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#### ORIGINAL RESEARCH

Glycyrrhetinic acid and E.resveratroloside act as potential plant derived compounds against dopamine receptor D3 for Parkinson's disease: a pharmacoinformatics study

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Abstract: Parkinson's disease (PD) is caused by loss in nigrostriatal dopaminergic neurons and is ranked as the second most common neurodegenerative disorder. Dopamine receptor D3 is considered as a potential target in drug development against PD because of its lesser side effects and higher degree of neuro-protection. One of the prominent therapies currently available for PD is the use of dopamine agonists which mimic the natural action of dopamine in the brain and stimulate dopamine receptors directly. Unfortunately, use of these pharmacological therapies such as bromocriptine, apomorphine, and ropinirole provides only temporary relief of the disease symptoms and is frequently linked with insomnia, anxiety, depression, and agitation. Thus, there is a need for an alternative treatment that not only hinders neurodegeneration, but also has few or no side effects. Since the past decade, much attention has been given to exploitation of phytochemicals and their use in alternative medicine research. This is because plants are a cheap, indispensable, and never ending resource of active compounds that are beneficial against various diseases. In the current study, 40 active phytochemicals against PD were selected through literature survey. These ligands were docked with dopamine receptor D3 using AutoDock and AutoDockVina. Binding energies were compared to docking results of drugs approved by the US Food and Drug Administration against PD. The compounds were further analyzed for their absorption, distribution, metabolism, and excretion-toxicity profile. From the study it is concluded that glycyrrhetinic acid and E.resveratroloside are potent compounds having high binding energies which should be considered as potential lead compounds for drug development against PD.

**Keywords:** AutoDock, AutoDockVina, molecular docking, parkinson's disease, glycyrrhetinic acid, E.resveratroloside

### Introduction

Parkinson's disease (PD) is the most common form of Parkinsonism. It is a neurodegenerative disorder which notably causes rigidity, bradykinesia, tremor, stooped posture, dementia, and depression in PD patients. Due to unclear etiology this disease is also known as Idiopathic Parkinsonism. But certain factors ie, environmental chemicals, drinking water chemistry, pesticide exposure, and rural living are responsible for PD.<sup>1–3</sup> Moreover, mutations in the *SNCA*, *PARK2*, *PINK1*, *PARK7*, and *LRRK2* genes are also found to be accompanied by PD.<sup>4</sup>

Degeneration of nigrostriatal pathway, noradrenergic locus ceruleus, motor vagal nucleus, the serotonergic raphe nuclei, cholinergic nucleus basalis of Meynert, pedunculopontine nucleus pars compacta, Edinger-Westphal nucleus, and many

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peptidergic brainstem nuclei are characteristic of PD.<sup>5</sup> However, it is believed that the primary cause of the disease is loss in the nigrostriatal dopaminergic neurons, formation of intraneuronal, proteinaceous cytoplasmic inclusions called Lewy bodies, inability to produce dopamine, mitochondrial respiration defect, and oxidative stress. Consequently, a striatal dopamine-deficiency syndrome occurs being responsible for the classical motor symptoms of PD.<sup>6,7</sup>

PD is a progressive disorder, affecting one in every 100 people older than 65 years of age. The symptoms usually appear when approximately 60% of the dopamine producing neurons are lost.<sup>8,9</sup> PD is ranked as the second most common neurodegenerative disorder after Alzheimer's disease.<sup>10</sup> Unfortunately, no therapeutic curative regimen has been devised yet.<sup>9</sup> One of the most prominent treatments for PD is the use of dopamine agonists which mimic the natural action of dopamine in the brain and stimulate dopamine receptors directly.<sup>11</sup> These dopamine agonists will meet the dopamine requirement of the brain and help in bringing about the restoration of normal functionality of the remaining dopaminergic neurons.

Biochemical and electrophysiological data suggest that dopamine acts on its very own special dopamine receptors. They show variance in their anatomical localization, functional substantiality, and pharmacological importance.<sup>12,13</sup> This is why they are differentiated into five types of dopamine receptors (D1–D5). Dopamine receptor D3 (DRD3) serves as a potential target for drug development as several experimental data suggest that DRD3 agonists are involved in neuroprotection and alleviating motor dysfunctions.<sup>14</sup> Furthermore, neuroprotection and neurorestoration are also seen in animal models by using DRD3 agonist.<sup>15</sup>

Dopamine carries out cell cycle modulation in both developing and adult brain and DRD3 receptor play a vital role in dopaminergic neuronal development. Activation of these receptor subtypes leads to propagation of neurogenesis in substantia nigra in animal models. Moreover, a persistent recovery in the locomotor functions was also seen.<sup>16</sup>

Drugs with DRD3 receptor proffering behavior, when acting on their receptors, causes an increase in production of dopamine neurotrophic factor in tissue culture. This factor can also be the autotrophic factor for the dopaminergic neurons. This protein is oxidant labile, therefore, the drugs having DRD3 receptor agonistic activity that results in the increase of this protein and also having the antioxidant profile will provide a unique therapeutic strategy.<sup>17</sup>

In early and moderate PD, the dopamine receptor agonists have shown effectiveness as symptomatic monotherapy.<sup>18</sup>

However, use of current pharmacological therapies such as bromocriptine, apomorphine, and ropinirole provides only temporary relief of the disease symptoms and is frequently linked with insomnia, anxiety, depression, and agitation.<sup>19</sup> Moreover, dose failure or wearing-off effects are common and high doses are required to overcome the problem. Usually elderly patients are highly sensitive to wearing-off effects.<sup>9,20,21</sup> Memory problems and confusion are associated with anticholinergics. Monoamine oxidase inhibitors precipitate many side effects when given in combination therapy. Psychiatric complications, pathological gambling, and depression is also reported with other anti-parkinsonian therapy.<sup>22,23</sup> Thus, there is a need for an alternative treatment that not only hinders neurodegeneration but also has few or no side effects.

