

Gelatin nanoparticle-mediated intranasal delivery of substance P protects against 6-hydroxydopamine-induced apoptosis: an in vitro and in vivo study

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Background: The aim of this study was to investigate the protective role of intranasally administered substance P-loaded gelatin nanoparticles (SP-GNPs) against 6-hydroxydopamine (6-OHDA)-induced apoptosis in vitro and in vivo, and to provide a new strategy for treating brain pathology, such as Parkinson's disease.

Methods: SP-GNPs were prepared by a water-in-water emulsion method, and their stability, encapsulating efficiency, and loading capacity were evaluated. PC-12 cells were used to examine the enhancement of growth and inhibition of apoptosis by SP-GNPs in vitro using MTT assays. In the in vivo study, hemiparkinsonian rats were created by intracerebroventricular injection of 6-OHDA. The rats then received intranasal SP-GNPs daily for 2 weeks. Functional improvement was assessed by quantifying rotational behavior, and the degree of apoptosis was assessed by immunohistochemical staining for caspase-3 in the substantia nigra region.

Results: PC-12 cells with 6-OHDA-induced disease treated with SP-GNPs showed higher cell viability than their untreated counterparts, and cell viability increased as the concentration of substance P (SP) increased, indicating that SP could enhance cell growth and inhibit the cell apoptosis induced by 6-OHDA. Rats with 6-OHDA-induced hemiparkinsonism treated with SP-GNPs made fewer rotations and showed less staining for caspase-3 than their counterparts not treated with SP, indicating that SP protects rats with 6-OHDA-induced hemiparkinsonism from apoptosis and therefore demonstrates their functional improvement.

Conclusion: Intranasal delivery of SP-GNPs protects against 6-OHDA-induced apoptosis both in vitro and in vivo.

Keywords: gelatin nanoparticles, intranasal delivery, substance P, 6-hydroxydopamine, apoptosis, Parkinson's disease

Introduction

A hydroxyl derivative of catecholamine, 6-hydroxydopamine (6-OHDA) is a neurotoxicant that activates apoptotic and proapoptotic factors, eg, caspase proteins, as well as transduction of Bax factor, leading to apoptosis and degeneration of dopamine neurons.¹⁻³ Studies have shown that 6-OHDA can induce apoptosis in PC-12 (adrenal pheochromocytoma) cells by activating apoptotic factors.² Also, rats intracerebroventricularly injected with 6-OHDA show apoptosis, degeneration, and death of dopaminergic neurons in the substantia nigra. The apoptosis occurs mainly due to a caspase family member-mediated protease cascade, and caspase-3 has a vital role in this process. If large numbers of dopaminergic neurons undergo apoptosis, the result is irreversible degenerative brain disease, ie, Parkinson's disease (PD), for which there is still no effective therapy.⁴

Substance P (SP), a member of the tachykinin peptide family, is involved in the regulation of many biological processes in the central and peripheral nervous

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systems (Figure 1).⁵ SP-containing neurons are widely distributed throughout the central and peripheral nervous systems, especially in the substantia nigra region.⁶ Most SP receptors are located within dopaminergic and cholinergic neurons in the basal ganglia, suggesting that SP may have a physiologically regulating effect on these neurons.⁷ Therefore, SP and its receptor may have a therapeutic use in PD, which is characterized by impaired dopaminergic transmission.

It has been reported that SP and dopamine are regulated via a positive feedback mechanism whereby binding of SP to its tachykinin neurokinin-1 receptor on dopamine neurons causes striatal release of dopamine, and by binding to its D1 receptor on striatal projection neurons, dopamine potentiates the release of SP within the substantia nigra.⁸ Previous research has shown that expression of SP is significantly decreased in the basal nuclei in both hemiparkinsonian rats and PD patients, indicating probable involvement of SP in regulating the pathogenesis of PD.⁷ In vitro experiments have demonstrated that SP can reduce anti-Fas-induced apoptosis in human tenocytes via regulation of neurokinin-1-specific and Akt-specific pathways.⁹ An vivo study suggested that SP can reduce apoptotic cell death by modulating the immune response in the early stages after spinal cord injury.¹⁰

