

Enhanced detection sensitivity of *Escherichia coli* O157:H7 using surface-modified gold nanorods

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Abstract: *Escherichia coli* O157:H7 (O157) is a Gram negative and highly virulent bacteria found in food and water sources, and is a leading cause of chronic diseases worldwide. Diagnosis and prevention from the infection require simple and rapid analysis methods for the detection of pathogens, including O157. Endogenous membrane peroxidase, an enzyme present on the surface of O157, was used for the colorimetric detection of bacteria by catalytic oxidation of the peroxidase substrate. In this study, we have analyzed the impact of the synthesized bare gold nanorods (AuNRs) and silica-coated AuNRs on the growth of *E. coli* O157. Along with the membrane peroxidase activity of O157, other bacteria strains were analyzed. Different concentrations of nanorods were used to analyze the growth responses, enzymatic changes, and morphological alterations of bacteria by measuring optical density, 3,3',5,5'-tetramethylbenzidine assay, flow cytometry analysis, and microscopy studies. The results revealed that O157 showed higher and continuous membrane peroxidase activity than other bacteria. Furthermore, O157 treated with bare AuNRs showed a decreased growth rate in comparison with the bacteria with surface modified AuNRs. Interestingly, silica-coated AuNRs favored the growth of bacteria and also increased membrane peroxidase activity. This result can be particularly important for the enzymatic analysis of surface treated AuNRs in various microbiological applicants.

Keywords: *E. coli*, O157:H7, endogenous membrane peroxidase, gold nanorod, silica-coated gold nanorod

Introduction

Consumption of microbial-contaminated food or water is still one of the main causes of death in this 21st century.¹ According to the report of the World Health Organization, approximately two million people died in 2005 because of consumption of microbial-contaminated food or water. The common pathogenic strain that released potent toxins and was responsible for complex food-borne diseases was *Escherichia coli* O157:H7 (O157). *E. coli* O157 is a serotype, Gram-negative, highly virulent anthroponozoonotic pathogen found in contaminated water, and foods like uncooked beef, unpasteurized milk, juice, and salami. This pathogen spreads from animals and could be fatal to humans.^{2,3} O157 strain causes life-threatening diseases such as hemolytic uremic syndrome (HUS), hemorrhagic colitis, renal failure, central nervous system failure, and even death.⁴ For treatment, antibiotics are given with caution since they produce kidney complications leading to HUS.⁵ Antidiarrheal agents reduce the bowel activity that leads to delayed antibacterial action on O157 which allow them to proliferate highly in the digestive tract. Antispasmodic drugs with analgesics are commonly prescribed for suppressing COX-2-dependent prostaglandins synthesis.⁶

O157 bacteria carry proteins, receptors, and enzymes on their surfaces, especially heme-binding enzyme catalase-peroxidases or membrane peroxidases, which are believed to be involved in the bacterial defense mechanisms against oxidative stresses.⁷

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Eukaryotic enzymes like endogenous membrane peroxidase have potential applications and are used as markers for monitoring the enzymatic activity in bacterial cells. Conventional methods used to detect bacteria include colony counting, immunology-based enzyme-linked immunosorbent assays, and polymerase chain reaction, which are time consuming and require special sample pretreatment.^{8–10} Biosensor-based detection techniques are sensitive, specific, and reliable, however, they require special chemical and fluidic attachments such as antibodies.¹¹

Emerging trends in nanotechnology open up materials to be applied in various aspects of biomedical applications, including antimicrobes,¹² cancer imaging and therapy,^{13,14} drug or DNA delivery,^{15,16} enzyme sensors,¹⁷ UV protector,¹⁸ and so forth. Au@Pt nanomaterials with hydrogen peroxide (H_2O_2) have been reported as peroxidase mimetic in immunoassays and glucose sensing.^{19,20} Several studies have reported the intrinsic peroxidase mimicking activity of nanostructured materials that can be used in traditional enzyme-linked immunosorbent assay method after replacing horseradish peroxidase with CeO_2 , Pt, carbon, and magnetic nanoparticles.^{21–24} Gold nanostructures have recently been useful in bio-applications because of their nontoxicity, ease of surface modification, polyvalent effects, and photothermal capacity.^{25–27} Zhao et al²⁸ and Ma et al²⁹ developed organic molecule-attached gold nanoparticles for bacteria lyses through cell membrane rupture, attachment with nucleic acids, and inhibition of protein synthesis. There are no existing reports that explain the interaction of metal nanostructures with *E. coli* O157 and its enzymatic activity without H_2O_2 .

Herein, we have analyzed the membrane peroxidase activity of *E. coli* O157, *Enterococcus faecalis*, *Bacillus subtilis*, and *Salmonella typhi*. And their continuous enzyme activity was compared. Then, the growth of *E. coli* O157 was assessed against the prepared bare gold nanorods (AuNRs) and silica-coated AuNRs (Si-AuNRs). Further, alterations in enzymatic activity of *E. coli* O157 in the presence of synthesized nanorods were studied. Finally, the morphological changes of bacteria before and after treatment with different nanorods were examined.

Materials and methods

Materials

Ready-made phosphate-buffered saline (PBS) solution, Lysogeny Broth (LB) broth (BD Biosciences, San Jose, CA, USA), and 3,3',5,5'-tetramethylbenzidine (TMB) were purchased from Sigma-Aldrich (St Louis, MO, USA). Cetyltrimethylammonium bromide (CTAB), 3-mercaptopropyl

trimethoxysilane (MPTMS), 3-aminopropyltrimethoxysilane (APDES), ammonium hydroxide (NH_4OH ; 28 wt% in water), silver nitrate ($AgNO_3$), sodium borohydride ($NaBH_4$), ascorbic acid, ethanol, hydrogen tetrachloroaurate(III) trihydrate ($HAuCl_4 \cdot 3H_2O$), methanol, and sulfuric acid (H_2SO_4) were purchased from Sigma-Aldrich.

Preparation of AuNRs

AuNRs were prepared by modified seed-mediated method described elsewhere.³⁰ In brief, 250 μ L of 0.01 M $HAuCl_4$ was mixed with 7.5 mL of 0.1 M CTAB and 600 μ L of ice-cold 0.01 M $NaBH_4$ under vigorous stirring at room temperature to make seed solution. Next, the growth solution was prepared by adding 400 μ L of 0.01 M $HAuCl_4$, 64 μ L of 0.1 M ascorbic acid, and 35.6 μ L of 0.1 M $AgNO_3$ in 9.5 mL of 0.1 M CTAB. Finally, 10 μ L of 2.5-hour-old seed solution was added to the growth solution, and the mixture was left for 24 hours at ambient temperature.

