oil volume fractions, characterized by closely packed oil droplets arranged in a polyhedral manner. This innovative method of emulsification stabilization holds great capacity for various food applications; however, the research in the realm of drug delivery remains limited, necessitating further exploration by the scientific community.¹⁰⁰

Composite Particles

The utilization of emulsifiers such as starch and cellulose to stabilize emulsions may be limited by certain properties of the particles. To overcome this limitation, composite particles can be created through interactions between particles with different properties, resulting in more stable PEs. For instance, solid biopolymer nanoparticles with prolonged coalescence lifetimes were successfully produced by leveraging effective associative interactions between proteins and polysaccharides.¹⁰¹ These composite particles, exemplified by lactoferrin nanoparticle-inulin polysaccharide,¹⁰² were utilized to stabilize the oil-water interface and shield protein particles from pepsin, thus enhancing the kinetic stability of the emulsions under gastric conditions. Moreover, Zhang et al demonstrated a sustainable approach by modifying the surface of bacterial cellulose through the physical adsorption of soybean isolate proteins onto cellulose protofibrils. The resulting complexes effectively stabilized PEs with diameters ranging from 10 µm to 40 µm, and subsequent in vivo digestion studies revealed enhanced resistance to lipid hydrolysis in these emulsions.⁶¹ These findings underscore the capacity of composite particles in the stabilization of PEs. Composite particles are increasingly being designed as multifunctional systems, offering simultaneous stabilization and active functionalities such as antibacterial effects, drug delivery, or environmental responsiveness. This has driven the exploration of advanced emulsion types, including multiple emulsions and double PEs, expanding their application capacity in the pharmaceutical field. Additionally, innovations in nanocomposite particles provide precise control over emulsion properties and enhanced interfacial performance, further advancing their role in complex formulation systems.

Functional Designs of PEs

Sustained or Controlled Drug Release

PEs have emerged as a reliable method for encapsulating and delivering active pharmaceutical ingredients, utilizing solid particles to create a protective barrier.⁷ Indeed, the solid particle film acts as a hindrance, slowing down the diffusion of therapeutic drugs from the emulsion to the target site. This controlled release mechanism offers numerous advantages, including prolonged drug persistence, protection against degradation, reduced administration frequency, and enhanced patient compliance.¹⁰³ For instance, a recent study demonstrated that pea isolate protein (PPI) could be transformed into nanoparticles at pH 3.0,¹⁰⁴ leading to the development of a stable gel-like emulsion with sustained β -carotene release properties through microfluidic emulsification.¹⁰⁵ Moreover, to overcome the disadvantages of high toxicity of antimicrobial peptides, high hemolysis of human cells, short circulating plasma half-life and poor stability in vivo, researchers have stabilized PEs with Parasin I-interacted lecithin and chitosan nanoparticles.¹⁰⁶ The results of the in vitro study indicated that the emulsion exhibited the ability to enhance membrane damage and coalescence area upon bacteria binding. Furthermore, the study revealed that the peptide-composite nanoparticles used to stabilize emulsions enabled the sustained release of Parasin I, ultimately boosting their antimicrobial efficacy. In addition, the sustained drug release properties of PEs are also beneficial to cancer treatment. For instance, oil-in-water lipid iodine

emulsion remains the preferred choice for localized chemotherapy in hepatocellular carcinoma treatment.¹⁰⁷ These findings underscore the benefits of utilizing PEs for the sustained or controlled release of active drugs, such as vitamins, antimicrobial peptides, and anticancer drugs.

Targeted Drug Delivery

Targeted delivery is a key benefit of drug delivery systems, offering the competence to minimize adverse effects and enhance therapeutic effectiveness by facilitating the accumulation of substances in specific tissues.¹⁰⁸ PEs play a crucial role in enabling precise drug delivery through mechanisms such as particle adsorption on interfaces, bonding with target cells, and increasing cellular binding of modified target ligands by expanding the contact area.¹⁰⁹ For instance, in the context of cancer vaccine administration, conventional liquid mixtures of immunostimulants, antigens, and target molecules often struggle with diffusion and degradation issues, hindering an optimal immune response.¹¹⁰ In contrast, PEs can be strategically formulated to encapsulate various components with diverse characteristics, such as size, hydrophobicity, or charges. To illustrate, Du et al utilized the recognition ability of mannose by antigen-presenting cell surface receptors to create mannose-based PEs for delivering complex vaccines aimed at synergistically combating tumors in conjunction with PD-1 antibodies.¹¹¹ This targeted approach not only amplified the cellular immune response but also heightened the anti-tumor effects synergistically. Insights gained from these findings shed light on the ability of PEs-based targeted drug delivery strategies. Moreover, the effectiveness of drug delivery from PEs is closely tied to the properties of the particles involved. For example, researchers have successfully stabilized β -carotene emulsions with PPI nanoparticles at pH 3.0 for intestinal targeting, leveraging the isoelectric point nature of protein particles to suit different parts of the digestive system.¹⁰⁵

Stimulus Responsive Drug Delivery Systems

In numerous application scenarios, the surface characteristics of solid particles can undergo various physical or chemical changes triggered by environmental factors like pH, temperature, specific reagents, light, and magnetic fields. These changes can impact the wettability, charge, or size of the particles and disrupt the arrangement of the emulsion's interface¹¹²(Figure 2). Consequently, PEs can serve as responsive delivery systems by striking a balance between droplet stability and controlled rupture, enabling researchers to achieve controlled or targeted drug release. For instance, in environments with weakly acidic conditions like inflammation sites, lysosomal compartments, or tumor tissues that necessitate in vivo treatment, researchers have harnessed the acid sensitivity and degradability of acetalized starch-based nanoparticles to enhance the efficacy of targeted drug delivery systems for anticancer medications, such as Cur.¹¹³ Moreover, studies have explored thermosensitive PEs, exclusively stabilized by SLNs, for drug encapsulation and delivery like ketoprofen. These SLNs exhibited controlled drug release upon experiencing specific temperature changes, such as partial melting at 37°C.¹¹⁴ The versatility of PEs, whether responsive to pH or temperature changes, introduces novel opportunities for precise drug delivery systems. Notably, nano delivery systems activated by external stimuli, like photothermal responses, offer distinct advantages over those reliant on internal triggers such as pH and heat, due to their ease of manipulation and precise treatment control. Furthermore, owing to its excellent tissue penetration capacity, nearinfrared radiation (NIR)-light responsive drug delivery systems present a promising avenue in cancer treatment.¹¹⁵ Recent research has explored the use of polydopamine (PDA) bowl-shaped mesoporous nanoparticles as Pickering stabilizers, resulting in pH-responsive emulsions with notable photothermal responsiveness under NIR light due to PDA integration.¹¹⁶ These properties hold significant potential for diverse biomedical applications, including drug delivery. It is important to note that the release of drugs is influenced by factors like the rupture and deformation of PEs, which could affect their stability. Hence, future stimulus-responsive PEs are likely to focus on achieving a balance between stability and effective drug release mechanisms.

Multilayer Structure

In cancer treatment, combining various medications through cocktail therapy has proven to be a promising strategy for combatting tumor regression and multidrug resistance.¹¹⁹ However, the challenge lies in effectively delivering multiple drugs with diverse hydrophobicity and targets in a spatiotemporal manner.¹⁰⁹ Hence, there is a demand for the

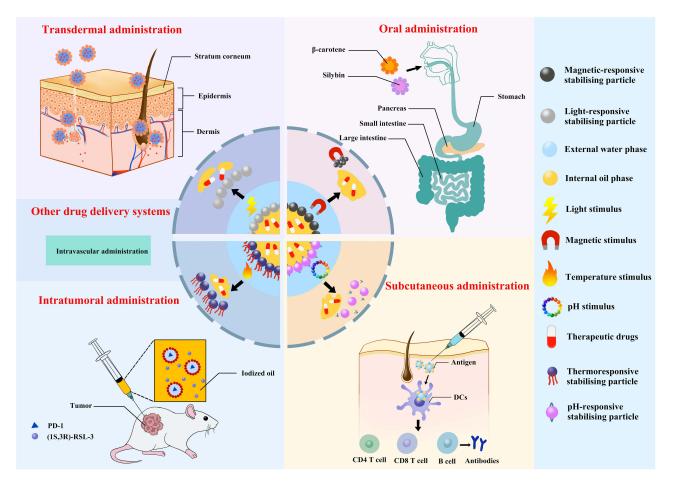


Figure 2 Schematic representation of stimulus-responsive drug release and routes of administration of PEs. The oral and intratumoral administration of PEs is referenced to Li et al, ¹¹⁷ Yi et al, ⁹² and Dai et al. ¹¹⁸

development of multi-drug delivery systems to enhance the bioavailability and effectiveness of treatments. PEs, known for their intricate structure, serve as an efficient multi-drug delivery system. The multilayer configuration of W/O/W or O/W/O emulsions offers a better solution for loading different components with varying hydrophobicity or charge.¹²⁰ For instance, a recent study by Tang et al introduced PEs loaded with betanin and Cur, where betanin and Cur were dissolved in the inner aqueous and oil phases, respectively, to create the W/O emulsion. The W/O emulsion was then combined with betanin-bovine serum albumin nanoparticles dispersed in the outer aqueous phase to form a composite W/O/W emulsion.¹²¹ The synergistic effect of betanin and curcumin in inhibiting A549 cell growth highlights the capability of this multilayer structure for co-delivering multiple drugs with diverse properties, suggesting its significance in future clinical applications. Furthermore, the multilayer structure of PEs can be further combined with active targeting ligands, stimuli-responsive designs, or sustained-release techniques to achieve precise and sequential drug delivery of multiple drugs.