Since the past decade much interest has been seen in exploitation of phytochemicals. Phytochemicals are considered as a potential alternative cure for various neurodegenerative diseases including PD. This is because plants are a cheap, indispensable, and never ending resource of active compounds that are beneficial against various diseases. These phytochemicals belong to various classes of compounds such as phenolics, alkaloids, terpenoids, lignans etc. Several phytochemicals having beneficial affects against PD have been reported in literature. Many studies have also revealed promising results in decreasing levels of free radicals which is a major cause of neurodegeneracy.<sup>24–28</sup> Therefore, the current study was designed to find potential lead compounds from different plants against PD that can be developed into commercial drugs.

In this investigation, various phytochemicals active against PD were identified through literature survey. They were screened through in silico docking analysis to find potential lead compounds for PD. The molecular docking tools, AutoDock and AutoDockVina (Scripps Research Institute, La Jolla, CA, USA) were used to dock 40 phytochemicals against DRD3. The ligands were also analyzed for their absorption, distribution, metabolism, and excretion-toxicity (ADMET) profile. ADMET profile determines the absorption, distribution, metabolism and excretion efficiency of the drug. It also predicts the risk of toxicity upon consumption of the compound.<sup>29</sup> Lead compounds having impressive pharmacokinetics and pharmacodynamic properties have a better chance to be developed into commercial drugs. In vitro analysis of ADMET properties is usually carried out during the end stage of drug development due to which several potent compounds fail to achieve drug status. In silico ADMET analysis not only decreases the cost of drug designing but also skips the risk of rejection of a compound.<sup>30</sup>

From the results of the current study it is concluded that glycyrrhetinic acid, and E.resveratroloside showed high binding energy against dopamine DRD3 when docked with AutoDock and AutoDockVina. They also possess a good ADMET profile and should be considered potential lead compounds for drug development against PD.

# Material and methods Preparation of data set

The three dimensional (3D) structure of DRD3 was retrieved from the protein data bank (PDB ID: 3PBL with a resolution [Å]: 2.89, *R*-Value: 0.245).<sup>31</sup> This structure was complexed with dopamine 2/dopamine 3 selective antagonist eticlopride (3-chloro-5-ethyl-N-{[(2S)-1-ethylpyrrolidin-2-yl] methyl}-6-hydroxy-2-ethoxybenzamide). Phytochemicals for the study were collected after extensive literature survey. All phytochemicals included in this study have been reported to be active against PD after in vitro analysis in different studies.<sup>32-34</sup> Their respective structures were obtained from the publicly available database, Pubchem.<sup>35</sup> Selected phytochemicals were sorted according to Lipinski's rule. Only those structures were selected for docking that strictly followed the Lipinski's rule (details for all selected compounds are given in Table S1).

# Protein and ligand optimization

In order to figure out the equilibrium configuration of biomolecules and solids, energy optimization methods were used. Crystal structure of DRD3 was further processed for molecular docking studies. For this, all the hetero-atoms and water molecules were removed from protein structure followed by energy minimization in order to remove all the bad steric clashes using the UCSF Chimera (Resource for Biocomputing, Visualization, and Informatics, University of California San Francisco, San Francisco, CA, USA) for 1,000 steepest descent steps at root-mean-square gradient of 0.02 with an update interval of 10 and using AMBER ff12SB force field.<sup>36,37</sup> The structure-data file two dimensional structures of phytochemicals were converted into MOL 3D structure using Open babel. It was followed by energy minimization through Hyperchem's (Gainesville, FL, USA) MM+ force field.

# Determination of binding site

Binding sites of proteins are often located in the structural pockets and cavities which show high affinity for selective drugs. The binding site of DRD3 was revealed by knowledge based studies.<sup>31</sup> The binding residues were further inspected by using Computed Atlas of Surface Topography of Proteins (CASTp) server and Q-Site Finder.<sup>38,39</sup> CASTp server uses the weighted Delaunay triangulation and the alpha complex for shape measurements. It provides identification and measurements of surface accessible pockets as well as interior inaccessible cavities, for proteins and other molecules. It measures analytically the area and volume of each pocket and cavity, both in solvent accessible surface and molecular surface. Q-Site Finder uses the interaction energy between the protein and a simple van der Waals probe to locate energetically favorable binding sites. Pocket Finder is another tool used for analysis of binding sites of a protein and uses Ligsite algorithm. Ligsite algorithm is designed to heuristically search protein for probable interaction of ligand and generate profiles of best active sites.<sup>40</sup>

