

Tacrolimus in preventing transplant rejection in Chinese patients – optimizing use

Chuan-Jiang Li^{1,*}
Liang Li^{2,*}

¹Department of Surgery, Nanfang Hospital, ²Department of Medical Genetics, School of Basic Medical Sciences, Southern Medical University, Guangzhou, People's Republic of China

*The authors contributed equally to this work

Abstract: Tacrolimus is a product of fermentation of *Streptomyces*, and belongs to the family of calcineurin inhibitors. It is a widely used immunosuppressive drug for preventing solid-organ transplant rejection. Compared to cyclosporine, tacrolimus has greater immunosuppressive potency and a lower incidence of side effects. It has been accepted as first-line treatment after liver and kidney transplantation. Tacrolimus has specific features in Chinese transplant patients; its in vivo pharmacokinetics, treatment regimen, dose and administration, and adverse-effect profile are influenced by multiple factors, such as genetics and the spectrum of primary diseases in the Chinese population. We reviewed the clinical experience of tacrolimus use in Chinese liver- and kidney-transplant patients, including the pharmacology of tacrolimus, the immunosuppressive effects of tacrolimus versus cyclosporine, effects of different factors on tacrolimus metabolism on Chinese patients, personalized medicine, clinical safety profile, and patient satisfaction and adherence. This article provides guidance for the rational and efficient use of tacrolimus in Chinese organ-transplant patients.

Keywords: tacrolimus, liver transplantation, kidney transplant, Chinese, personalized medicine

Introduction

Organ transplantation is the treatment of choice for patients with end-stage organ failure. Kidney and liver transplantation accounts for greater than 90% of all large-organ transplants in the People's Republic of China (PRC). According to data published by the Chinese Scientific Registry of Kidney Transplantation, 6,471 kidney transplantations were performed in the PRC in 2013. The China Liver Transplant Registry gathered data on 20,818 patients who underwent liver transplantation between February 2005 and December 2011. Although the PRC is the second-largest country after the United States (US) with regard to the annual number of organ transplants, only 10,000 transplants are actually performed per year in the PRC. However, approximately 300,000 patients require organ transplantation annually; many die due to shortage of donor organs. Increasing the success and long-term survival of grafts is an efficient way to offset donor organ shortage. Graft rejection is the primary cause of chronic loss of graft function in the PRC.^{1,2} The key to controlling graft rejection is appropriate immunosuppression. Immune-induction therapy, either intravenous antilymphocyte globulin or anti-IL-2 receptor monoclonal antibody and methylprednisolone pulse therapy, is frequently used in the perioperative period. Maintenance therapy includes dual-, triple-, or quadruple-therapy regimens, including cyclosporine A (CsA) or tacrolimus (FK506) as the main components (CsA or tacrolimus plus mycophenolate mofetil [MMF] plus prednisone) to prevent acute rejection.³ Tacrolimus was first used in organ transplantation in the PRC in the late 1990s.⁴ With greater immunosuppression and a

Correspondence: Liang Li
Department of Medical Genetics, School of Basic Medical Sciences, Southern Medical University, 1023 Shatai Road, Baiyun District, Guangzhou, Guangdong 510515, People's Republic of China
Tel +86 20 6164 8510
Email liliang@smu.edu.cn

more favorable safety profile over CsA, tacrolimus gradually became the preferred immunosuppressant after liver and kidney transplantation, in place of CsA. This article reviews the clinical utility of tacrolimus in the PRC.

Pharmacology, mode of action, and pharmacokinetics of tacrolimus

In 1984, tacrolimus, a macrolide lactone antibiotic, was isolated from the fermentation broth of *Streptomyces tsukubaensis* and was considered a novel immunosuppressant.⁵ In vitro animal experiments subsequently revealed that tacrolimus had a strong immunosuppressive effect.^{6,7} In 1989, the use of tacrolimus for the prevention of rejection in liver transplantation was published.⁸ In 1994, the US Food and Drug Administration approved the use of tacrolimus in liver transplantation.^{9,10} Its use has since expanded to other organ types, and today it is the most widely used posttransplant immunosuppressant medication.

There are two main patterns of graft rejection following organ transplantation: acute and chronic rejection. During the process of graft rejection, T cells, B cells, and cytokines play important roles. T cells recognize the antigens of donors and are activated to proliferate, differentiate, and secrete cytokines. These cytokines stimulate B cells to produce anti-graft antibodies. These cytokines also help cytotoxic T cells to develop cytotoxicity against the graft. The antigens of the graft are recognized by T-cell receptors; this stimulation induces generation of the second messengers of T cells. The activation of second messengers results in a sustained high level of intracellular calcium. Calcium activates calcineurin phosphatase, which changes the cytoplasmic component of nuclear factor and transports it into the nucleus. Then, transcription factor is formed, which can activate *IL-2* gene transcription. *IL-2* can induce T-cell proliferation and activation. Activated T cells secrete different types of cytokines. *IL-2* also increases cytotoxic T-cell activity.¹¹ Tacrolimus can form a complex with immunophilin FK-binding protein 12. This complex strongly inhibits calcineurin phosphatase activity and inhibits *IL-2* expression. Subsequently, T-cell activation and cytokine secretion are inhibited. This is the mechanism by which tacrolimus prevents allograft rejection (Figure 1).

Tacrolimus is absorbed in the gastrointestinal tract, reaching peak blood concentrations in 1–3 hours, with a mean bioavailability of approximately 20%–25%.¹² Tacrolimus mainly binds to erythrocytes. In the plasma, 99% of tacrolimus is bound to α_1 -acid glycoprotein. The blood/plasma tacrolimus-distribution ratio is about 20:1. Tacrolimus is the substrate of cytochrome P450 (CYP)-3A isoenzymes and P-glycoprotein.

Tacrolimus is metabolized mainly by CYP3A4 and CYP3A5 in the liver and intestinal epithelium.¹³ P-glycoprotein, also known as multidrug-resistance protein (MDR)-1 or adenosine triphosphate-binding cassette subfamily B member 1, is an adenosine triphosphate-dependent efflux pump. It is expressed in intestinal epithelium cells, limiting the absorption of tacrolimus, and in the bile canalicular membrane of hepatocytes, mediating the biliary elimination of tacrolimus.¹⁴

Tacrolimus has wide interindividual variability in its pharmacokinetics. The maximum area under the curve (AUC) of tacrolimus was almost four times higher than the minimum AUC after the first oral dose in combination with MMF and prednisone among Chinese renal transplant recipients.¹⁵ In addition, it is characterized by a narrow therapeutic index. Underdosage of tacrolimus may result in graft rejection, while overdosage may result in toxicity.^{16,17} In order to avoid adverse effects, therapeutic drug monitoring (TDM) of tacrolimus whole-blood trough concentrations is necessary. In Europe and the United States, AUC-based tacrolimus TDM has been widely used. AUC, the best marker of exposure to tacrolimus, is calculated based on a limited sampling strategy using Bayesian estimation. The dose of tacrolimus is adjusted to reach the AUC target.¹⁸ In the PRC, most hospitals have been using trough blood concentrations for routine dose adjustment of tacrolimus. The proposed initial dose of tacrolimus is approximately 0.10–0.20 mg/kg per day in adult liver-transplant recipients, and 0.15–0.30 mg/kg per day in adult kidney-transplant recipients, and is taken orally twice daily, 1 hour before meals or 2 hours after meals.¹⁹

The recommended target trough blood concentration for Chinese renal transplant recipients is 12–15 ng/mL for the first month after transplantation, 8–12 ng/mL for the second month, 6–10 ng/mL for the third month, and 5–10 ng/mL as the sustained concentration after the third month.²⁰ The recommended target trough blood concentration for Chinese liver-transplant recipients is 10–12 ng/mL during the first 3 months after transplantation, 8–10 ng/mL within 3–6 months, 6–8 ng/mL within 6–12 months, and 4–6 ng/mL as the sustained concentration after 1 year.²¹ For pediatric patients, tacrolimus dosing is based on body-surface area. The recommended starting dose for tacrolimus in Chinese pediatric renal allograft recipients is 4.7–5.6 mg/m² per day, with a maintenance dose of 0.93–1.56 mg/m² per day.²² Owing to interindividual variability, it can take several weeks for the dosage to achieve therapeutic efficacy; transplant recipients have a significant risk of side effects during this period. Therefore, it is paramount to achieve a stable maintenance dose as soon as possible after transplantation.²³

