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ORIGINAL RESEARCH

Size-dependent antimicrobial properties of CuO nanoparticles against Gram-positive and -negative bacterial strains

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Background: CuO is one of the most important transition metal oxides due to its captivating properties. It is used in various technological applications such as high critical temperature superconductors, gas sensors, in photoconductive applications, and so on. Recently, it has been used as an antimicrobial agent against various bacterial species. Here we synthesized different sized CuO nanoparticles and explored the size-dependent antibacterial activity of each CuO nanoparticles preparation.

Methods: CuO nanoparticles were synthesized using a gel combustion method. In this approach, cupric nitrate trihydrate and citric acid were dissolved in distilled water with a molar ratio of 1:1. The resulting solution was stirred at 100°C, until gel was formed. The gel was allowed to burn at 200°C to obtain amorphous powder, which was further annealed at different temperatures to obtain different size CuO nanoparticles. We then tested the antibacterial properties using well diffusion, minimum inhibitory concentration, and minimum bactericidal concentration methods.

Results: XRD spectra confirmed the formation of single phase CuO nanoparticles. Crystallite size was found to increase with an increase in annealing temperature due to atomic diffusion. A minimum crystallite size of 20 nm was observed in the case of CuO nanoparticles annealed at 400°C. Transmission electron microscopy results corroborate well with XRD results. All CuO nanoparticles exhibited inhibitory effects against both Gram-positive and -negative bacteria. The size of the particles was correlated with its antibacterial activity.

Conclusion: The antibacterial activity of CuO nanoparticles was found to be size-dependent. In addition, the highly stable minimum-sized monodispersed copper oxide nanoparticles synthesized during this study demonstrated a significant increase in antibacterial activities against both Gram-positive and -negative bacterial strains.

Keywords: CuO, nanoparticles, X-ray diffraction, FTIR, antimicrobial activity

Introduction

The unique, unusual and interesting physical, chemical, and biological properties of nanometer-sized materials have recently attracted a great deal of interest in the scientific community. As the size of materials is reduced to the nanometer regime the resulting properties change noticeably. Considerable efforts have been made to characterize and describe the physical and chemical properties of metal oxide nanomaterials because of their significant applications in numerous technological fields. ¹⁻⁴ The oxides of transition metals are an important class of semiconductors that have wider applications in magnetic storage media, solar energy transformation, electronics, and catalysis. 5-12 Among various transition metal oxides, copper oxide (CuO) has attracted greater attention due

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to its fascinating properties such as the basis of high critical temperature (T) superconductors. CuO is a semiconducting compound with a narrow band gap and is used for photoconductive and photothermal applications.¹³ Reports on the preparation and characterization of nanocrystalline CuO are few compared to those on some other transition metal oxides, such as zinc oxide, titanium dioxide, tin dioxide, and iron oxide. Some methods for the preparation of nanocrystalline CuO have been reported such as the sonochemical method,14 sol-gel technique,15 one-step solid state reaction method at room temperature,16 electrochemical method,17 and thermal decomposition of precursors.¹⁸ Copper can also be used as an antimicrobial agent, and CuO nanoparticles have been investigated previously for enhancing antibacterial properties. 19-22 Here we report a novel gel-combustion method, controlling the size of the synthesized nanoparticles and its effect on antimicrobial characteristics. The bactericidal property of such nanoparticles depends on their size, stability, and concentration added to the growth medium, since this provides greater retention time for bacterium nanoparticles interaction. In general, bacterial cells are in the micron-sized range. Most bacterial cells have cellular membranes that contain pores in the nanometer range. A unique property of crossing the cell membrane can potentially be attributed to synthesized nanoparticles through such bacterial pores. However, to make this possible, it is important to overcome challenges and prepare/ design nanoparticles which are stable enough to significantly restrict bacterial growth while crossing the cell membrane.

To realize the potential of CuO nanoparticles to act as antimicrobial agents, we synthesized different sized CuO nanoparticles by controlling the annealing temperature during the gel-combustion synthesis. Furthermore, the antibacterial activities of CuO nanoparticles against two Gram-positive bacteria (*Staphylococcus aureus* and *Bacillus subtilis*) and two Gram-negative bacteria (*Pseudomonas aeruginosa* and *Escherichia coli*) were investigated.