Intracerebroventricular administration of SP to rats with 6-OHDA-induced disease can increase the dopamine content in the brain and help to restore the dopamine deficit, with the positive effects seen being more prominent in the nigrostriatal system than in the mesocorticolimbic dopaminergic system.¹¹ Further, hemiparkinsonian rats pretreated with SP fragments¹² or an SP receptor antagonist¹³ show increased levels of dopamine and its metabolites in the corpus striatum,

as well as clear functional recovery. However, the current research focuses on the pharmacological effects of SP given by invasive intracerebroventricular injection, which can result in a high local concentration of SP in the brain, and it has been confirmed that a high level of SP in the brain can induce serious neuroinflammation and further aggravate illness,^{8,14,15} so intracerebroventricular injection is not a safe or practical strategy for PD patients who need continuous treatment.

Intranasal administration has been reported to be an efficient and noninvasive way to delivery biologics directly into the brain.¹⁶ Gelatin nanoparticles (GNPs) are a type of gelatin-cored nanostructured lipid carrier prepared by a water-in-water emulsion method and have good stability and strong penetrating ability, encapsulating efficiency, and loading capacity, as well as bioactivity.¹⁷ It has been reported that GNPs are a suitable carrier for targeted delivery, making it possible to delivery therapeutics to a focal zone effectively without compromising drug stability or concentration.^{18,19} Theoretically, a novel strategy combining GNP-loaded therapeutics and the nasal olfactory pathway might maximize the potential efficacy of SP in the treatment of PD.

In the present study, we investigated whether intranasally administered SP-GNPs could maximize the ability of SP to protect against 6-OHDA-induced apoptosis in vitro and in vivo. SP-GNPs were prepared by a water-in-water emulsion method and were found to have good stability, encapsulating efficiency, and loading capacity. The protective effect of SP-GNPs on PC-12 cells with 6-OHDA-induced disease was assessed by MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] assay, and the inhibition of apoptosis and neuroprotective effect of SP-GNPs in rats

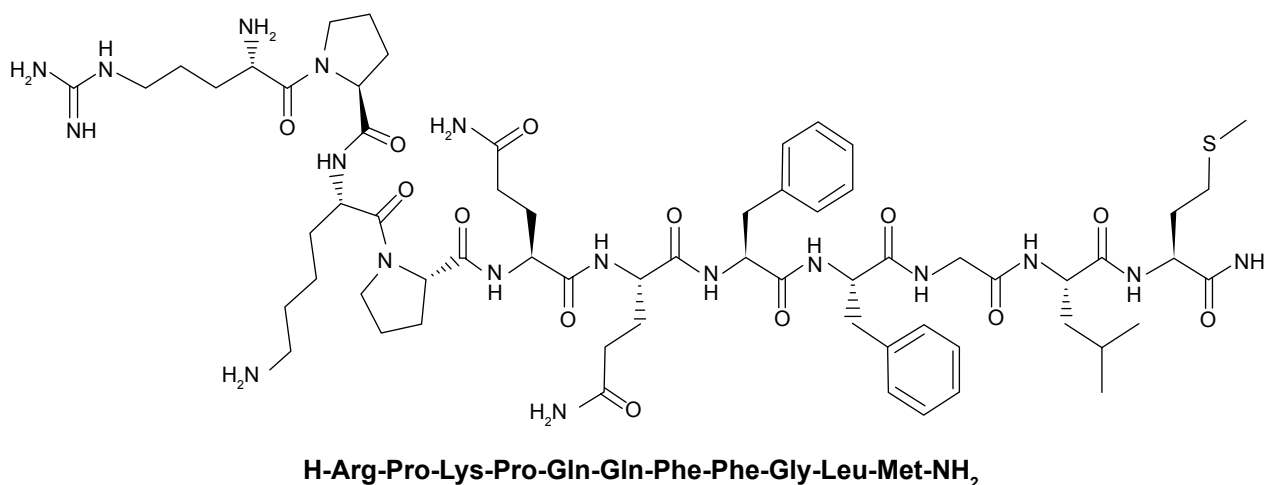


Figure 1 Molecular structure of substance P.

with 6-OHDA-induced hemiparkinsonism were evaluated by behavioral assessment and immunohistochemical staining for caspase-3.