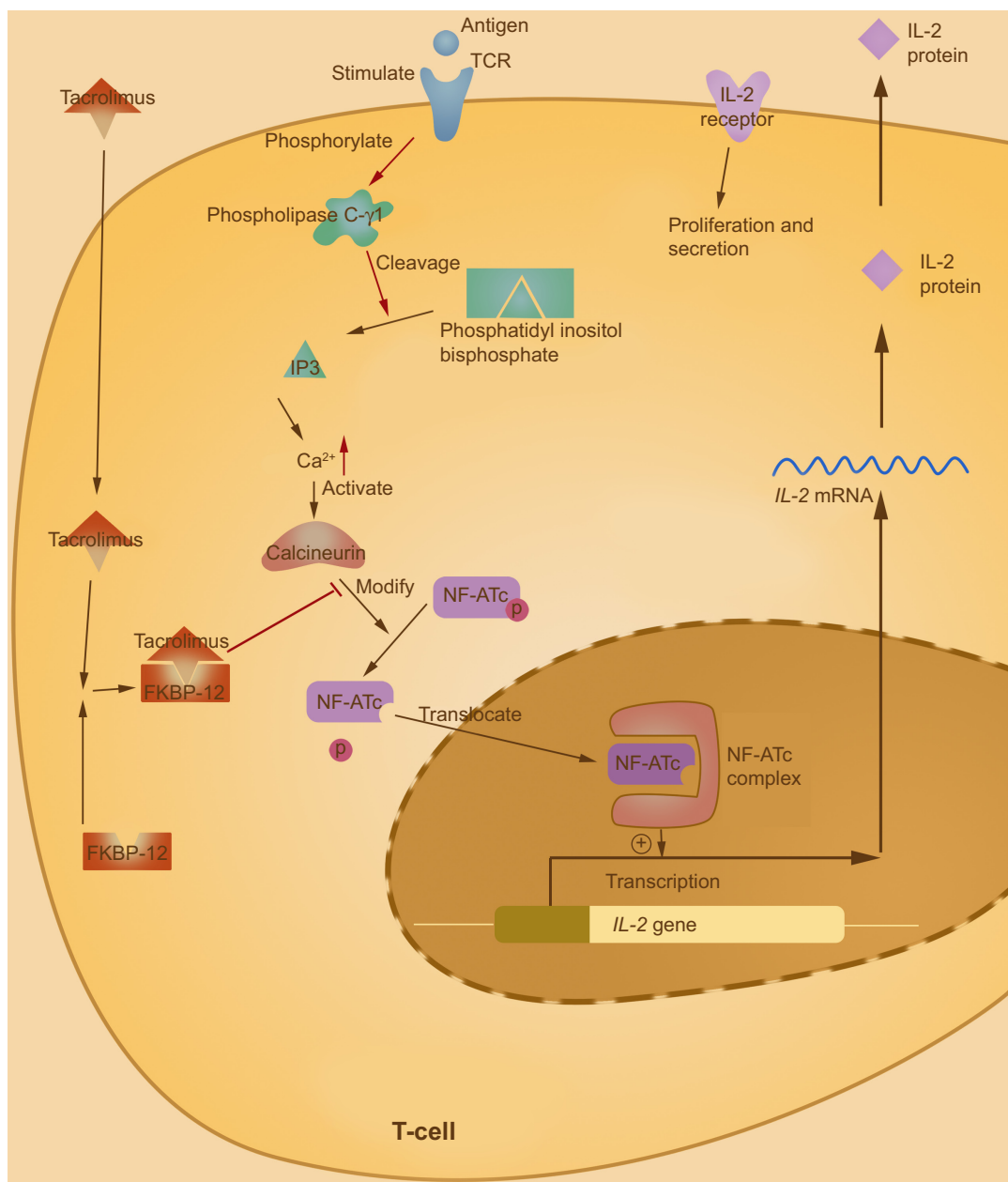


Figure 1 Mechanism of action of tacrolimus.

Abbreviations: FKBP-12, immunophilin FK-binding protein 12; IP3, inositol 1,4,5-triphosphate; mRNA, messenger ribonucleic acid; NF-ATc, nuclear factor of activated T cells; p, phosphate; TCR, T-cell receptor.

Several factors can affect tacrolimus pharmacokinetics. In a study of 142 Chinese renal transplant recipients, tacrolimus maintenance dose negatively correlated with hematocrit, hemoglobin, total bilirubin, and indirect bilirubin, and positively correlated with body weight.¹⁸ In another study of 262 Chinese adult liver-transplant recipients, clearance of tacrolimus significantly correlated with hematocrit and total plasma protein.²⁴ In addition, it has been reported that grapefruit juice can increase tacrolimus concentration in Chinese liver-transplant patients.²⁵

Furthermore, drug–drug interactions can also affect tacrolimus pharmacokinetics.^{26,27}

In addition to these factors, genetic factors play an important role in the variability of tacrolimus pharmacokinetics. A series of studies revealed that *CYP3A5* 6986A>G (*CYP3A5**3) had a significant impact on tacrolimus pharmacokinetics in Chinese renal transplant recipients. Patients with *CYP3A5**3/*3 had higher C₀/D (dose-corrected tacrolimus trough concentrations) compared with patients with *CYP3A5**1/*3 or *CYP3A5**1/*1.^{28–30} Two studies found that

*CYP3A4*1G* was significantly associated with tacrolimus pharmacokinetics in Chinese renal transplant recipients.^{31,32} Patients with *CYP3A4*1/*1* had higher C_0/D and lower tacrolimus clearance compared with patients with *CYP3A4*1/*1G* or *CYP3A4*1G/*1G*. Li et al³³ found that *CYP3A4*18B* may be partly responsible for the large interindividual variability of tacrolimus blood levels in Chinese renal transplant patients during the first month after transplantation. However, in a study of 227 Chinese renal transplant patients, Zhu et al³⁰ found no significant association between the C_0/D of tacrolimus and *CYP3A4*18B* genotypes when they were classified by the different *CYP3A5* genotypes.

Regarding the relationship between *MDR1* polymorphisms and tacrolimus pharmacokinetics, there were inconsistent results in Chinese renal transplant recipients. Several studies revealed that the *MDR1 3435C>T* had a significant effect on the C_0/D of tacrolimus ($P<0.05$). *MDR1 3435CC* patients displayed a lower tacrolimus level per dose than *MDR1 3435CC/TT* patients.^{34–36} Nevertheless, other studies failed to find a significant association between the *MDR1 3435T>C* polymorphism and tacrolimus pharmacokinetics.^{19,37,38} An outline of the genetic polymorphisms affecting tacrolimus pharmacokinetics in Chinese renal transplant patients is presented in Table 1.

The liver is the organ in which tacrolimus is metabolized. Therefore, donor genetics may also affect tacrolimus pharmacokinetics in Chinese liver-transplant patients. Several studies revealed significantly higher tacrolimus C_0/D in donors with *CYP3A5*3/*3* compared with *CYP3A5*1/*3* or *CYP3A5*1/*1* ($P<0.05$), and no significant difference of tacrolimus C_0/D was observed in recipients with different *CYP3A5* genotypes.^{39–41} However, other studies reported that recipient *CYP3A5*3* was also significantly associated with tacrolimus C_0/D .^{42–44} A series of studies reported that recipient *MDR1 3435T>C* significantly affected tacrolimus pharmacokinetics in Chinese liver-transplant patients. A lower daily tacrolimus dose or higher C_0/D was observed in recipients carrying the *MDR1 3435T* allele, compared with *3435CC* recipients at different times after transplantation.^{40,45–47} Correspondingly, two other studies found that recipient *MDR1 3435T>C* was not associated with tacrolimus C_0/D in Chinese liver-transplant patients.^{41,43} Moreover, no correlation has been observed between tacrolimus daily dose or C_0/D and donor *MDR1 3435T>C* in any of the published studies until now.^{40,41,45,46} Li et al⁴¹ reported that the *IL-10 1082G>A* of recipients can affect tacrolimus pharmacokinetics. Recipients with the homozygous variant *IL-10 -1082AA* genotype had higher tacrolimus C_0/D compared with the *IL-10 -1082GG* and *GA* genotype, and the *IL-10 1082G>A* of the donors had no significant association

with tacrolimus C_0/D . Nevertheless, another study found that the *IL-10 1082G>A* of donors can affect tacrolimus C_0/D , and the same variant of recipients did not significantly influence tacrolimus C_0/D .⁴⁶ In addition to these genetic variants, it has been reported that donor *TLR4 rs1927907G>A* and *IL6 rs1800796 G>C* were also closely associated with tacrolimus elimination in Chinese liver-transplant patients.^{42,44} An outline of the genetic polymorphisms affecting tacrolimus pharmacokinetics in Chinese liver-transplant patients is presented in Table 2.

