

Microwave-assisted fibrous decoration of mPE surface utilizing *Aloe vera* extract for tissue engineering applications

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Abstract: Developing multifaceted, biocompatible, artificial implants for tissue engineering is a growing field of research. In recent times, several works have been reported about the utilization of biomolecules in combination with synthetic materials to achieve this process. Accordingly, in this study, the ability of an extract obtained from *Aloe vera*, a commonly used medicinal plant in influencing the biocompatibility of artificial material, is scrutinized using metallocene polyethylene (mPE). The process of coating dense fibrous *Aloe vera* extract on the surface of mPE was carried out using microwaves. Then, several physicochemical and blood compatibility characterization experiments were performed to disclose the effects of corresponding surface modification. The Fourier transform infrared spectrum showed characteristic vibrations of several active constituents available in *Aloe vera* and exhibited peak shifts at far infrared regions due to aloe-based mineral deposition. Meanwhile, the contact angle analysis demonstrated a drastic increase in wettability of coated samples, which confirmed the presence of active components on glazed mPE surface. Moreover, the bio-mimic structure of *Aloe vera* fibers and the influence of microwaves in enhancing the coating characteristics were also meticulously displayed through scanning electron microscopy micrographs and Hirox 3D images. The existence of nanoscale roughness was interpreted through high-resolution profiles obtained from atomic force microscopy. And the extent of variations in irregularities was delineated by measuring average roughness. *Aloe vera*-induced enrichment in the hemocompatible properties of mPE was established by carrying out in vitro tests such as activated partial thromboplastin time, prothrombin time, platelet adhesion, and hemolysis assay. In conclusion, the *Aloe vera*-glazed mPE substrate was inferred to attain desirable properties required for multifaceted biomedical implants.

Keywords: Metallocene polyethylene, fibrous coating, physicochemical properties, blood compatibility

Introduction

Aloe vera is one of the familiar plants that has been unceasingly used in the day-to-day life of human beings. Its nutrients and active constituents have helped humankind to overcome a spectrum of ailments ranging from wound healing to malignancy for several centuries. Basically, it is a short-stemmed, perennial, drought-resisting, succulent plant that contains hundreds of potentially active substances in the form of essential amino acids, glucomannans, minerals, lipids, vitamins, polysaccharides, major polypeptides, proteins, and antioxidants.^{1,2} These substances have gained *Aloe vera* a favorable reputation in cosmetic, health care, and food industries. At present, the total market value of *Aloe vera* and its nutrients is about 35 billion US dollars, and it is being cultivated in several countries for commercial purposes.³ On the other hand,

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extensive research performed to scrutinize the therapeutic aptitude of *Aloe vera* has revealed its habit of encouraging tissue regeneration or rehabilitation by improving oxygen and blood supply, shielding microbial attacks, and providing essential nourishments.⁴⁻⁷ Recent works reported on the use of *Aloe vera* in the synthesis of modern materials such as nanoparticles, nanocomposites, and nanofibers have proved its importance in the field of wound healing, tissue regeneration, and cancer drug delivery.⁸⁻¹³ Inspired by its profound background, in this work, the extract obtained from *Aloe vera* is utilized to develop an antifouling and biocompatible material for multipurpose biomedical implants for tissue engineering applications.

Bio-implants are man-made systems intended to assist or replace the functions of biological parts. Nowadays, the need for durable bio-implants has been mounting steadily in response to the increase in incidence of age-related chronic illnesses like osteoarthritis, cardiovascular diseases, and congenital and neurological disorders. Because of this, the global market of bio-implants ranging from cardiovascular to urological disorders is expected to rise from 94.1 billion US dollars in 2012 to over 134.3 billion US dollars by 2017, assuming compound annual growth rate of 7.3%. Further, the fact of an increasing ageing population around the world is anticipated to highly influence the statistics mentioned earlier. According to Centers for Disease Control and Prevention, the United States accommodated more than 40 million people in the age group of 65 years and older in 2010. If it continues in this manner, the total population of United States will comprise >20% of aged people by 2030.^{14,15} Apart from this, the poor lifestyle habits are creating a platform for young people to become prey for several age-related ailments. So, sooner or later, the above-mentioned causes will create huge demand for reliable bio-implants, especially in the cardiovascular, dental, and orthopedic fields.