Materials and methods

Materials and animals

All the materials and equipment used in this study were commercially available. SP was purchased from GL Biochem Ltd (Shanghai, People's Republic of China). The protocols and procedures were approved by the local animal experimentation ethics committee. Male Sprague-Dawley rats weighing around 300–320 g were provided by the Laboratory Animal Services Centre at our university. Two to three animals were housed per stainless steel cage on a 12-hour light/12-hour dark cycle in an air-conditioned room at 22°C, and checked daily by the animal care staff. Standard commercial rat chow (Prolab RMH 2500, PMI Nutrition International LLC, Brentwood, MO, USA) and water were available *ad libitum*.

Preparation and characterization of SP-GNPs and blank GNPs

SP-GNPs and blank GNPs were prepared using water-in-water emulsion and freeze-drying techniques.^{20,21} Briefly, a high concentration of SP was dissolved in 1 mL of 20% w/v Poloxamer 188-grafted heparin copolymer solution. This solution was added to 2 mL of 2.0% w/v gelatin solution to produce a homogeneous mixture. Under sonication (110 W, 15°C) using a probe sonicator, D,L-glyceraldehyde was injected into the mixed solution until its final concentration reached 0.1% w/v to initiate the cross-linking reaction. The mixture was bathed at 5°C under magnetic stirring at 2,500 rpm for 5 hours to form a suspension of SP and GNPs. The suspension was lyophilized to obtain a powder containing SP and polymeric GNPs. Next, the lyophilized powder was dispersed in a solution containing soy phosphatidylcholine, trehalose, and cholesterol. By sonication (90 W, 20 seconds) at 25°C, the suspension was then lyophilized to obtain a powder containing SP-GNPs, which were reconstituted in double-distilled water to form a 2 mg/mL SP-GNP suspension for administration. Blank GNPs (using gelatin solution instead of SP gelatin solution during preparation) was also prepared for the subsequent experiment.

The morphologies of the SP-GNPs and blank GNPs were determined using a scanning electron microscope (X-650, Hitachi, Tokyo, Japan). The particle size and zeta potential were determined by dynamic light scattering using a Nicomp™ 380 ZLS zeta potential/particle sizer (PSS Nicomp, Santa Barbara, CA, USA).

To determine the encapsulating efficiency of the SP-GNPs and blank GNPs, approximately 1.5 mL of the SP-GNP dispersion were placed in a microtube and centrifuged at 10,000 g for 40 minutes. The supernatant was then collected and diluted for determination of SP content using an enzyme-linked immunosorbent assay kit; this experiment was performed in triplicate. Drug encapsulation efficiency (%) = (total amount of drug – amount of drug in supernatant)/total amount of drugs added initially × 100%.

Experiment in vitro

Cell culture

Male rat PC-12 cells were used for the *in vitro* study. The PC-12 cells were cultured at 37°C in high-glucose Dulbecco's Modified Eagle's Medium with 10% fetal bovine serum and 1% penicillin–streptomycin in a humidified incubator containing 5% CO₂. Cells in the logarithmic growth phase were harvested with trypsin for further experiments.

MTT assay

The ability of SP-GNPs to impede the growth of PC-12 cells with 6-OHDA-induced disease was confirmed by MTT assay (run in triplicate). PC-12 cells were cultured in a 96-well plate for 24 hours at a density of 5,000 cells per well. With blank PC-12 cells as the control, 100 μM of 6-OHDA was added to the cells for 24 hours to induce cell apoptosis, after which blank GNPs and different concentrations of SP-GNPs were incubated for another 24 hours. Next, 10 μL of MTT 5 mg/mL were added to each well and incubated for 4 hours; 100 μL of formazan solution was then added to each well, followed by incubation for a further 4 hours to dissolve the crystals that developed in each well. The plate was then put into a microplate reader to measure the optical density at 526 nm and quantify the extent of cell viability. The higher the amount of cell viability in each well, the less the degree of apoptosis.