Currently, two studies have individualized the first oral dose of tacrolimus on the basis of the *CYP3A5* genotype in Chinese renal transplant patients. Zhang et al⁴⁸ conducted a study of 76 Chinese renal transplant recipients within two successive periods. In the first period, 0.1 mg/kg/day of tacrolimus was the initial dose prescribed for 28 recipients, regardless of their *CYP3A5* genotype. In the second period, another 48 recipients were prescribed the following doses according to their genotypes: 0.08 mg/kg/day for *CYP3A5*3/*3* (*CYP3A5* non-expressor) and 0.15 mg/kg/day for *CYP3A5*1/*3* (*CYP3A5* expresser). Adjustment of the initial dosage of tacrolimus was documented to improve the proportion of patients achieving target drug blood levels in the early postoperative stage: from 46.7% to 81.8% of the **1/*3* group, and from 46.2% to 73.1% of the **3/*3* group on the third day. Another prospective study, by Chen et al⁴⁹ also confirmed the efficiency of *CYP3A5* genotype-guided use of tacrolimus in Chinese renal transplant recipients. The study consisted of two parts. In the first part, 120 patients received routine initial doses of tacrolimus (0.06 mg/kg every 12 hours). Pharmacokinetic data from these patients were used to fit a regression equation that predicted tacrolimus initial dose on the basis of the *CYP3A5* genotypes. In the second part, patients received initial doses of tacrolimus according to the equation derived from the first part. The results demonstrated that genotype-guided dosing could significantly decrease the incidence of out-of-range tacrolimus levels. Therefore, initial tacrolimus-dosage selection based on *CYP3A5* genotype can improve the proportion of patients achieving target trough blood concentrations in Chinese renal transplant recipients.

However, besides genetic factors, other factors can also affect tacrolimus pharmacokinetics. Li et al¹⁹ developed a tacrolimus-dosing model to predict the initial dosage for Chinese renal transplant patients based on both genetic and clinical factors. However, this model has not been evaluated in clinical practice. Determining additional genetic factors of variability among Chinese individuals in response to tacrolimus and developing the tacrolimus-dosing model are very important to improve therapeutic and clinical efficacy in Chinese transplant patients.

Table 1 Effect of different gene polymorphisms on tacrolimus pharmacokinetics in Chinese renal transplant patients

References	Study subjects	Polymorphisms	Findings
Zhao et al ²⁸	Kidney recipients (n=30)	<i>CYP3A5</i> *3 and *6	Patients with the <i>CYP3A5</i> *3/*3 polymorphisms required less tacrolimus to reach target concentrations compared to those with the <i>CYP3A5</i> *1 allele.
Chen et al ²⁹	Kidney recipients (n=67)	<i>CYP3A5</i> *3	<i>CYP3A5</i> *3 influenced the blood concentrations of tacrolimus. This study suggested that the initial dosage be chosen according to the <i>CYP3A5</i> *3 genotype.
Zhang et al ⁴⁸	Kidney recipients (n=28)	<i>CYP3A5</i> *3	<i>CYP3A5</i> *3 played an important role in influencing tacrolimus blood levels. Initial tacrolimus dosage selection based on <i>CYP3A5</i> *3 genotype could improve drug blood levels in the early stage following renal transplantation.
Chen et al ⁴⁹	Kidney recipients (n=120)	<i>CYP3A5</i> *3	<i>CYP3A5</i> expressers (<i>CYP3A5</i> *1/*3 and <i>CYP3A5</i> *1/*1) needed more tacrolimus to reach therapeutic concentration window.
Zhu et al ³⁰	Kidney recipients (n=227)	<i>CYP3A5</i> *3 and <i>CYP3A4</i> *18B	Patients with <i>CYP3A5</i> *1/*1 and *1/*3 genotypes were significantly lower than those with <i>CYP3A5</i> *3/*3 genotypes. No significant association was found between the <i>C₀/D</i> of tacrolimus and <i>CYP3A4</i> *18B genotypes when they were classified by two <i>CYP3A5</i> genotypes.
Zuo et al ³¹	Kidney recipients (n=161)	<i>CYP3A5</i> *3 and <i>CYP3A4</i> *1G	<i>CYP3A5</i> *3, <i>CYP3A4</i> *1G, and hematocrit were determining factors in the apparent clearance of tacrolimus.
Li et al ³³	Kidney recipients (n=83)	<i>CYP3A5</i> *3 and <i>CYP3A4</i> *18B	<i>CYP3A5</i> *3 and <i>CYP3A4</i> *18B may be partly responsible in large interindividual variability of tacrolimus blood levels in Chinese renal transplant patients during the first month after transplantation.
Rong et al ³⁷	Kidney recipients (n=63)	<i>CYP3A5</i> *3 and <i>MDR1</i> 3435C>T	Renal transplant recipients who were <i>CYP3A5</i> *1 carriers required a higher dose of tacrolimus than <i>CYP3A5</i> *3/*3, indicating a significantly lower dose-adjusted <i>AUC₀₋₁₂</i> of tacrolimus. In contrast, <i>MDR1</i> 3435C>T polymorphism was not an important factor in tacrolimus pharmacokinetics.
Zhang et al ³⁸	Kidney recipients (n=118)	<i>CYP3A5</i> *3 and <i>MDR1</i> 3435C>T	<i>CYP3A5</i> *3 polymorphisms were associated with tacrolimus pharmacokinetics and dose requirements in renal transplant recipients.
Li et al ¹⁹	Kidney recipients (n=142)	<i>CYP3A5</i> *3, <i>MDR1</i> -129T>C, 1236C>T, 2677G>T/A, 3435C>T, and <i>NR1I2</i> -25385C>T	<i>CYP3A5</i> *3, body weight, hematocrit, hemoglobin, and total bilirubin significantly influenced the maintenance tacrolimus dose. <i>MDR1</i> and <i>NR1I2</i> gene polymorphisms were not significantly associated with maintenance tacrolimus dose.
Wu et al ³⁴	Kidney recipients (n=63)	<i>CYP3A5</i> *3, <i>CYP3A4</i> -44G>A, <i>MDR1</i> 1236C>T, 2677G>T/A, and 3435C>T	Tacrolimus <i>C₀/D</i> was larger in <i>CYP3A5</i> *3 and <i>CYP3A4</i> nonexpressers than in expressers. Wild-type homozygotes for <i>MDR1</i> C3435T had a slightly lower dose-adjusted <i>C₀/D</i> compared with heterozygotes. No significant association was found between <i>MDR1</i> 1236C>T, 2677G>T, or haplotypes and tacrolimus pharmacokinetics.
Li et al ³²	Kidney recipients (n=240)	Seventeen polymorphisms of <i>CYP3A5</i> , <i>CYP3A4</i> , <i>COMT</i> , <i>IL-10</i> , and <i>POR</i>	<i>CYP3A5</i> *3, <i>CYP3A4</i> *1G, <i>CYP3A4</i> rs4646437 T>C, and <i>IL-10</i> rs1800871 C>T might be potential polymorphisms affecting the interindividual variability in tacrolimus metabolism among Chinese renal transplant recipients.
Wang et al ³⁵	Kidney recipients (n=86)	<i>MDR1</i> 2677G>T/A and 3435C>T	<i>MDR1</i> 2677G>T/A and 3435C>T were correlated with the whole blood concentration of tacrolimus.
Li et al ³⁶	Kidney recipients (n=66)	<i>MDR1</i> 3435C>T	<i>MDR1</i> 3435C>T was associated with the tacrolimus concentration/dose ratio.

Abbreviations: AUC, area under the concentration–time curve; *C₀/D*, dose-corrected tacrolimus trough concentrations.