Preparation of surface modified AuNRs

Si-AuNR was synthesized by adding a mixture of 10 mM APDES and MPTMS to the prepared AuNR, stirred continuously at 800 rpm for 12 hours. Next, ammonium hydroxide at pH 9 was added and stirred vigorously for 1 hour. The thickness of the silica layer around Au was controlled by the amount of APDES added. The mixture was centrifuged, and the particles were washed several times with ethanol.

Particle size distributions before and after surface modification were calculated using field emission scanning electron microscopy (FE-SEM; JEOL-JSM-7500F, JEOL, Tokyo, Japan) images. Likewise, Zeta-potential measurements of the AuNRs and Si-AuNRs were taken using a Malvern ZetaSizer 3000HS instrument (Malvern Instruments, Malvern, UK).

Bacterial growth kinetics

The bacteria strains were obtained as a gift from Bong Hyun Chung's laboratory. Bacteria were cultured aerobically in LB broth and grown at 37°C for 4–6 hours with continuous agitation at 200 rpm. The organisms were harvested by centrifugation at 4,000 rpm (4°C) for 5 minutes, washed twice with PBS and then re-suspended in double-distilled water. The cell growth was determined by measuring optical density (OD) using spectrophotometer at every 1 hour interval with three replications.

Endogenous membrane peroxidase activity

The enzyme activities in the bacteria were assessed using TMB assay.³¹ Approximately 80 μ L of the peroxidase substrate

was incorporated in 20 μL of bacteria ($10^7/\text{mL}$) per well of a clear and black microtiter plates. The bacteria were incubated at 37°C for 30 minutes with continuous shaking at 80 rpm. After the incubation period, 50 μL of the 2N H_2SO_4 was added to stop further reaction and the produced blue color was read at OD of 450 nm using PerkinElmer Victor 3 spectrophotometer (PerkinElmer Inc., Waltham, MA, USA). Peroxidase activity was also measured for other bacteria such as *B. subtilis*, *S. typhi*, and *E. faecalis* for comparing with *E. coli* O157.

In each bacteria sample, continuous membrane peroxidase activity toward TMB substrate was further analyzed after repeated washing. The samples were washed five times and the OD was measured at 450 nm.

Impact of nanoparticle on *E. coli* O157 growth

After reaching required confluence, bacterial cells were incubated at 37°C in LB broth containing different concentrations (100–500 $\mu\text{g}/\text{mL}$) of bare AuNRs and Si-AuNRs. After incubation, samples from each nanorod-treated bacterial strain were collected at every hour to monitor the growth of *E. coli* O157 by measuring OD at 600 nm up to 8 hours. The test was performed in triplicate.

Flow cytometry analysis of nanoparticle-treated *E. coli* O157

For flow cytometry analysis, *E. coli* O157 was grown at a density of 10^7 cells/mL. The cells were then incubated with bare AuNRs and Si-AuNRs. The cells were harvested, washed with ice-cold PBS by centrifugation and fixed with 70% cold ethanol. Resulting cell suspension was then treated with the equal parts mixture of propidium iodide (PI) and SYTO9. The cell-lysis pattern was analyzed, and the number of live cells after nanorods treatment was evaluated using flow cytometry (BD FACSCalibur; BD Biosciences). Cell membrane of live cells is impermeable to PI because of its intact nature whereas PI leaks into dead cells with compromised membranes and produces red fluorescence. SYTO9 binds to nucleic acid and produce green fluorescence. The unique fluorescence pattern directly relates to the degree of bacterial mortality.

Enzymatic activity of *E. coli* O157 in the presence of nanoparticles

TMB substrate (80 μL) was added in the 96-well plate with 20 μL of *E. coli* O157 ($10^7/\text{mL}$) per well containing AuNRs and Si-AuNRs. The treated bacterial cells were incubated

at 37°C for 30 minutes with continuous shaking at 80 rpm. Bacteria without nanorods were kept as control. Then, 50 μL of the stop solution was added to cease further reaction. OD of the nanorod-treated samples were measured at 450 nm.

Morphology analysis of *E. coli* O157 by electron microscopy

Morphology of nanorod-treated bacteria after TMB assay was analyzed and compared against control cells using FE-SEM. AuNRs and Si-AuNRs-treated bacteria samples were placed on small pieces of cover slips. Different nanorods-treated *E. coli* O157 were fixed using 4% glutaraldehyde buffered with 0.2 M sodium phosphate (pH 7). The samples were dried to critical-point with CO_2 and sputter coated. Then, they were analyzed under the FE-SEM.

Results

Fabrication and characterization of AuNRs

To investigate the response of nanorods on *E. coli* O157, we fabricated AuNRs and Si-AuNRs. The short AuNRs with $\text{AR} \approx 2.5$ were prepared by a seed-mediated growth method in a single-component surfactant (CTAB) solution. The CTAB stabilized AuNRs as shown in Figure 1A were modified with a silica layer using a sol-gel hydration and condensation method. After coating, AuNRs were purified in ethanol and finally dispersed in water without forming aggregates. When the mixture of MPTMS and APDES was used, all the AuNRs were coated by silica layer without forming Au-free silica particles (Figure 1B). In the 12-hour mediation process, the amine and mercapto functional groups were attached onto the Au surfaces.³² FE-SEM analysis clearly depicts the existence of amorphous silica layer around AuNRs to form Si-AuNRs. The average particle size distributions of AuNR were measured by averaging 100 particles of each of three batches from the FE-SEM images using ImageJ software (developed by the National Institutes of Health, Bethesda, MD, USA). The size distribution of bare AuNR was calculated to be 49.8×19.5 nm whereas the Si-AuNR was 71.3×32.1 nm due to the thick silica shell around the gold core. Also, the bare AuNRs showed a zeta potential value of +52 mV. After surface modification, the charge reversal of -33.7 mV was obtained for the Si-AuNRs. These high zeta potential values of both bare AuNRs and Si-AuNRs supported the good colloidal stability. Generally, the particles in suspension having a large negative/positive zeta potential will tend to repel each other, and there will be no possibility aggregation thus they are considered more