Delivery Routes of PEs

PEs have shown significant applications in the pharmaceutical field, particularly in drug delivery systems. When administered via different routes, such as oral, subcutaneous, intravascular, or intratumoral, the characteristics of the stabilizing particles (size, surface properties, and biodegradability) must be carefully optimized to enhance therapeutic efficacy and ensure safe delivery. For instance, Pickering particles for oral administration should be suitable for strongly acidic environment and protect bioactive drugs.¹²² Furthermore, PEs encapsulating chemotherapeutic agents for

intravascular and intratumoral administration may use particle stabilizers that respond to the tumor environment and release the drugs.³⁵

Oral Administration

PEs are primarily used for the oral delivery of hydrophobic as well as poorly bioavailable drugs in the expectation of increasing the stability and improving the aqueous solubility of orally administered drugs. Poorly water-soluble drugs present challenges in formulating oral drug products due to their inconsistent bioavailability and limited dissolution in the gastrointestinal tract.¹²³ Lipid-based formulations have been explored to enhance the oral bioavailability of such drugs, but the high surfactant content in these formulations may cause gastrointestinal irritation, leading to reduced drug effectiveness and bioavailability.¹²⁴ PEs offer a novel drug delivery approach using solid particles instead of surfactants, which can minimize irritation. For instance, Eudragit[®] RL100 nanoparticles have been utilized to stabilize PEs, extending the efficacy of drugs like ketoprofen and reducing the dosing frequency.¹²⁵ Furthermore, recent advancements in PEs involve utilizing solid particles to create a protective barrier at the water-oil interface, safeguarding the drug core from degradation in particularly acidic environments or under various stress conditions. Li et al successfully enhanced the bioavailability of β -carotene by encapsulating it in PEs stabilized with chitosan hydrochloride-carboxymethyl starch nanogels¹¹⁷ (see Table 2). The Pickering particles should be biocompatible and able to withstand or respond to the acidic environment of the stomach and the enzymatic conditions of the gastrointestinal tract. For instance, Hu et al hypothesized that media-milled black rice particle-stabilized PEs would be a good oral delivery vehicle for 5-demethylnobiletin (5-DMN), which has the potential to enhance 5-DMN absorption and metabolic conversion in the colon (Figure 3).¹²⁶ Black rice particles are high in dietary fiber and phenolic compounds, which could be selectively broken down by the colonic microbiota. They found that black rice-stabilized PEs encapsulated 5-DMN targeting the colon could alleviate dextran sulfate sodium-induced weight loss and diarrhea. However, comprehensive data on the pharmacokinetics and bioavailability of PEs remain limited, with in vitro lipolysis models commonly employed to assess their suitability for oral drug delivery. Moreover, utilizing refractory drug nanocrystals to stabilize emulsions for oral drug delivery systems can increase drug loading capacity without the need for surfactants or polymer stabilizers.⁹² Besides, the elucidation of the in vivo degradation of PEs, encompassing particle stabilization and emulsion digestion, holds paramount importance. The employment of a pioneering bioimaging methodology reliant on aggregation-induced burst fluorescent probes presents a promising avenue to address this intricate issue.^{127,128}

Transdermal Administration

PEs demonstrate promising capabilities in the topical administration of antibiotics for skin infections and in the transdermal delivery of lipophilic drugs, as evidenced by research findings. Topical drug delivery is a challenging field with many advantages, including extended administration duration, reduced frequency of dosing, and minimized systemic side effects.¹³⁰ While conventional emulsions with surfactants may compromise the skin barrier, PEs offer a promising solution due to their low toxic, non-carcinogenic nature and excellent dermal compatibility. PEs could enhance the skin permeation of drugs by augmenting the moisture level within the stratum corneum, while certain solid particles may induce changes in the structure of the stratum corneum proteins.¹⁴³ Recent research by Frelichowska et al demonstrated that PEs effectively enhance the delivery of lipophilic drugs like retinol into the skin, particularly within the stratum corneum, when compared to traditional emulsions.¹²⁹ These nanoparticlestabilized emulsions exhibit unique characteristics suitable for targeting the stratum corneum or enabling controlled drug release. This phenomenon could be attributed to the capacity of PEs for coalescence in contrast to surfactantsstabilized droplets, potentially stemming from differences in droplet flexibility. Moreover, in the context of skin barrier damage and associated bacterial infections, topical application of antibiotics via PEs shows capability for treating infections and preventing further complications.¹³⁰ While PEs hold promise for sustained antibiotic delivery to prevent wound infections, further research is needed to investigate their skin penetration and safety profiles in vivo.8 Techniques like confocal laser scanning microscopy were employed to track particles and investigate the penetration capability of emulsion droplets through the stratum corneum into the deeper layers of the skin. This

Delivery routes		Emulsion		Drug /localization	Functional designs	References
	Туре	Particles	Droplet size			
Oral	O/W	Eudragit RL100 nanoparticles	≈220 nm	Ketoprofen/ oil	Controlled Release	Dieng et al, 2020 ¹²⁵
	O/W	Chitosan hydrochloride/carboxymethyl starch complex nanogels	≈5-8 μm	β-carotene/ oil	-	Li et al, 2020 ¹¹⁷
	O/W	Silybin nanocrystal	≈24.2–30.4 μm	Silybin/ NPs	-	Yi et al, 2017 ⁹²
	O/W	Pea protein isolate protein	≈3-8 μm	β-carotene/ oil	Stimulus response and targeted delivery	Shao et al, 2016 ¹⁰⁵
Transdermal	O/W	Silica particles	≈2-4 μm	Retinol/ oil	Sustained release	Frelichowska et al, 2009 ¹²⁹
	W/O	Aluminum starch octenyl succinate particles	-	Minocycline hydrochloride/ water	Controlled release	Marto et al, 2019 ¹³⁰
	W/O	Chitosan/collagen peptides nanoparticles	≈7.63–15.72 μm	-	-	Sharkawy et al, 2021 ¹³¹
	O/W	Carboxymethyl chitosan - sodium alginate nanoparticles	≈5 µm	Curcumin/ oil	Controlled release	Wu et al, 2022 ¹³²
Subcutaneous injections	O/W	PLGA-PEG-mannose nanoparticles	≈Iµm	Lipopeptide ovalbumin/ oil- water interface	Targeted delivery	Du et al, 2022 ¹¹¹
	O/W	Alum microgels	≈4 μm	SARS-CoV-2 spike protein/ oil-water interface	Targeted delivery	Peng et al, 2020 ¹³³
Intravascular administration	O/W	Poly(N-isopropylacrylamide-co-allylamine) nanogels	≈1.5 μm	Paclitaxel/ oil	Sustained release and targeted delivery	Chen et al, 2011 ²⁷
	O/W	CTAB and LA	≈294-502 nm	CTAB and LA/ NPs	Targeted delivery and stimulus response	Chen et al, 2023 ¹³⁴
	W/O	PLGA nanoparticles	-	Oxaliplatin/ water	Sustained release	Deschamps et al, 2018 ¹³⁵
	O/W	Poly (N-isopropylacrylamide-co-acrylic acid) nanogels	≈I-I0 μm	-	Stimulus response	Li et al, 2021 ¹³⁶

Table 2 Examples of Different Delivery Routes for PEs

Intratumoral administration	W/O	PLGA nanoparticles	≈37-47 µm	lpilimumab/ water
	O/W	СТАВ	≈91.72–96.44 nm	Paclitaxel/ oil
Other drug	O/W	Chitin nanocrystals	≈3.5 µm	-
delivery systems	O/W	Galactose-functionalized hydroxyethyl starch-grafted polycaprolactone nanoparticles	≈140 nm	Doxorubicin and indocyanine green/ oil
	O/W	Poly(N-isopropylacrylamide-co-acrylic acid) nanogel	≈I50 nm	Doxorubicin/ oil