Table 2 Effect of different gene polymorphisms on tacrolimus pharmacokinetics in Chinese liver transplant patients

References	Study subjects	Polymorphisms	Findings
Yu et al ³⁹	Liver recipients and donors (n=53)	<i>CYP3A5</i> *3	The tacrolimus C_0/D in patients with <i>*1/*1</i> (<i>*1/*3</i>) genotype donors was significantly lower than in patients with <i>*3/*3</i> genotype donors at 2 weeks ($P<0.036$) and 1 month ($P<0.021$).
Wei-lin et al ⁴⁰	Liver recipients and donors (n=50)	<i>CYP3A5</i> *3 and <i>MDR1</i> 3435C>T	The tacrolimus C_0/D were obviously lower in recipients carrying <i>MDR1</i> 3435CC genotype. For <i>CYP3A5</i> genotype, recipients who received organs from <i>CYP3A5</i> *3/*3 donors had higher C_0/D , but the donors' <i>ABCB1</i> and recipients' <i>CYP3A5</i> genotypes did not affect the recipients' pharmacokinetics.
Li et al ⁴¹	Liver recipients and donors (n=70)	<i>TNFα</i> -308G>A, <i>IL-10</i> -1082G>A, <i>CYP3A5</i> *3, <i>MDR1</i> 2677G>T/A, and 3435C>T	A significantly higher tacrolimus C_0/D was observed in recipients with the homozygous variant <i>IL-10</i> -1082AA genotype, compared with the homozygous wild type <i>IL-10</i> -1082GG genotype. The <i>CYP3A5</i> *3 variant in donors significantly influenced tacrolimus C_0/D during the first 3 weeks posttransplantation.
Shi et al ⁴³	Liver recipients (n=216)	<i>CYP3A5</i> *3, <i>CYP3A4</i> *22, <i>MDR1</i> 3435C>T, and 1236C>T	Recipients <i>CYP3A5</i> *3 affected daily dose requirements, concentration, and nephrotoxicity of tacrolimus.
Wang et al ⁴²	Liver recipients and donors (n=96)	<i>CYP3A5</i> *3 and <i>TLR4</i> rs1927907G>A	Both donor and recipient <i>CYP3A5</i> *3 allele A and donor <i>TLR4</i> rs1927907 allele A were associated with a lower C_0/D during the early stage after transplantation.
Chen et al ⁴⁴	Liver recipients and donors (n=96)	<i>CYP3A5</i> *3 and <i>IL-6</i> rs1800796G>C	Both donor and recipient <i>CYP3A5</i> *3 allele A showed association with lower C_0/D ratios, while donor <i>IL-6</i> rs1800796 allele G showed an association with higher C_0/D ratios. Donor <i>CYP3A5</i> *3 allele A, <i>IL-6</i> rs1800796 allele C, and recipient <i>CYP3A5</i> *3 allele A were associated with fast tacrolimus metabolism.
Jin et al ⁴⁵	Liver recipients and donors (n=50)	<i>MDR1</i> 3435C>T	Tacrolimus dose requirement and dose-adjusted trough levels were correlated with recipient's <i>MDR1</i> 3435C>T polymorphism. No significant differences were found in tacrolimus doses or dose-adjusted trough levels according to the donor's <i>MDR1</i> genotype.
Zhang et al ⁴⁶	Liver recipients and donors (n=53)	<i>IL-10</i> -1082G>A, -819C>T, -592C>A; <i>CYP3A5</i> *3; <i>MDR1</i> 1236C>T, 2677G>T/A, and 3435C>T	Donor <i>CYP3A5</i> *3/*3 was associated with higher tacrolimus C_0/D . In the first 2 weeks, the tacrolimus C_0/D ratios of the recipients with donors who were <i>CYP3A5</i> nonexpressers and had a low <i>IL-10</i> production genotype (-819TT, -592AA) were higher than those with donors who were <i>CYP3A5</i> nonexpressers and had a high <i>IL-10</i> production genotype (-819CC or CT, -592CC or AC).
Yu et al ⁴⁷	Liver recipients (n=62)	<i>MDR1</i> 1236C>T, 2677G>T/A, and 3435C>T	Recipients with C/C genotype at 3435C>T had a little lower tacrolimus C_0/D compared to those with C/T and T/T genotypes ($P=0.024$).

Abbreviation: C_0/D , dose-corrected tacrolimus trough concentrations.

Efficacy studies

With the improvement in technology and accumulation of clinical experience, kidney and liver transplantation has progressed, with increased survival rates of grafts and Chinese transplant recipients. Chen et al⁵⁰ analyzed the data of 1,806 Chinese renal transplant recipients between February 1984 and December 2003. The overall 1-, 5-, 10-, 15-, and 20-year patient-survival rates were 92.28%, 87.20%, 78.60%, 63.45%, and 47.59%, respectively. The 1-, 5-, 10-, 15-, and 20-year graft survival rates were 84.61%, 73.64%, 57.31%,

46.77%, and 31.18%, respectively. The overall half-life of transplanted kidneys was 11.94 ± 0.84 years. In addition, some researchers compared the clinical efficacy of kidney transplant during the different periods. From 1983 to 1989, the 1-, 5-, and 10-year patient-survival rates were 87.8%, 68.3%, and 53.4%, respectively. From 1990 to 1998, the corresponding patient-survival rates were 90.6%, 84.9%, and 79.1%, respectively. The corresponding patient-survival rates were 98.9%, 94.0%, and 89.6% between 1999 and 2012.⁵¹ Patient-survival rates increased with time. Hu et al⁵² analyzed

502 Chinese patients who underwent liver transplantation from 2000 to 2007, of whom 60.1% had malignant liver tumors. The overall 3-year survival rate in this group was 68.7%. Wang et al²¹ performed a retrospective study in a single medical center of 255 liver-transplant patients whose primary disease was hepatocellular carcinoma (HCC). The 1-, 2-, and 3-year patient-survival rates were 84.6%, 66.2%, and 52.6%, respectively; the 1-, 2-, and 3-year recurrence-free survival rates were 67.4%, 53.8%, and 47.3%, respectively. Recurrence of HCC is an important factor that influences long-term survival of Chinese liver-transplant patients.

Advancements in organ transplantation not only result from improvements in surgical technique but also benefit from the utility of immunosuppressive agents. It is particularly important for improving long-term survival of graft organs to choose a rational initial and maintenance immunosuppressive therapy regimen. So far, calcineurin inhibitors, including CsA and tacrolimus, are still the basic components of immunosuppressive regimens. There have been studies that suggested that tacrolimus is associated with a lower incidence of early stage rejection and fewer liver-toxicity events compared to CsA, therefore reducing the risk of chronic loss of graft function and cardiovascular disease while prolonging survival of patients and graft organs.^{53,54} The statistical data are shown in Table 3.

Yu et al²⁰ performed a prospective study to investigate the efficacy of tacrolimus in Chinese kidney-transplant patients. Patients were randomly divided into tacrolimus (n=40) and CsA (n=50) groups. The incidence of rejection in the tacrolimus group decreased by 10% compared to the CsA group, and there were fewer toxicity events associated with tacrolimus. Acute and refractory rejection after transplant surgery is usually steroid-resistant. Favorable curative effects cannot be obtained by high-dose steroid pulse therapy in such patients. In this study, it was demonstrated that tacrolimus could reverse acute and refractory rejection in most cases. Another study also indicated the incidence of acute rejection within 1 year after renal transplantation in the tacrolimus group was 4.0%, while it was 15.6% in the CsA group ($P<0.01$).⁵⁵

Cheung et al⁵⁶ performed a prospective randomized trial with paired kidney analysis to compare the efficacy of tacrolimus and CsA in reducing the incidence of 6-year postoperative long-term rejection of 76 Chinese kidney-transplant patients. The rates of acute rejection were significantly different between groups: 18.4% (7 of 38, tacrolimus group) versus 42.1% (16 of 38, CsA group) ($P=0.03$). Chronic allograft nephropathy (CAN) is the main reason for graft dysfunction. Ji et al⁵⁷ conducted a 3-year follow-up study in 31 patients with pathologically diagnosed CAN. The authors suggested that

conversion from a CsA-based regimen to a tacrolimus-based regimen could effectively improve renal function and delay the progression of CAN. This is comparable to another study including 73 patients with CAN conducted by Peng et al⁵⁸ after 1 year of conversion, the level of serum creatinine in the CsA-tacrolimus group (n=43) was significantly lower than in the CsA group (n=30) ($194.8\pm 42.5\ \mu\text{mol/L}$ versus $245.4\pm 52.8\ \mu\text{mol/L}$, $P<0.01$). Meanwhile, the glomerular filtration rate in the CsA-tacrolimus group was significantly higher than in the CsA group. This indicated that tacrolimus improved renal function more effectively than CsA.