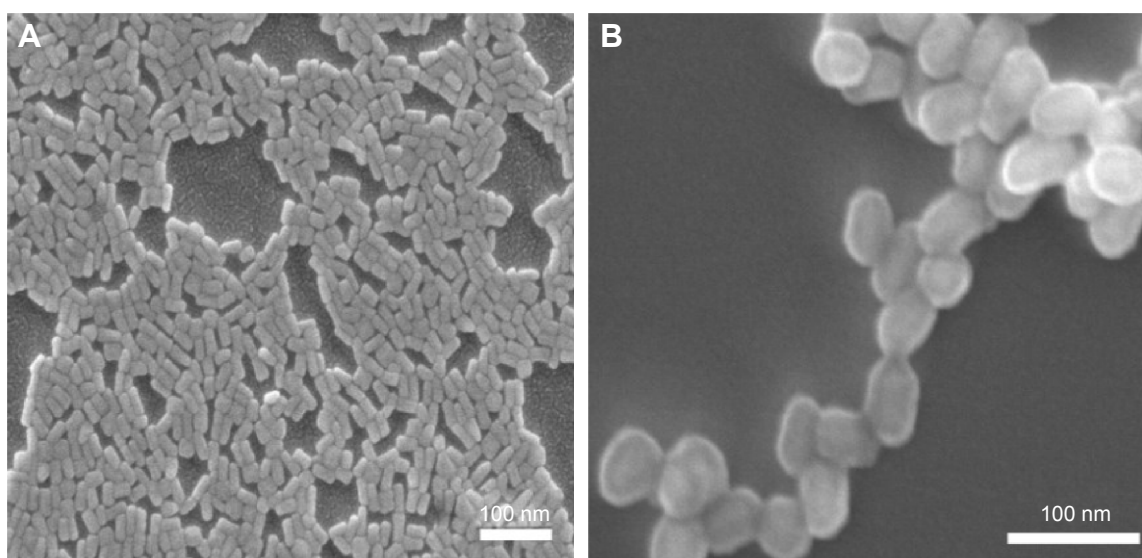


Figure 1 FE-SEM characterization of nanorods synthesized by modified seed-mediated method.

Notes: (A) Bare AuNRs and (B) surface-modified Si-AuNRs.

Abbreviations: FE-SEM, field emission scanning electron microscopy; AuNRs, gold nanorods; Si-AuNRs, silica-coated gold nanorods.

stable in nature.^{33,34} Hence, the zeta potential values confirm the formation of silica layer with better stability, which was free from aggregation.

Growth kinetics of bacteria

Comparison among the regular growth rate of different bacterial cells is shown in Figure 2. The obtained results showed that at 10 hours, *B. subtilis* showed higher growth, and *E. coli* O157 showed slower growth compared to *E. faecalis* and *S. typhi*, which showed normal growth rate at the same incubation conditions. Since all bacteria showed similar

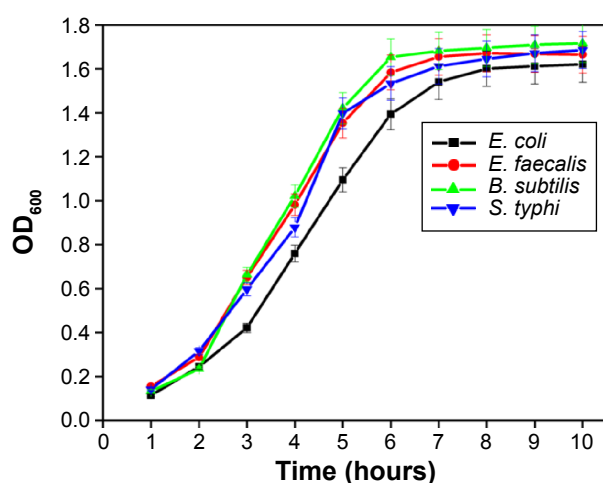


Figure 2 Bacterial dynamic growth curve of *E. coli* O157, *E. faecalis*, *B. subtilis*, and *S. typhi* in LB medium.

Abbreviations: OD, optical density; LB, Lysogeny Broth; *E. coli* O157, *Escherichia coli* O157; *E. faecalis*, *Enterococcus faecalis*; *B. subtilis*, *Bacillus subtilis*; *S. typhi*, *Salmonella typhimurium*.

growth phase, these bacteria were used to analyze EMP activities.

Membrane peroxidase activity

The membrane peroxidase activity of various bacteria was evaluated with TMB substrate because it is sensitive at low hydrogen peroxide concentrations and cost effective compared to other substrates. TMB develops insoluble navy blue color (Figure 3A) after reacting with 2N H₂SO₄, which is the characteristic identification for peroxidase-labeled molecules. Hence, the results shown in Figure 3B confirmed the presence of peroxidase molecules on the surface of bacteria. When the number of bacteria increases, the enzymatic activity proportionally increased by giving enhanced signals in the assay. Compared to other three microorganisms, *E. coli* O157 showed higher enzyme activity in TMB assay. Relatively, *E. faecalis* showed significant peroxidase activity followed by *B. subtilis* which showed very less enzymatic activity and no enzymatic activity was observed in *S. typhi*.

After TMB assay for the first instance, the same bacteria samples were washed with water to remove the traces of the stop solution, and they were then analyzed again for continuous membrane peroxidase activity. Interestingly, we found that only *E. coli* O157 showed continuous enzymatic activity even after being washed more than five times with water compared with other bacteria (Figure 4). The inset picture depicts the visible color changes of the continuous enzymatic activity, where *E. coli* O157 produced yellow color (before adding stop solution) after repeated washings compared with