Materials and methods Synthesis of CuO nanoparticles

In a typical synthesis procedure, $\text{Cu(NO}_3)_2 \cdot 3\text{H}_2\text{O}$ and citric acid were dissolved in distilled water with a molar ratio of 1:1. The solution was stirred with a magnetic stirrer at 100°C . Stirring continued until gel formation (approximately 1 hour). Afterwards, the gel was allowed to burn at 200°C . A light fluffy mass was obtained as a result of combustion, which was further annealed for 2 hours at varying temperatures, 400°C , 500°C , 600°C , and 700°C , to obtain the highly crystalline CuO nanoparticles.

Characterization of CuO nanoparticles

Synthesized CuO nanoparticles were characterized by X-ray diffraction (XRD), Fourier-transform infrared spectroscopy (FTIR), Raman spectroscopy, and transmission electron microscopy (TEM). Crystallinity, structure, and crystallite size of CuO nanoparticles were determined by XRD technique using a Rigaku-Miniflex X-ray diffractometer (Rigaku Corporation, Tokyo, Japan) with Cu-K α radiations (λ =0.15406 nm) in 2 θ range from 20° to 80°. TEM analysis was carried out using a 200 kV JEOL transmission electron microscope (JEOL Ltd, Tokyo, Japan). FTIR spectra of the samples were obtained using a PerkinElmer FTIR spectrophotometer (PerkinElmer Inc, Waltham, MA).

Determination of antimicrobial activity by the well-diffusion method

Antimicrobial activities of the synthesized CuO nanoparticles of different sizes were determined using Gram-negative bacteria (E. coli and P. aeruginosa) and Gram-positive bacteria (B. subtilis and S. aureus) following a modified Kirby Bauer disc diffusion method.²³ In brief, the bacteria were cultured in Müller-Hinton broth at $35^{\circ}C \pm 2^{\circ}C$ on an orbital shaking incubator (Remi, India) at 160 rpm. A lawn of bacterial culture was prepared by spreading 100 µL culture broth, having 106 CFU/mL of each test organism on solid nutrient agar plates. The plates were allowed to stand for 10–15 minutes. to allow for culture absorption. The 8 mm size wells were punched into the agar with the head of sterile micropipette tips. Wells were sealed with one drop of molten agar (0.8% nutrient agar) to prevent leakage from the bottom of the plate. Using a micropipette, 100 µL (50 µg) of the nanoparticles solution sample was poured into each of five wells on all plates. After incubation at $35^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 24 hours, the size of the zone of inhibition was measured. A solvent blank was run as a negative control whereas the antibiotic (tetracycline) was used as a positive control.

Determination of minimal inhibitory/ bactericidal concentrations (MIC/MBC)

Minimum inhibitory concentrations (MIC) and minimum bactericidal concentrations (MBC) of CuO nanoparticles were determined by the broth dilution method which conformed to the recommended standards of the National Committee for Clinical Laboratory Standards (NCCLS; now renamed the Clinical and Laboratory Standards Institute, CLSI, 2000). Tetracycline was used as a positive control. A dilution series with 10 mL nutrient broth medium containing $10{\text -}100\,\mu\text{g/mL}$ copper oxide nanoparticles was prepared.

Each set was inoculated aseptically with 50 µL of respective bacterial suspension (approximately 10⁶ CFU/mL). The bacteria were plated onto solid nutrient agar plates. The lowest concentration inhibiting bacterial growth was defined as the MIC. In contrast the minimal concentration which completely inhibited the bacterial growth was defined as the minimum bactericidal concentration (MBC).²⁴ Each experiment was repeated three times, and the resulting bacterial growth on three plates corresponding to a particular sample were averaged and reported.

Results

Structural analysis

The typical XRD pattern of the CuO nanoparticles annealed at 400°C is shown in Figure 1. The peak positions of the sample exhibited the monoclinic structure of CuO which was confirmed from the International Centre for Diffraction Data (ICDD) card No 801916. Further, no other impurity peak was observed in the XRD pattern, showing the single phase sample formation. The crystalline size was calculated using the Scherrer formula, $D = 0.9 \lambda / \beta \cos \theta$, where λ is the wavelength of X-ray radiation, β is the full width at half maximum (FWHM) of the peaks at the diffracting angle θ . Crystallite size calculated by the Scherrer formula was found to be 20 nm. Lattice parameters calculated by powder X software (Cheng Dong, Institute of Physics, Chinese Academy of Sciences, Beijing, China) were found to be a = 4.688 Å, b = 3.427 Å, c = 5.132 Å. These values are in good agreement with the standard values reported by the ICDD Card No 801916. In order to investigate the effect of

