

Gold and silver nanoparticles from *Trianthema decandra*: synthesis, characterization, and antimicrobial properties

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Background: There is an increasing commercial demand for nanoparticles due to their wide applicability in various markets, including medicine, catalysis, electronics, chemistry, and energy. In this report, a simple and ecofriendly chemical reaction for the synthesis of gold and silver nanoparticles from *Trianthema decandra* (Aizoaceae) has been developed.

Methods and results: On treatment of aqueous solutions containing chloroauric acid or silver nitrate with root extract of *T. decandra*, stable gold or silver nanoparticles were rapidly formed. The kinetics of reduction of gold and silver ions during the reaction was analyzed by ultraviolet-visible spectroscopy. Field emission-scanning electron microscopy showed formation of gold nanoparticles in various shapes, including spherical, cubical, triangular, and hexagonal, while silver nanoparticles were spherical. The size of the gold nanoparticles was 33–65 nm and that of the silver nanoparticles was 36–74 nm. Energy dispersive x-ray and Fourier transform infrared spectroscopy confirmed the presence of metallic gold and metallic silver in the respective nanoparticles. The antimicrobial properties of the synthesized nanoparticles were analyzed using the Kirby-Bauer method. The results show varied susceptibility of microorganisms to the gold and silver nanoparticles.

Conclusion: It is believed that phytochemicals present in *T. decandra* extract reduce the silver and gold ions into metallic nanoparticles. This strategy reduces the cost of production and the environmental impact. The silver and gold nanoparticles formed showed strong activity against all microorganisms tested.

Keywords: *Trianthema decandra*, gold, silver, nanoparticles, antimicrobial activity

Introduction

Nanoparticles are a special group of materials with unique features and extensive applications in diverse fields.¹ Metal nanoparticles, such as those containing gold and silver have recognized importance in chemistry, physics, and biology because of their unique optical, electrical, and photothermal properties.² The ease of synthesizing gold and silver nanoparticles and their affinity for binding many biological molecules makes them attractive candidates for study. Various physical and chemical methods have been reported over the last two decades for the synthesis of gold and silver nanoparticles, of which chemical approaches, such as chemical reduction, electrochemical techniques, and photochemical reduction, are most widely used. The green method of nanoparticle synthesis employing plant extracts is a simple and viable alternative to chemical procedures and physical methods.³ Chemical and physical methods are harmful because the chemicals used are toxic, flammable, and are not disposed easily of in the environment.⁴ In recent years, biosynthesis of nanoparticles has received considerable attention due

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to the growing need to develop clean and nontoxic chemicals, environmentally friendly solvents, and renewable materials.⁵ Utilization of biological systems such as plant extracts for the synthesis of nanoparticles has advantages over chemical and physical methods, because these systems are cost-effective, environment friendly, single-step in nature, and easily scaled up for large-scale synthesis.⁶

Studies have shown that the size, morphology, stability, and physicochemical properties of metal nanoparticles are strongly influenced by the experimental conditions,^{7–9} and play pivotal roles in controlling the physical, chemical, optical, and electronic properties of these nanoscopic materials. This is particularly important for noble metals, such as gold and silver, which have strong surface plasmon resonance oscillations.¹⁰

Several researchers have reported the use of natural products, inactivated plant biomass, and as living plants such as *Aloe vera*,¹¹ *Carica papaya*,¹² *Magnolia kobus*, *Diospyros kaki*,¹³ *Medicago sativa*,^{14,15} *Azadirachta indica*,¹⁶ *Pelargonium graveolens*,¹⁷ *Chilopsis linearis*,¹⁸ and *Tamarindus indica*¹⁹ for the production of silver and gold nanoparticles (Table 1). Large amounts of nanoparticles can be easily synthesized from plants, and the majority of these are nontoxic. They have been used in pharmaceutical drug-based industries for treating lymphocytic leukemia, detecting DNA, inhibiting bacteria and fungi, and preventing burns and open wound infections.^{20,21}

Trianthema decandra, a prostrate, glabrous, succulent annual belonging to the Aizoaceae family, is found as a weed throughout most of India.²² The plant has various medicinal properties and has been reported to have antioxidant and antimicrobial properties.²³ In the present study, we describe the biosynthesis of gold and silver nanoparticles from aqueous root extract of *T. decandra* for the first time. Chloroauric acid and silver nitrate were reduced to metallic gold and metallic silver respectively, on reaction with root extract of *T. decandra*, resulting in synthesis of gold and silver nanoparticles. The gold and silver nanoparticles formed were

characterized using ultraviolet-visible, scanning electron microscopy, energy dispersive x-ray, and Fourier transform infrared spectroscopy. Antimicrobial assays using gold and silver nanoparticles demonstrated activity against all the microorganisms tested.

Materials and methods

Plant materials

T. decandra L was collected from Salem district, Tamil Nadu, India, during June 2011. The plant was taxonomically identified and authenticated by the Botanical Survey of India, Coimbatore, and voucher specimen BSI/SRC/5/23/10-11/Tech 975 was deposited at the Plant Biotechnology Laboratory, Sri Ramaswamy Memorial University, for future reference.

Chemicals

Gold (III) chloride trihydrate (HAuCl_4) and silver nitrate (AgNO_3) were purchased from Sigma-Aldrich, St Louis, MO. Double-distilled deionized water was used.

Preparation of plant extracts

Roots of *T. decandra* were washed with distilled water to remove dirt and soil. The shade-dried materials were powdered and passed through a 40-mesh sieve. Next, 20 g of coarsely powdered material was extracted with 100 mL of double-distilled water. The mixture was filtered through Whatman No 1 filter paper. The extract was stored at 4°C until further use.