# Computational docking strategy between DRD3 and phytochemicals

All computational docking studies were carried out using AutoDock 4.0 and AutoDockVina<sup>41</sup> installed in a single machine running on an OptiPlex 980 (Dell, Round Rock, TX, USA), Corei3-550/3.2 card GT-203 (Intel, Santa Clara, CA, USA) with LINUX (Ubuntu, Canonical Group Limited, London, UK) as an operating system.<sup>42</sup> Automated dockings were performed using the AutoDock 4.0 tool to locate the proper binding orientations and conformations of various ligands. For DRD3 binding precisely, ligands and receptors were subjected to polar hydrogen atoms and Gasteiger charges and all torsions were permissible to rotate while docking. Grid maps were created using the AutoGrid program (Scripps Research Institute). The dimensions of the grid were defined with points separated by 0.375A°. A random approach of starting positions, torsions, and orientations was applied for all ligands and default settings of the AutoDock program were used for translation, quaternion, and torsion steps. Minimization was done by using default parameters by Lamarckian genetic algorithm. The standard docking protocol consisted of 100 runs with an initial population of 150 randomly placed samples for rigid and flexible ligand docking with energy evaluations 2.5×105, mutation rate 0.02, crossover rate of 0.80, maximum number of 27,000 iterations, and an elitism value of 1 used as standard. Root-mean-square tolerance of 1.0A° was used for cluster analysis on the docked results. The cluster having a higher number of confirmation and a low binding energy was selected as the docked pose of the respective ligand; the binding energy of each cluster being the mean binding energy of sum of confirmations present in the cluster.

AutoDockVina was employed to find the most probable ligands for target protein, and ligands were docked at DRD3. For this purpose, blind docking was performed by covering the whole protein under the grid to detect if there is any other binding site available other than provided by PDB crystal structure. Gasteiger charges were added to convert protein and all ligand molecules into PDBQT files. Protein was prepared with identical steps used for AutoDock 4 docking experiment. All ligands were docked by AutoDockVina and results were further analyzed in conjunction with results by AutoDock tools using Chimera. Potential ligands were sorted on the basis of making highest interactions with binding pocket and minimum binding energy values. The comparison of results from both of the tools was considered based on the minimum binding energy.

## Drug-likeness and ADMET analysis

In-silico analysis of drug likeness was performed in order to check the potential DRD3 ligands for their ability to follow Lipinski's rule of five.<sup>43</sup> This was done by uploading ligands to Mcule,<sup>44</sup> Molsoft, and Molinspiration server (http://www.molinspiration.com/cgi-bin/properties) for calculation of their molecular properties (Table 1). The chemical structures of potential ligands were submitted to admetSAR (http://lmmd.ecust.edu.cn:8000/) server for in-silico prediction of ADME-Tox (absorption, distribution, metabolism, excretion and toxicity) properties.<sup>45</sup> Furthermore, online server Osiris Property Explorer (http://www.organic-chemistry.org/prog/peo/) was used to predict the tumorigenic, reproductive, and mutagenic risks (Table 2).

## **Results and discussion**

Molecular docking enables a scientist to virtually screen a number of candidate compounds based on their binding ability and binding orientation with a target molecule. It also allows one to select compounds with strong affinity for the target site. In the current study, phytochemicals have been docked in silico with DRD3 to find the best dopamine agonist candidates that can hinder neurodegeneration in PD. Saponins, flavonoids, alkaloids, lignin, and diarylheptanoids have shown high binding energy when docked with DRD3. Specifically, glycyrrhetinic acid (-7.11 and -12.7 kcal/mol) and E.resveratroloside (-7.03 and -11.7 kcal/mol) have shown stronger binding at the receptor's binding site in experiments when analyzed using AutoDock and AutoDockVina respectively (Tables 3 and 4).

### Binding residues' analysis

We performed binding site analysis of our protein through CASTp and Pocket Finder. CASTp results were analyzed through special plugin of Chimera for better understanding of active site. It was observed that the pockets provided by CASTp were similar to those found in DRD3 crystal structure. It also provided other pockets with measurements. An in-depth analysis of pockets demonstrated that the pocket having residues similar to those found in DRD3 crystal structure could be the most potent active site. The results of CASTp were also compared with Pocket Finder results. The pockets generated through Pocket Finder were analyzed and compared with crystal structure of DRD3. It demonstrated residues already observed in crystal structure and also some additional potent active sites' residues involved in the binding site of protein are shown in (Figure 1). The crystal structure and computational analysis of binding site support that Asp110, Ile183, Ser192, Phe346, His349, Thr369, Tyr373 act as active site residues in 3D crystal structure of human DRD3.31

### Molecular interactions

The location of drug binding is a crucial site for designing viable drug molecules against any disease. Therefore, the

Ligand	Glycyrrhetinic acid	E.resveratroloside	Genkwanin	Protopine	Naringenin	Pergolide	Magnolo
MW	470	390	284.26	353.36	272.25	341.4	266.3
LogP	5.64	1.99	2.87	2.49	2.5	4.2	4.22
TPSA	74.3	138.5	79	57.23	86	44.3	40.4
Reactivity	136	100	78	97	71.5	101.6	84.13
n ON	4	8	5	6	5	2	2
n OHNH	2	5	2	0	3	I	2
n rotb	I	5	2	0	I	I	5
n atoms	80	50	33	45	32	48	38
n violations	0	0	0	0	0	0	0

Table I Lipinski's rule of five drug-likeness properties of potential compounds by using Molsoft, Mcule, and Molinspiration web-servers

Abbreviations: MW, molecular weight; LogP, lipophilic efficiency; n ON, hydrogen bond acceptor; n OHNH, number of hydrogen bond donor; TPSA, topological polar surface area; n violations, number of Lipinski's rule of five violations; n rotb, number of rotatable bonds; n atoms, number of atoms.