Basically, the implants are made of metals, ceramics, polymers, and composites, which are collectively called biomaterials. Polymers have evolved as the most fascinating choice because of their commendable mechanical properties and easy-to-tailor nature. At present, polymers have a spectrum of applications in the medical arena, ranging from artificial prostheses to drug delivery systems. Though polymers have reasonable advances when compared with their fellow materials, they fail on several occasions due to the problem of blood compatibility.^{16,17} In addition, rejection of biomaterials is also reported to occur due to cytotoxicity and microbial attacks. So, an ideal material is one that can combat the expected complications, and this property is termed as biocompatibility. In fact, to improve the biocompatibility

of a material, several modification techniques have been formulated and demonstrated.^{18,19}

In a different trend, the ability of *Aloe vera* in influencing the biocompatibility of a material is demonstrated using metallocene polyethylene (mPE). It is one of the polyolefin polymers synthesized using a metallocene catalyst, and mPE has been utilized for several basic applications in clinical industry. It is reported to possess commendable mechanical properties; further, it is cost-effective and displays excellent film density, which makes it a suitable replacement choice for several expensive medical materials in practice.²⁰⁻²² However, the problem of biocompatibility associated with mPE is hampering its exploitation in the field of biomedical implants. Accordingly, recent efforts made to solve this problem by manipulating surface modification techniques like radiation (microwave) and acid etching have yielded encouraging results and also expressed its capability to act as a promising biomaterial.^{23,24} In this study, an effort has been made to improve biocompatibility and to impart the integrity of *Aloe vera* on the mPE surface to make it suitable for tissue engineering applications.

The aptitude of artificial materials to promote and support the tissue rehabilitation mechanism is highly desired. So, the surface tailoring techniques are always anticipated to create a wholesome platform to satisfy all the aspects of biocompatibility. This can be achieved by exploiting common biological substances like proteins, lipids, peptides, polysaccharides, antioxidants, etc. Scientists have already demonstrated the role of these biomolecules in enhancing the quality of several polymeric materials by delaying clot formation, reducing platelet adhesion followed by activation, and shielding microbial attacks.²⁵⁻²⁹ On the other hand, *Aloe vera* extract utilized in this work is a natural storehouse of several vital substances required to promote and guide the regeneration mechanism. Therefore, the competence of *Aloe vera* extract in repairing the surface characteristics and enhancing the blood compatibility of mPE was scrutinized.

Experimental section

Materials

Chemicals required for performing activated partial thromboplastin time (APTT), prothrombin time (PT), hemolysis, and platelet adhesion tests were purchased from Sigma-Aldrich (St Louis, MO, USA) and utilized without further purification. Sheets of mPE were kindly offered by Indian Institute of Technology, Kharagpur, India. Blood samples and freshly prepared platelet-rich plasma (PRP) required for performing blood compatibility assays were obtained from Dindigul Blood Bank, Dindigul, India. Blood was taken from healthy

adult human volunteers according to the predetermined protocol of Dindigul Blood Bank. Further, all the procedures involved in the usage of blood were approved and overviewed by the Institutional Ethical Committee at PSNA College of Engineering and Technology, Dindigul, India.

Isolation and preparation of *Aloe vera* extract

Initially, fresh succulent leaves of *Aloe vera* were carefully handpicked and washed with distilled water. Then, the skinny upper part and the yellowish latex part were peeled and discarded. This was followed by the separation of semitransparent whitish gel with the help of a metal ladle. Finally, the gel was blended into a concentrated fibrous extract, which was used for coating mPE samples without further alteration.