Synthesis of gold and silver nanoparticles

For the synthesis of gold and silver nanoparticles, 10 mL of 1 mM aqueous HAuCl_4 and AgNO_3 solutions were separately added to 5 mL, 10 mL, and 15 mL of aqueous root extract and incubated in the dark. The resulting colloidal solutions of gold and silver were then analyzed using ultraviolet-visible spectroscopy.

Table 1 Nanoparticles synthesized from different plants and their size range

Number	Plant material used	Metal nanoparticle	Particle size (nm)	Reference
1	<i>Aloe vera</i>	Ag and Au	15.2	11
2	<i>Carica papaya</i>	Ag	60–80	12
3	<i>Magnolia kobus</i>	Au	5–300	13
4	<i>Diospyros kaki</i>	Au	5–300	13
5	<i>Medicago sativa</i>	Ag	86–108	14
6	<i>Medicago sativa</i>	Au	15–200	15
7	<i>Azadirachta indica</i>	Ag and Au	50–100	16
8	<i>Pelargonium graveolens</i>	Au	21–70	17
9	<i>Chilopsis linearis</i>	Au	1.1	18
10	<i>Tamarindus indica</i>	Au	20–40	19

Ultraviolet-visible spectroscopy

The colloidal solutions of gold and silver were diluted by adding 2.9 mL of deionized water to 0.1 mL of sample. Bioreduction of HAuCl_4 and AgNO_3 was recorded as a function of time using water as a reference at 3-hour intervals from zero hour up to 24 hours using an ultraviolet-visible spectrophotometer (Perkin-Elmer, Boston, MA) in a scanning range of 200 nm to 800 nm.

Purification of gold and silver nanoparticles

To remove excess gold and silver ions, the gold and silver colloids were centrifuged at 10,000 rpm for 15 minutes and washed at least three times with deionized water. A dried powder of the gold and silver nanoparticles was obtained by freeze-drying.

Fourier transform infrared spectroscopy

The air-dried powder form of the synthesized gold and silver nanoparticles was ground with KBr pellets and subjected to Fourier transform infrared spectroscopy.

FE-SEM and energy dispersive x-ray spectroscopy

The gold and silver nanoparticles formed were characterized using high resolution field emission scanning electron microscopy (FE-SEM). The samples were prepared by simple drop coating of the suspension of gold and silver nanoparticles separately on a carbon-coated copper grid, by simply dropping a very small amount of the sample on the grid, with excess solution removed using blotting paper. The film on the scanning electron microscopy grid was then allowed to dry under a mercury lamp for 5 minutes. Energy dispersive x-ray spectroscopy was then performed using the Hitachi S-3400 N FE-SEM instrument equipped with a Thermo energy dispersive x-ray spectroscopy attachment.

Antimicrobial activity

The antimicrobial activity of the gold and silver nanoparticles formed was assayed using the standard Kirby-Bauer disc diffusion method.²⁴ Bacterial suspensions of *Staphylococcus aureus* (MTCC 29213), *Streptococcus faecalis* (MTCC 0459), *Enterococcus faecalis* (MTCC 2729), *Escherichia coli* (MTCC 443), *Pseudomonas aeruginosa* (MTCC 1035), *Proteus vulgaris* (MTCC 1771), *Bacillus subtilis* (MTCC 121), *Yersinia enterocolitica* (MTCC 840), and the fungus, *Candida albicans* (MTCC 183), grown overnight were swabbed on separate nutrient agar plates using L-rods. Whatman filter paper (No 1) discs 6 mm in diameter were

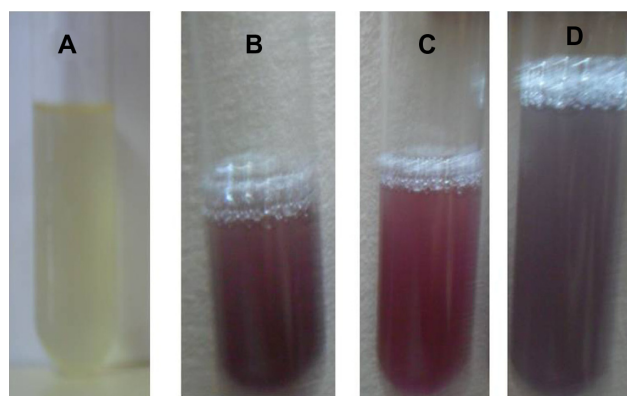


Figure 1 Optical photograph of colloidal solution of (A) *Trianthema decandra* root extract, (B) HAuCl_4 solution reduced with 5 mL of *T. decandra* extract, (C) HAuCl_4 solution reduced with 10 mL of *T. decandra* extract, and (D) HAuCl_4 solution reduced with 15 mL of *T. decandra* extract.

separately impregnated with 20 μL of a 10 mg/mL solution of gold and silver nanoparticles in dimethylsulfoxide. The discs were evaporated at 37°C for 24 hours and then impregnated on plates. Chloramphenicol and nystatin were used as positive controls for bacteria and the fungus respectively, while dimethylsulfoxide served as a negative control. Triplicates were maintained and the plates were incubated at 37°C for 24–48 hours. Diameters of the zones of inhibition were measured using a meter ruler, and the mean value for each organism was recorded and expressed in millimeters.

Statistical analysis

The data were analyzed by one-way analysis of variance followed by the Duncan's multiple range test.