The population with hepatitis B virus (HBV) accounts for about 7% of the total Chinese population. The ratio of HBV-carrier kidney-transplant patients is even higher. Liu et al⁵⁹ indicated that tacrolimus treatment caused less liver function impairment than CsA in HBV-carrier kidney-transplant recipients. In addition, another study suggested that a tacrolimus-based triple regimen in kidney-transplant patients with delayed graft function was associated with fewer adverse reactions and facilitated renal function recovery more effectively.⁶⁰ Tacrolimus was also associated with a lower rejection rate in liver transplantation, and could reverse refractory transplant rejections, so tacrolimus should be the first choice for Chinese patients after liver transplantation.^{4,61} Ye et al⁶² investigated the relationship between serum tacrolimus concentration and clinical efficacy. They found that tacrolimus with 10–15 ng/mL whole blood concentrations not only effectively controlled rejection but also caused fewer adverse reactions.

In conclusion, tacrolimus is a highly effective immunosuppressive agent, and is suitable for Chinese liver- and kidney-transplant patients. Tacrolimus is superior to CsA in controlling rejection, reducing adverse reactions, and prolonging graft survival.

Safety and tolerability

Tacrolimus has been used in Chinese organ-transplant practice for nearly 20 years, and has a good safety and tolerability profile. The short-term adverse effects of tacrolimus include posttransplantation diabetes mellitus, nephrotoxicity, neurotoxicity, and gastrointestinal effects. Symptoms include tremor, headache, diarrhea, and nausea; hyperglycemia and elevation of serum creatinine are additional effects. Xu et al⁶³ analyzed data from 887 Chinese kidney-transplant patients with normoglycemia prior to transplantation. The 3-month and 1-, 3-, 5-, 10-, and 20-year cumulative incidence of hyperglycemia was 10.4%, 11.4%, 13.4%, 15.2%, 22.7%, 27.9%, and 38.3%, respectively. Among them, 61.6% of patients developed posttransplantation diabetes mellitus. The utility of tacrolimus was a risk factor

Table 3 The efficacy and safety of tacrolimus compared with cyclosporine

References	Study subjects	Research methods	Transplant type	Follow-up time	Findings
Yu et al ²⁰	90 cadaveric renal transplant recipients	Randomized into tacrolimus (n=40) and CsA (n=50) groups after cadaveric renal transplantation	Kidney transplantation	12 months	Tacrolimus had a better safety profile and efficacy and fewer side effects than CsA.
Wang et al ⁵⁵	57 cadaveric renal transplant recipients	Randomized into tacrolimus (n=25) and CsA (n=32) groups after cadaveric renal transplantation	Kidney transplantation	Mean follow-up of 12.1 months (7–17 months)	Dramatic reduction in the incidence and severity of acute allograft rejection in patients treated with tacrolimus.
Cheung et al ⁵⁶	76 patients received cadaveric kidneys from 38 donors	Each pair of kidneys was randomly assigned to a separate group and received triple-immunosuppressive therapy with either tacrolimus or Neoral CsA	Kidney transplantation	Mean follow-up duration was 6.1±1.8 years	Tacrolimus-based therapy provided adequate immunosuppression with better renal function and less acute rejection compared with CsA-based therapy.
Ji et al ⁵⁷	31 patients with a histological diagnosis of CAN	Conversion from a CsA-based regimen to a tacrolimus-based regimen	Kidney transplantation	36 months	Conversion from a CsA-based regimen to a tacrolimus-based regimen was an effective choice for salvage of patients with abnormal graft renal function induced by CAN.
Peng et al ⁵⁸	73 renal transplantation patients with CAN proved by allograft biopsy	Patients were either converted to tacrolimus (tacrolimus group, n=43) or remained on their initial CsA-based immunosuppression (CsA group, n=30)	Kidney transplantation	12 months	Conversion from CsA to tacrolimus was an effective and safe alternative therapy for delaying the progression of renal dysfunction induced by CAN.
Liu et al ⁵⁹	109 recipients carrying HBV	Randomized into tacrolimus group (52 cases) and CsA group (57 cases) after kidney transplantation	Kidney transplantation	2 years	For HBV-carrying renal transplant recipients, tacrolimus as the primary choice of immunosuppressant could be more effective and safer than CsA.
Liu et al ⁶⁰	27 patients who developed DGF due to acute renal tubular cell necrosis	Randomized into tacrolimus (n=15) and CsA (n=12) groups after cadaveric renal transplantation	Kidney transplantation	6 months	Tacrolimus-based triple-combined immunosuppressive regimen was the optimal combination of immunosuppressants in cadaveric renal transplant recipients experiencing DGF.
Lo et al ⁶¹	94 LTx performed in 92 patients	The results of 44 LTx performed after November 1996 using a double regimen of tacrolimus and steroids were compared with those of 50 LTx given a triple regimen of CsA, steroids, and azathioprine before this period	LTx	12 months	Tacrolimus offered superior immunosuppression, and will replace CsA as the primary baseline drug in patients after LTx.
Lo et al ⁴	61 patients after orthotopic LTx	13 patients were converted from CsA to tacrolimus. 12 patients received a double regimen of primary tacrolimus-based immunosuppression	LTx	Median follow-up of 12 months (range 1–47 months)	Tacrolimus was highly effective as rescue therapy for resistant rejection, and primary therapy with tacrolimus resulted in a low rejection rate.
Ye et al ⁶²	50 LTx patients	Tacrolimus high-concentration (20 patients), tacrolimus mid-concentration (20), CsA-tacrolimus (5), and tacrolimus-CsA (5) groups	LTx	12 months	Tacrolimus was a highly effective immunosuppressive agent, and the mid-concentration protocol was a better one.

Abbreviations: CAN, chronic allograft nephropathy; CsA, cyclosporine A; DGF, delayed graft function; HBV, hepatitis B Virus; LTx, liver transplantation(s).

for developing diabetes (relative risk 1.835, 95% confidence interval 1.181–2.851; $P=0.007$). Ling et al⁶⁴ analyzed data from 125 Chinese liver-transplant patients without a history of diabetes. Multivariate logistic regression analysis indicated that a serum tacrolimus concentration of more than 10 ng/mL 1 month after transplantation was an independent risk factor for predicting posttransplantation diabetes mellitus (odds ratio 3.264, $P=0.017$). Therefore, serum glucose levels should be strictly monitored in organ-transplant patients. For patients with tacrolimus-associated diabetes, if the condition cannot be controlled by standard drug therapy, conversion to CsA should be considered. Tacrolimus is a calcineurin inhibitor with less nephrotoxicity compared to CsA. However, patients taking tacrolimus still have a risk of developing renal impairment. Lai et al⁶⁵ conducted a study of 124 liver-transplant patients. Of these, 102 patients took tacrolimus as immunosuppressive drug and nine (8.8%) patients developed renal adverse effects. Calcineurin inhibitor-associated nephrotoxicity can be significantly reduced by decreasing the dose of tacrolimus and increasing the dose of MMF.

Tacrolimus has the potential to induce neurotoxicity, particularly when used in pediatric patients. Xie et al⁶⁶ concluded their single-center clinical experience, and suggested that in pediatric transplant patients, tacrolimus-induced neurotoxicity is manifested by epilepsy that occurs within 2 weeks after transplant. High blood tacrolimus concentration can induce epileptic seizures; the adverse effects of tacrolimus are closely associated with blood concentrations.⁶⁷

Adverse effects always occur when blood tacrolimus concentrations are higher than the therapeutic concentration—15 ng/mL. However, most adverse effects are transient, and resolve when the treatment is withdrawn. Therefore, the TDM of tacrolimus trough concentrations is necessary. Transplant recipients require long-term immunosuppressive treatment, along with long-term survival of the graft organ. Therefore, they have a higher risk of developing infections and cancers. Chen et al⁵⁰ analyzed data from 1,806 renal transplant patients; 252 patients died, with 146 (57.9%) from infection and 10 (4.0%) from malignant tumors. Pneumonia is the most common of transplant-associated infections. Zhang et al⁶⁸ reported in 386 kidney transplant patients who received tacrolimus-based immunosuppression an incidence of pneumonia of 7.25%, most of which occurred within 6 months after transplant. Infections were usually associated with tacrolimus overdose. In addition, the expressive status of CYP3A5 was also an independent risk factor for the development of infections in Chinese pediatric liver-transplant patients.⁶⁹

Combination therapy with steroids significantly affected infection and tumor-recurrence rates of transplant patients

who received tacrolimus-based immunosuppressive therapy. In a study conducted by Hu et al⁵² a total of 502 liver-transplant recipients were divided into the following groups: tacrolimus with basiliximab induction and steroid-avoidance group, tacrolimus with 14 days of steroid-withdrawal group, tacrolimus with 3 months of steroid-withdrawal group, and tacrolimus with 6 months of steroid-withdrawal group. The incidence of HBV infection in these groups was 20.5%, 30.5%, 56.1%, and 62.2%, respectively. The 3-year HCC-recurrence rate was lowest in the basiliximab-induction group and the steroid-avoidance group (12.8%, $P=0.037$). These results indicated that a basiliximab-induction and steroid-avoidance immunosuppressive protocol can reduce HBV infection and HCC recurrence after liver transplantation.