ADMET	Glycyrrhetinic acid	E.resveratroloside	Genkwanin	Protopine	Naringenin	Pergolide	Magnolol
BBB	+	+	_	+	+	+	+
Human Intestinal	+	+	+	+	+	+	+
Absorption							
Caco-2 permeable	+	-	+	+	+	+	+
Aqueous solubility	-4.09	-2.45	-3.17	-4.72	-2.64	-4.05	-4.53
P-gp							
Substrate	+	+	+	+	+	+	_
Inhibitor	_	_	_	_	_	_	_
CYP450 Substrate							
CYP450 2C9	_	-	_	_	_	_	_
CYP450 2D6	_	_	_	+	_	_	_
CYP450 3A4	+	-	-	+	_	_	_
CYP450 Inhibitor							
CYP450 1A2	-	-	+	+	+	+	+
CYP450 2C9	_	_	+	_	+	_	+
CYP450 2D6	_	_	_	+	_	+	_
CYP450 2C19	_	_	+	+	+	_	+
CYP450 3A4	_	_	+	_	+	_	+
CYP IP	Low	Low	High	Low	High	Low	High
ROCT	_	_	_	_	_	+	_
HERG Inhibition							
HERG-I	Weak	Weak	Weak	Weak	Weak	Weak	Weak
HERG-II	_	_	_	_	_	+	_
AMES Toxicity	_	_	+	+	_	_	_
, Mutagenic	_	_	_	_	+	_	_
Tumorigenic	_	_	_	_	_	_	_
Reproductive	_	+	_	_	_	+	_
effective							
Carcinogens	_	_	_	_	_	_	_
Biodegradation	_	+	_	+	_	_	_
Irritant	-	-	_	_	_	_	_
RAT, LD50 mol/kg	2.3773	2.1382	2.5626	2.3873	3.511	2.8857	2.0106
FT, pLC50 mol/kg	High, 0.6994	High, 0.9172	High, 0.5913	High, 1.0291	High, 0.7217	High, 1.1094	High, 0.14
TPT, pIGL50 mol/kg	High, 0.9506	High, 0.3495	High, 1.1785	High, 0.4287	High, 0.6757	High, 0.7614	High, 1.9675

Notes: RAT, LD<sub>50</sub> is the lethal dosage of drug when tested on mice; FT (fish toxicity) is environmental risk assessment of drug based on fish and TPT (*Tetrahymena pyriformis* toxicity) as environmental indicators.

**Abbreviations:** ADMET, absorption, distribution, metabolism, and excretion-toxicity; DRD3, dopamine receptor D3; BBB, blood–brain barrier penetration; HIA, human intestinal absorption; Caco-2, Caco-2 permeability; CYP, cytochrome P; IP, inhibitory promiscuity; ROCT, renal organic cation transportation; HERG, human ether-a-go-go-related genes inhibition; RAT, rat acute toxicity; P-gp, permeability glycoprotein; pLC50, lethal concentration, 50%; pIGL50, blood glucose; LD<sub>co</sub>, lethal dose, 50%; +, present; –, not present.

binding site was further confirmed with the help of CASTp and Pocket Finder. It was observed that DRD3 crystal structure shows valuable information of interacting residues, located in the binding pocket such as Asp110, Ile183, Ser192, Phe346, His349, Thr369, and Tyr373, respectively which play an important role in phytochemical binding. Our docking result with phytochemicals showed that many potential compounds were actively involved in hydrogen bonding with five polar residues such as, aspartic acid, serine, histidine, threonine, and tyrosine. Other crucial hydrophobic interactions were also found to exist that increase binding of phytochemicals with protein binding pocket (Table 5). In the docking results however, Thr369 was frequently seen to be a common residue amongst phytochemicals with high binding energies in the docking results. This is seen to be followed by Tyr373, Asp110, and Ile183 residues which also occur frequently. Thr369 and Tyr373 were also found to be common amongst the binding site of synthetic drugs used in the study namely bromocriptine, apomorphine, and ropinirole. Further, docking studies revealed the results with a binding score up to a maximum of -7.11 kcal/mol, which is higher than the binding energy range of

Ligands-PubChem ID	Inhibition	Intermolecular	Binding energy	Docking energy
	constant (µM)	energy (kcal/mol)	G (kcal/mol)	(kcal/mol)
Glycyrrhetinic acid-10114	6.11	-8.65	-7.11	-12.7
E.resveratroloside-6481477	11.65	-8.34	-7.03	-11.7
Genkwanin-5281617	20.64	-6.95	-6.89	-9.5
Protopine-4970	17.94	-6.67	-6.59	-9.6
Naringenin-439246	48.83	-7.07	-6.21	-9.8
Pergolide-47811	41.34	-7.05	-6.15	-8.3
Magnolol-72300	31.77	-8.16	-6.14	-9.1
Bromocriptine-31101	15.36	-7.71	-5.81	-8.7
Apomorphine-6005	11.99	-6.81	-5.71	-8.3
Ropinirole-5095	263.84	-6.76	-4.88	-8

Notes: The table demonstrates a comparison against our computationally selected drugs with US Food and Drug Administration (FDA) drugs that are currently being used in market. The bold font corresponds to FDA approved drugs.