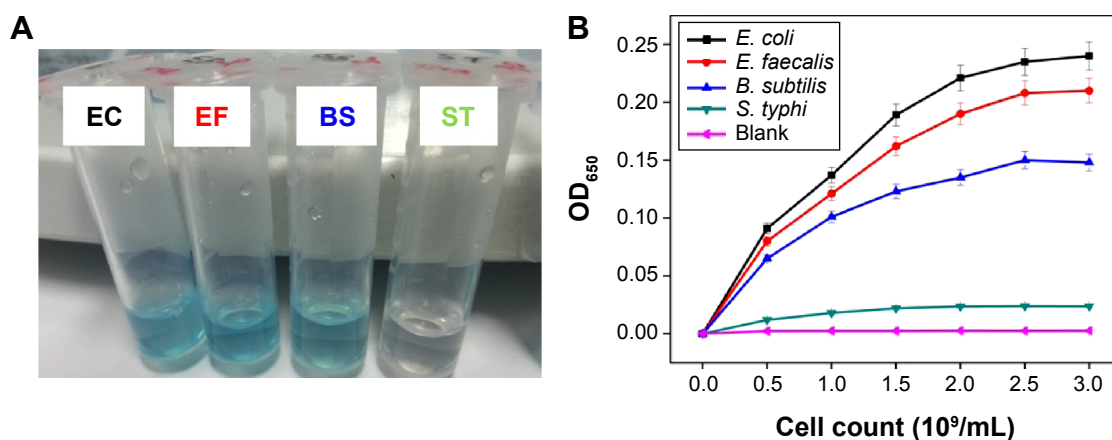


Figure 3 Endogenous membrane peroxidase activity measurement using TMB assay.

Notes: (A) Color development of bacteria samples (EC – *E. coli* O157, EF – *E. faecalis*, BS – *B. subtilis*, and ST – *S. typhi*, respectively) in the presence of TMB substrate. (B) Optical density measurement of enzymatic active *E. coli* O157, *E. faecalis*, *B. subtilis*, and *S. typhi* samples.

Abbreviations: OD, optical density; TMB, 3,3',5,5'-tetramethylbenzidine; *E. coli* O157, *Escherichia coli* O157; *E. faecalis*, *Enterococcus faecalis*; *B. subtilis*, *Bacillus subtilis*; *S. typhi*, *Salmonella typhimurium*.

other bacterial pathogens, which were colorless after washings. The possible reason can be that, *E. coli* O157 could internalize the TMB substrate thus producing repeated color. But, the exact reason for these enzymatic activity differences in bacteria are yet to be identified.

Effect of the nanorods on bacterial growth

E. coli O157 growth was tested to evaluate the impact of bare and surface-modified AuNRs on it, as this strain showed the

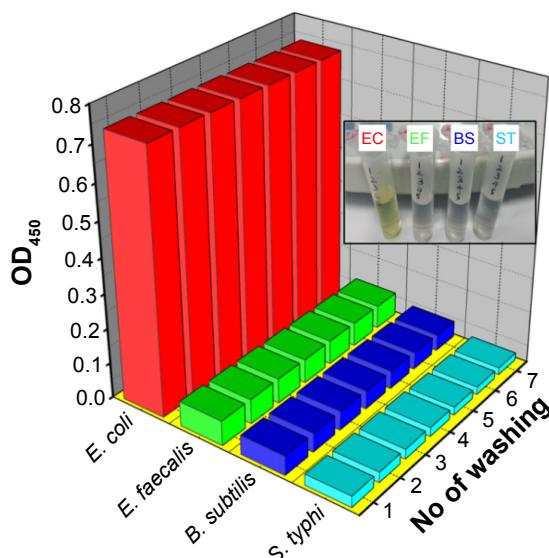


Figure 4 Repeated membrane peroxidase analysis of bacteria samples.

Notes: OD₄₅₀ values of the bacteria samples after repeated washings without the addition of stop solution. Inset: Photographs of the corresponding bacterial suspensions (EC – *E. coli* O157, EF – *E. faecalis*, BS – *B. subtilis*, and ST – *S. typhi*, respectively).

Abbreviations: OD, optical density; *E. coli* O157, *Escherichia coli* O157; *E. faecalis*, *Enterococcus faecalis*; *B. subtilis*, *Bacillus subtilis*; *S. typhi*, *Salmonella typhimurium*.

highest enzyme activity. The comparative growth condition results between AuNRs- and Si-AuNRs-treated bacteria cells at different concentrations are shown in Figure 5. The growth rate of bacteria was calculated by using its exponential phase as described by Maurer-Jones et al.³⁵ Obtained results (Figure 5A) clearly indicate that the AuNRs exposed cells showed more toxicity with no further cell growth at all concentrations whereas, Si-AuNRs-treated bacteria showed enhanced growth rate compared to control experiment, which resembles the nontoxic nature of the nanoparticles (Si-AuNRs). In detail, even at higher concentrations, viability was not altered for Si-AuNR-treated bacteria, but bare AuNR caused a dose-dependent decrease in the growth of O157. The growth or cell lysis was calculated by plotting the absorbance (OD₆₀₀) over time in the exponential growth phase of bacteria. It was noted that initial high absorbance values occurred from the scattering of nanoparticles in the sample (Figure 5A). Based on the followed calculations from Maurer-Jones et al.³⁵ it was apparent that the rate of growth of Si-AuNRs exposed bacteria was higher for lower concentrations, than that of control (Figure 5B). In contrast, significant decrease in growth rate was observed for all five concentrations of AuNRs-treated bacteria against control (Figure 5C).

Live/dead analysis of nanoparticle treated *E. coli* O157

Flow cytometric analysis (Figure 6) was adopted to analyze the difference in bacterial viability behavior in the presence of bare AuNRs and Si-AuNRs. Here, membrane-permeating PI-mediated red fluorescence subject to dead cell staining and green fluorescence by SYTO 9 subject to a total live-cell