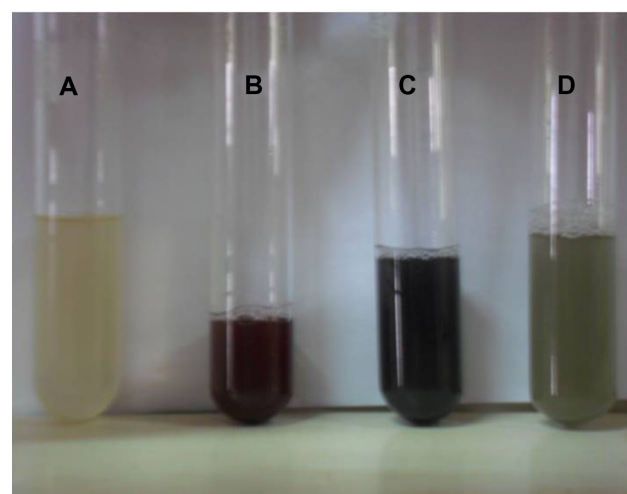


Figure 2 Optical photograph of colloidal solution of (A) *Trianthema decandra* root extract, (B) AgNO_3 solution reduced with 5 mL of *T. decandra* extract, (C) AgNO_3 solution reduced with 10 mL of *T. decandra* extract, and (D) AgNO_3 solution reduced with 15 mL of *T. decandra* extract.

Results and discussion

Synthesis and characterization of gold and silver nanoparticles

Nanoparticles of noble metals are characterized by the presence of bright colors produced by oscillations of the surface electron cloud of these particles. The formation of gold and silver nanoparticles was observed by monitoring the change in color, ie, a light yellow solution turning to ruby pink when HAuCl_4 was used and a pale yellow solution

turning to dark grey when AgNO_3 was used. The collective oscillations of electrons in a nanoparticle upon interaction with light of suitable energy causes the nanoparticles to attain the color specific to a particular metal. Particle size, shape, and aggregation depend strongly on the frequency of oscillations.²⁵

The reduction kinetics of aqueous chloroaurate ions and silver ions leading to synthesis of gold and silver nanoparticles was compared using different amounts,

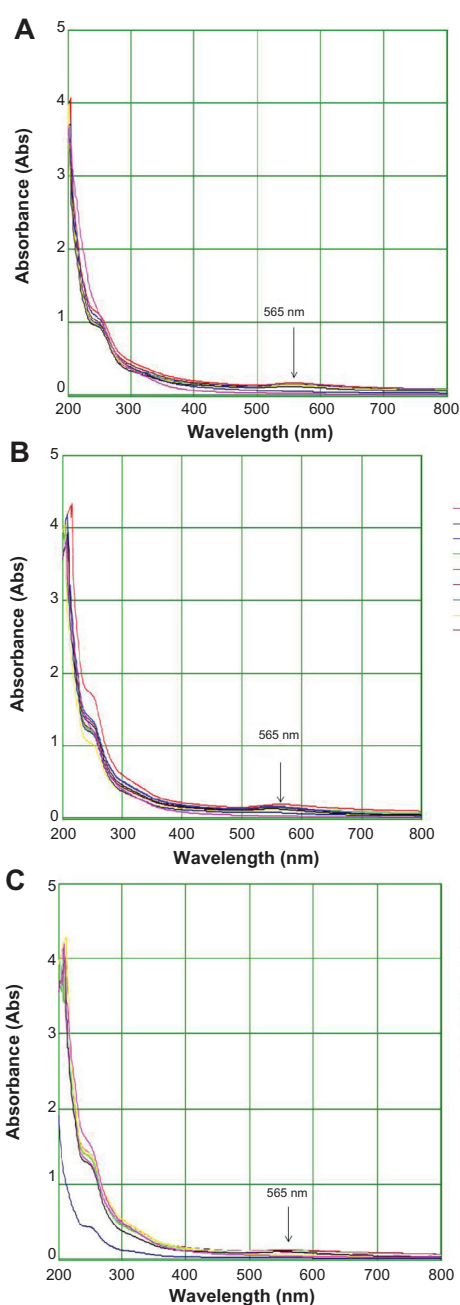


Figure 3 Ultraviolet-visible bioreduction kinetics in the 20–800 nm range for colloidal HAuCl_4 solution with 5 mL of *Trianthema decandra* root extract (A), 10 mL of *T. decandra* root extract (B), and 15 mL of *T. decandra* root extract (C).