In the PRC, calcineurin inhibitors are widely used in combination with mycophenolic acid (MPA), an antimetabolite immunosuppressant. It is reported that tacrolimus can affect the metabolism of MPA. Therefore, the MPA concentration should also be monitored when combined with calcineurin inhibitors, in order to improve the safety of transplant recipients.⁷⁰

In spite of the potential adverse effects of tacrolimus, it is still the most important immunosuppressive agent for Chinese transplant patients, due to its high efficacy and low toxicity. Some strategies can be adopted to control transplant rejection effectively and to the maximum extent possible limit the toxicity of tacrolimus, including optimizing dosage and blood concentration, adopting appropriate combination regimens and personalizing regimens for each patient.

Patient satisfaction and adherence

The success of organ transplantation significantly improves quality of life in end-stage organ failure. However, patients confront certain challenges, such as economic burden and side effects, during the course of their lifetime. Psychological issues may also negatively influence treatment adherence. Approximately 20% of patients are unable to follow up, or cannot follow instructions to adjust drug dosage; some change or stop therapy on their own, resulting in adverse effects.⁷¹ Weng et al⁷² found that there was a significant difference in adherence between patients with once- or twice-daily dosage. The tacrolimus sustained-release capsule is administered only once daily, and its efficacy in preventing acute rejection is comparable with tacrolimus. There are no significant differences in the safety profiles between the formulations.⁷³ The once-daily tacrolimus formulation can significantly improve patient adherence; however, in order to maintain a similar tacrolimus blood concentration, the once-daily dosage of tacrolimus should increase by 30%.⁷⁴

Special features

Since the genetic backgrounds of various ethnic groups are different, the gene variations that can affect the metabolism of tacrolimus are different in various ethnic groups. It has been reported that *COMT*rs2239393, *COMT*rs4646312, and *POR**28 were associated with tacrolimus trough blood concentrations in American renal transplant patients.^{75,76} However, the association between these variants and tacrolimus trough concentration was not found in Chinese renal transplant recipients.³² There are reports that *CYP3A4**22 can affect tacrolimus pharmacokinetics in European renal transplant patients,^{77,78} whereas *CYP3A4**22 has not yet been found in Chinese renal transplant patients.³² *CYP3A5* 6986A>G (*CYP3A5**3) is a well-known genetic variation that can significantly affect the metabolism of tacrolimus. The gene frequency of this variant also varies with ethnic group. It was reported that the frequency of *CYP3A5* expressers (*CYP3A5**1/*1 and *1/*3) was 85% in African-Americans. In Asians, the frequency is 56%, and in Caucasians the frequency is about 16%.⁷⁹ The variation of genetic background in different ethnic groups is a key factor leading to the different tacrolimus dosages used in Chinese and Western transplant recipients.

In Chinese adult liver-transplant patients, 78.47% have HBV, whereas HCV accounts for approximately 60% of liver transplants performed in the US.⁸⁰ Due to the unprecedented proportion of patients with HBV-associated HCC, the application of the Milan criteria is limited in Chinese liver-transplant patients. Niu et al⁸¹ found that HCC-recurrence rates at postoperative years 1, 2, 3, and 4 in the sirolimus group were 13.3%, 36.7%, 43.3%, and 53.3%, respectively, and were 38.7%, 67.7%, 74.2%, and 77.4%, respectively, in the tacrolimus group. Sirolimus significantly reduced tumor recurrence and increased survival for Chinese liver transplant recipients with HCC beyond Milan criteria. Considering this situation, Jia et al⁸² established the Hangzhou criteria, which are suitable for Chinese transplant recipients with HCC. Based on this standard, tacrolimus is still a first-choice immunosuppressive agent for patients in the first month after liver transplantation. After the first postoperative month, sirolimus, which has antitumor effects, can be added to the immunosuppressive regimen and other immunosuppressive agents can be withdrawn gradually. Without decreasing postoperative or tumor-free survival, the Hangzhou criteria effectively expands the indication range for liver cancer-associated liver transplantation. Compared to the Milan criteria, the Hangzhou criteria increase the number of HCC patients who are eligible for liver transplantation by 37.5%. Therefore, more HCC patients can benefit from application of the Hangzhou criteria.

Conclusion

In summary, tacrolimus has been used in Chinese organ transplantation for over 10 years, and is the first-choice calcineurin inhibitor. The safety and efficacy of tacrolimus in preventing posttransplant acute rejection, as well as improving graft-organ survival, have been validated in many clinical trials. Personalized use of tacrolimus has gradually gained the attention of researchers. Owing to a narrow therapeutic range and large individual variability in its pharmacokinetics, the personalized use of tacrolimus is a challenge in clinical practice. To improve tacrolimus efficacy, reduce the incidence of side effects, and prolong graft survival, the appropriate initial and maintenance dose of tacrolimus, rational combination of tacrolimus with other agents, and optimization of immunosuppressive regimens are very important. We should evaluate patients' immune status according to genetic background, liver function, nutritional state, complications such as infection and cancer, and TDM, to find the optimal balance between efficacy and adverse effects, and to personalize the initial and maintenance dose of tacrolimus further for Chinese patients. Conversion from a general therapy regimen for all patients to personalized regimens is the upcoming trend for tacrolimus use in Chinese organ transplantation.

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Disclosure

The authors report no conflicts of interest in this work.

References

1. El-Zoghby ZM, Stegall MD, Lager DJ, et al. Identifying specific causes of kidney allograft loss. *Am J Transplant*. 2009;9:527–535.
2. Bucuvalas J. Long-term outcomes in pediatric liver transplantation. *Liver Transpl*. 2009;15 Suppl 2:S6–S11.
3. Yu LX, Xia RF, Deng WF, et al. 15 year clinical experience in ATG induction after renal transplantation. *Zhonghua Qiguan Yizhi Zazhi*. 2013;34:466–468.
4. Lo CM, Fan ST, Liu CL, et al. Use of FK506 as primary or rescue therapy after liver transplantation in Hong Kong. *Transplant Proc*. 1998;30:3587–3588.
5. Kino T, Hatanaka H, Hashimoto M, et al. FK-506, a novel immunosuppressant isolated from a *Streptomyces*. I. Fermentation, isolation, and physico-chemical and biological characteristics. *J Antibiot (Tokyo)*. 1987;40:1249–1255.
6. Kino T, Hatanaka H, Miyata S, et al. FK-506, a novel immunosuppressant isolated from a *Streptomyces*. II. Immunosuppressive effect of FK-506 in vitro. *J Antibiot (Tokyo)*. 1987;40:1256–1265.