O/W

OSA-modified starch particles

Tselikas et al, 2020¹³⁷

Xu et al, 2016¹³⁸

Yu et al, 2024¹³⁹

Hu et al, 2017¹⁴⁰

Shang et al, 2019¹⁴¹

Zheng et al, 2022¹⁴²

Sustained release

Sustained release

Targeted delivery and

Targeted delivery and

Stimulus response and

stimulus response

stimulus response

controlled release

_

Nile red/ oil

_

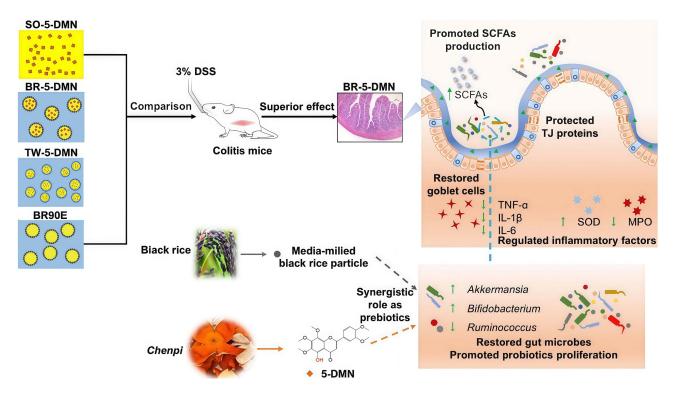


Figure 3 A milled black rice particle-stabilized PEs (BR-5-DMN) has been developed as a delivery vehicle for 5-DMN to treat colitis. Reprinted with permission from Hu H, Wang Y, Zhao D-G, Lu X. Oral Delivery of 5-Demethylnobiletin by Media-Milled Black Rice Particle-Stabilized PEs Alleviates Colitis in Mice: Synergistic Effects of Carrier and Loaded Bioactive. J Agric Food Chem. 2025;73(2):1257–1272. Copyright © 2025, American Chemical Society.¹²⁶

method was utilized to assess the potential entry of the topically administered drug into the bloodstream, enabling the evaluation of in vivo skin penetration and safety of PEs.¹³¹ However, the application of PEs to transdermal drug delivery has some limitations because of emulsions with strong mobility. Consequently, certain researchers have explored the application of PEs in conjunction with a gel matrix containing Cur to address skin wound treatment, thereby extending drug retention on the skin.¹³² In addition, the potential for irritation caused by particles and emulsions on the skin, mucous membranes, and damaged skin is a critical consideration for ensuring the safety of drug delivery systems, which is essential for the advancement of future research.

Subcutaneous Administration

Subcutaneous injections of PEs are predominantly utilized in vaccines. A significant obstacle in vaccine development involves producing highly effective and safe adjuvants to assist antigens to elicit strong humoral and cellular immune responses.¹⁴⁴ Research on adjuvants has particularly concentrated on utilizing particulate carriers that imitate the characteristics of microorganisms or diseased cells.¹⁴⁵ PEs, which utilize solid particles as stabilizers, have shown promise in maintaining the deformability and migration of presented antigens while offering high biosafety and antigenloading capabilities.¹⁴⁶ To improve vaccine delivery efficiency, Du et al have developed a multicomponent cancer vaccine delivery system by incorporating target molecules, immunostimulants, and antigens into these emulsions.¹¹¹ In the B16-MUC1 tumor model, the large interfacial area of PEs enhanced interactions between antigens and antigenpresenting cells (APCs), thereby improving antigen uptake and promoting immune activation. Notably, no signs of inflammation or toxicity were observed in the treated mice, indicating a favorable biosafety profile and effective tumor suppression. Furthermore, adjuvants, in the form of solid particles, can stabilize PEs, as demonstrated by Peng and collaborators, who created an aluminum-stabilized PEs by adsorbing alum within squalene/water, based on the nature of the microgel of alum.¹³³ This approach not only addresses the limitations of alum in eliciting T-cell responses but also enables effective co-loading of antigens and adjuvants to enhance immune reactions. Analogously, Guo et al designed PEs loaded with adeno-associated viruses (AAVs), stabilized by biomineralized manganese nanoparticles and aluminum

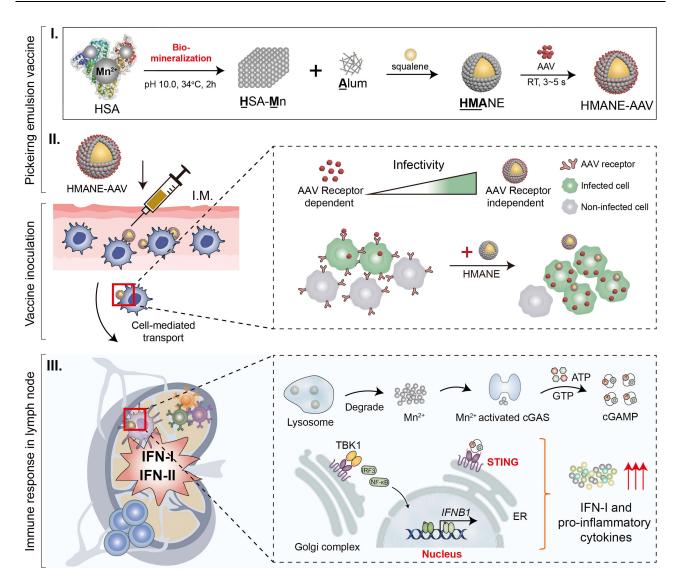


Figure 4 Schematic illustration of a biomineralized manganese nanoparticles and aluminum hydroxide stabilized PEs vaccine for enhancing AAV transduction and immune response. Reprinted from Guo Z, Wu F, Guo C, et al. Metalloparticle-engineered Pickering emulsion displaying AAV-vectored vaccine for enhancing antigen expression and immunogenicity against pathogens. Adv Mater. 2025;37(8):2412627.. *Copyright* © 2025 John Wiley & Sons, Inc.¹⁴⁷

hydroxide.¹⁴⁷ This platform conferred AAVs with favorable in vivo distribution kinetics and improved the safety of AAVs vaccine, ultimately enhancing AAVs infection efficiency in APCs (Figure 4). Furthermore, compared to the AAVs vaccination dose, the PEs with one-fifth AAVs induced effective immune responses. Stimuli-responsive PEs offer a promising approach for the controlled delivery of antigens and adjuvants tailored for cancer treatment requirements. For instance, researchers have successfully created PEs stabilized with Fe₃O₄ cellulose nanocomposites via ultrasound-assisted in situ co-precipitation.¹⁴⁸ This magnetic emulsion shows promise as a smart nanotherapeutic carrier for biomedical and drug delivery applications. Coating this Fe₃O₄-stabilized emulsion with antigens allowed for lymph node retention of the vaccine via subcutaneous delivery and external stimulation, thereby achieving strong activation of APCs.

Intravascular Administration

Intravascular administration of PEs is mainly used as the form of nanodroplets, especially in the treatment of tumors. Recent researchers have demonstrated that nanodroplets can passively target tumor tissues through enhanced penetration

and retention effects.⁶ For instance, utilizing the temperature-sensitive properties of poly(N-isopropylacrylamide), researchers have developed nanoscale droplets encapsulated with PTX,²⁷ inducing higher accumulation of the drug at the tumor site and demonstrating the capability of PEs for cancer therapy in the realm of nanomedicine. However, the tightness of the arrangement of nanoparticles at the water-oil interface may have an effect on the stability of the nanoemulsion. Consequently, more stable nano-PEs loaded with active molecules could be created using deformable or soft building blocks as nanoparticle emulsifiers, which were able to form stronger interfacial barriers to resist dropletdroplet coalescence.²⁷ Notably, once nanocarriers enter cells via endocytosis, they are typically captured by lysosomes, reducing the efficiency of drug delivery.¹⁴⁹ To address this issue, various delivery vectors with different lysosome escape mechanisms have been designed.^{150,151} For instance, PEs stabilized by hybridized nanoparticles (HNPs) composed of cetyl trimethylamine bromide (CTAB) and linoleic acid (LA) possessed the lysosomal escape ability. In an acidic environment, HNPs disintegrated the emulsion upon protonation, with CTAB disrupting lysosomal membranes and LA acting on lysosomal and mitochondrial membranes through interactions involving lysosomal iron ions, leading to tumor cell death.¹³⁴ In vivo studies in a CT26 melanoma tumor-bearing mouse model demonstrated that PEs stabilized by HNPs had no potential off-target toxicity in major organs, supporting the good biocompatibility of this formulation. If employed in the transportation of anticancer medications, these nanocarriers have the possibility to generate a more potent anti-tumor effect. While scholars have investigated the safety profile of the emulsion in healthy cells and animal models, establishing its remarkable tumor-targeting specificity with minimal off-target tissue accumulation, the cellular toxicity of CTAB warrants careful consideration. Further experimentation is imperative to delineate the safety parameters comprehensively.¹⁵²