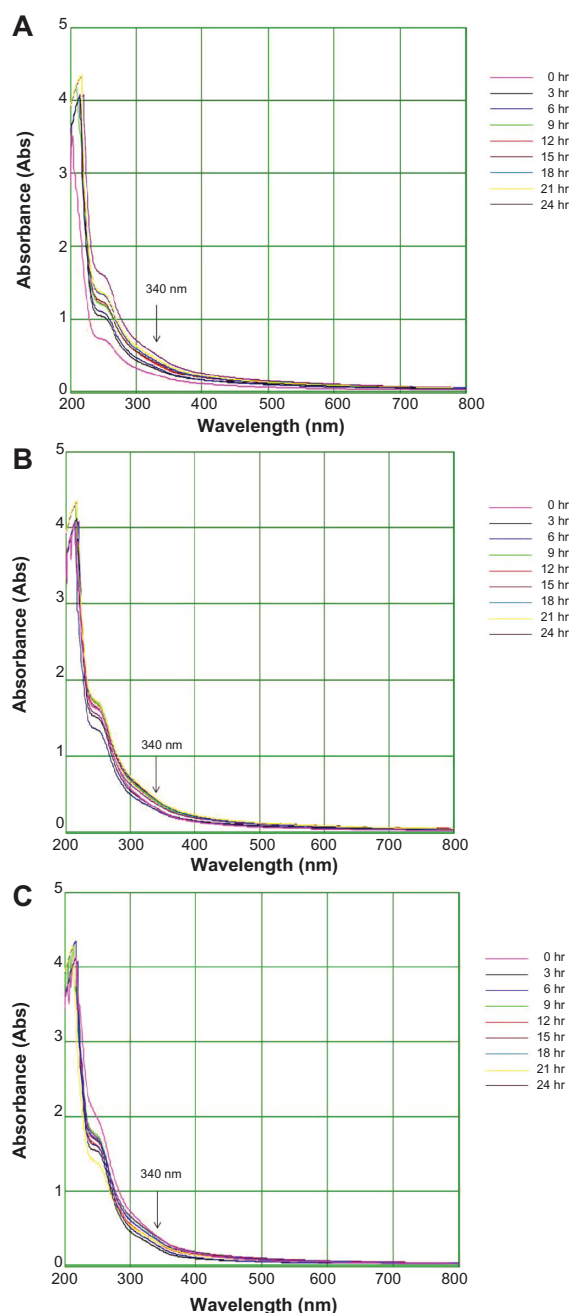


Figure 4 Ultraviolet-visible bioreduction kinetics in the 200–800 nm range for colloidal AgNO_3 solution with 5 mL of *Trianthema decandra* root extract (A), 10 mL of *T. decandra* root extract (B), and 15 mL of *T. decandra* root extract (C).

ie, 5 mL, 10 mL, and 15 mL of root extract of *T. decandra* (Figures 1 and 2). Reduction of chloraurate ions was more rapid than that of silver ions. Gold nanoparticles were synthesized within 3 hours of reaction with the root extract and the components thereof, while more than a 6-hour reaction period was required for synthesis of the silver counterpart. Preliminary phytochemical screening of aqueous extracts of *T. decandra* showed the presence of various secondary metabolites.²⁶ The bioreduction achieved in the present study is attributed to the metabolites present in the plant. Synthesis of gold nanoparticles was pronounced when 15 mL of root extract was used for bioreduction, while 10 mL of the same root extract was sufficient for production of silver nanoparticles.

The colloidal gold solution showed strong (0.15 AU), weak (0.04 AU), and moderate (0.08 AU) absorption peaks with 15 mL, 5 mL, and 10 mL of extract respectively, at 575 nm (Figure 3). The results also indicate a surface plasmon resonance gold band at 530–575 nm. The colloidal silver solution showed strong (0.81 AU), weak (0.23 AU), and moderate (0.31 AU) absorbance peaks with 10 mL, 5 mL, and 15 mL of extract respectively at 340 nm (Figure 4). Nanoparticles may grow in a process involving rapid bioreduction and strongly influence surface plasmon resonance in the water extract.²⁷ The results of the present study are in accordance with previous reports, including that by Vijayakumar et al²⁷ on *Artemisia nilagirica*, indicating occurrence of a silver band at 340–400 nm. Further characterization using Fourier

transform infrared, scanning electron, and energy dispersive x-ray spectroscopy was carried out for gold nanoparticles formed with 15 mL and silver nanoparticles formed with 10 mL of *T. decandra* root extract.

The functional groups present in gold nanoparticles detected using Fourier transform infrared spectroscopy had absorption peaks located at 1649.33 cm^{-1} and 1020.12 cm^{-1} in the region of 1000–1800 cm^{-1} , characteristic of gold atoms (Figure 5). Two absorption peaks located at 1649.33 cm^{-1} and 1020 cm^{-1} are indicative of C=C stretch and aromatic C–C bonds, respectively. Distinct peaks in the range of 2344.87 cm^{-1} and 2364.35 cm^{-1} correspond to C≡N stretch, while peaks at 2925.92 cm^{-1} and 3422.38 cm^{-1} correspond to C–H and O–H stretching vibration, respectively.

Fourier transform infrared spectroscopy analysis of the silver nanoparticles showed absorption peaks of reduced silver at 1653.96 cm^{-1} and 1027.44 cm^{-1} in the region of 1000–1800 cm^{-1} (Figure 6). Two absorption peaks located at 1653 cm^{-1} are associated with the stretch vibration of C=C, and the single absorbance peak located at 1027 cm^{-1} is assigned to C–N stretching vibrations of amine. Further distinct peaks in the region of 2343.97–2362.27 cm^{-1} correspond to C≡N stretch for nitrile groups, while 3447.86 cm^{-1} corresponds to O–H stretching vibration.

Scanning electron micrographs (Figure 7) for the gold nanoparticles reveal numerous spherical, triangular, hexagonal, and cubic structures produced by reduction of gold ions with sizes ranging between 33.7 nm and 99.3 nm.