7. Ochiai T, Nakajima K, Nagata M, et al. Effect of a new immunosuppressive agent, FK 506, on heterotopic cardiac allotransplantation in the rat. *Transplant Proc.* 1987;19:1284–1286.
8. Starzl TE, Todo S, Fung J, Demetris AJ, Venkataramman R, Jain A. FK 506 for liver, kidney, and pancreas transplantation. *Lancet.* 1989;2:1000–1004.
9. [No authors listed]. Randomized trial comparing tacrolimus (FK506) and cyclosporine in prevention of liver allograft rejection. European FK506 Multicenter Liver Study Group. *Lancet.* 1994;344:423–428.
10. [No authors listed]. A comparison of tacrolimus (FK506) and cyclosporine for immunosuppression in liver transplantation. The US Multicenter FK506 Liver Study Group. *N Engl J Med.* 1994;331:1110–1115.
11. Pattison JM, Krensky AM. New insights into mechanisms of allograft rejection. *Am J Med Sci.* 1997;313:257–263.
12. Wallemacq PE, Furlan V, Möller A, et al. Pharmacokinetics of tacrolimus (FK506) in paediatric liver transplant recipients. *Eur J Drug Metab Pharmacokinet.* 1998;23:367–370.
13. de Jonge H, de Loo H, Verbeke K, Vanrenterghem Y, Kuypers DR. In vivo CYP3A4 activity, CYP3A5 genotype, and hematocrit predict tacrolimus dose requirements and clearance in renal transplant patients. *Clin Pharmacol Ther.* 2012;92:366–375.
14. Masuda S, Inui K. An up-date review on individualized dosage adjustment of calcineurin inhibitors in organ transplant patients. *Pharmacol Ther.* 2006;112:184–198.
15. Chen YH, Zheng KL, Chen LZ, et al. Clinical pharmacokinetics of tacrolimus after the first oral administration in combination with mycophenolate mofetil and prednisone in Chinese renal transplant recipients. *Transplant Proc.* 2005;37:4246–4250.
16. Staatz C, Taylor P, Tett S. Low tacrolimus concentrations and increased risk of early acute rejection in adult renal transplantation. *Nephrol Dial Transplant.* 2001;16:1905–1909.
17. Kuypers DR, Claes K, Evenepoel P, Maes B, Vanrenterghem Y. Clinical efficacy and toxicity profile of tacrolimus and mycophenolic acid in relation to combined long-term pharmacokinetics in de novo renal allograft recipients. *Clin Pharmacol Ther.* 2004;75:434–447.
18. Saint-Marcoux F, Woillard JB, Jurado C, Marquet P. Lessons from routine dose adjustment of tacrolimus in renal transplant patients based on global exposure. *Ther Drug Monit.* 2013;35:322–327.
19. Li L, Li CJ, Zheng L, et al. Tacrolimus dosing in Chinese renal transplant recipients: a population-based pharmacogenetics study. *Eur J Clin Pharmacol.* 2011;67:787–795.
20. Yu L, Wang Y, Fu SJ, Cheng XJ. Clinical experience with Prograf (tacrolimus, FK 506) in Chinese patients after renal transplantation. *Transplant Proc.* 2000;32:1709–1710.
21. Wang ZX, Song SH, Teng F, et al. A single-center retrospective analysis of liver transplantation on 255 patients with hepatocellular carcinoma. *Clin Transplant.* 2010;24:752–757.
22. Yang S, Wu Z, Wu W, et al. Characteristics of long-term immunosuppressive therapy in Chinese pediatric renal transplant patients: a single-center experience. *Transplant Proc.* 2009;41:4169–4171.
23. Wallemacq P, Armstrong VW, Brunet M, et al. Opportunities to optimize tacrolimus therapy in solid organ transplantation: report of the European consensus conference. *Ther Drug Monit.* 2009;31:139–152.
24. Zhang XQ, Wang ZW, Fan JW, et al. The impact of sulfonylureas on tacrolimus apparent clearance revealed by a population pharmacokinetics analysis in Chinese adult liver-transplant patients. *Ther Drug Monit.* 2012;34:126–133.
25. Liu C, Shang YF, Zhang XF, et al. Co-administration of grapefruit juice increases bioavailability of tacrolimus in liver transplant patients: a prospective study. *Eur J Clin Pharmacol.* 2009;65:881–885.
26. Zuo XC, Zhou YN, Zhang BK, et al. Effect of CYP3A5*3 polymorphism on pharmacokinetic drug interaction between tacrolimus and amlodipine. *Drug Metab Pharmacokinet.* 2013;28:398–405.
27. Xin HW, Li Q, Wu XC, et al. Effects of *Schisandra sphenanthera* extract on the blood concentration of tacrolimus in renal transplant recipients. *Eur J Clin Pharmacol.* 2011;67:1309–1311.
28. Zhao Y, Song M, Guan D, et al. Genetic polymorphisms of CYP3A5 genes and concentration of the cyclosporine and tacrolimus. *Transplant Proc.* 2005;37:178–181.
29. Chen JS, Li LS, Cheng DR, et al. Effect of CYP3A5 genotype on renal allograft recipients treated with tacrolimus. *Transplant Proc.* 2009;41:1557–1561.
30. Zhu L, Song HT, Wang QH, Wu WZ, Yang SL, Tan JM. [Effect of CYP3A4*18B, CYP3A5*3 gene polymorphism on dosage and concentration of tacrolimus in renal transplant patients]. *Yao Xue Xue Bao.* 2012;47:878–883. Chinese.
31. Zuo XC, Ng CM, Barrett JS, et al. Effects of CYP3A4 and CYP3A5 polymorphisms on tacrolimus pharmacokinetics in Chinese adult renal transplant recipients: a population pharmacokinetic analysis. *Pharmacogenet Genomics.* 2013;23:251–261.
32. Li CJ, Li L, Lin L, et al. Impact of the CYP3A5, CYP3A4, COMT, IL-10 and POR genetic polymorphisms on tacrolimus metabolism in Chinese renal transplant recipients. *PLoS One.* 2014;9:e86206.
33. Li DY, Teng RC, Zhu HJ, Fang Y. CYP3A4/5 polymorphisms affect the blood level of cyclosporine and tacrolimus in Chinese renal transplant recipients. *Int J Clin Pharmacol Ther.* 2013;51:466–474.
34. Wu P, Ni X, Wang M, Xu X, Luo G, Jiang Y. Polymorphisms in CYP3A5*3 and MDR1, and haplotype modulate response to plasma levels of tacrolimus in Chinese renal transplant patients. *Ann Transplant.* 2011;16:54–60.
35. Wang W, Zhang XD, Ma LL, et al. [Relationship between MDR1 gene polymorphism and blood concentration of tacrolimus in renal transplant patients]. *Zhonghua Yi Xue Za Zhi.* 2005;85:3277–3281. Chinese.
36. Li D, Gui R, Li J, Huang Z, Nie X. Tacrolimus dosing in Chinese renal transplant patients is related to MDR1 gene C3435T polymorphisms. *Transplant Proc.* 2006;38:2850–2852.
37. Rong G, Jing L, Deng-Qing L, Hong-Shan Z, Shai-Hong Z, Xin-Min N. Influence of CYP3A5 and MDR1(ABCB1) polymorphisms on the pharmacokinetics of tacrolimus in Chinese renal transplant recipients. *Transplant Proc.* 2010;42:3455–3458.
38. Zhang X, Liu ZH, Zheng JM, et al. Influence of CYP3A5 and MDR1 polymorphisms on tacrolimus concentration in the early stage after renal transplantation. *Clin Transplant.* 2005;19:638–643.
39. Yu S, Wu L, Jin J, et al. Influence of CYP3A5 gene polymorphisms of donor rather than recipient to tacrolimus individual dose requirement in liver transplantation. *Transplantation.* 2006;81:46–51.
40. Wei-lin W, Jing J, Shu-sen Z, et al. Tacrolimus dose requirement in relation to donor and recipient ABCB1 and CYP3A5 gene polymorphisms in Chinese liver transplant patients. *Liver Transpl.* 2006;12:775–780.
41. Li D, Zhu JY, Gao J, Wang X, Lou YQ, Zhang GL. Polymorphisms of tumor necrosis factor- α , interleukin-10, cytochrome P450 3A5 and ABCB1 in Chinese liver transplant patients treated with immunosuppressant tacrolimus. *Clin Chim Acta.* 2007;383:133–139.
42. Wang Z, Wu S, Chen D, et al. Influence of TLR4 rs1927907 locus polymorphisms on tacrolimus pharmacokinetics in the early stage after liver transplantation. *Eur J Clin Pharmacol.* 2014;70:925–931.
43. Shi Y, Li Y, Tang J, et al. Influence of CYP3A4, CYP3A5 and MDR-1 polymorphisms on tacrolimus pharmacokinetics and early renal dysfunction in liver transplant recipients. *Gene.* 2013;512:226–231.
44. Chen D, Fan J, Guo F, Qin S, Wang Z, Peng Z. Novel single nucleotide polymorphisms in interleukin 6 affect tacrolimus metabolism in liver transplant patients. *PLoS One.* 2013;8:e73405.
45. Jin J, Wu LH, Wang WL, Yu SF, Yan S, Zheng SS. Impact of multidrug resistance 1 gene polymorphism on tacrolimus dose and concentration-to-dose ratio in Chinese liver transplantation recipients. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi.* 2005;22:616–620.
46. Zhang X, Wang Z, Fan J, Liu G, Peng Z. Impact of interleukin-10 gene polymorphisms on tacrolimus dosing requirements in Chinese liver transplant patients during the early posttransplantation period. *Eur J Clin Pharmacol.* 2011;67:803–813.
47. Yu X, Xie H, Wei B, et al. Association of MDR1 gene SNPs and haplotypes with the tacrolimus dose requirements in Han Chinese liver transplant recipients. *PLoS One.* 2011;6:e25933.