temperature on CuO nanoparticles, samples were further annealed at 500°C, 600°C, and 700°C. Figure 2 exhibits the XRD spectra of CuO nanoparticles annealed at different temperatures. It is clear from Figure 2 that the intensity of crystalline peaks increases with temperature, indicating an improvement in the samples crystallinity. Simultaneously, the peaks become narrower as the temperature increases resulting in the increase of crystallite size. The variation of crystallite size and lattice parameters with temperature was calculated and the results are presented in Table 1. It can be seen from Table 1 that crystallite size and lattice parameters increase with the increase in annealing temperature. The increase in crystallite size with temperature can be attributed to atomic diffusion. From an atomic perspective, diffusion is the stepwise migration of atoms from lattice site to lattice site. In fact, the atoms in solid materials are in constant motion, rapidly changing positions. For an atom to make such a move, the atom must have sufficient energy to break bonds with its neighbor atoms and then cause some lattice distortion during the displacement. As the temperature increases, the atoms gain sufficient energy for diffusive motion and hence increase in size takes place.

Figure 3A and B show the TEM micrographs of CuO nanoparticles sintered at 400°C and 700°C, respectively, while Figure 3C and D exhibit the size distribution of CuO nanoparticles sintered at 400°C and 700°C, respectively. Average particle sizes obtained from TEM images were found to be 20 ± 1.24 nm and 28.9 ± 1.22 nm for the samples sintered at 400°C and 700°C, respectively. The average particle sizes determined by TEM are closely matched

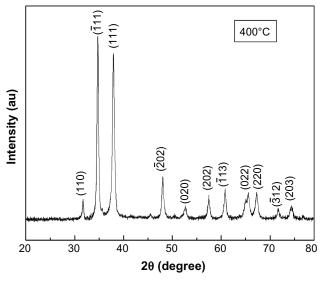


Figure 1 XRD spectra of CuO nanoparticles annealed at 400°C. **Abbreviations:** XRD, X-ray diffraction; CuO, copper oxide; AU, units of intensity.

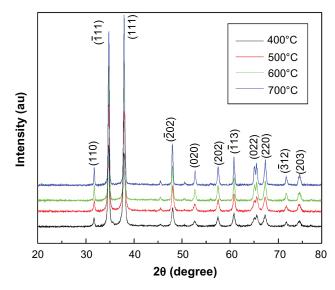


Figure 2 XRD spectra of CuO nanoparticles annealed at different temperatures.

Abbreviations: XRD, X-ray diffraction; CuO, copper oxide; AU, units of intensity.

Table I Variation of crystallite size and lattice parameters with annealing temperature

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Temperature	a	b	С	Crystallite
(°C)	(Å)	(Å)	(Å)	size (nm)
400	4.688	3.427	5.132	20
500	4.711	3.429	5.133	21
600	4.715	3.430	5.135	25
700	4.719	3.432	5.136	27

to the crystallite size calculated from XRD results. TEM results also confirm the increase in the particle size with sintering temperature which corroborates well with the XRD results.

FTIR spectra were recorded in solid phase using the KBr pellets technique in the range of 3500–400 cm⁻¹. FTIR spectra of CuO nanoparticles treated at 400°C, 500°C, 600°C, and 700°C are shown in Figure 4. FTIR spectra exhibit only three vibrations: occurring at approximately 480 cm⁻¹, 530 cm⁻¹, and 580 cm⁻¹ for all the samples, which can be attributed to the vibrations of Cu-O, confirming the formation of highly pure CuO nanoparticles. A weak band at around 2300 cm⁻¹

may be attributed to the vibrations of atmospheric ${\rm CO_2}$. These assignments are in agreement with the values available in literature. $^{25-27}$

The Raman spectrum of CuO nanocrystals sintered at 400°C is shown in Figure 4, which exhibits three one-phonon Raman scattering bands at approximately 277.5 cm $^{-1}$ ($A_{\rm g}$), 329.9 cm $^{-1}$ ($B_{\rm g}$), and 610.8 cm $^{-1}$ ($B_{\rm g}$), respectively. 28 The Raman spectra showed the presence of all three Raman active phonons ($A_{\rm g}+2B_{\rm g}$) of CuO confirming the formation of CuO nanoparticles. The high crystalline nature of CuO nanoparticles is reflected by the significant intensity of Raman bands.