Experiment in vivo

Rat model of hemiparkinsonism

The rats were anesthetized with pentobarbital sodium 60 mg/kg and then injected with 12 μL of 6-OHDA solution into the right striatum (or vehicle for sham animals) using stereotaxic apparatus (Figure 2).^{22,23} Gentamicin was then given to prevent infection.

Four weeks after injection of the 6-OHDA solution, rodent behavior was evaluated by counting the number of apomorphine-induced rotations to determine if the rat model

of hemiparkinsonism had been successfully created. The rats were injected with apomorphine 0.5 mg/kg subcutaneously, and both contralateral and ipsilateral full-body rotations were recorded in the following 30 minutes. At least seven full-body contralateral rotations per minute were considered to indicate a successful hemiparkinsonian (PD) model, and these rats were used in the following experiment.

Effect of SP-GNPs in hemiparkinsonian rats

The day after the behavior evaluation, the sham rats and PD rats were randomized into five groups ($n=8$ per group) and started on daily treatment for 2 weeks. Group 1 comprised sham rats receiving intranasal phosphate-buffered saline and group 2 comprised PD rats receiving intranasal blank GNPs. Groups 3, 4, and 5 comprised PD rats receiving intranasal SP-GNPs at different concentrations (Table 1).

Two hours after the end of 2 weeks of daily treatment, the experimental rats were injected subcutaneously with apomorphine 0.5 mg/kg to evaluate the extent of their neurorecovery. All contralateral and ipsilateral full-body rotations were recorded during the 30 minutes following injection of apomorphine. The fewer the number of rotations in the hemiparkinsonian rats, the better the neurorecovery was deemed to be.

All rats were euthanized at this point, and their brains were collected for coronal sectioning across the substantia nigra region. The brain tissues were then embedded in paraffin and sectioned for immunohistochemical staining.

Immunohistochemical staining

As one of the endpoint shear enzymes in apoptosis, caspase-3 plays a critical role in the apoptotic cascade.²⁴ Immunohistochemical staining with anti-caspase-3 antibody was used to evaluate levels of apoptosis in the substantia nigra region in hemiparkinsonian rats treated or not treated with SP. Image-Pro Plus version 6.0 was used to quantify the number of cells, the areas stained, and the degree of staining. The better the

protective effect against 6-OHDA-induced apoptosis, the lower rates of caspase-3 staining and apoptosis in brain sections from the PD rats.

Statistical analysis

Statistically significant differences across multiple groups were determined using one-way analysis of variance with the Newman–Keuls post hoc test. Statistically significant differences between individual groups was determined using the Mann–Whitney *U*-test. All testing was done using Statistical Package for the Social Sciences version 19 software (SPSS Inc, Chicago, IL, USA). A difference was considered to be statistically significant at $P<0.05$.

Results

Physicochemical properties and bioactivity of SP-GNPs and blank GNPs

Scanning electron micrographs showed that the SP-GNPs and blank GNPs were uniform in shape and size (Figure 3). Characterization for the SP-GNPs and blank GNPs is shown in Table 2. Dynamic light scattering showed the average particle size of the blank GNPs to be 136 ± 1.32 nm.

The polydispersity index (PDI) indicates the distribution of particle size. Low PDI values were observed for the SP-GNPs and the blank GNPs (Table 2), indicating that both were monodispersed stable systems. After loading with SP, the mean diameters of the nanoparticles and liposomes increased, but were still below 200 nm (Table 2).

The zeta potential is an important indicator of the physical stability of nanoparticles. Nanoparticles with a high absolute zeta potential value are electrically stable while those with a low absolute zeta potential value tend to be less electrically stable. As shown in Table 2, both the SP-GNPs and the blank GNPs had a strong negative surface charge, indicating that coating with phospholipids makes these nanoparticles more stable.