Microwave-assisted *Aloe vera* coating

Metallocene polymer sheet was cut into identical square-shaped samples of dimension 2 cm × 2 cm and washed with distilled water and 70% ethanol to get rid of foreign particles (if any) on the surface. Before coating, the samples were treated with microwaves; the ability of microwaves in accelerating the chemical process and their role in the development of homogeneous coating have been documented already.^{30–33} However, in this work, we followed a unique protocol to facilitate effective coating of *Aloe vera* on mPE samples by energizing the polymer surface and improving its roughness with the help of microwaves.²³

Initially, samples to be coated were exposed to microwave of 800 W and 2,450 MHz produced from Samsung ME711K microwave oven for a period of 60 seconds.

Then the samples were submerged in sterilized petri dishes filled with 10–15 mL of prepared *Aloe vera* extract. To smoothen the coating process, *Aloe vera*-filled vessels were placed on the rocking shaker providing seesaw motion at constant speed and maintained under optimum conditions. The samples were removed at two different times, that is, one set of samples was taken out from the petri dish after 12 hours, while the other set was retrieved after 24 hours. Both *Aloe vera*-glazed samples were dried at room temperature overnight to eliminate the moisture present on the surface. Further, for all surface and blood compatibility characterization studies, samples were freshly prepared according to the described protocol. The results of 12 hours *Aloe vera*-coated (A-12 h-mPE) and 24 hours *Aloe vera*-coated (A-24 h-mPE) samples were compared with pristine-control mPE. The process of extracting, preparing, and coating *Aloe vera* extract on mPE substrates is presented in Figure 1.

Surface characterization studies

Fourier transform infrared (FTIR) spectroscopic analysis was performed for both control and coated samples using IRTracer-100 (Shimadzu) system, and the spectrum was recorded over the range of 400–4,000 cm^{-1} at 40 scans per minute. The changes in wettability of mPE surfaces after coating with *Aloe vera* were demonstrated using VCA Optima contact angle measurement unit. A water droplet of size 2 μL was used; since the surfaces were highly hydrophilic, the photographs were taken immediately after placing the droplet (within 10 seconds) using ultrafast mode. Further, the degree of angle formed was analyzed using computer integrated software. Scanning electron microscopy (SEM) was utilized to display the presence of *Aloe vera* coating on

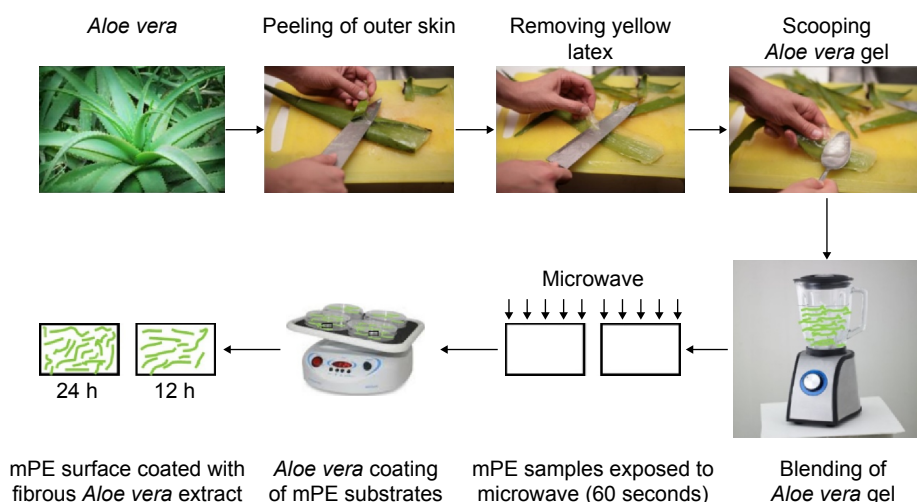


Figure 1 Steps involved in coating of *Aloe vera* extract on mPE samples.