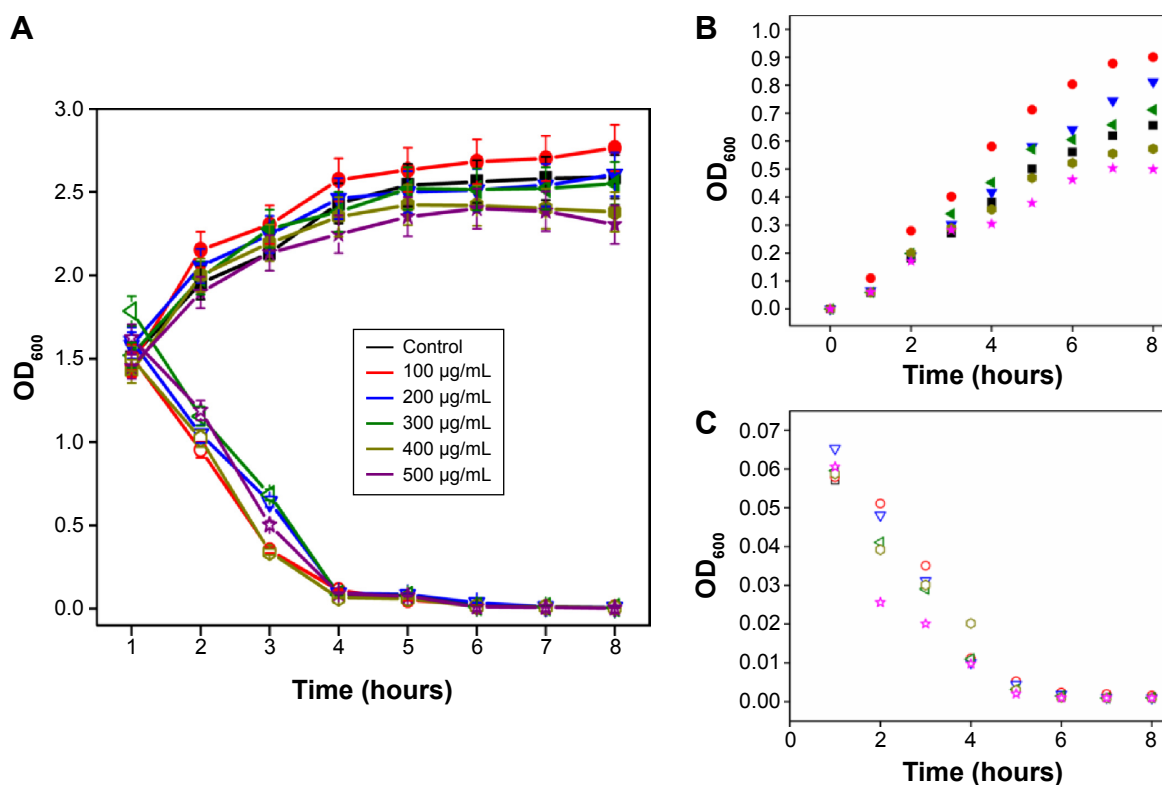


Figure 5 Growth rate of *E. coli* O157 in the presence of uncoated and surface coated AuNRs.

Notes: (A) Exponential growth phase and lysis rate of bacteria in the presence of Si-AuNRs and AuNRs, respectively. (B) Bacteria growth curve values subtracted with OD at time 0 hour considered as scattering values of used Si-AuNRs. (C) Lysis rate of bacteria samples in the presence of AuNRs. Open symbols ($\square$, $\circ$, ∇ , $\triangleleft$, $\star$, $\odot$) represent control and 100, 200, 300, 400 and 500 $\mu\text{g/mL}$ concentrations of bare AuNRs; and closed symbols ($\blacksquare$, $\bullet$, $\blacktriangledown$, $\blacktriangleleft$, $\blackstar$, $\bullet$) represent control and 100–500 $\mu\text{g/mL}$ concentrations of Si-AuNRs, respectively.

Abbreviations: OD, optical density; *E. coli* O157, *Escherichia coli* O157; AuNRs, gold nanorods; Si-AuNRs, silica-coated gold nanorods.

staining, resulted in a distinctive fluorescence pattern for the bacteria exposed to different AuNRs. The corresponding signals were denoted as PI fluorescence in FL1 channel and SYTO 9 fluorescence in FL3 channel, respectively. Figure 6A shows the flow cytometric patterns of *E. coli* O157, which was exposed to different doses of AuNRs and Si-AuNRs. Initially, the untreated control cells showed unique one population (region 1) with none or less dead cells (region 2). However, after AuNR treatment, bacteria exhibited increased fluorescence intensity in FL1 channel showing the cell lysis. The data demonstrate that when the bare AuNRs concentration was increased to 200 $\mu\text{g/mL}$ or higher, there was an increased damage in bacteria due to injured cells followed by total cell death, which was recorded with higher fluorescence density in region 2. Thus, it exhibits the growth inhibition for the bare AuNRs-treated bacteria cells. In contrast, Si-AuNRs-treated bacteria up to 400 $\mu\text{g/mL}$ concentration showed only single population indicating low mortality which was similar to the control that caused non-separated unique population at FL3 channel (SYTO 9 fluorescence) in region 1. At the highest concentration of Si-AuNRs (500 $\mu\text{g/mL}$), there was a minimal cell lysis observed with more injured cells in region 1 which

was comparatively the lower cell lysis rate than bare AuNRs. Hence, with increasing dose levels, there was no specific cell lysis observed for Si-AuNRs even at higher concentrations. This indicates that the exposed Si-AuNRs were nontoxic to the bacteria. Figure 6B depicts the corresponding statistical analysis values of the flow cytometry results. Increasing the AuNRs, concentration from 100 $\mu\text{g/mL}$ to 500 $\mu\text{g/mL}$ resulted in a markedly higher lysis rate from approximately 98% to 2%, respectively. On the other hand, increasing the concentration of Si-AuNRs did not affect the bacteria viability. More specifically, the lethal rate of bacteria at 500 $\mu\text{g/mL}$ was only 20% with approximately 80% live bacteria. Altogether, the bare AuNRs-exposed bacteria resulted in higher cell death whereas Si-AuNRs-treated bacteria showed a higher number of live cells even at the highest concentration of Si-AuNRs. This reveals that surface-coated AuNRs can enhance the cell growth, which could be useful to detect the bacteria without lysis.