The encapsulation efficiency and loading capacity of the SP-GNPs were $93.3\pm1.4\%$ and $5.2\pm0.02\%$, respectively (Table 2).

MTT assays

Table 3 shows the ability of different concentrations of SP-GNPs to limit the growth of PC-12 cells with 6-OHDA-induced disease. When compared with untreated PC-12 cells with 6-OHDA-induced disease, those treated with blank GNPs showed slightly higher but not significantly different cell viability, whereas their counterparts treated with SP-GNPs did demonstrate significantly higher cell viability

Table 1 Rat groups designed for 2 weeks of daily experimental treatment ($n=8$ per group)

Group	Rat model	Intranasal treatment	SP dosage ($\mu\text{g/day}$)
1	Sham	PBS	
2	PD	Blank GNPs	
3	PD	SP-GNPs	50
4	PD	SP-GNPs	75
5	PD	SP-GNPs	100

Abbreviations: PD, Parkinson's disease; PBS, phosphate-buffered saline; SP, substance P; GNPs, gelatin nanoparticles.

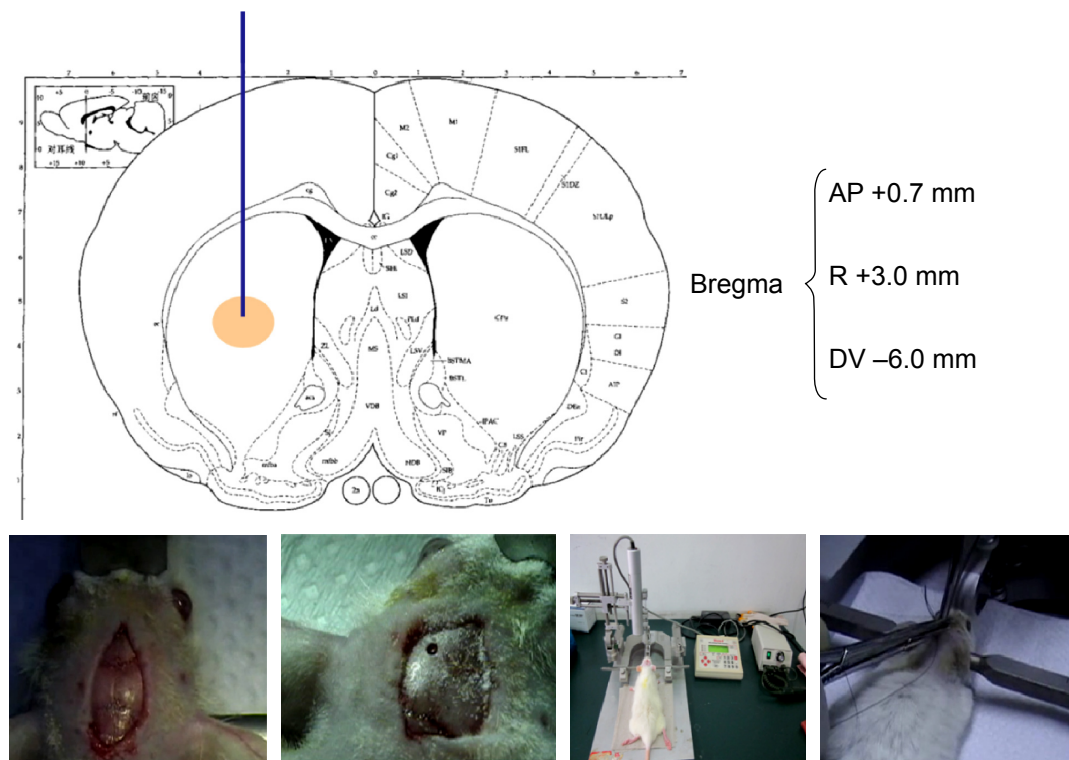


Figure 2 Rat model of hemiparkinsonism.

Note: The anesthetized SD rats were placed on stereotaxic apparatus and then injected with 12 μ L 6-OHDA solution (or vehicle for sham animals) in the right-side striatum.