Antimicrobial properties

In this study, the copper oxide nanoparticles showed remarkable antibacterial activity against both Gram-positive ($B.\ subtilis$ and $S.\ aureus$) and Gram-negative ($E.\ coli$ and $P.\ aeruginosa$) bacteria (Table 2). The extent of inhibition of bacterial growth observed in this study was found to be variable and size-dependant. The smallest CuO nanoparticles (particle size 20 ± 1.24 nm) synthesized at the lowest

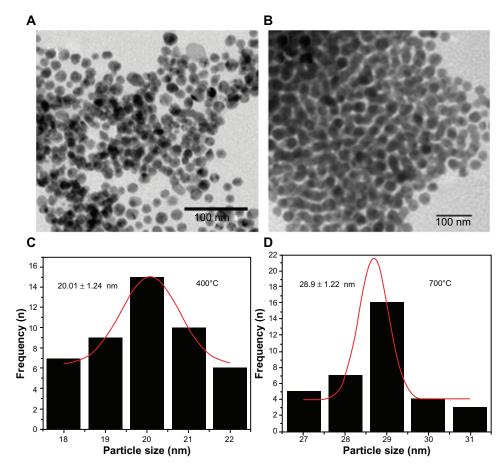


Figure 3 TEM image of CuO nanoparticles.

Notes: TEM image of CuO nanoparticles annealed at (A) 400°C (B) 700°C.

Abbreviations: TEM, transmission electron microscopy; CuO, copper oxide; n, number.

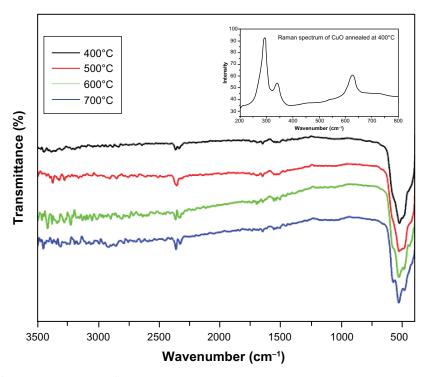


Figure 4 FTIR spectra of CuO nanoparticles annealed at different temperatures. **Note:** Inset shows Raman spectrum of CuO nanoparticles annealed at 400°C. **Abbreviations:** FTIR, Fourier-transform infrared spectroscopy; CuO, copper oxide.

temperature of 400°C, showed a significant inhibitory effect against both Gram-negative and -positive bacteria as compared to the CuO samples sintered at higher temperatures (Tables 3 and 4). One unique observation was that CuO nanoparticles synthesized at 400°C with the smallest particle size demonstrated the maximum zone of inhibition in the case of B. subtilis, which was 20% more than the zone of inhibition observed for tetracycline (Table 2). While comparing the effect of nanoparticles synthesized at varying temperature ranges, the greatest inhibitory effect was recorded for those CuO particles, which were synthesized at 400°C against P. aeruginosa strain. Furthermore, the smaller particles synthesized at 400°C had a zone of inhibition radii twice that of particles produced at 700°C. While comparing the effect of nanoparticles on bacterial strains, CuO nanoparticles were more toxic to E. coli regardless of the particle size except in one case (Tables 3 and 4). In all cases, the smaller the CuO nanoparticles, the lower the MIC and MBC values. In the case of the MIC, the most pronounced difference amongst all strains except S. aureus was between 400°C and 500°C nanoparticles. In case of S. aureus, the largest increase in MIC was observed for nanoparticles synthesized at temperatures between 500°C and 600°C (Table 3). For E. coli and S. aureus, the MIC values for particles synthesized at 700°C were

threefold higher than those recorded for the nanoparticles synthesized at a comparatively smaller temperature (400°C). In general, the CuO nanoparticles had a less pronounced effect on other bacterial strains, but these effects were still twofold higher than those determined for 700°C synthesized particles. As can be seen in Table 4, the MBC values were even more drastically dependent on the particle size. Again we observed that *E. coli* and *S. aureus* had threefold lower values for the 400°C MBC than those synthesized at 700°C.