Abbreviation: mPE, metallocene polyethylene.

mPE substrates. For that, samples were sputter coated with gold and imaged through Hitachi Tabletop Microscope (TM3000) coupled with an energy dispersive spectrometry analysis system, at a magnification of 1,500 \times –2,000 \times . In addition, real-time color images of coated samples and the thickness of *Aloe vera* coating were determined using Hirox-3D digital microscope (KH-8700). Hirox microscope offers broad range of magnifications from 1 \times to 7,000 \times , and it also produces 3D images of the scanned area for a better perspective. The images of controls and extract-coated samples were taken at 100 \times , 500 \times , and 1,000 \times , respectively, using MGS-2500 REZ lens in standard resolution of 1,200 \times 1,600. Moreover, the distribution of crests and troughs on polymer surfaces was illustrated using built-in 3D profilometry software. Finally, the SPA3800N atomic force microscopy (AFM) was used to perform nanometric surface characterization of mPE substrates. Scanning was performed in the contact mode over an area of 10 \times 10 μ m at a rate of 1.50 Hz. Furthermore, SPI3800 software was used to obtain average roughness (Ra) and root mean square (RMS) values of mPE substrates.

In vitro blood compatibility studies

Coagulation assay

Activated partial thromboplastin time

APTT is a first-line and basic blood compatibility test used to detect the capability of a biomaterial to delay the activation of the coagulation pathway through the intrinsic mechanism, that is, induced by foreign bodies. First, four substrates (two control and two *Aloe vera*-coated substrates) were incubated at 37°C with 100 μ L of platelet-poor plasma obtained from centrifuged tri-sodium citrate added whole blood. Then, it was activated by adding rabbit brain cephalin (100 μ L) for 5 minutes. Finally, 0.025 M of calcium chloride was added to the mixture, and the time taken for the appearance of the initial clot was detected by using a steel hook and stopwatch.

Prothrombin time

On the other hand, PT was calculated to display *Aloe vera*-coated mPE-induced activation of the extrinsic pathway. Like the APTT assay, initially, the samples were preincubated with platelet-poor plasma (100 μ L) at 37°C. Then 0.9% NaCl-thromboplastin (Factor III; 100 mL)-containing Ca²⁺ ions was added, and the mixture was slowly stirred using a steel hook until the inception of a clot; the time taken for clot formation was measured using a stopwatch.

Platelet adhesion study

Adhesion of platelets on polymer surfaces is another threat since this eventually releases clotting factors that end up

activating coagulation pathways. This test was performed to scrutinize the ability of *Aloe vera*-coated surfaces to impede platelet adhesion and activation. Here, PRP was utilized, and the samples were incubated in freshly isolated PRP at 37°C for an hour. After that, to fix the adhered platelets on the surface, the samples were incubated with 2% Triton-PSB buffer for 30 minutes at 37°C, and it was allowed to dry before imaging. Finally, the morphology of adhered platelets and formation of clumps were imaged using a light microscope at 40 \times .

Hemolysis assay

To exhibit the percentage of red blood cell (RBC) damage caused by the mPE surface, hemolysis assay (HA) was performed using citrated whole blood. To start with, both control and *Aloe vera*-coated samples were equilibrated in physiological saline (0.9% w/v, 37°C, 30 minutes). Then, it was incubated with a mixture of aliquots of citrated blood (3 mL) and diluted saline. Subsequently, the samples were incubated in a mixture of blood and distilled water (4:5) to cause complete hemolysis and in physiological saline solution, respectively. The above mixtures were considered as positive (blood: distilled water) and negative (physiological saline) controls, in which samples were incubated for a period of 60 minutes at 37°C. After retrieving the sample, the mixtures were centrifuged, and the clear supernatant was collected for further analysis. Finally, the absorbance of samples was measured at 542 nm, which directly relates the degree of damage done to RBCs.³⁴

Statistical analysis

All the quantitative experiments were done three times (n=3), and the statistical significance was predicted through one-way ANOVA method. Moreover, the results were expressed as mean \pm standard deviation (SD).