Impact of the nanorods on the membrane peroxidase activity of O157

E. coli O157 in the presence of bare AuNRs and Si-AuNRs was tested in a dose- and time-dependent manner to analyze the

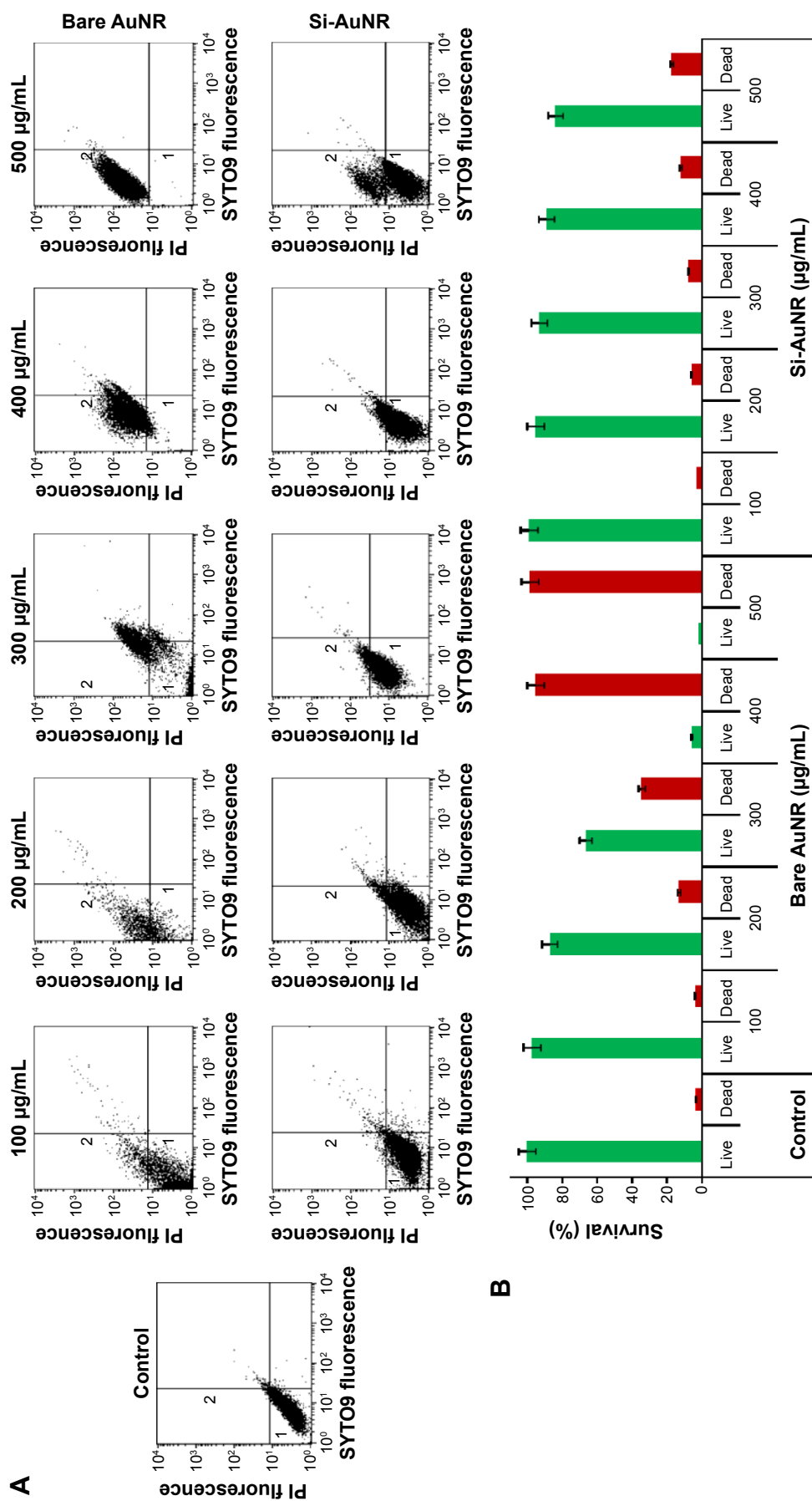


Figure 6 Flow cytometry analysis of AuNRs-treated bacteria. **Notes:** (A) Fluorescence density plots of *E. coli* O157 in response to staining with SYTO9 and PI after being treated at different concentrations of AuNRs and Si-AuNRs. Two gate regions define the position of live (region 1) and dead (region 2) bacterial populations, respectively. (B) Corresponding quantitative survival rate of bacterial cells according to the different concentration of nanorods. **Abbreviations:** *E. coli* O157, *Escherichia coli* O157; AuNRs, gold nanorods; Si-AuNRs, silica-coated gold nanorods; PI, propidium iodide.

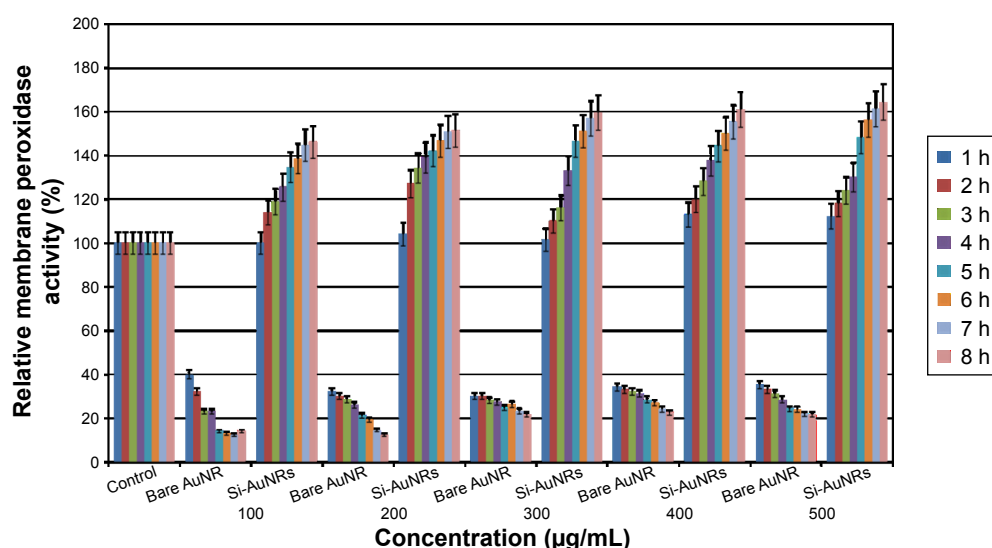


Figure 7 Specific membrane peroxidase activity of *E. coli* O157 in the presence of AuNRs and Si-AuNRs compared to control cells.
Abbreviations: *E. coli* O157, *Escherichia coli* O157; AuNRs, gold nanorods; Si-AuNRs, silica-coated gold nanorods; h, hours.

membrane peroxidase activity. Figure 7 depicts the effect of nanoparticles and alterations in the enzymatic activity of bacteria. Bare AuNRs-treated bacteria showed reduced membrane peroxidase activity with increasing time as compared to the control cells. At the starting time, the enzymatic activity was higher but when the incubation time reached 8 hours, the peroxidase activity was reduced. An opposite trend in membrane peroxidase activity has been observed for Si-AuNRs-exposed bacteria. When the concentration of Si-AuNRs increased from 100 µg/mL to 500 µg/mL, the enzyme activity also increased comparatively against the control. Interestingly, the time dependency also enhanced the membrane peroxidase activity to twofold compared to the control, which could be due to the formation of new bacteria cells.