Abbreviations: AP, distance after the fontanelle; R, movement to the right side; DV, depth value.

($P < 0.05$), indicating that SP-GNPs can decrease the extent of apoptosis caused by 6-OHDA and enhance cell growth. In the meantime, cell viability increased as the concentration of SP increased, suggesting that within a certain range of concentrations, the degree of inhibition of apoptosis achieved by SP is concentration-dependent.

Behavioral evaluation of PD rats after 2 weeks of treatment

The number of apomorphine-induced rotations following 2 weeks of daily treatment with SP-GNPs in each experimental group were consistent with the dopamine levels in the diseased brain. As seen in Table 4, the PD rats that received

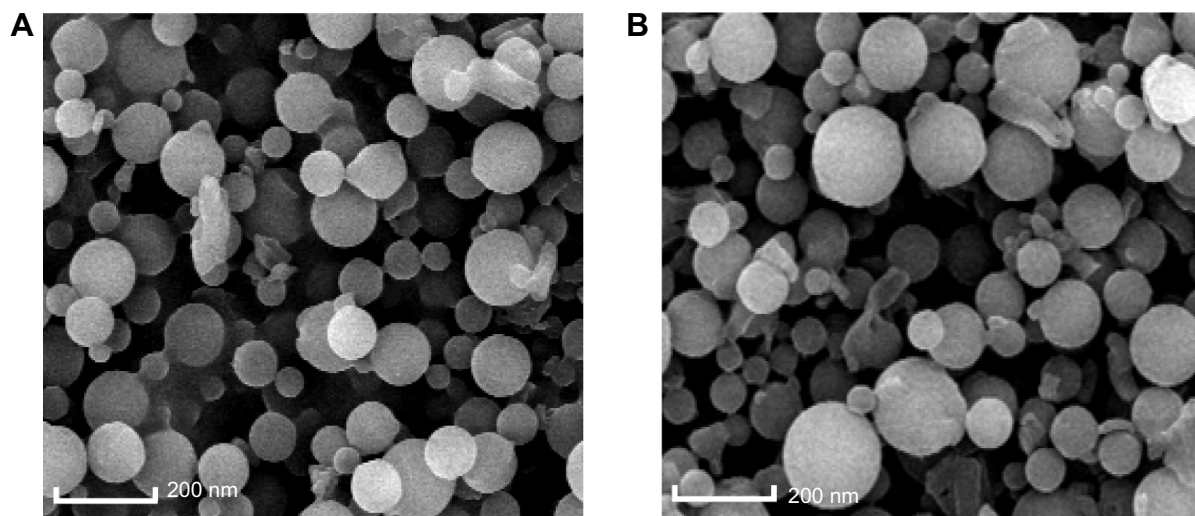


Figure 3 Scanning electron micrographs of blank GNPs (A) and SP-GNPs (B).

Abbreviations: SP, substance P; GNPs, gelatin nanoparticles.

Table 2 Characterization of gelatin nanoparticles and liposomes with or without substance P (n=3)

Formulation	Particle size (nm)	PDI	Zeta potential (mV)	EE (%)	Loading capacity (%)
Blank GNPs	166±1.32	0.145±0.021	-37.6±1.4	–	–
SP-GNPs	172±1.52	0.107±0.013	-29.6±1.2	93.3±1.4	5.2±0.02

Abbreviations: SP, substance P; GNPs, gelatin nanoparticles; EE, encapsulation efficiency; PDI, polydispersity index.

intranasal SP-GNP treatment made fewer rotations than those that received blank GNPs, demonstrating that SP could protect dopaminergic neurons against 6-OHDA-induced apoptosis and aid recovery of diseased dopaminergic neurons in rats with PD. Meanwhile, the PD rats that received daily intranasal treatment with 75 µg or 100 µg of SP showed a significant decrease in the number of apomorphine-induced rotations when compared with hemiparkinsonian rats ($P<0.05$), while PD rats that received 50 µg of SP per day intranasally showed a trend towards improvement ($P>0.05$), indicating that the extent of the protective effect of SP against 6-OHDA-induced apoptosis has a positive relationship with the concentration of SP.