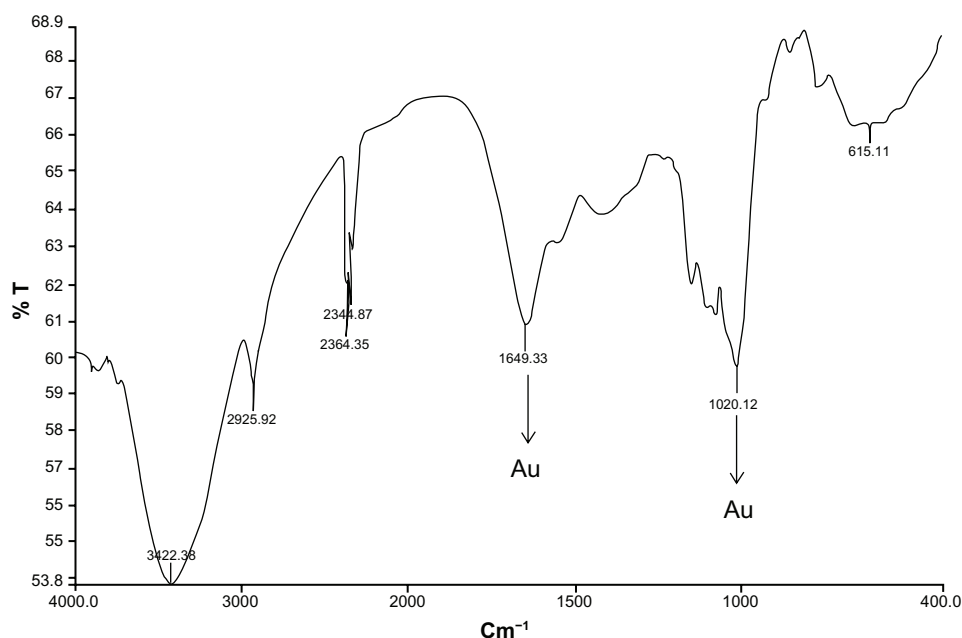


Figure 5 Fourier transform infrared absorption spectra for nanoparticles synthesized by bioreduction of HAuCl_4 ions using *Trianthema decandra* root extract.

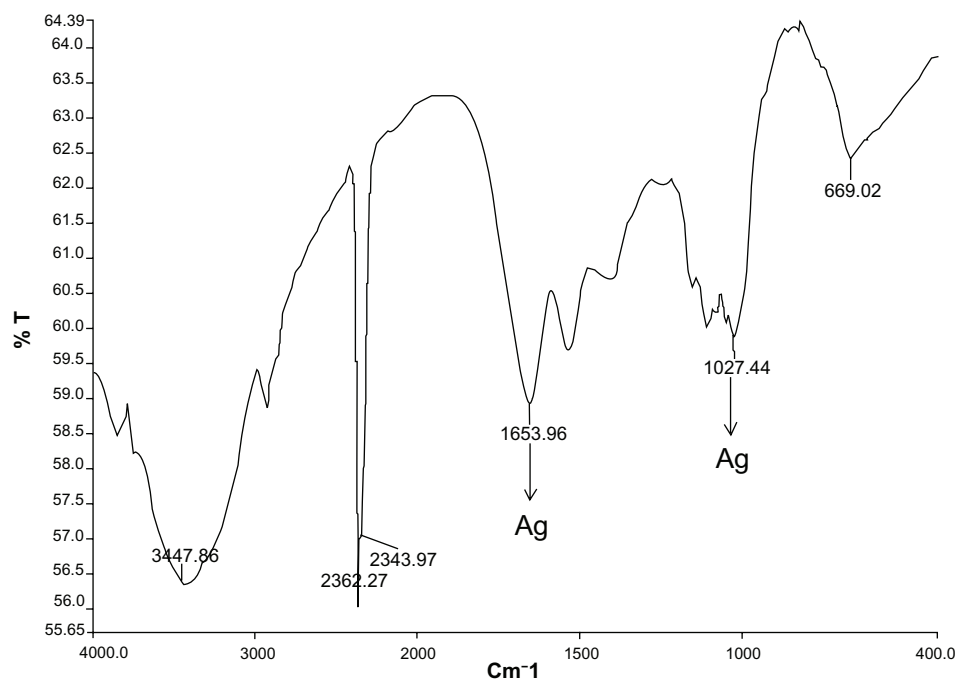


Figure 6 Fourier transform infrared absorption spectra for nanoparticles synthesized by bioreduction of AgNO_3 ions using *Trianthema decandra* root extract.