Morphology of nanorods-treated *E. coli* O157

The morphological changes of the bacteria in the presence of nanorods were analyzed using FE-SEM study (Figure 8). The TMB assayed, bare AuNRs-treated bacteria showed disintegrated cell structure in Figure 8Bi compared to intact, proximally populated control cells (Figure 8Ai and Aii). More precisely, the enlarged view clearly depicts the ruptured, irrecoverable, disordered population in bare AuNRs-treated bacteria cells (AuNRs as white dots; Figure 8Bii). On the other hand, Si-AuNRs-treated bacteria showed enhanced growth with the formation of new populations as shown in Figure 8Ci. Attachment of Si-AuNRs on the surface of bacteria with enhanced formation of new colony was observed (Figure 8Cii). These observations corroborate the significant

impact of the uncoated and coated nanorods on bacterial growth and enzymatic activity.

Discussion

This experiment was conducted for evaluating the impact of bare AuNRs and Si-AuNRs on growth and enzymatic activity of *E. coli* O157, a highly virulent bacterial strain. AuNRs were prepared by modified seed-mediated method and silica-coated method using the reaction mixture of MPTMS and APDES. Electron microscopy confirmed the formation of silica layer around each AuNR without aggregation. Both bare and surface-coated AuNRs were applied and their impacts on the bacterial growth and enzymatic activity were analyzed. Since O157 showed the highest enzymatic activity, it was selected to evaluate the different concentration (100–500 µg/mL) effects of bare AuNRs and Si-AuNRs. The result of cell growth revealed that bare AuNRs inhibited the growth of O157 compared to Si-AuNRs. In contrast, Si-AuNR had a positive relationship with the bacteria growth. At the starting point, the number of cells incubated was 1×10^7 /mL. After 8 hours of incubation, the number of cells in the control group increased by onefold in their log phase. Interestingly, the bacterial cells treated with surface modified-AuNRs showed increased growth rate with control. After 8 hours, cells treated with 100 µg/mL showed approximately threefold increase in the number of cells compared with that of the control group whereas, cells treated with the maximum 500 µg/mL showed a decrease in their growth at total 8 hours. This trend may be a function of nonspecific surface bonding or adsorption of broth components onto the

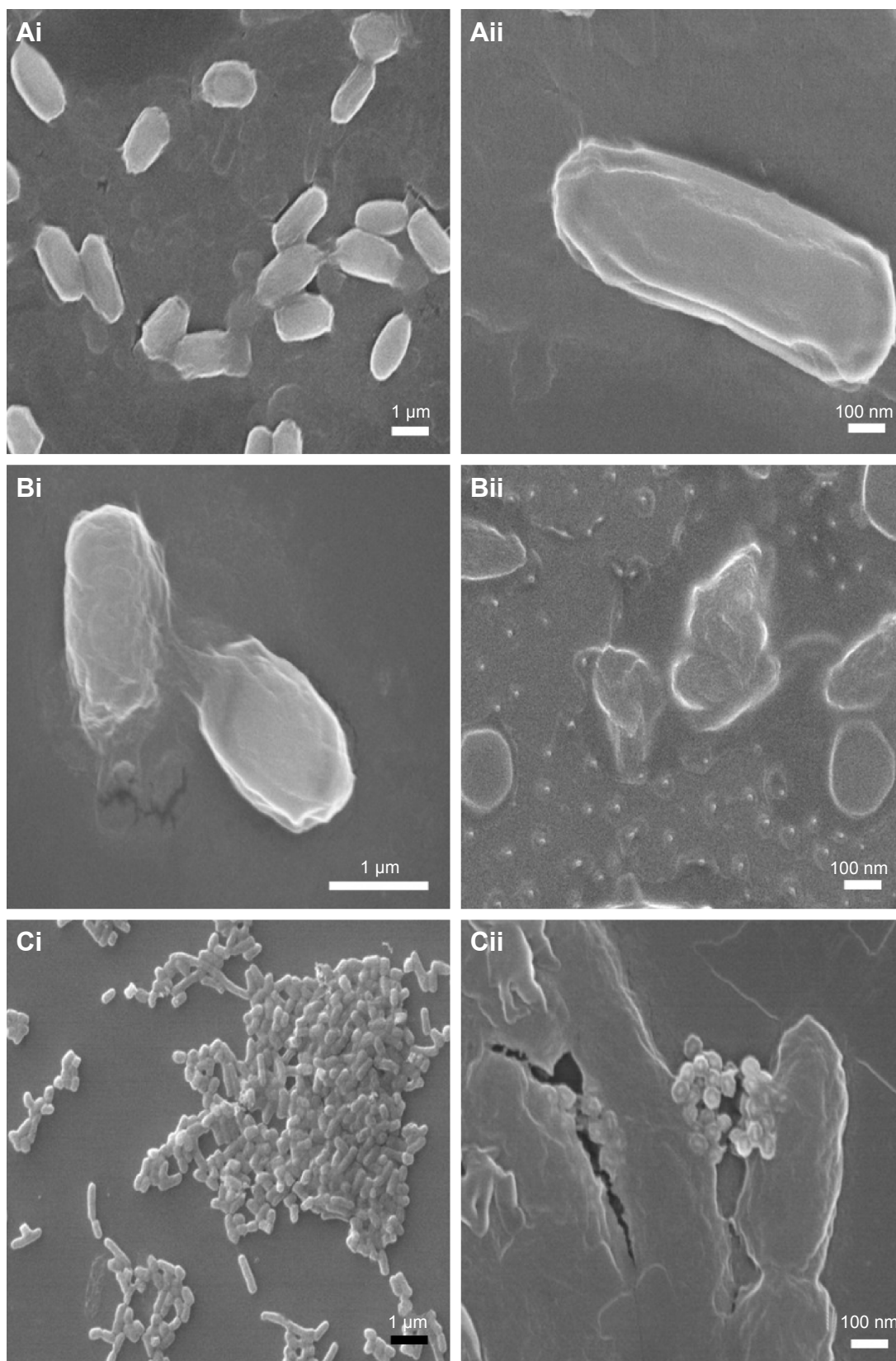


Figure 8 Morphological analyses of *E. coli* O157 after TMB assay using FE-SEM.