US Food and Drug Administration (FDA) approved drugs (-4.88 to -5.81 kcal/mol). Analyses showed that the top nine compounds with considerable high affinity for DRD3 share a somewhat common active site. After careful observation of molecular interactions, it was recognized that the H-bond interactions with Thr369 and Tyr373 were reported in many phytochemicals that were showing binding energies greater than -6.00 kcal/mol, such as, glycyrrhetinic acid, E.resveratroloside, genkwanin, protopine, naringenin, pergolide, magnolol, honokiol, and hirsutanone. However, glycyrrhetinic acid and E.resveratroloside neatly fit into the binding pocket of DRD3 hence (Figure 1), justifying their high binding values as compared to other compounds.

### Comparative studies with AutoDockVina

The present comparative docking of phytochemicals with DRD3 using two different tools confirmed that AutoDockVina

showed high binding energies as compared to AutoDock Tool. This is due to the fact that Vina uses a gradient optimization method in its local optimization procedure and has better accuracy of the binding mode prediction. Our results suggested that binding energy between ligand and target molecule has been enhanced in AutoDockVina. Binding energies calculated using AutoDock tool for glycyrrhetinic acid, E.resveratroloside, genkwanin, protopine, naringenin, pergolide, and magnolol were in the range (-6.14 to -7.11 tokcal/mol). However, this range of binding energy increased (-8.3 to -12.7 kcal/mol) in the case of AutoDockVina and showed strong binding (Table 6). A varying trend of binding energies was seen amongst the phytochemicals docked in both tools. Flavonoids were observed to rank higher when docked in AutoDockVina as compared to AutoDock where a mixed trend of saponins, lignins, phenols, and alkaloids was seen to rank high.

Ligands-PubChem ID	Binding energy G (kcal/mol)	Binding site interacting residues	No of H-bonds interactions	No of hydrophobic interactions	Total no of bonds
Glycyrrhetinic acid-10114	-12.7	Tyr373, Thr369	2	10	12
E.resveratroloside-6481477	-11.7	Thr369, Phe345, His349, Ile183, Asp110, Ser192	2	6	8
Curcumin-969516	-10.9	lle183, His349, Phe345, Thr369, Asp110	0	11	11
Hirsutanonol-9928190	-10.9	Phe345, Ile183, Ser192	0	8	8
Glabridin_124052	-10.7	Ser192, Phe345, Asp110, lle183	2	8	10
Alloin-313325	-10.6	lle183, Tyr373	0	8	8
Diacerein-26248	-10.3	Tyr373, Thr369	3	10	13
Bromocriptine-31101	-7.9	Ser192, Thr369	0	10	10
Apomorphine-6005	-7.7	Aspl10, Thr369	I.	9	10
Ropinirole-5095	-7.4	Ser192, Phe346, Thr369, Tyr373	0	5	5

**Table 4** AutoDockVina estimated binding energy (G) and different molecular interactions of phytochemicals in the binding site of dopamine receptor D3

**Note:** The bold font corresponds to US Food and Drug Administration approved drugs. **Abbreviation:** No, number.

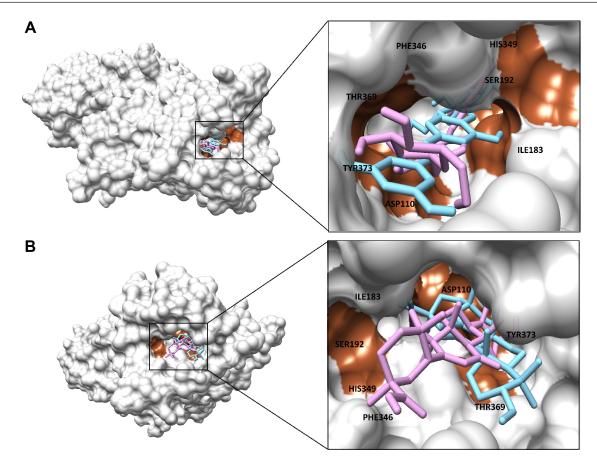


Figure 1 Molecular surface representation of binding pocket of dopamine receptor D3. Notes: Binding pocket residues within the deep groove are labeled as black text and signified with brown surface. Ligands (stick view) are shown with colors as evaluated by AutoDock 4.2 (in pink) and AutoDockVina (in sky blue) in both software programs. (A) Confirmation of E.resveratroloside; (B) confirmation of glycyrrhetinic acid.