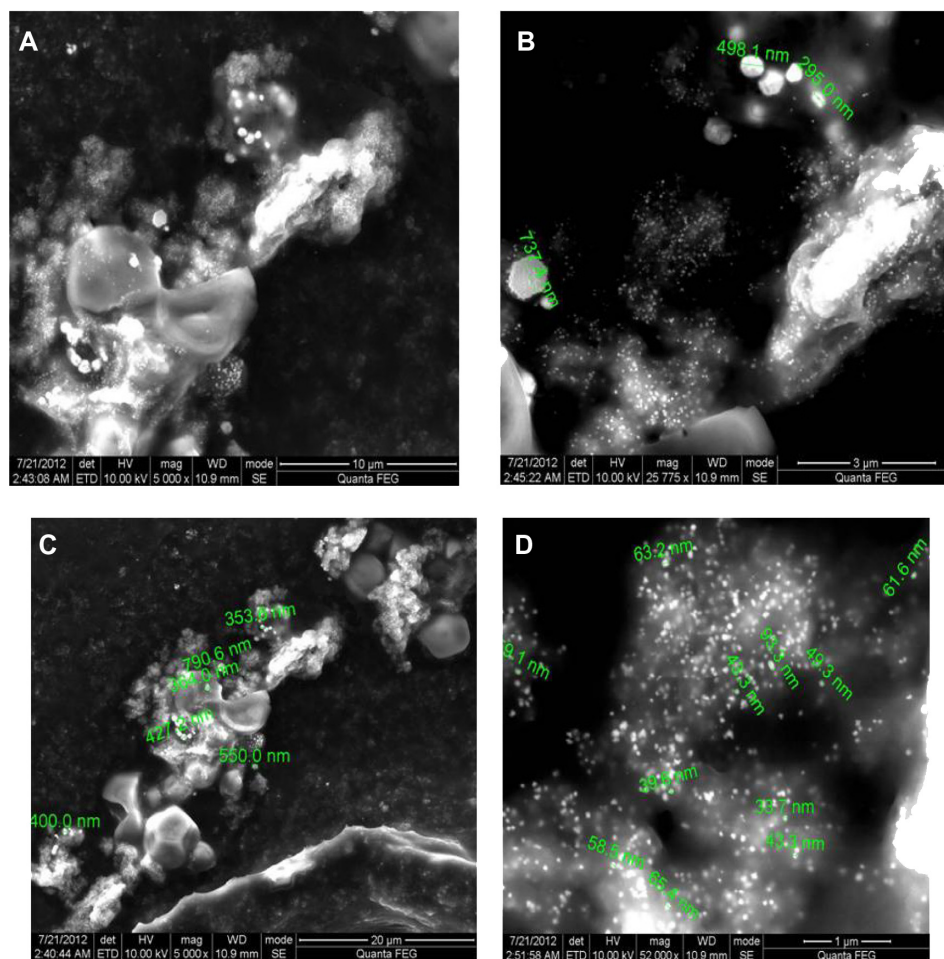


Figure 7 Scanning electron micrographs of gold nanoparticles observed from the reaction of Au^{3+} cations with *Trianthema decandra* root extract.

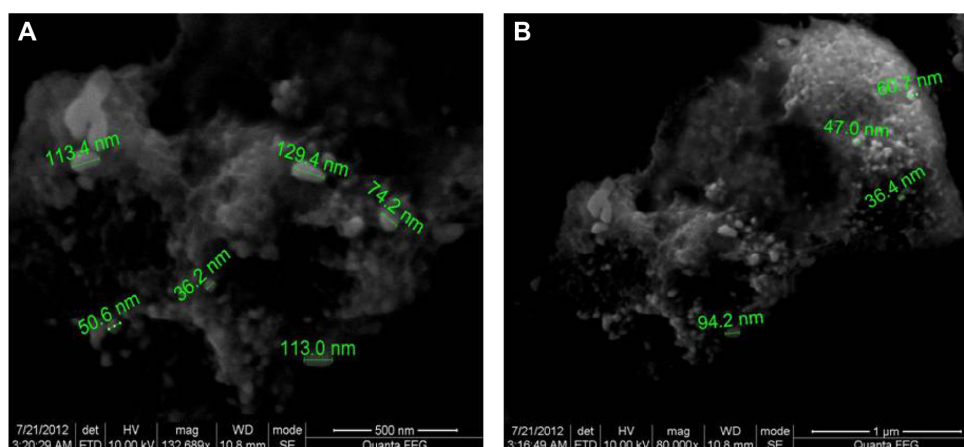


Figure 8 Scanning electron micrographs of silver nanoparticles observed from reaction of Ag^+ cations with *Trianthema decandra* root extract.

An individual gold nanohexagon with an average edge length of 737 nm is also seen in Figure 7B.

Very recently, Rai et al have demonstrated that the presence of halide ions and modulation of temperature can control the morphology of biologically synthesized gold triangles using lemongrass leaf extract.²⁸ In the present study, nanoparticles resembling triangles with sizes ranging between 33.7 nm and 99.3 nm were produced during bioreduction of HAuCl_4 , which could be attributed to the chloride ions present in it (Figure 7D).

Spherical silver nanoparticles with diameters of 36–94 nm are depicted in the scanning electron micrographs (Figure 8). The variation in particle sizes, such as 36, 47, 60, and 94 nm, is possibly due to the fact that the nanoparticles are being formed at different times.

Water-soluble heterocyclic compounds, including saponin, were mainly responsible for the reduction of silver ions or chloraurate ions to metallic silver and metallic gold, respectively. Further, it has been shown that the presence of chloride ions during nanosynthesis promotes the growth of nanotriangles. In the present study, reduction of chloraurate ions produced nanotriangles. The presence of gold atoms in the gold nanoparticles was further confirmed by energy dispersive x-ray spectroscopy. The optical absorption peak was observed at approximately 2.2 keV, which is typical of gold nanocrystallite absorption due to surface plasmon resonance. The current energy dispersive x-ray spectroscopic profile of gold nanoparticles of *T. decandra* (Figure 9) showed strong gold atom signals around 0.30, 2, 2.2, 2.4, and 9.7 keV. Similarly, the presence of silver atoms in the silver nanoparticles was confirmed using energy dispersive x-ray spectroscopy (Figure 10). The optical absorption peak was observed at 3 keV, which is typical for silver nanocrystallite

absorption due to surface plasmon resonance. In an earlier study, individual spherical silver nanoparticles synthesized using alfalfa showed absorption peaks in the range of 2.5–4 keV.²⁹ The current profile of energy dispersive x-ray spectroscopy for silver nanoparticles of *T. decandra* indicate strong silver atom signals around 0.20, 3, 3.2, and 3.4 keV.

Antimicrobial activity of gold and silver nanoparticles

Antimicrobial activity of the synthesized gold and silver nanoparticles was studied using eight different bacteria, namely *S. aureus*, *S. faecalis*, *E. faecalis*, *E. coli*, *P. aeruginosa*, *P. vulgaris*, *B. subtilis*, *Y. enterocolitica*, and a fungus, *C. albicans*.