48. Zhang J, Zhang X, Liu L, Tong W. Value of CYP3A5 genotyping on determining initial dosages of tacrolimus for Chinese renal transplant recipients. *Transplant Proc.* 2010;42:3459–3464.
49. Chen SY, Li JL, Meng FH, et al. Individualization of tacrolimus dosage basing on cytochrome P450 3A5 polymorphism – a prospective, randomized, controlled study. *Clin Transplant.* 2013;27:E272–E281.
50. Chen LZ, Chen GD, Wang CX, et al. Outcome of cadaveric kidney transplantation: analysis of 1,806 cases. *Chin J Urol.* 2006;27:166–170.
51. Shu KH, Ho HC, Wen MC, et al. Changing pattern of mortality in renal transplant recipients: a single-center, 30-year experience. *Transplant Proc.* 2014;46:442–444.
52. Hu AB, Wu LW, Tai Q, Zhu XF, He XS. Safety and efficacy of four steroid-minimization protocols in liver transplant recipients: 3-year follow-up in a single center. *J Dig Dis.* 2013;14:38–44.
53. First MR. Improving long-term renal transplant outcomes with tacrolimus: speculation vs evidence. *Nephrol Dial Transplant.* 2004;19 Suppl6:vi17–vi22.
54. Beckebaum S, Klein C, Varghese J, et al. Renal function and cardiovascular risk profile after conversion from ciclosporin to tacrolimus: prospective study in 80 liver transplant recipients. *Aliment Pharmacol Ther.* 2009;30:834–842.
55. Wang XH, Tang XD, Xu D. Tacrolimus vs CyA Neoral in combination with MMF and steroids after cadaveric renal transplantation. *Transplant Proc.* 2000;32:1702–1703.
56. Cheung CY, Chan HW, Liu YL, Chau KF, Li CS. Long-term graft function with tacrolimus and cyclosporine in renal transplantation: paired kidney analysis. *Nephrology (Carlton).* 2009;14:758–763.
57. Ji SM, Li LS, Sha GZ, Chen JS, Liu ZH. Conversion from cyclosporine to tacrolimus for chronic allograft nephropathy. *Transplant Proc.* 2007;39:1402–1405.
58. Peng LK, Xie XB, Peng FH, et al. [Slowing progression of chronic allograft nephropathy by conversion from cyclosporin A to tacrolimus]. *Zhong Nan Da Xue Xue Bao Yi Xue Ban.* 2007;32:59–62. Chinese.
59. Liu XY, Yu LX, Fu SJ, et al. [Application of tacrolimus and cyclosporine A in HBV-carrying renal transplant recipients]. *Nan Fang Yi Ke Da Xue Xue Bao.* 2007;27:1090–1092. Chinese.
60. Liu B, Lin ZB, Ming CS, et al. Randomized trial of tacrolimus in combination with mycophenolate mofetil versus cyclosporine with mycophenolate mofetil in cadaveric renal transplant recipients with delayed graft function. *Transplant Proc.* 2003;35:87–88.
61. Lo CM, Fan ST, Liu CL, et al. More effective immunosuppression with the use of FK506 after liver transplantation. *Transplant Proc.* 2000;32:2269–2270.
62. Ye QF, Ruzig MA, Gong NQ. Administration of tacrolimus in 50 liver transplantation patients. *Hepatobiliary Pancreat Dis Int.* 2002;1:492–494.
63. Xu Y, Liang JX, Liu B, et al. Prevalence and long-term glucose metabolism evolution of post-transplant diabetes mellitus in Chinese renal recipients. *Diabetes Res Clin Pract.* 2011;92:11–18.
64. Ling Q, Xie H, Lu D, et al. Association between donor and recipient TCF7L2 gene polymorphisms and the risk of new-onset diabetes mellitus after liver transplantation in a Han Chinese population. *J Hepatol.* 2013;58:271–277.
65. Lai W, Lu SC, Wang ML, et al. Mycophenolate mofetil-based calcineurin inhibitor reduced immunosuppressive protocol for the improvement of renal dysfunction after liver transplantation. *Zhonghua Yi Xue Za Zhi.* 2009;89:1529–1532.
66. Xie M, Rao W, Sun LY, et al. Tacrolimus-related seizure after pediatric liver transplantation – a single-center experience. *Pediatr Transplant.* 2014;18:58–63.
67. Wu Z, Meng Q, Xia Y, Zhang F, You W. A retrospective analysis of the safety and efficacy of low dose tacrolimus (FK506) for living donor liver transplant recipients. *J Biomed Res.* 2013;27:305–309.
68. Zhang YX, Yu LX, Fu SJ, Ye JS, Liu XY. [Clinical study of pulmonary infection in kidney transplantation recipients taking new immunosuppressant]. *Nan Fang Yi Ke Da Xue Xue Bao.* 2008;28:1037–1040. Chinese.
69. Xue F, Han L, Chen Y, et al. CYP3A5 genotypes affect tacrolimus pharmacokinetics and infectious complications in Chinese pediatric liver transplant patients. *Pediatr Transplant.* 2014;18:166–176.
70. Huang HF, Yao X, Chen Y, et al. Cyclosporine A and tacrolimus combined with enteric-coated mycophenolate sodium influence the plasma mycophenolic acid concentration – a randomised controlled trial in Chinese live related donor kidney transplant recipients. *Int J Clin Pract Suppl.* 2014:4–9.
71. Butler JA, Roderick P, Mullee M, Mason JC, Peveler RC. Frequency and impact of nonadherence to immunosuppressants after renal transplantation: a systematic review. *Transplantation.* 2004;77:769–776.
72. Weng FL, Israni AK, Joffe MM, et al. Race and electronically measured adherence to immunosuppressive medications after deceased donor renal transplantation. *J Am Soc Nephrol.* 2005;16:1839–1848.
73. Posadas Salas MA, Srinivas TR. Update on the clinical utility of once-daily tacrolimus in the management of transplantation. *Drug Des Devel Ther.* 2014;8:1183–1194.
74. Ma MK, Kwan LP, Mok MM, Yap DY, Tang CS, Chan TM. Significant reduction of tacrolimus trough level after conversion from twice daily Prograf to once daily Advagraf in Chinese renal transplant recipients with or without concomitant diltiazem treatment. *Ren Fail.* 2013;35:942–945.
75. Jacobson PA, Oetting WS, Brearley AM, et al. Novel polymorphisms associated with tacrolimus trough concentrations: results from a multicenter kidney transplant consortium. *Transplantation.* 2011;91:300–308.
76. Elens L, Hesselink DA, Bouamar R, et al. Impact of POR*28 on the pharmacokinetics of tacrolimus and cyclosporine A in renal transplant patients. *Ther Drug Monit.* 2014;36:71–79.
77. Elens L, van Schaik RH, Panin N, et al. Effect of a new functional CYP3A4 polymorphism on calcineurin inhibitors' dose requirements and trough blood levels in stable renal transplant patients. *Pharmacogenomics.* 2011;12:1383–1396.
78. Kurzawski M, Dąbrowska J, Dziewanowski K, Domański L, Perużyńska M, Drożdżik M. CYP3A5 and CYP3A4, but not ABCB1 polymorphisms affect tacrolimus dose-adjusted trough concentrations in kidney transplant recipients. *Pharmacogenomics.* 2014;15:179–188.
79. Zan SJ, Zhu LQ, Duan WY, Zhang Y. Effects of race on the pharmacokinetics of tacrolimus in transplantation patients. *China Pharm.* 2013;24:2438–2441.
80. Saidi RF. Current status of liver transplantation. *Arch Iran Med.* 2012;15:772–776.
81. Niu YJ, Liu Y, Wang LT, et al. Impact of sirolimus and tacrolimus on the tumor recurrence after liver transplantation due to HCC beyond Milan criteria: randomized controlled clinical trial. *Chin J Organ Transplant.* 2014;35:99–102.
82. Jia JJ, Lin BY, He JJ, et al. “Minimizing tacrolimus” strategy and long-term survival after liver transplantation. *World J Gastroenterol.* 2014;20:11363–11369.

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