Both AutoDock 4.0 and AutoDockVina use Monte-Carlo algorithm. In our experiment, we tried to find the most probable small molecules that may act against DRD3 to cure PD. Docking results were sorted and ranked on the basis of their binding energy values. The first ten compounds from each of AutoDock 4.0 and AutoDockVina results were chosen for further analysis. The comparison showed that four compounds were common that have higher binding energy values for DRD3. Except binding energy values, these four compounds were found to make more interactions

Ligands–PubChem ID	Binding site interacting residues	No of H-bonds interactions	No of hydrophobic interactions	No of polar interactions	No of non-polar interactions	Total no of bonds
Glycyrrhetinic acid-10114	Tyr373, Thr369	2	11	3	4	20
E.resveratroloside-6481477	Asp110, lle183, Ser192, His349, Thr369, Tyr373	2	6	4	3	15
Genkwanin-5281617	Asp110, Ile183, Ser192, His349, Tyr373	I	10	2	5	18
Protopine-4970	Phe346, Thr369	I	9	2	8	20
Naringenin-439246	Asp110, Ile183, Ser192, His349	0	8	5	3	16
Pergolide-47811	lle183, His349, Thr369, Tyr373	I	9	4	2	16
Magnolol-72300	Asp110, Ile183, Ser192, Phe346, Thr369	I	10	3	2	16
Bromocriptine-31101	His349, Thr369	1	11	2	3	17
Apomorphine-6005	Asp110, Thr369, Tyr373	3	9	3	4	19
Ropinirole-5095	Asp110, lle183, Ser192, Phe346, Thr369, Tyr373	I	12	I	5	19

<b>Table 5</b> Phytochemicals displaying different types of molecular interactions with dopamine reception
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**Note:** The bold font corresponds to US Food and Drug Administration approved drugs. **Abbreviation:** No, number.

with DRD3. These compounds were also compared with FDA approved drugs. The comparison with drugs showed that the most potent compounds, glycyrrhetinic acid and E.resveratroloside, showed the highest binding energy values when analyzed using both software and also presented strong interactions with DRD3 (Table 6).

Although differing in precision and binding affinities, glycyrrhetinic acid and E.resveratroloside were popularly recognized as lead compounds in both the software programs (Figure 1A, B). Both ligands were found to bind inside a deep groove surrounded by seven amino acid residues with considerably high binding energy values regardless of the tool being used. Figures 1 and 2 illustrate a detailed interaction of glycyrrhetinic acid and E.resveratroloside with DRD3 inside a deep groove.

#### ADMET analysis

The molecular structures of potential ligands, glycyrrhetinic acid, E.resveratroloside, genkwanin, protopine, naringenin, pergolide, and magnolol were submitted to Mcule, Molsoft, Molinspiration, admetSAR, and Osiris property servers to determine their different properties including drug likeness and ADMET properties. All these potential compounds followed the Lipinski's rule of five without any violation with respect to an octanol-water partition coefficient (LogP  $\leq$ 5), molecular weight ( $\leq$ 500 KDa), number of H-bond donors ( $\leq$ 5), number of H-bond acceptors ( $\leq$ 10), molecular refractivity (40–130) as tabulated in Table 1. In ADMET assessment, different pharmacokinetic and pharmacodynamic parameters were considered such as aqueous solubility,<sup>46</sup> human intestinal absorption,<sup>47</sup> blood—brain barrier penetration, Caco-2 permeability, cytochrome P450 inhibition,<sup>48</sup> cytochrome P (CYP) inhibitory promiscuity, renal organic cation transportation, human ether-a-go-gorelated genes inhibition, rat acute toxicity, fish toxicity, Tetrahymena pyriformis toxicity, AMES toxicity, tumorigenic, reproductive, and mutagenic risks. The results have been summarized in Table 2. The bioavailability and toxicity risks of the potential compounds were predicted based on their ADMET properties. Interestingly, the analysis performed on admetSAR and Osiris property explorer revealed that only glycyrrhetinic acid, E.resveratroloside, and pergolide had no substantial ADMET properties that could cause adverse effects in humans. Whereas genkwanin, naringenin, and magnolol have the potential to show adverse effects in recipients. The analysis displays high CYP inhibitory promiscuity, as they inhibit most of the cytochrome P450 isoforms containing CYP450 1A2, 2C9, 2C19, and 3A4.49 The cytochrome P450 superfamily shows a significant role in metabolizing the drug and its clearance in the liver. Therefore, the inhibition of cytochrome P450 isoforms might affect the drug metabolism and elevate the toxicity level.<sup>50</sup> The analysis also inferred that, except pergolide, none of the potential compounds were potential compounds of the human ether-a-go-go-related gene. In fact, other than pergolide, none of the compounds were carcinogenic, tumorigenic or irritant to humans. Most interestingly, it was noticed that both lead compounds, glycyrrhetinic acid and E.resveratroloside, indicated the most favorable ADMET properties. They were seen to show negative results for AMES toxicity, carcinogenicity, mutagenicity, and tumorigenicity. Both compounds also showed promising human

Table 6 Comparison of estimated binding energies (G) of first ten compounds from each of AutoDock 4.0 and AutoDockVina