Furthermore, hepatic artery injection embolization serves as a temporary reservoir for chemotherapeutic agents, allowing a controlled drug release into hepatic tumor tissues, thus minimizing systemic exposure.¹⁵³ Lipiodol, a radiopaque contrast agent, preferentially accumulates in liver tumor tissues and was adjuvanted with chemotherapeutic drugs to create stable PEs.¹³⁵ Li et al developed temperature-sensitive nanogels with a shell-and-core structure using two gel materials, resulting in Pickering gel emulsions with a temperature-sensitive sol-gel phase transition in vivo, enhancing intravascular embolization properties and long-term angiography of lipiodol in orthotopic liver cancer mice.¹³⁶ The Pickering gel emulsion has been shown to have 7.0 times higher mechanical strength than hairy p(NIPAM-acrylic acid) nanogels and better plugging properties through creep testing. In a recent study, calcium phosphate nanoparticles-stabilized lipiodol PEs were prepared for hepatocellular carcinoma, which encapsulated a clinically approved vascular disrupting agent. The emulsion responded to the tumor microenvironment and released combretastatin A4-phosphate to disrupt tumor nutrient delivery.¹⁵⁴ Therefore, these studies hold great promise for potential clinical translation of vascular embolization lipiodol PEs.^{12,154}

Overall, when formulating PEs for intravascular drug delivery systems, it is imperative to prioritize factors such as sterility and non-pyrogenicity to ensure both the safety and regulatory compliance of the system, thereby optimizing its therapeutic efficacy.¹⁵⁵

Intratumoral Administration

PEs are recognized for their suitability in delivering drugs within tumors as they exhibit excellent stability and the capability to regulate drug release effectively. Numerous research studies have revealed that administering low-selective chemotherapeutic agents systemically may lead to severe side effects due to dosage constraints. Over the past few decades, there have been significant advancements in the technology of directly injecting long-acting liquid formulations into tumors, effectively achieving high concentrations within the targeted tumor area.¹⁵⁶ Local administration of immunotherapy emerges as a promising strategy to mitigate the autoimmune and inflammatory toxicity associated with systemic delivery, thereby enhancing the therapeutic effectiveness of such treatments. Consequently, researchers developed a radiopaque PEs by dispersing an anti-CTLA-4 antibody in lipiodol with PLGA nanoparticles as stabilizers. The results demonstrated that the anti-CTLA-4 antibody's structure remained intact and retained its anti-tumor efficacy in vivo when released from this PEs.¹³⁷ In addition, Dai and his team designed a W/O PEs

gel loaded with PD-1 antibody in the internal phase and iron death inducer (1S, 3R)-RSL-3 (RSL-3) in the continuous phase, which promoted iron death of the tumor cells and activated the body's immune response upon intratumoral administration¹¹⁸ (Figure 2). Furthermore, numerous studies have explored the incorporation of small molecule inhibitors and anti-tumor drugs into PEs, which, when administered intratumorally, collaborate to enhance anti-tumor efficacy.^{6,36,83} These findings underscore the ability of intratumoral injections to effectively target the entire tumor and its microenvironment, showcasing significant promise in the field of oncology. PEs, with their unique slow-release property, deformability, and capacity to transport various medications, present a promising approach for intratumoral administration and controlled release of a diverse array of antineoplastic agents, showing great capability for therapeutic use in oncology.

Other Drug Delivery Systems Based on PEs Preparation Methods

It is challenging to achieve the encapsulation of two or more drugs in the same carrier using traditional nanocarriers. PEs, with their unique structural features, can fulfill this requirement. Researchers have utilized PEs' droplets as templates to develop multi-drug co-delivery systems, especially for tumor treatment. The fundamental concept of this method involves utilizing solid particles to stabilize oil-in-water emulsions, then fixing the solid particles through cross-linking or volatile oil phases to create hollow structures resembling microspheres.¹³⁹ These structures hold great ability as carriers for drug delivery. For instance, Hu et al employed galactose-functionalized hydroxyethyl starch-polycaprolactone nanoparticles as templates to create carbon nanotubes encapsulating DOX/indocyanine green.¹⁴⁰ These nanotubes demonstrated strong tumor-targeting capabilities through intravenous injection, enhanced tumor penetration, and facilitated photothermal effects. The researchers also identified these nanocolloids as a beneficial drug delivery system for actively targeting tumors in imaging-guided combination therapies for conditions like hepatocellular carcinoma.¹⁴⁰ Nanocapsules present an appealing approach to achieving highly efficient anti-cancer drug delivery. Traditional techniques like emulsion-diffusion and nanoprecipitation, which involve small-molecule surfactants, often result in high toxicity issues. To address this concern, researchers have utilized the Pickering technique to produce polymer nanogels for stabilizing nanocapsules, incorporating a cyclic peptide for enhanced tumor targeting and disulfide bond for stimulus-response. The capsules containing these are proposed as a great nanocarrier for precise drug release and targeted delivery.¹⁴¹ Furthermore, hydrogel-based drug delivery systems offer significant advantages in safeguarding bioactive compounds from degradation and improving their bioavailability. When integrated with PEs, a novel stimuli-responsive drug delivery system is created, characterized by improved stability and controlled drug release kinetics. For example, a hydrogel composed of carboxymethyl cellulose, sodium alginate, and knotted cold gel was utilized as a carrier for loading PEs containing hydrophobically active drugs, representing a promising immobilization and pH-responsive controlled drug release approach.142

Conclusion

PEs, stabilized by solid particles, offer a superior alternative to surfactant-based systems in pharmaceutics due to their enhanced stability, biocompatibility, and reduced toxicity. Unlike conventional emulsions, PEs enable controlled drug release, protect labile therapeutics, and facilitate co-delivery of multiple agents—critical for complex applications like cancer therapy and vaccine development. The versatility of particle stabilizers allows tailored formulations for diverse routes, including oral, transdermal, and intratumoral delivery.

However, despite their potential, there is a lack of commercial products and clinical applications for PEs, requiring more input from researchers in this field. Patent applications for PEs mainly focus on methods of preparing emulsions using solid particles, as well as applications in detergents and cosmetics. Only a limited number of patents exist for PEs intended for pharmaceutical and food purposes. Moreover, there is a lack of detailed in vivo toxicity studies of PEs, and more researchers are needed to fill the gaps in toxicity testing. Optical microscopy provides a simple and rapid way to observe PEs, but there are limitations in observing interfacial, phase, and morphological features of solid particles. Confocal laser scanning microscopy with appropriate dyes has been identified as one of the most suitable methods for evaluating emulsion properties.^{157,158} Some scholars are utilizing

neural network models to assess emulsion properties through object detection algorithms.¹⁵⁹ Moreover, research often lacks comparisons between different dosage forms or formulations against PEs for drug delivery in disease treatment. The industrial-scale production of PEs is still in its early stages, presenting challenges in predicting cost-effectiveness.⁸ Future studies are likely to focus on the preparation of versatile and stable PEs using safe and stable particles to cater to diverse delivery requirements.

Acknowledgment

This work was supported by the National Natural Science Foundation of China (82003683), the Sichuan Science and Technology Program (2023NSFSC1684), and the Fundamental Research Funds for the Central Universities, Southwest Minzu University (ZYN2024155).

Disclosure

The authors report no conflicts of interest in this work.