Both gold and silver nanoparticles exhibited excellent activity against *Y. enterocolitica*, *P. vulgaris*, *E. coli*, *S. aureus*, and *S. faecalis*. Zones of inhibition measuring 13.5–15.5 mm

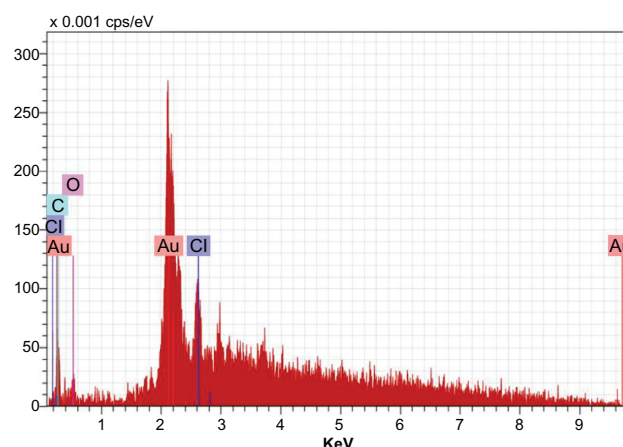


Figure 9 Energy dispersive x-ray spectroscopy of gold nanoparticles synthesized by *Trianthema decandra* root extract.

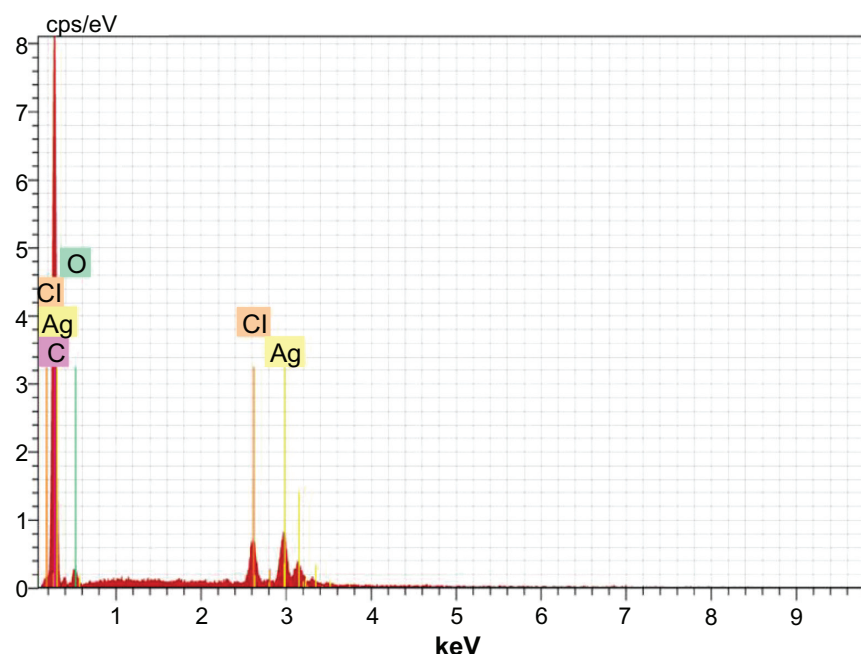


Figure 10 Energy dispersive x-ray spectroscopy of silver nanoparticles synthesized by *Trianthema decandra* root extract.

were recorded for the gold nanoparticles, while their silver counterparts recorded zones of 15.5–20.5 mm. Moderate activity was recorded against the other three bacteria tested (Table 2, Figures 11 and 12). The growth inhibition of *C. albicans* was double in silver nanoparticles when compared with gold nanoparticles. The antibacterial effect was more pronounced in Gram-negative bacteria than Gram-positive ones. The antimicrobial activity of colloidal silver particles is influenced by the particle dimensions.³⁰ Silver has long been recognized as having an inhibitory effect on microbes present in medical and industrial processes. The most important application of silver and silver nanoparticles is in medical industry, such as in topical ointments to prevent infection

against burns and open wounds.³¹ Gold nanoparticles are harmful to bacteria and fungi. They bind closely to the surface of microorganisms, causing visible damage to cells, with complete destruction of flagella, stimulated production of biofilm, and aggregation within the biofilm.³²

Comparison of gold and silver nanoparticle sizes indicates the latter to be smaller than the former. We attribute the higher activity of silver nanoparticles to their small size. The smaller particles have greater antimicrobial effects.³³ Further, the antimicrobial activity in the present study was much higher when compared with previous reports. Silver nanoparticles from *Polyalthia longifolia*, *Balsamodendron mukul*, *Melia azadirachta*, *Berberis aristata*, *Acorus calamus*,

Table 2 Mean zone of inhibition of synthesized gold and silver nanoparticles from *T. decandra*

	AuNPs	AgNPs	Positive control	Negative control	Water extract ²³
Diameter of zone of inhibition (mm)					
<i>Enterococcus faecalis</i>	10.5 ± 0.7 ^c	14.0 ± 1.4 ^b	19.5 ± 0.7 ^a	—	—
<i>Staphylococcus aureus</i>	14.5 ± 0.7 ^b	13.5 ± 0.7 ^b	29.0 ± 0.7 ^a	—	—
<i>Streptococcus faecalis</i>	13.5 ± 0.7 ^c	15.5 ± 0.7 ^b	19.5 ± 0.7 ^a	—	—
<i>Bacillus subtilis</i>	9.5 ± 0.7 ^c	12.0 ± 1.4 ^b	25.5 ± 0.7 ^a	—	2 ± 0.7 ^d
<i>Yersinia enterocolitica</i>	15.5 ± 0.7 ^c	17.5 ± 0.7 ^b	29.0 ± 1.4 ^a	—	3 ± 0.7 ^d
<i>Proteus vulgaris</i>	15.0 ± 1.4 ^c	20.5 ± 0.7 ^b	25.0 ± 1.4 ^a	—	3 ± 0.7 ^d
<i>Escherichia coli</i>	9.5 ± 0.7 ^c	15.5 ± 0.7 ^b	25.5 ± 0.7 ^a	—	4 ± 0.7 ^d
<i>Pseudomonas aeruginosa</i>	11.5 ± 0.7 ^c	14.5 ± 0.7 ^b	27.5 ± 0.7 ^a	—	3 ± 0.7 ^d
<i>Candida albicans</i>	8.5 ± 0.7 ^c	14.5 ± 0.7 ^b	22.5 ± 0.7 ^a	—	2 ± 0.7 ^d