Results and discussion

The prime duty of a bio-implant is to provide a virtual environment to stimulate and support the regeneration of a desired tissue or an organ system in the human body. To achieve this goal, the artificial material should possess optimum wettability, chemical functionalities, surface energy, roughness, conductivity, and reactivity, which are collectively termed as physicochemical properties. Meanwhile, the range and influence of these factors on promoting rehabilitation have been demonstrated by copious research. So, any type of manufacturing and surface modification technique proposed is essentially directed toward satisfying optimum requirements, thereby achieving the anticipated outcome. Furthermore,

these surface parameters are also reported to greatly influence crucial aspects of biocompatibility such as blood component interaction and cellular compatibility. Hence, to illustrate the ability of *Aloe vera*-coated mPE to produce these essential features, the following studies were performed.

Physicochemical characterization studies

Fourier transform infrared spectroscopy

FTIR is the basic characterization test used to analyze and confirm the presence of appropriate functional groups. The recorded FTIR spectrum is displayed in Figure 2(A), in which the FTIR outline of mPE, used *Aloe vera* extract, A-12 h-mPE and A-24 h-mPE substrates, are compared. Similar patterns of peaks with different intensities were noted for both control and coated samples at wavelengths 2,916 cm^{-1} , 2,848 cm^{-1} , 1,735 cm^{-1} , 1,467 cm^{-1} , 1,371 cm^{-1} , 1,236 cm^{-1} , 719 cm^{-1} , and 607 cm^{-1} , respectively. The exhibited twin bands at 2,916 cm^{-1} and 2,848 cm^{-1} represent the presence of C–H stretch for the alkanes group. The characteristic absorption peak at 1,735 cm^{-1} was for C=O stretch of esters; the peak at 1,467 cm^{-1} shows the presence of alkane groups (C–H

bending), and the existence of N–O stretch is inferred from the peak produced at 1,371 cm^{-1} . Further, the absorption band in the region of 1,236 cm^{-1} express C–O stretch of carboxylic acids (controls), and the crest at 1,018 cm^{-1} indicates the C–O stretch in secondary alcohol. Finally, the peaks at 719 cm^{-1} and 607 cm^{-1} displays C–H bend of aromatics and alkynes group, respectively.

Though there was no major change in the functional groups, coated mPE samples displayed characteristic absorption spectrum of *Aloe vera* at 1,735 cm^{-1} and 1,236 cm^{-1} , indicating the presence of O-acetyl esters; the peaks formed at 1,467 cm^{-1} and 719 cm^{-1} revealed the essence of aromatic vibrations of components in *Aloe vera* gel.^{8,11} Meanwhile, the *Aloe vera*-coated substrates were also observed to possess shifts in the peaks at far regions of the IR spectra ($<600 \text{ cm}^{-1}$), which is displayed clearly in Figure 2B and C. The peaks found at 424 cm^{-1} , 439 cm^{-1} , 451 cm^{-1} , 480 cm^{-1} , 516 cm^{-1} of control were shifted to 422 cm^{-1} , 435 cm^{-1} , 447 cm^{-1} , 476 cm^{-1} , 514 cm^{-1} for A-12 h-mPE and 426 cm^{-1} , 437 cm^{-1} , 453 cm^{-1} , 470 cm^{-1} , 501 cm^{-1} for A-24 h-mPE, respectively. These shifts were suspected to appear because of minerals