Notes: Panels (Ai and Aii) represent the untreated control bacteria and its magnified image. (Bi) AuNRs-treated bacteria, (Bii) disoriented cells after treatment. (Ci) Enhanced bacteria growth toward Si-AuNRs, (Cii) specific attachment of Si-AuNRs with *E. coli* cells by maintaining complete cell morphology.

Abbreviations: FE-SEM, field emission scanning electron microscopy; TMB, 3,3',5,5'-tetramethylbenzidine; *E. coli* O157, *Escherichia coli* O157; AuNRs, gold nanorods; Si-AuNRs, silica-coated gold nanorods.

surface of Si-AuNRs. As described previously, three major factors are responsible for dynamic nano–bio interfaces, 1) physicochemical composition of nanoparticles, 2) modifications in the interface of nanoparticle in the surrounding medium, and 3) contact area of the nanoparticle interface with biological components. In specific, AuNRs after surface modifications may form increased interaction with biological substrates.^{36,37} Likewise, from our findings Si-AuNRs resulted in higher cell viability even at higher concentrations than uncoated AuNRs.

The results were further established by flow cytometric analysis. The cells treated with Si-AuNRs observed with an increased SYTO9-mediated green fluorescence intensity in a dose-dependent manner describing healthy cells, whereas PI-mediated red fluorescence was increased at FL1 channel with increasing dose of bare AuNRs, indicating a gradual shift in cell population from FL3 channel to FL1 for damaged bacteria. In contrast, PI-mediated red fluorescence increased at FL1 channel with increasing dose of bare AuNRs, indicating a gradual shift from FL3 channel to FL1. These results clearly demonstrate that the Si-AuNRs significantly maintained the bacteria alive whereas bare AuNRs were toxic to the cells and caused cell lysis. The morphological analysis also supported the viability differences of O157 toward bare AuNRs and Si-AuNRs.

In total, possibility of nanoparticle-mediated cell death was because of inherent toxicity produced by CTAB present in the AuNRs. The number of bare AuNRs-treated cells decreased gradually with time and nearly came to 0 after 8 hours. Although surface CTAB was removed by washing, still residual amount and desorption of CTAB might cause cell lysis. Furthermore, strong positive surface charge of AuNRs might also be associated with the decreased growth rate in bacteria.

In contrast, Si-AuNRs favored bacterial growth. This can be hypothesized by the hydrophilic and nontoxic nature of silica. Delving further into it, silica layer can provide many advantages over free cell growth. Previous reports demonstrated that silica matrices facilitated *E. coli* growth as it increased metabolic activity and plasmid stability, and provided a nontoxic environment.^{38–40} It is supported by the results we obtained by evaluating the enzymatic activity of the bacteria. Among the four different bacteria, *E. coli* O157 demonstrated the highest level of membrane peroxidase activity which is inconsistent with previous report.³¹ When we incorporated nanorods in the media, peroxidase activity greatly varied depending on the types of nanorods. Enzymatic activity was found quite low in case of AuNRs-treated bacteria. This can be correlated with the growth inhibition of

O157 by AuNRs. Decreased bacterial growth resulted in a decrease in the endogenous membrane peroxidase activity. Also, TMB might be repulsed by the positive charge of AuNRs as TMB tends to bind to the negatively charged particles because of its net positive charge resulting from amine groups. On the other hand, Si-AuNRs significantly increased the membrane peroxidase activity in dose- and time-dependent manner. It also increased in dose- and time-dependent manner. After 8 hours incubation, approximately 10%–15% increase in enzyme activity was observed at the dose 500 µg/mL compared with that of control. At dose 100 µg/mL, a 25% increase was observed after 8 hours than the activity at 1 hour. Similarly, the other concentrations of Si-AuNRs showed approximately 40%–50% increase in enzyme activity at 8 hours compared with that at 1 hour. Since, our method shows better results, the membrane peroxidase activity of the bacteria could be enhanced with surface-modified AuNRs after directly using with the substrate TMB. Here, the main difference is silica-coated AuNRs showed enhanced visible color change compared with only bacteria with TMB. Moreover, TMB tends to be attracted more to the negatively charged silica-coated AuNRs via electrostatic attraction, and favorable conformational changes were reported once the enzyme was bound to nanoparticles, causing increased TMB signals. Also, silica-coated AuNR acts as an artificial enzyme along with natural membrane peroxidase present in the bacteria, which increased the sensitivity of detection with TMB thus resulting in enhanced TMB signals in our methods. In addition, silica covering could provide large specific surface area for more TMB loading by the interaction with protein molecules present in the bacteria.^{28,38} But, Gao et al presented an interesting report for the high TMB signals.⁴¹ Increase in enzyme concentration or H₂O₂ with TMB substrate creates multiple nucleation centers to form different self-assembled nanostructures by producing different colors due to charge-transfer complex.⁴² In corroboration with our results, the more viable Si-AuNRs-treated O157 could form high enzyme concentration thus producing large signals compared to control bacteria after reacting with TMB substrate. This capability of nanoparticle-mediated enzymatic response may also create new opportunities in the preparation of smart-sensors and as a powerful bio-analytical tool.

The results of these experiments indicate that Si-AuNRs can be used as bacterial growth enhancers for a rapid detection. They can also be used for increasing the sensitivity of TMB assay for the detection of *E. coli* O157. Si-AuNRs may also be potentially applied as biocatalyst and for metabolite production, and for constructing biosensors.

Conclusion

Here, we have demonstrated the efficacy of bare and surface-modified AuNRs on highly pathogenic bacterial strains for their growth and corresponding enzyme activity. Bare AuNRs inhibited the growth and decreased the enzyme activity whereas surface-modified AuNRs favored excellent bacterial growth, higher metabolic activity and enhanced membrane peroxidase signal. The effects were observed in a dose- and time-dependent manner. The results suggest that silica coating on AuNRs makes them promising agents for potential bacterial growth and increasing detection sensitivity for other applications.

Since, nanoparticle interactions toward bacteria are relatively new field, this study presents an important approach to analyzing their enzymatic activity. That is, a clear understanding of the bare and surface-coated AuNRs, impact on bacteria is gained, and this explores beneficial information to make safer nanoparticles without toxicity.

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Disclosure

The authors report no conflicts of interest in this work.

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