Table 2 Antimicrobial activity of copper oxide (CuO) nanoparticles against two Gram-positive and two Gram-negative bacteria

Treatment	Zone of inhibition (mm)				
(100 μL)	Escherichia coli	Pseudomonas aeruginosa	Bacillus subtilis	Staphylococcus aureus	
CuO (400°C) ^a	20	21	24	22	
CuO (500°C) ^b	17	20	22	20	
CuO (600°C)°	15	12	20	18	
CuO (700°C) ^d	14	10	15	12	
Tetracycline ^e	20	18	20	22	

Note: **eindicate the copper oxide nanoparticles synthesized at different temperatures was used in an antimicrobial experiment on nutrient agar plates in Figure 5.

Table 3 MIC of copper oxide nanoparticles (annealed at different temperatures) against different laboratory bacterial strains

Microorganisms	MIC of copper nanoparticles (µg/mL)				
	400°C	500°C	600°C	700°C	
E. coli	20 ± 3	35 ± 5	45 ± 4	65 ± 6	
P. aeruginosa	28 ± 4	40 ± 3	50 ± 5	55 ± 5	
B. subtilis	30 ± 5	50 ± 4	65 ± 4	70 ± 5	
S. aureus	25 ± 4	42 ± 2	70 ± 5	75 ± 4	

Note: Each value in mean \pm standard deviation.

Abbreviation: MIC, minimum inhibitory concentration.

As mentioned above, unlike the MIC values, each temperature dependent synthesis had more variable MBC values (Table 4); however, for the *S. aureus*, the largest MBC impact was from the 400°C to 500°C transition, unlike the MIC.

Discussion

A few studies have been performed to elucidate the mechanism of bactericidal action of nanoparticles. For example, Tsao et al suggested that the exposure of Gram-positive bacteria to carboxyfullerene nanoparticles resulted in the puncturing of the bacteria leading to cell death.²⁹ Another proposed way in which the membrane can be compromised is the alteration of membrane lipid components.³⁰ It is, however, difficult to distinguish between the bactericidal activity of nanoparticles from the ions released by the nanoparticles themselves.31 Previously, Ruparelia et al estimated the concentration of released ions for 10 mg of copper nanoparticles suspended in 100 mL nutrient media and distilled water.³² They found that the concentration of Cu²⁺ ions released in nutrient media was 17 mgL⁻¹ after 24 hours of incubation in a rotary shaker, while in distilled water under the same conditions over a period of 24 hours, the concentration of ions released was 0.5 mgL⁻¹. These results indicate that the nutrient media can facilitate the release of Cu2+ ions. The considerably greater release of Cu²⁺ ions in the nutrient media is possibly due to the interaction of the media chloride ions with the oxide layer of the nanoparticles.³² For oxidized

Table 4 MBC of copper oxide nanoparticles (annealed at different temperatures) against different laboratory bacterial strains

Microorganisms	MBC of copper nanoparticles (µg/mL)				
	400°C	500°C	600°C	700°C	
E. coli	30 ± 2	45 ± 5	60 ± 3	95 ± 4	
P. aeruginosa	35 ± 2	50 ± 3	75 ± 5	85 ± 5	
B. subtilis	45 ± 3	62 ± 4	85 ± 4	95 ± 5	
S. aureus	32 ± 5	75 ± 2	90 ± 5	100 ± 4	

Note: Each value in mean \pm standard deviation. **Abbreviation:** MBC, minimum bactericidal concentration. copper particles embedded in an inert, Teflon-like matrix, Cioffi et al demonstrated significant antimicrobial activity due to the release of ions.³³ Consequently, the bactericidal effects observed in this study might have been influenced by the release of Cu2+ ions in solution. The presence of nanoparticles in suspension would ensure continuous release of ions into the nutrient media.³³ However, the preceding studies did not correlate their findings with the size of the nanoparticles and hence, the precise mechanisms of how nanoparticles act as biocidal agent needs to be explained. There are, however, a few mechanisms of nanoparticle toxicity suggested by other works. For example, copper ions released by the nanoparticles may attach to the negatively charged bacterial cell wall and rupture it, thereby leading to protein denaturation and cell death.³⁴ Copper ions inside the bacterial cells may bind to deoxyribonucleic acid molecules and become involved in cross-linking within and between the nucleic acid strands, resulting in the disorganized helical structure. In addition, copper ion uptake by the bacterial cells has also been found to damage important biochemical processes. 35,36

It is clear from Table 2 (maximum zone of inhibition against B. subtilis and S. aureus) that CuO nanoparticles have shown greater antimicrobial activity against B. subtilis and S. aureus. The variation in the sensitivity or resistance to both Gram-positive and -negative bacteria populations could be due to the differences in the cell structure, physiology, metabolism, or degree of contact of organisms with nanoparticles. For example, greater sensitivity among Grampositive bacteria such as B. subtilis and S. aureus to the CuO nanoparticles has been attributed to the greater abundance of amines and carboxyl groups on their cell surface and greater affinity of copper towards these groups.³⁷ Alternatively, Gram-negative bacteria like E. coli have a special cell membrane structure which possesses an important ability to resist antimicrobial agents.³⁸ Furthermore, other factors such as nanoparticle diffusion rates may also affect bacterial strain differently. Nevertheless, further studies are required to confirm this and it is beyond the scope of this manuscript.