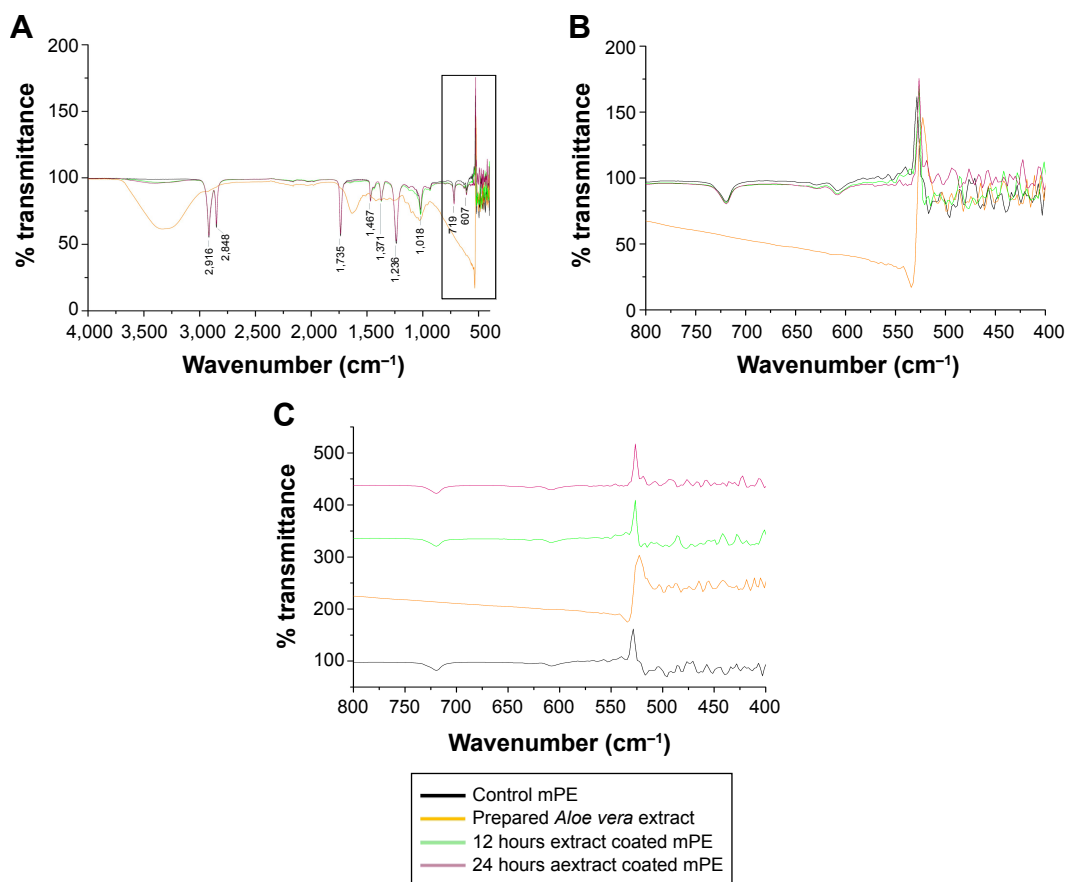


Figure 2 Recorded FTIR spectrum of control and *Aloe vera*-coated samples.

Notes: (A) Spectrum from 4000–500 cm^{-1} , (B) spectrum from 800–400 cm^{-1} , (C) individual spectrum of substrates at 800–400 cm^{-1} .

Abbreviations: mPE, metallocene polyethylene; FTIR, fourier transform infrared.

present in *Aloe vera* gel, which can be clearly concluded from the FTIR fingerprint of extract prepared and the works documented. Additionally, the peaks formed in between $400\text{--}500\text{ cm}^{-1}$ and $500\text{--}600\text{ cm}^{-1}$ demonstrated Si–O bending and C–H stretch appropriately.^{35,36}

Hence, the reported characteristic absorption spectrum and shifts in peaks depict the existence of *Aloe vera* coating on microwave-treated mPE samples. The above interpretations also illustrated the presence of active components of *Aloe vera* gel and chemical functionalities (cell adhesion–assisting oxygen groups), which are crucial for mimicking the natural environment and supporting the recovery process.