AutoDock 4		AutoDockVina			
Ligands-PubChem ID	AutoDock 4 binding energy G (kcal/mol)	Ligands-PubChem ID	AutoDockVina binding energy G (kcal/mol)		
Glycyrrhetinic acid-10114	-7.11	Glycyrrhetinic acid-10114	-12.7		
E.resveratroloside-6481477	-7.03	E.resveratroloside-6481477	-11.7		
Genkwanin-5281617	-6.89	Curcumin-969516	-10.9		
Protopine-4970	-6.59	Hirsutanonol-9928190	-10.9		
Naringenin-439246	-6.21	Glabridin_124052	-10.7		
Pergolide-47811	-6.15	Alloin-313325	-10.6		
Magnolol-72300	-6.14	Diacerein-26248	-10.3		
Honokiol-72303	-6.04	Hesperetin-72281	-10.2		
Hirsutanone-637394	-6.01	Hirsutanone-637394	-10.1		
Hesperetin-72281	-6.01	Luteolin-5280445	-10.1		
Bromocriptine-31101	-5.81	Bromocriptine-31101	-7.9		
Apomorphine-6005	-5.71	Apomorphine-6005	-7.7		
Ropinirole-5095	-4.88	Ropinirole-5095	-7.4		

Note: The bold font corresponds to US Food and Drug Administration approved drugs.

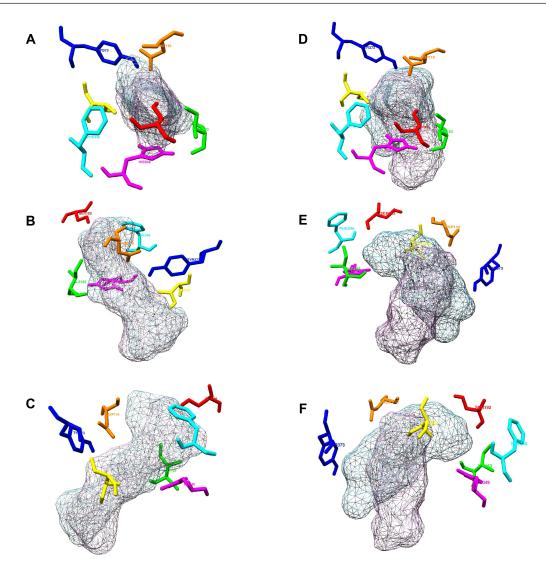


Figure 2 Dopamine receptor D3 and ligands' interactions.

Notes: The seven binding residues of receptor are displayed (stick view) and ligands are represented in mesh form as evaluated by AutoDock 4.2 (in pink) and AutoDockVina (in sky blue). Binding residues are colored as: Asp110 in orange, Ile183 in green, Ser192 in red, Phe346 in cyan, His349 in magenta, Thr369 in yellow, and Tyr373 in blue. (A) top view, (B) back view, and (C) front view of E.resveratroloside. (D) Top view, (E) back view, and (F) front view of glycyrrhetinic acid.

intestinal absorption, blood-brain barrier ability and solubility, and potential non-inhibitor of CYP450 1A2, 2C9, 2D6, 2C19, 3A4, as these factors help in metabolizing and in flushing out the drugs from the body.<sup>49</sup>

# Conclusion

The ascription of all the unique benefits to the DRD3 agonists is currently under investigation and there is a possibility of an increase in therapeutic index of regimes using these agonists. However, an increase in potency of drugs is evident using DRD3 agonists. Anti-parkinsonian efficacy has also dramatically increased using DRD3 agonists via the autotrophic factor pathway. A profound therapeutic leap seems to be promising using these agonists. In the current study 40 phytochemicals were retrieved from a literature survey and docked using AutoDock and AutoDockVina against DRD3 to find potent lead compounds for PD treatment. The compounds were also assessed for their ADMET properties. From this study it can be concluded that Thr369, Tyr373, Asp110, and Ile183 are likely target sites for designing drugs against PD. Also, glycyrrhetinic acid and E.resveratroloside have been identified as potential compounds that should be scrutinized for use against PD as potent drugs. These two compounds have shown strong binding affinity for DRD3. The ligand-receptor complex of glycyrrhetinic acid and E.resveratroloside shows strong hydrogen bonding as well as van der Waals forces between the two. It is apparent that this bonding firmly grips the ligand into the receptor's binding pocket. We performed an in-depth analysis of protein binding site and comparative docking to ascertain best docked poses of ligands. The best ligands we found were based on making high interactions with protein residues and had minimum energy values. The ligand docking results were compared with docking results of FDA approved drugs. Interestingly, a wide difference in affinity values was observed between glycyrrhetinic acid, E.resveratroloside, and FDA approved drugs. A consensus of result was observed when docking results of AutoDock and AutoDockVina were evaluated. Further, glycyrrhetinic acid and E.resveratroloside also exhibited good ADMET profiles and can be considered safe for development into a commercial drug. These results are certainly enlightening that glycyrrhetinic acid and E.resveratroloside can be considered as a template for future drug designing against PD.

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## Disclosure

The authors declare that they have no conflict of interest.