Immunohistochemical staining of caspase-3 in the substantia nigra

As seen in Figure 4, immunohistochemical staining for caspase-3 in the diseased substantia nigra was limited in the sham group but extensive in the PD group, indicating that caspase-3 is rarely expressed in normal circumstances but is expressed in large amounts in the presence of PD. Less staining was seen in the SP-GNP groups than in the blank GNP group, suggesting that SP-GNPs can inhibit the expression of caspase-3 and reduce neuronal apoptosis, thus helping the diseased neurons to recover. Further, PD rats receiving 75 µg or 100 µg of intranasal SP per day showed significantly less caspase-3 staining than PD rats that did not receive SP ($P<0.05$), while PD rats receiving 50 µg of intranasal SP per day showed slightly lower level of caspase-3 staining than PD rats ($P>0.05$), indicating that the higher the concentration

of SP, the better the effect in protecting against 6-OHDA-induced neuronal apoptosis.

Discussion

6-OHDA is a neurotoxin that activates the apoptotic cascade in the central nervous system, leading to apoptosis and degeneration of dopaminergic neurons, which culminates in cell apoptosis in vitro and PD in vivo. SP, a member of the tachykinin peptide family, has been shown to play an important role in protecting against neurotoxin-induced apoptosis.

Studies show that drugs or particles smaller than 300 nm can bypass the blood–brain barrier, can be absorbed through the mucous membrane in the nasal olfactory region, and can be delivered into the brain directly through the cribriform plate, beyond which they exert their therapeutic effects in specific regions inside the brain.^{25–27} In earlier studies, nanoparticles were used as intranasal carriers for therapeutics to enable effective treatment of brain disorders, such as cerebral ischemia²⁸ and PD.²⁹ It is reported that nanoparticles administered intranasally can penetrate the brain through several pathways: the olfactory pathway, in which particles are taken up by the olfactory epithelium and the olfactory bulb; the trigeminal pathway, in which particles are delivered along the trigeminal nerve system; the vascular pathway, in which particles are absorbed into the capillaries underlying the nasal mucosa; and other pathways, such as cerebrospinal fluid and the lymphatic system.^{30–32} However, because of the mucociliary clearance mechanism in the nose, particles cannot be lodged in the nasal cavity for a long period, which

Table 3 Results of MTT assay on PC-12 cells

Group	Cell viability
Sham	1.0000±0.5397
6-OHDA disease	0.3562±0.3173
Blank GNPs	0.4036±0.3087
1 nM SP-GNPs	0.4817±0.2431*
10 nM SP-GNPs	0.5239±0.2345*
100 nM SP-GNPs	0.5966±0.2551**

Notes: * $P<0.05$, ** $P<0.01$ versus group with 6-OHDA disease.

Abbreviations: SP, substance P; GNPs, gelatin nanoparticles; 6-OHDA, 6-hydroxydopamine; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; PC-12, adrenal pheochromocytoma.

Table 4 Apomorphine-induced rotations after 2 weeks of daily treatment with substance P (n=8 per group)

Group	Rat model	Intranasal treatment	SP dosage (µg/day)	Rotations per minute
1	Sham	PBS		0
2	PD	Blank GNPs		7.6±0.5
3	PD	SP-GNPs	50	6.8±1.1
4	PD	SP-GNPs	75	6.5±1.3*
5	PD	SP-GNPs	100	6.3±1.7*

Note: * $P<0.05$ versus group 2.

Abbreviations: PD, Parkinson's disease; PBS, phosphate-buffered saline; SP, substance P; GNPs, gelatin nanoparticles.