Figures 5 and 6 exhibit the zone of inhibition of CuO nanoparticles synthesized at different temperatures (a–d) and positive control, a known antibiotic tetracycline (e), against two Gram-negative bacteria [(A) *E. coli* (B) *P. aeruginosa*], and two Gram-positive bacteria [(C) *B. subtilis* (D) *S. aureus*]. Figure 5 clearly indicates that the copper oxide nanoparticles inhibit the growth of both Gram-negative and -positive bacteria and the zone of inhibition decreases with the increase in annealing temperature from 400°C–700°C. Figure 6 shows

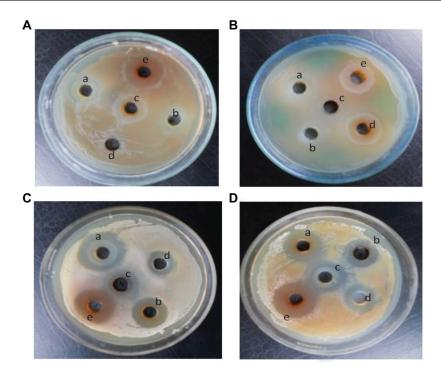


Figure 5 Zone of inhibition of copper oxide nanoparticles.

Notes: Zone of inhibition of copper oxide nanoparticles synthesized at different temperatures (a–d) and positive control, a known antibiotic tetracycline (e) against two Gram-negative bacteria (A) Escherichia coli and (B) Pseudomonas aeruginosa, and two Gram-positive bacteria (C) Bacillus subtilis and (D) Staphylococcus aureus.

that the zone of inhibition is maximum when the particle size is minimum (20 ± 1.24 nm). These results demonstrate the excellent antimicrobial behavior of CuO nanoparticles synthesized at low temperature. Broadly, interactions between the negative charges of microorganisms and the

positive charge of nanoparticles produces an electromagnetic attraction between the microbe and effective levels of active nanoparticles. Such interactions lead to oxidation of surface molecules of microbes resulting in their death. Biodestructive effects such as degradation of deoxyribonucleic acid was

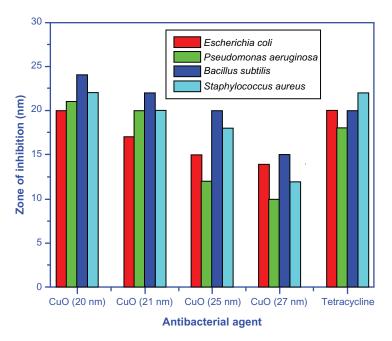


Figure 6 Bar graph representing the zone of inhibition for CuO nanoparticles and tetracycline against Gram-positive and -negative bacteria. Abbreviation: CuO, copper oxide.

observed following exposure of Gram-positive bacteria to silver and copper nanoparticles by other works,^{39,40} and are in agreement with present findings.

Conclusion

We have successfully synthesized CuO nanoparticles using a gel combustion route. XRD spectra confirmed the formation of single phase CuO nanoparticles. Crystallite size was found to increase with the increase in annealing temperature. Minimum crystallite size of 20 \pm 1.24 nm was observed in the case of CuO nanoparticles annealed at 400°C. TEM results corroborate well with XRD results. FTIR and Raman spectra also validated the purity of CuO nanoparticles. Antibacterial activity experiments performed on various microorganisms clearly demonstrated the higher effectiveness of CuO nanoparticles annealed at 400°C against bacterial growth due to smaller particle size of this sample compared to other samples. Zone of inhibition for all the microorganisms reached a maximum point using CuO nanoparticles annealed at 400°C. Moreover, minimum inhibitory concentration and minimum bactericidal concentration of CuO nanoparticles annealed at 400°C was lowest for all the bacterial strains.

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Disclosure

The authors report no conflicts of interest in this work.

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