References

- 1. Wilson RJ, Li Y, Yang GZ, et al. Nanoemulsions for drug delivery. Particuology. 2022;64:85–97. doi:10.1016/j.partic.2021.05.009
- 2. Marto J, Gouveia L, Jorge IM, et al. Starch-based Pickering emulsions for topical drug delivery: a QbD approach. *Colloid Surf B-Biointerfaces*. 2015;135:183–192.
- 3. Wei YS, Niu ZC, Wang FQ, et al. A novel Pickering emulsion system as the carrier of tocopheryl acetate for its application in cosmetics. *Mater Sci Eng C-Mater Biol Appl.* 2020;109:110503.
- 4. Zhu F. Starch based Pickering emulsions: fabrication, properties, and applications. Trends Food Sci Technol. 2019;85:129-137.
- 5. Binks BP. Particles as surfactants—similarities and differences. Curr Opin Colloid Interface Sci. 2002;7(1-2):21-41.
- 6. Jia L, Pang M, Fan M et al. A pH-responsive Pickering nanoemulsion for specified spatial delivery of immune checkpoint inhibitor and chemotherapy agent to tumors. *Theranostics*. 2020;10(22):9956–9969. doi:10.7150/thno.46089
- 7. Harman CLG, Patel MA, Guldin S, et al. Recent developments in Pickering emulsions for biomedical applications. *Curr Opin Colloid Interface Sci.* 2019;39:173–189.
- Marto J, Ascenso A, Simoes S, et al. Pickering emulsions: challenges and opportunities in topical delivery. *Expert Opin Drug Deliv.* 2016;13 (8):1093–1107. doi:10.1080/17425247.2016.1182489
- 9. Wang D, Zhang L, Yang WH, et al. Arginine-loaded nano-calcium-phosphate-stabilized lipiodol Pickering emulsions potentiates transarterial embolization-immunotherapy. Adv Sci. 2025;12(6):13.
- Gu P, Zhu Y, Xu P, et al. Poria cocos polysaccharide-loaded Alum Pickering emulsion as vaccine adjuvant to enhance immune responses. Colloids Surf B Biointerfaces. 2024;244:114144. doi:10.1016/j.colsurfb.2024.114144
- 11. Liang L, Zhu J, Zhang Z, et al. Pickering emulsion stabilized by tea seed cake protein nanoparticles as lutein carrier. *Foods*. 2022;11(12):1712. doi:10.3390/foods11121712
- 12. Xia X, Li Y, Huang R, et al. A lipiodol Pickering emulsion stabilized by iron-doped carbon nanozymes for liver transarterial chemoembolization. *Adv Sci.* 2025;12(5):2410873. doi:10.1002/advs.202410873
- Lin X, Li S, Yin J, et al. Anthocyanin-loaded double Pickering emulsion stabilized by octenylsuccinate quinoa starch: preparation, stability and in vitro gastrointestinal digestion. Int J Biol Macromol. 2020;152:1233–1241. doi:10.1016/j.ijbiomac.2019.10.220
- Pan Y, Ma X, Sun J, et al. Fabrication and characterization of anthocyanin-loaded double Pickering emulsions stabilized by β-cyclodextrin. Int J Pharm. 2024;655:124003. doi:10.1016/j.ijpharm.2024.124003
- 15. Yang F, Wang Y, He H, et al. Construction of highly stable, monodisperse water-in-water Pickering emulsions with full particle coverage using a composite system of microfluidics and helical coiled tube. *Colloids Surf B Biointerfaces*. 2024;242:114079.
- Candiani A, Diana G, Martoccia M, et al. Microencapsulation of a Pickering oil/water emulsion loaded with vitamin D3. *Gels*. 2023;9(3):255. doi:10.3390/gels9030255
- Albert C, Beladjine M, Tsapis N, et al. Pickering emulsions: preparation processes, key parameters governing their properties and potential for pharmaceutical applications. J Control Release. 2019;309:302–332. doi:10.1016/j.jconrel.2019.07.003
- Saffarionpour S. Nanocellulose for stabilization of Pickering emulsions and delivery of nutraceuticals and its interfacial adsorption mechanism. Food Bioprocess Technol. 2020;13(8):1292–1328.
- 19. Ghosh S, Tran T, Rousseau D. Comparison of Pickering and network stabilization in water-in-oil emulsions. Langmuir. 2011;27(11):6589–6597.
- 20. Finkle P, Draper HD, Hildebrand JH. The theory of emulsification. J Am Chem Soc. 1923;45(12):2780-2788.
- Dapčević Hadnađev T, Dokić P, Krstonošić V, et al. Influence of oil phase concentration on droplet size distribution and stability of oil-in-water emulsions. Eur J Lipid Sci Technol. 2013;115(3):313–321.
- 22. Lu S, Yang DJ, Wang M, et al. Pickering emulsions synergistic-stabilized by amphoteric lignin and SiO₂ nanoparticles: stability and pH-responsive mechanism. *Colloid Surf A-Physicochem Eng Asp.* 2020;585:124158.
- 23. Binks BP, Clint JH. Solid wettability from surface energy components: relevance to Pickering emulsions. Langmuir. 2002;18(4):1270–1273.
- 24. Binks BP, Lumsdon SO. Pickering emulsions stabilized by monodisperse latex particles: effects of particle size. Langmuir. 2001;17 (15):4540-4547.
- 25. Destributs M, Gineste S, Laurichesse E, et al. Pickering emulsions: what are the main parameters determining the emulsion type and interfacial properties? *Langmuir*. 2014;30(31):9313–9326. doi:10.1021/la501299u

- 26. Zhang YX, Bao Y, Zhang WB, et al. Factors that affect Pickering emulsions stabilized by mesoporous hollow silica microspheres. J Colloid Interface Sci. 2023;633:1012–1021.
- Chen HB, Zhu HD, Hu JD, et al. Highly compressed assembly of deformable nanogels into nanoscale suprastructures and their application in nanomedicine. ACS Nano. 2011;5(4):2671–2680.
- 28. Scholz P, Keck CM. Nanoemulsions produced by rotor-stator high speed stirring. Int J Pharm. 2015;482(1-2):110-117.
- Athanasiou KA, NG G, Agrawal CM. Sterilization, toxicity, biocompatibility and clinical applications of polylactic acid/polyglycolic acid copolymers. *Biomaterials*. 1996;17(2):93–102. doi:10.1016/0142-9612(96)85754-1
- 30. Bai L, Greca LG, Xiang WC, et al. Adsorption and assembly of cellulosic and lignin colloids at oil/water interfaces. *Langmuir*. 2019;35 (3):571–588.
- 31. Gombotz WR, Wee SF. Protein release from alginate matrices. Adv Drug Del Rev. 2012;64:194-205.
- 32. Kragh-Hansen U. Structure and ligand binding properties of human serum albumin. Dan Med Bull. 1990;37(1):57-84.
- Pang L, Huang Z, Li R, et al. Reducing the allergenicity of whey proteins while improving their functional properties and bioactivity using combined enzymes. *Food Chem.* 2025;481:144052.
- 34. Yang J, Liu X, Yang H, et al. Combination application of Pickering emulsion and BLS protein nanoparticles enhances vaccine efficacy. Int J Biol Macromol. 2024;283:137635. doi:10.1016/j.ijbiomac.2024.137635
- Ben-Haim AE, Shalev N, Amalraj AJJ, et al. Nanocarriers for cancer-targeted delivery based on Pickering emulsions stabilized by casein nanoparticles. Int J Biol Macromol. 2025;298:140822.
- Wang C, Zhang L, Yang Z, et al. Self-fueling ferroptosis-inducing microreactors based on pH-responsive Lipiodol Pickering emulsions enable transarterial ferro-embolization therapy. *Natl Sci Rev.* 2023;11(1):nwad257.
- Yaremenko AV, Zelepukin IV, Ivanov IN, et al. Influence of magnetic nanoparticle biotransformation on contrasting efficiency and iron metabolism. J Nanobiotechnol. 2022;20(1):535.
- 38. Dickinson E. Hydrocolloids as emulsifiers and emulsion stabilizers. *Food Hydrocoll*. 2009;23(6):1473–1482.
- 39. Dong H, Ding QJ, Jiang YF, et al. Pickering emulsions stabilized by spherical cellulose nanocrystals. Carbohydr Polym. 2021;265:118101.
- 40. Kalashnikova I, Bizot H, Cathala B, et al. New Pickering emulsions stabilized by bacterial cellulose nanocrystals. *Langmuir*. 2011;27 (12):7471-7479.
- Bouhoute M, Taarji N, Felipe LD, et al. Microfibrillated cellulose from shells as sustainable solid particles for O/W Pickering emulsions. Carbohydr Polym. 2021;251:116990. doi:10.1016/j.carbpol.2020.116990
- 42. Wang C, Fu X, Tang CH, et al. Octenylsuccinate starch spherulites as a stabilizer for Pickering emulsions. Food Chem. 2017;227:298-304.
- 43. Ma YX, Li M, Chen ZD, et al. Construction of butyric acid modified porous starch for stabilizing Pickering emulsions: encapsulation of paclitaxel. *Food Hydrocoll*. 2023;142:108858.
- Alehosseini E, Jafari SM, Tabarestani HS. Production of D-limonene-loaded Pickering emulsions stabilized by chitosan nanoparticles. Food Chem. 2021;354:129591. doi:10.1016/j.foodchem.2021.129591
- Shomali Z, Fatehi P. Carboxyalkylated lignin nanoparticles with enhanced functionality for oil-water Pickering emulsion systems. ACS Sustain Chem Eng. 2022;10(50):16563–16577.
- Dai L, Li YT, Kong FG, et al. Lignin-based nanoparticles stabilized Pickering emulsion for stability improvement and thermal-controlled release of-resveratrol. ACS Sustain Chem Eng. 2019;7(15):13497–13504.
- Richter AR, Feitosa JPA, Paula HCB, et al. Pickering emulsion stabilized by cashew gum- poly-L-lactide copolymer nanoparticles: synthesis, characterization and amphotericin B encapsulation. *Colloid Surf B-Biointerfaces*. 2018;164:201–209.
- Deschamps F, Isoardo T, Denis S, et al. Biodegradable Pickering emulsions of lipiodol for liver trans-arterial chemo-embolization. Acta Biomater. 2019;87:177–186. doi:10.1016/j.actbio.2019.01.054
- 49. Heidari F, Jafari SM, Ziaiifar AM, et al. Preparation of Pickering emulsions stabilized by modified silica nanoparticles via the Taguchi approach. *Pharmaceutics*. 2022;14(8):1561. doi:10.3390/pharmaceutics14081561
- Tang Q, Xie XL, Li CJ, et al. Medium-chain triglyceride/water Pickering emulsion stabilized by phosphatidylcholine-kaolinite for encapsulation and controlled release of curcumin. *Colloid Surf B-Biointerfaces*. 2019;183:110414.
- Huang ZY, Koubek JT, Sellinger A, et al. Pickering emulsions of self-assembled lead sulfide quantum dots with janus-ligand shells as nanoreactors for photocatalytic reactions. ACS Appl Nano Mater. 2022;5(3):3183–3187.
- 52. Xie CY, Meng SX, Xue LH, et al. Light and magnetic dual-responsive Pickering emulsion micro reactors. *Langmuir.* 2017;33 (49):14139–14148. doi:10.1021/acs.langmuir.7b03642
- Sun Y, Zhong M, Liao Y, et al. Pickering emulsions stabilized by reassembled oleosome protein nanoparticles for co-encapsulating hydrophobic nutrients. Food Hydrocoll. 2023;138:108445.
- Xu BM, Liu CK, Sun HY, et al. Oil-in-water Pickering emulsions using a protein nano-ring as high-grade emulsifiers. Colloid Surf B-Biointerfaces. 2020;187:110646.
- 55. Feng YM, Lee Y. Surface modification of zein colloidal particles with sodium caseinate to stabilize oil-in-water Pickering emulsion. *Food Hydrocoll*. 2016;56:292–302.
- Wang XL, Liao ZG, Zhao GW, et al. Curcumin nanocrystals self-stabilized Pickering emulsion freeze-dried powder: development, characterization, and suppression of airway inflammation. Int J Biol Macromol. 2023;245:125493. doi:10.1016/j.ijbiomac.2023.125493
- Li GY, Lee WJ, Tan CP, et al. Tailored rigidity of W/O Pickering emulsions using diacylglycerol-based surface-active solid lipid nanoparticles. Food Funct. 2021;12(23):11732–11746. doi:10.1039/d1fo01883c
- Wang ZC, Gao SJ, Liu XX, et al. Programming self-assembly of tobacco mosaic virus coat proteins at Pickering emulsion interfaces for nanorod-constructed capsules. ACS Appl Mater Interfaces. 2017;9(33):27383–27389. doi:10.1021/acsami.7b08186
- 59. Zhu YZ, Teng OH, Sun DS, et al. Convenient synthesis of uncovered imprinted microspheres by spore-stabilized Pickering emulsion polymerization and their enhanced recognition of spiramycin. RSC Adv. 2019;9(60):34772-34783. doi:10.1039/c9ra07745f
- Firoozmand H, Rousseau D. Microbial cells as colloidal particles: Pickering oil-in-water emulsions stabilized by bacteria and yeast. Food Res Int. 2016;81:66–73.
- Zhang XZ, Liu YL, Wang YX, et al. Surface modification of cellulose nanofibrils with protein nanoparticles for enhancing the stabilization of O/W Pickering emulsions. *Food Hydrocoll*. 2019;97:105180. doi:10.1016/j.foodhyd.2019.105180