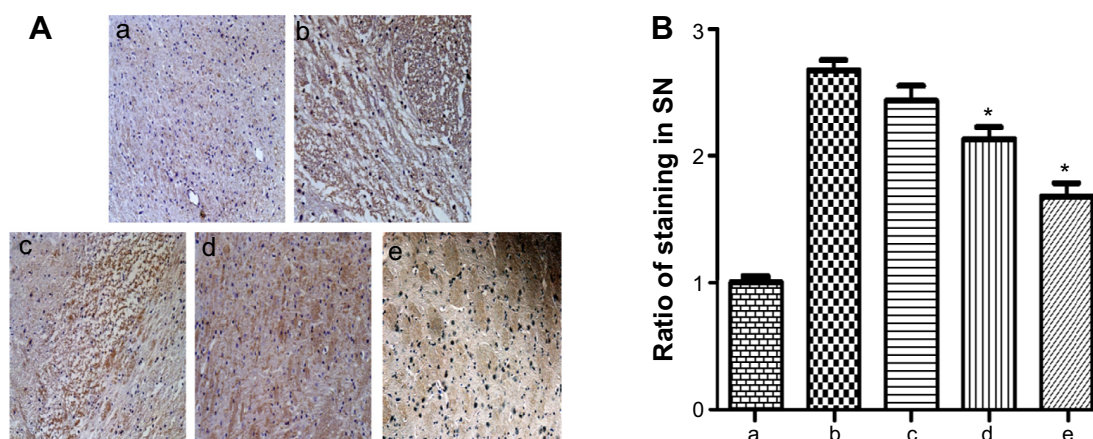


Figure 4 Immunohistochemical staining of caspase-3 in the SN region.

Notes: (A) Results of caspase-3 staining in SN region. (B) Quantification of caspase-3 staining in SN region. (a) Sham group. (b) PD rats intranasally administered blank GNP. (c) PD rats intranasally administered SP-GNP 50 µg/day. (d) PD rats intranasally administered SP-GNP 75 µg/day. (e) PD rats intranasally administered SP-GNP 100 µg/day. **Abbreviations:** PD, Parkinson's disease; SN, substantia nigra; SP, substance P; GNP, gelatin nanoparticle.

limits the application of intranasal administered drug-loaded particles.^{27,33}

In recent years, gelatin and nonionic surfactants (such as Poloxamer 188) have been used to prepare nanoparticles due to their biocompatibility, biodegradability, low immunogenicity, and amenability for surface modification.^{34–36} Nanoparticles modified with gelatin have a negative charge that can reduce mucociliary clearance, extend the residence time at the site of delivery, and enhance the therapeutic effect when administered intranasally.^{17–19} In a previous study, we found that gelatin nanostructured lipid carrier-mediated intranasal delivery of basic fibroblast growth factor could enhance functional recovery in hemiparkinsonian rats.³⁷

PC-12 cells, a monoamine cell line derived from a pheochromocytoma in the adrenal medulla of a male rat, can express tyrosine hydroxylase and synthesize dopamine intracellularly, so are widely used in the study of PD models in vitro.³⁸ In our in vitro experiment, we used 6-OHDA to trigger apoptosis and then added SP-GNPs at different concentrations to investigate the effect of SP-GNPs on growth of PC-12 cells. It is evident from the results of these investigations that SP can decrease apoptosis and enhance cell growth to a considerable degree. Further, within a certain range of concentrations, the degree of inhibition of cell apoptosis increases as the concentration of SP increases, with cells growing better and in larger numbers at higher SP concentrations.

In our in vivo experiment, SP-GNPs were administered intranasally to rats with 6-OHDA-induced hemiparkinsonism, and these rats showed more functional improvement and less apoptosis than their counterparts that were not treated with intranasal SP-GNPs. Intranasal administration of SP-GNPs inhibited 6-OHDA-induced apoptosis and

improved symptoms of hemiparkinsonism. With increasing concentrations of SP, rats with hemiparkinsonism showed more functional improvement, with further decreases in levels of apoptosis, indicating that the strength of the neuroprotective effect had a positive relationship with the SP concentration in the brain.

As a noninvasive strategy, GNP-mediated intranasal delivery of SP protects against 6-OHDA-induced apoptosis, and might constitute a practical therapy for PD patients in the future.

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Disclosure

The authors report no conflicts of interest in this work.

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