Notes: Values are expressed as the mean ± standard error of the mean (n = 3), and values followed by same letter are not significantly different at the $P < 0.05$ as determined by the Duncan's multiple range test. Positive control was chloramphenicol/nystatin; negative control was dimethyl sulfoxide; —, indicates no inhibition.

Abbreviations: AuNPs, gold nanoparticles; AgNPs, silver nanoparticles.

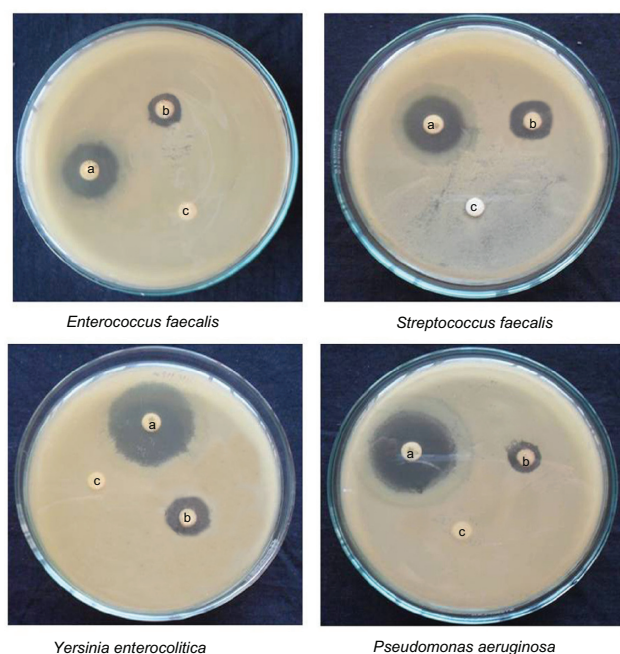


Figure 11 Activity of gold nanoparticles against different microorganisms depicting zones of inhibition of (A) positive control, (B) gold nanoparticles, and (C) dimethylsulfoxide control.

Embelia ribes, *Psoralea corylifolia*, *Smilax china*, *Glycyrrhiza glabra*, and *Andropogon muricatus* showed activity against *E. coli*, *P. aeruginosa*, *S. aureus*, and *C. albicans*, with zones of inhibition measuring less than 8 mm in diameter.^{30,33} The results of the present study are indicative of pronounced antibacterial effects. They provide an important basis for use

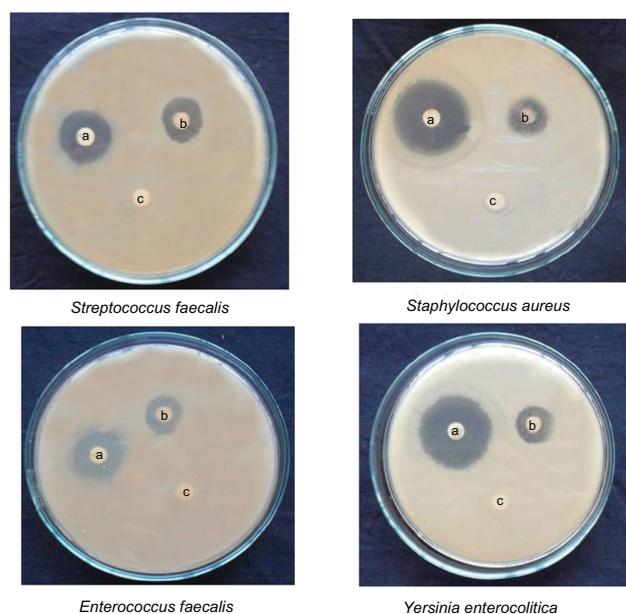


Figure 12 Activity of silver nanoparticles against different micro-organisms depicting zones of inhibition of (A) positive control, (B) silver nanoparticles, and (C) dimethylsulfoxide control.

of gold and silver nanoparticles synthesized from the root extract of *T. decandra* in the treatment of human infections associated with the microorganisms used in this study. These might prove to be important leads for the development of new antimicrobial drugs.

Conclusion

To summarize, we achieved successful green synthesis of gold and silver nanoparticles using *T. decandra* root extract. Silver and gold nanoparticles were synthesized under ambient conditions, and characterization of synthesized nanoparticles was carried out by ultraviolet-visible, Fourier transform infrared, scanning electron and energy dispersive x-ray spectroscopy. It is believed that phytochemicals present in *T. decandra* extract reduce the silver and gold ions into metallic nanoparticles. This strategy reduces the cost of production and the environmental impact. The silver and gold nanoparticles formed showed strong activity against all microorganisms tested.

Acknowledgments

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Disclosure

The authors declare no conflicts of interest in this work.

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