- 62. Low LE, Tan LTH, Goh BH, et al. Magnetic cellulose nanocrystal stabilized Pickering emulsions for enhanced bioactive release and human colon cancer therapy. *Int J Biol Macromol.* 2019;127:76–84. doi:10.1016/j.ijbiomac.2019.01.037
- 63. Li WX, Ju BZ, Zhang SF. Novel amphiphilic cellulose nanocrystals for pH-responsive Pickering emulsions. *Carbohydr Polym*. 2020;229:115401. doi:10.1016/j.carbpol.2019.115401
- Abeer MM, Amin MCIM, Martin C. A review of bacterial cellulose-based drug delivery systems: their biochemistry, current approaches and future prospects. J Pharm Pharmacol. 2014;66(8):1047–1061.
- 65. Ji CY, Wang YX. Nanocellulose-stabilized Pickering emulsions: fabrication, stabilization, and food applications. Adv Colloid Interface Sci. 2023;318:102970.
- 66. Li C, Li YX, Sun PD, et al. Pickering emulsions stabilized by native starch granules. Colloid Surf A-Physicochem Eng Asp. 2013;431:142–149.
- 67. Saari H, Fuentes C, Sjöö M, et al. Production of starch nanoparticles by dissolution and non-solvent precipitation for use in food-grade Pickering emulsions. *Carbohydr Polym.* 2017;157:558–566. doi:10.1016/j.carbpol.2016.10.003
- 68. Guida C, Aguiar AC, Cunha RL. Green techniques for starch modification to stabilize Pickering emulsions: a current review and future perspectives. *Curr Opin Food Sci.* 2021;38:52–61.
- 69. Yu SW, Xu XL, Feng JF, et al. Chitosan and chitosan coating nanoparticles for the treatment of brain disease. Int J Pharm. 2019;560:282-293.
- Ho KW, Ooi CW, Mwangi WW, et al. Comparison of self-aggregated chitosan particles prepared with and without ultrasonication pretreatment as Pickering emulsifier. *Food Hydrocoll*. 2016;52:827–837.
- 71. Souza EMC, Ferreira MRA, Soares LAL. Pickering emulsions stabilized by zein particles and their complexes and possibilities of use in the food industry: a review. *Food Hydrocoll*. 2022;131:20.
- 72. Wang LJ, Yin SW, Wu LY, et al. Fabrication and characterization of Pickering emulsions and oil gels stabilized by highly charged zein/chitosan complex particles (ZCCPs). *Food Chem*. 2016;213:462–469. doi:10.1016/j.foodchem.2016.06.119
- 73. Meng W, Sun H, Mu T, et al. Chitosan-based Pickering emulsion: a comprehensive review on their stabilizers, bioavailability, applications and regulations. *Carbohydr Polym*. 2023;304:120491. doi:10.1016/j.carbpol.2022.120491
- 74. Xiao Q, Chen ZZ, Xie XJ, et al. A novel Pickering emulsion stabilized solely by hydrophobic agar microgels. *Carbohydr Polym*. 2022;297:120035. doi:10.1016/j.carbpol.2022.120035
- 75. Sathawong S, Sridach W, Techato KA. Lignin: isolation and preparing the lignin based hydrogel. J Environ Chem Eng. 2018;6(5):5879–5888.
- Zhang W, Shen JD, Gao P, et al. An eco-friendly strategy for preparing lignin nanoparticles by self-assembly: characterization, stability, bioactivity, and Pickering emulsion. *Ind Crops Prod.* 2022;188:115651.
- 77. Calabrese V, Courtenay JC, Edler KJ, et al. Pickering emulsions stabilized by naturally derived or biodegradable particles. *Curr Opin Green Sustainable Chem.* 2018;12:83–90.
- Banik BL, Fattahi P, Brown JL. Polymeric nanoparticles: the future of nanomedicine. Wiley Interdiscip Rev-Nanomed Nanobiotechnol. 2016;8 (2):271–299. doi:10.1002/wnan.1364
- Grabowski N, Hillaireau H, Vergnaud J, et al. Surface coating mediates the toxicity of polymeric nanoparticles towards human-like macrophages. Int J Pharm. 2015;482(1-2):75–83.
- Jana U, Mohanty AK, Pal SL, et al. Felodipine loaded PLGA nanoparticles: preparation, physicochemical characterization and in vivo toxicity study. Nano Convergence. 2014;1(1):31.
- Zhou J, Wang L, Qiao X, et al. Pickering emulsions stabilized by surface-modified Fe3O4 nanoparticles. J Colloid Interface Sci. 2012;367:213–224. doi:10.1016/j.jcis.2011.11.001
- Song XY, Pei YQ, Zhu W, et al. Particle-stabilizers modified from rice starches differing in amylose content. Food Chem. 2014;153:74–80. doi:10.1016/j.foodchem.2013.12.046
- Li Z, Liu X, Xiao J, et al. Ultrastable iodinated oil-based Pickering emulsion enables locoregional sustained codelivery of hypoxia inducible factor-1 inhibitor and anticancer drugs for tumor combination chemotherapy. ACS Biomater Sci Eng. 2024;10(4):2270–2281.
- 84. Kettiger H, Sen Karaman D, Schiesser L, et al. Comparative safety evaluation of silica-based particles. Toxicol Vitro. 2015;30(1):355-363.
- Zhang T, Xu JM, Chen JH, et al. Protein nanoparticles for Pickering emulsions: a comprehensive review on their shapes, preparation methods, and modification methods. *Trends Food Sci Technol.* 2021;113:26–41.
- 86. Lim S, Salentinig S. Protein nanocage-stabilized Pickering emulsions. Curr Opin Colloid Interface Sci. 2021;56:101485.
- Zeng T, Wu ZL, Zhu JY, et al. Development of antioxidant Pickering high internal phase emulsions (HIPEs) stabilized by protein/polysaccharide hybrid particles as potential alternative for PHOs. *Food Chem.* 2017;231:122–130. doi:10.1016/j.foodchem.2017.03.116
- Yan XJ, Ma CC, Cui FZ, et al. Protein-stabilized Pickering emulsions: formation, stability, properties, and applications in foods. *Trends Food Sci Technol*. 2020;103:293–303.
- Huang G, Liu GC, Xu ZJ, et al. Stability, rheological behavior and microstructure of Pickering emulsions co-stabilized by soy protein and carboxymethyl chitosan. *Food Hydrocoll*. 2023;142:108773.
- Yan SZ, Regenstein JM, Zhang S, et al. Edible particle-stabilized water-in-water emulsions: stabilization mechanisms, particle types, interfacial design, and practical applications. *Food Hydrocoll*. 2023;140:108665.
- 91. Zhang JF, Zhang J, Wang S, et al. Development of an oral compound Pickering emulsion composed of nanocrystals of poorly soluble ingredient and volatile oils from traditional Chinese medicine. *Pharmaceutics*. 2018;10(4):170. doi:10.3390/pharmaceutics10040170
- 92. Yi T, Liu C, Zhang J, et al. A new drug nanocrystal self-stabilized Pickering emulsion for oral delivery of silybin. *Eur J Pharm Sci.* 2017;96:420–427. doi:10.1016/j.ejps.2016.08.047
- Wang Z, Dai B, Tang XH, et al. Fabrication and in vitro/vivo evaluation of drug nanocrystals self-stabilized Pickering emulsion for oral delivery of quercetin. *Pharmaceutics*. 2022;14(5):897. doi:10.3390/pharmaceutics14050897
- 94. Sheng YZ, Yu Q, Huang YP, et al. Pickering emulsions enhance oral bioavailability of curcumin nanocrystals: the effect of oil types. *Pharmaceutics*. 2023;15(5):1341. doi:10.3390/pharmaceutics15051341
- Kalaycioglu GD, Aydogan N. Preparation and investigation of solid lipid nanoparticles for drug delivery. Colloid Surf A-Physicochem Eng Asp. 2016;510:77–86.
- 96. Tenorio-Garcia E, Araiza-Calahorra A, Rappolt M, et al. Pickering water-in-oil emulsions stabilized solely by fat crystals. *Adv Mater Interfaces*. 2023;10(31):2300190.