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# Supplementary material

Table SI Molecular properties of compounds under study

Ligands name-PubChem ID	Molecular weight	XLog3	H-bond donor	H-bond acceptor	Molecular formula
Glycyrrhetinic acid-10114	470.6838 [g/mol]	6.4	2	4	C <sub>30</sub> H <sub>46</sub> O <sub>4</sub>
E.resveratroloside-6481477	406.38328 [g/mol]	0.2	5	8	C <sub>20</sub> H <sub>22</sub> O,
Genkwanin-5281617	284.26348 [g/mol]	2.1	2	5	$C_{16}H_{12}O_{5}$
Protopine-4970	353.36856 [g/mol]	2.8	0	6	C <sub>20</sub> H <sub>19</sub> NO <sub>5</sub>
Naringenin-439246	272.25278 [g/mol]	2.4	3	5	C <sub>15</sub> H <sub>12</sub> O <sub>5</sub>
Pergolide-47811	314.48814 [g/mol]	4.2	I	2	C <sub>19</sub> H <sub>26</sub> N <sub>2</sub> S
Magnolol-72300	266.33432 [g/mol]	5	2	2	
Honokiol-72303	266.33432 [g/mol]	5	2	2	$C_{18}H_{18}O_{2}$
Hirsutanone-637394	328.3591 [g/mol]	3.1	4	5	$C_{19}H_{20}O_{5}$
Hesperetin-72281	302.27876 [g/mol]	2.4	3	6	C <sub>16</sub> H <sub>14</sub> O <sub>6</sub>
Bromocriptine-31101	654.5945 [g/mol]	3.8	3	6	$C_{32}H_{40}BrN_5O_5$
Apomorphine-6005	267.32238 [g/mol]	2.3	2	3	$C_{17}H_{17}NO_{2}$
Glabridin 124052	324.3704 [g/mol]	3.9	2	4	C <sub>20</sub> H <sub>20</sub> O <sub>4</sub>
Imperatorin-10212	270.27996 [g/mol]	3.4	0	4	
Apigenin-5280443	270.2369 [g/mol]	1.7	3	5	
Luteolin-5280445	286.2363 [g/mol]	1.4	4	6	
Lisuride-28864	338.44664 [g/mol]	2.7	2	2	C <sub>20</sub> H <sub>26</sub> N <sub>4</sub> O
Polydatin-5281718	390.38388 [g/mol]	1.7	6	8	C <sub>20</sub> H <sub>22</sub> O <sub>8</sub>
Emodin 3220	270.2369 [g/mol]	2.7	3	5	C <sub>15</sub> <sup>20</sup> H <sub>10</sub> O <sub>5</sub>
Diacerein-26248	368.29378 [g/mol]	1.9	I	8	C <sub>19</sub> H <sub>12</sub> O <sub>8</sub>
Rhein-10168	284.22042 [g/mol]	2.2	3	6	C <sub>I</sub> H <sup>2</sup> O
Coumarins 323	146.14274 [g/mol]	1.4	0	2	C,H,Ô2
Hirsutanonol-9928190	346.37438 [g/mol]	2	5	6	C <sub>19</sub> H <sub>22</sub> O <sub>6</sub>
Ropinirole-5095	260.37456 [g/mol]	2.7	I	2	C <sub>16</sub> H <sub>24</sub> N <sub>2</sub> O
Kaempferol 5280863	286.2363 [g/mol]	1.9	4	6	
Genistein 5280961	270.2369 [g/mol]	2.7	3	5	
Elemicin-10248	208.25364 [g/mol]	2.5	0	3	C <sub>12</sub> H <sub>16</sub> O <sub>3</sub>
Pelargonidin-440832	271.24484 [g/mol]	1.2	4	4	C <sub>15</sub> H <sub>11</sub> O <sub>5</sub> <sup>+</sup>
Oregonin-14707658	478.489 [g/mol]	0.5	7	10	$C_{24}H_{30}O_{10}$
Herniarin-10748	176.16872 [g/mol]	1.9	0	3	$C_{10}H_8O_3$
Quercetin 5280343	302.2357 [g/mol]	1.5	5	7	$C_{15}H_{10}O_{7}$
Isoliquiritigenin 638278	256.25338 [g/mol]	3.2	3	4	$C_{15}H_{12}O_{4}$
Arecoline-2230	155.19432 [g/mol]	0.3	0	3	$C_8H_{13}NO_2$
Isohamnetin 5281654	316.26228 [g/mol]	1.9	4	7	$C_{16}H_{12}O_{7}$
Pilocarpine 5910	208.25694 [g/mol]	1.1	0	3	$C_{11}H_{16}N_{2}O_{2}$
Resveratrol-445154	228.24328 [g/mol]	3.1	3	3	$C_{14}H_{12}O_3$
Catechin-9064	290.26806 [g/mol]	0.4	5	6	$C_{15}H_{14}O_{6}$
Curcumin-969516	368.3799 [g/mol]	3.2	2	6	$C_{21}H_{20}O_{6}$
Zingerone 31211	194.22706 [g/mol]	0.8	-	3	$C_{11}H_{14}O_{3}$
Myricetin 5281672	318.2351 [g/mol]	1.2	6	8	$C_{15}H_{10}O_{8}$
Asarone-636822	208.25364 [g/mol]	3	0	3	$C_{12}H_{16}O_{3}$
Alloin-313325	418.39398 [g/mol]	_0.1	7	9	$C_{12}H_{16}O_{3}$ $C_{21}H_{22}O_{9}$
Mannitol 6251	182.17176 [g/mol]	-3.I	6	6	$C_{21} + C_{22} + C_{6} + C_$

Note: The bold font corresponds to US Food and Drug Administration approved drugs.

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