- 97. Bibi M, Din F, Anwar Y, et al. Cilostazol-loaded solid lipid nanoparticles: bioavailability and safety evaluation in an animal model. J Drug Deliv Sci Technol. 2022;74:103581.
- 98. Nkanga CI, Steinmetz NF. The pharmacology of plant virus nanoparticles. Virology. 2021;556:39–61. doi:10.1016/j.virol.2021.01.012
- 99. Jeevanandam J, Pal K, Danquah MK. Virus-like nanoparticles as a novel delivery tool in gene therapy. *Biochimie*. 2019;157:38–47. doi:10.1016/j.biochi.2018.11.001
- 100. Cheng T, Zhang G, Sun F, et al. Study on stabilized mechanism of high internal phase Pickering emulsions based on commercial yeast proteins: modulating the characteristics of Pickering particle via sonication. Ultrason Sonochem. 2024;104:106843. doi:10.1016/j.ultsonch.2024.106843
- Ribeiro EF, Morell P, Nicoletti VR, et al. Protein- and polysaccharide-based particles used for Pickering emulsion stabilisation. Food Hydrocoll. 2021;119:106839.
- Sarkar A, Ademuyiwa V, Stubley S, et al. Pickering emulsions co-stabilized by composite protein/ polysaccharide particle-particle interfaces: impact on gastric stability. *Food Hydrocoll*. 2018;84:282–291.
- Wang W, Liu X, Gao X, et al. Characterization, digestive properties and glucose metabolism regulation of curcumin-loaded Pickering emulsion. Carbohydr Polym. 2025;356:123408. doi:10.1016/j.carbpol.2025.123408
- 104. Liang HN, Tang CH. pH-dependent emulsifying properties of pea [Pisum sativum (L.)] proteins. Food Hydrocoll. 2013;33(2):309–319. doi:10.1016/j.foodhyd.2013.04.005
- 105. Shao Y, Tang CH. Gel-like pea protein Pickering emulsions at pH 3.0 as a potential intestine-targeted and sustained-release delivery system for β-carotene. Food Res Int. 2016;79:64–72. doi:10.1016/j.foodres.2015.11.025
- 106. Cai LY, Cao MJ, Regenstein J. Slow-release and nontoxic Pickering emulsion platform for antimicrobial peptide. J Agric Food Chem. 2020;68 (28):7453–7466. doi:10.1021/acs.jafc.0c00874
- Llovet JM, Lencioni R, Di Bisceglie AM, et al. EASL-EORTC clinical practice guidelines: management of hepatocellular carcinoma. *J Hepatol.* 2012;56(4):908–943.
- Ciftei F, Özarslan AC, Kantarci İC, et al. Advances in drug targeting, drug delivery, and nanotechnology applications: therapeutic significance in cancer treatment. *Pharmaceutics*. 2025;17(1):121. doi:10.3390/pharmaceutics17010121
- 109. Ming Y, Xia Y, Ma G. Aggregating particles on the O/W interface: tuning Pickering emulsion for the enhanced drug delivery systems. Aggregate. 2022;3(2):e162.
- Ni QQ, Zhang FW, Liu YJ, et al. A bi-adjuvant nanovaccine that potentiates immunogenicity of neoantigen for combination immunotherapy of colorectal cancer. Sci Adv. 2020;6(12):eaaw6071. doi:10.1126/sciadv.aaw6071
- 111. Du YQ, Song TT, Wu J, et al. Engineering mannosylated Pickering emulsions for the targeted delivery of multicomponent vaccines. *Biomaterials*. 2022;280:121313. doi:10.1016/j.biomaterials.2021.121313
- 112. Tang JT, Quinlan PJ, Tam KC. Stimuli-responsive Pickering emulsions: recent advances and potential applications. *Soft Matter*. 2015;11 (18):3512–3529. doi:10.1039/c5sm00247h
- Zhang QM, Zhao QF, Zhu BB, et al. Acetalized starch-based nanoparticles stabilized acid-sensitive Pickering emulsion as a potential antitumor drug carrier. Int J Biol Macromol. 2023;244:125393. doi:10.1016/j.ijbiomac.2023.125393
- Dieng SM, Anton N, Bouriat P, et al. Pickering nano-emulsions stabilized by solid lipid nanoparticles as a temperature sensitive drug delivery system. Soft Matter. 2019;15(40):8164–8174. doi:10.1039/c9sm01283d
- 115. Yang GB, Liu JJ, Wu YF, et al. Near-infrared-light responsive nanoscale drug delivery systems for cancer treatment. *Coord Chem Rev.* 2016;320:100–117.
- Acter S, Vidallon MLP, King JP, et al. Photothermally responsive Pickering emulsions stabilised by polydopamine nanobowls. J Mater Chem B. 2021;9(43):8962–8970. doi:10.1039/d1tb01796a
- 117. Li XM, Li XH, Wu ZZ, et al. Chitosan hydrochloride/carboxymethyl starch complex nanogels stabilized Pickering emulsions for oral delivery of β-carotene: protection effect and in vitro digestion study. *Food Chem.* 2020;315:126288. doi:10.1016/j.foodchem.2020.126288
- Dai X, Zhang J, Bao X, et al. Induction of tumor ferroptosis-dependent immunity via an injectable attractive Pickering emulsion gel. Adv Mater. 2023;35(35):2303542.
- Wu JY, Chen J, Feng YJ, et al. An immune cocktail therapy to realize multiple boosting of the cancer-immunity cycle by combination of drug/ gene delivery nanoparticles. Sci Adv. 2020;6(40):eabc7828. doi:10.1126/sciadv.abc7828
- 120. Deng L, Liao J, Liu W, et al. Study of Zn2+ complexation driven zein-based W/O/W Pickering emulsions for enhancing curcumin bioaccessibility and fluorouracil bioactivity. *Food Hydrocoll*. 2025;164:111243.
- 121. Tang XY, Wang ZM, Meng HC, et al. Robust W/O/W emulsion stabilized by genipin-cross-linked sugar beet pectin-bovine serum albumin nanoparticles: co-encapsulation of betanin and curcumin. J Agric Food Chem. 2021;69(4):1318–1328. doi:10.1021/acs.jafc.0c05212
- 122. Li H, Li J, Su T, et al. Construction of anti-bubble microcapsule based on Pickering emulsion stabilization system and its protective effect on probiotics. *LWT*. 2025;217:117405.
- 123. Sitovs A, Cernisova V, Mohylyuk V. Dissolution-permeation systems in biopharmaceutical research: a graphical review on the current state-ofthe-art. J Drug Deliv Sci Technol. 2025;107:106814.
- 124. Jadhav NV, Pawar MA, Vavia PR. Pickering dry emulsion system for improved oral delivery of fenofibrate. *AAPS Pharm Sci Tech*. 2022;23 (6):168.
- 125. Dieng SM, Omran Z, Anton N, et al. Pickering nano-emulsions stabilized by Eudragit RL100 nanoparticles as oral drug delivery system for poorly soluble drugs. *Colloid Surf B-Biointerfaces*. 2020;191:111010.
- 126. Hu H, Wang Y, Zhao D-G, et al. Oral delivery of 5-demethylnobiletin by media-milled black rice particle-stabilized Pickering emulsion alleviates colitis in mice: synergistic effects of carrier and loaded bioactive. J Agric Food Chem. 2025;73(2):1257–1272. doi:10.1021/acs. jafc.4c08558
- 127. Tai Z, Huang Y, Zhu Q, et al. Utility of Pickering emulsions in improved oral drug delivery. Drug Discov Today. 2020;25(11):2038–2045.
- 128. Zhou Y, Hua J, Ding D, et al. Interrogating amyloid aggregation with aggregation-induced emission fluorescence probes. *Biomaterials*. 2022;286:121605. doi:10.1016/j.biomaterials.2022.121605
- Frelichowska J, Bolzinger MA, Pelletier J, et al. Topical delivery of lipophilic drugs from o/w Pickering emulsions. Int J Pharm. 2009;371 (1–2):56–63. doi:10.1016/j.ijpharm.2008.12.017

- 130. Marto J, Duarte A, Simoes S, et al. Starch-based Pickering emulsions as platforms for topical antibiotic delivery: in vitro and in vivo studies. *Polymers*. 2019;11(1):108. doi:10.3390/polym11010108
- 131. Sharkawy A, Barreiro MF, Rodrigues AE. New Pickering emulsions stabilized with chitosan/collagen peptides nanoparticles: synthesis, characterization and tracking of the nanoparticles after skin application. *Colloid Surf A-Physicochem Eng Asp.* 2021;616:126327.
- 132. Wu XR, Zhang QA, Wang ZM, et al. Investigation of construction and characterization of carboxymethyl chitosan- sodium alginate nanoparticles to stabilize Pickering emulsion hydrogels for curcumin encapsulation and accelerating wound healing. Int J Biol Macromol. 2022;209:1837–1847. doi:10.1016/j.ijbiomac.2022.04.157
- 133. Peng S, Cao FQ, Xia YF, et al. Particulate alum via Pickering emulsion for an enhanced COVID-19 vaccine adjuvant. Adv Mater. 2020;32 (40):2004210.
- 134. Chen Y, Wang SB, Ma Q, et al. Utilizing endosomal capture for tumor therapy via membrane-lytic mechanism-based Pickering emulsion. J Control Release. 2023;354:523–537. doi:10.1016/j.jconrel.2023.01.035
- 135. Deschamps F, Harris KR, Moine L, et al. Pickering-emulsion for liver trans-arterial chemo-embolization with oxaliplatin. *Cardiovasc Interv Radiol.* 2018;41(5):781–788.
- Li H, Qian K, Zhang HS, et al. Pickering gel emulsion of lipiodol stabilized by hairy nanogels for intra-artery embolization antitumor therapy. Chem Eng J. 2021;418:129534.
- 137. Tselikas L, de Baere T, Isoardo T, et al. Pickering emulsions with ethiodized oil and nanoparticles for slow release of intratumoral anti-CTLA4 immune checkpoint antibodies. J Immunother Cancer. 2020;8(1):e000579. doi:10.1136/jitc-2020-000579
- 138. Xu YR, Asghar S, Li HY, et al. Preparation of a paclitaxel-loaded cationic nanoemulsome and its biodistribution via direct intratumoral injection. *Colloid Surf B-Biointerfaces*. 2016;142:81–88.
- Yu SM, Peng GN, Jiao JL, et al. Chitin nanocrystals-stabilized emulsion as template for fabricating injectable suspension containing polylactide hollow microspheres. Carbohydr Polym. 2024;337:122176.
- Hu H, Xiao C, Wu HL, et al. Nanocolloidosomes with selective drug release for active tumor-targeted imaging-guided photothermal/chemo combination therapy. ACS Appl Mater Interfaces. 2017;9(48):42225–42238.
- 141. Shang XX, Liu Q, Qin T, et al. Fabrication of cRGD-modified reduction-sensitive nanocapsule via Pickering emulsion route to facilitate tumor-targeted delivery. *Int J Nanomed*. 2019;14:3361–3373.
- 142. Zheng WX, Zhang HZ, Wang J, et al. Pickering emulsion hydrogel based on alginate-gellan gum with carboxymethyl chitosan as a pH-responsive controlled release delivery system. *Int J Biol Macromol.* 2022;216:850–859. doi:10.1016/j.ijbiomac.2022.07.223
- 143. Ma JQ, Wang YC, Lu R. Mechanism and application of chitosan and its derivatives in promoting permeation in transdermal drug delivery systems: a review. *Pharmaceuticals*. 2022;15(4):459. doi:10.3390/ph15040459
- 144. D'Amico C, Fontana F, Cheng RY, et al. Development of vaccine formulations: past, present, and future. *Drug Deliv Transl Res.* 2021;11 (2):353–372. doi:10.1007/s13346-021-00924-7
- 145. Cui L, Yao H, Xue F, et al. A tumor-targeting nano-adjuvant for in situ vaccine based on ultrasound therapy. Aggregate. 2024;5(3):e504.
- 146. Xia YF, Wu J, Wei W, et al. Exploiting the pliability and lateral mobility of Pickering emulsion for enhanced vaccination. *Nat Mater.* 2018;17 (2):187–194. doi:10.1038/nmat5057
- Guo Z, Wu F, Guo C, et al. Metalloparticle-engineered Pickering emulsion displaying AAV-vectored vaccine for enhancing antigen expression and immunogenicity against pathogens. Adv Mater. 2025;37(8):2412627.
- Low LE, Tey BT, Ong BH, et al. Palm olein-in-water Pickering emulsion stabilized by Fe₃O₄-cellulose nanocrystal nanocomposites and their responses to pH. Carbohydr Polym. 2017;155:391–399. doi:10.1016/j.carbpol.2016.08.091
- Degors IMS, Wang CF, Rehman ZU, et al. Carriers break barriers in drug delivery: endocytosis and endosomal escape of gene delivery vectors. Acc Chem Res. 2019;52(7):1750–1760.
- Butt AM, Abdullah N, Rani NNIM, et al. Endosomal escape of bioactives deployed nanocarriers: insights into the design of polymeric micelles. *Pharm Res.* 2022;39(6):1047–1064. doi:10.1007/s11095-022-03296-w
- Ma YT, Fenton OS. Tannic acid lipid nanoparticles can deliver messenger RNA payloads and improve their endosomal escape. Adv Therap. 2023;6(6):2200305.
- 152. Han H-W, Joe A, Jang E-S. Reduced cytotoxicity of CTAB-templated silica layer on gold nanorod using fluorescence dyes and its application in cancer theranostics. *J Ind Eng Chem.* 2021;96:202–212.
- 153. Cheng Z, Xu S, Lei Y, et al. A novel protein-polysaccharide oral nanoemulsion targeting activated hepatic stellate cells to enhance the therapeutic effect of pirfenidone on fibrosis after transarterial chemoembolization for liver cancer. *Adv Funct Mater.* 2025;2411665.
- Zhang L, Wang D, Zhang L-Z, et al. Pickering emulsion with tumor vascular destruction and microenvironment modulation for transarterial embolization therapy. *Biomaterials*. 2025;316:123018.
- 155. Domańska IM, Oledzka E, Sobczak M. Sterilization process of polyester based anticancer-drug delivery systems. Int J Pharm. 2020;587:119663. doi:10.1016/j.ijpharm.2020.119663
- 156. Kim DY, Kwon DY, Kwon JS, et al. Synergistic anti-tumor activity through combinational intratumoral injection of an in-situ injectable drug depot. *Biomaterials*. 2016;85:232–245. doi:10.1016/j.biomaterials.2016.02.001
- 157. Xue Y, Li XF, Dong JF. Interfacial characteristics of block copolymer micelles stabilized Pickering emulsion by confocal laser scanning microscopy. J Colloid Interface Sci. 2020;563:33–41. doi:10.1016/j.jcis.2019.12.016
- 158. Zembyla M, Lazidis A, Murray BS, et al. Stability of water-in-oil emulsions co-stabilized by polyphenol crystal-protein complexes as a function of shear rate and temperature. *J Food Eng.* 2020;281:109991.
- 159. Huang ZY, Ni Y, Yu Q, et al. Deep learning in food science: an insight in evaluating Pickering emulsion properties by droplets classification and quantification via object detection algorithm. *Adv Colloid Interface Sci.* 2022;304:102663. doi:10.1016/j.cis.2022.102663

International Journal of Nanomedicine

Dovepress Taylor & Francis Group

Publish your work in this journal

The International Journal of Nanomedicine is an international, peer-reviewed journal focusing on the application of nanotechnology in diagnostics, therapeutics, and drug delivery systems throughout the biomedical field. This journal is indexed on PubMed Central, MedLine, CAS, SciSearch[®], Current Contents[®]/Clinical Medicine, Journal Citation Reports/Science Edition, EMBase, Scopus and the Elsevier Bibliographic databases. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit http:// www.dovepress.com/testimonials.php to read real quotes from published authors.

Submit your manuscript here: https://www.dovepress.com/international-journal-of-nanomedicine-journal

🖪 💥 in 🔼 5947