Contact angle measurement

Wettability is one of the important surface parameters, which plays a pivotal role in controlling biomolecule adsorption and cellular interaction. In this work, the wettability was measured for both control and coated samples; the experiment was repeated three times, and the contact angle is expressed in mean \pm SD, as displayed in Figure 3. The mean contact angle calculated for mPE was 88.43° ; the A-12 h-mPE samples had a mean contact angle of 68.13° , whereas the A-24 h-mPE samples showed a drastic increase in wettability by yielding a mean contact angle value of 32.93° only. So, from the results, we can clearly infer that after extract coating, mPE surfaces become super-hydrophilic with contact angle values reduced by more than half compared with control, exclusively in the case of A-24 h-mPE substrates.

This significant improvement is suspected to be due to the water-loving constituents present in *Aloe vera* gel,

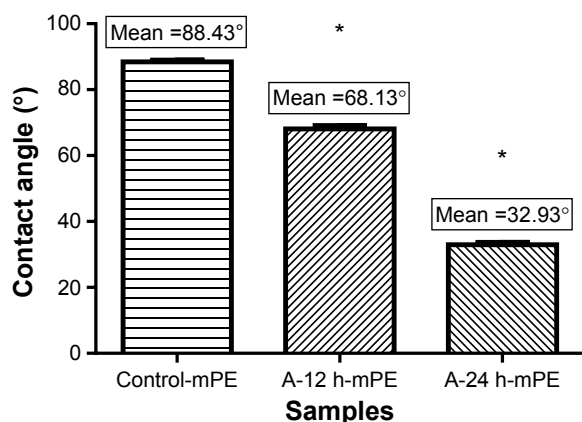


Figure 3 Mean contact angle values of control and coated samples.

Note: *Differences between the mean values is significant with respect to control ($P < 0.05$).

Abbreviations: mPE, metallocene polyethylene; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

and it correlates with the results of previous studies.^{8,11} As mentioned earlier, *Aloe vera* is a natural store house of several polysaccharides, minerals, vitamins, amino acids, and proteins that are basically hydrophilic. The recorded contact angle falls in the optimum range of $70^\circ\text{--}30^\circ$, which has previously been reported in several works^{37,38} of research. A wide variety of cells like fibroblasts, human umbilical vein endothelial cells, osteoblasts, and cardiomyocytes were noted to adhere, proliferate, and differentiate expressively in this range.³⁷ On the other hand, surfaces with higher wettability also allows the displacement of undesired proteins adhered (like fibrinogen) by cell adhesive serum proteins, such as fibronectin and vitronectin, in contrast to hydrophobic surfaces.³⁸ This property of controlling and translating protein adsorption is greatly needed to avoid the complication of blood component activation.

Furthermore, the increased wettability indirectly exhibits the changes in surface energy of mPE samples. In general, surfaces with high energy allow uniform biofilm formation, which in turn produces better spreading of water droplets.³⁹ Accordingly, coated samples were observed to facilitate spreading of water droplets, which supports the fact of increased surface energy. The spreading of water droplets on control and *Aloe vera*-coated samples is illustrated in Figure 4. Therefore, the results of contact angle studies confirm the presence of *Aloe vera* coating and active biomolecules, as stated in the FTIR test. They also express the possible exploitation of *Aloe vera*-coated mPE substrates for multipurpose biomedical applications.

SEM analysis

The SEM micrograph offers a quick view on surface topographical characteristics like roughness, presence of pits, coating, etching, etc. The micrographs were pictured for all samples and the same is compared in Figure 5. The SEM image of untouched mPE recorded at $1,500\times$ showed a clear surface indicating the absence of crests and troughs which ultimately disclosing its poor roughness. In contrast, the 60-second microwave-treated control imaged at $2,000\times$ showed bright white spots depicting the improvement in roughness and formation of pits; these surfaces were reported to improve coating characteristics. Basically, rougher surfaces support better adhesion due to the fact that there is an increase in surface area between coating and substrate. Unlike a flat surface, the area of physical contact is documented to be better in rough surfaces, which ultimately results in mechanical interlocking of coating with substrate, thus enhancing the glazing strength as well.^{40,41}

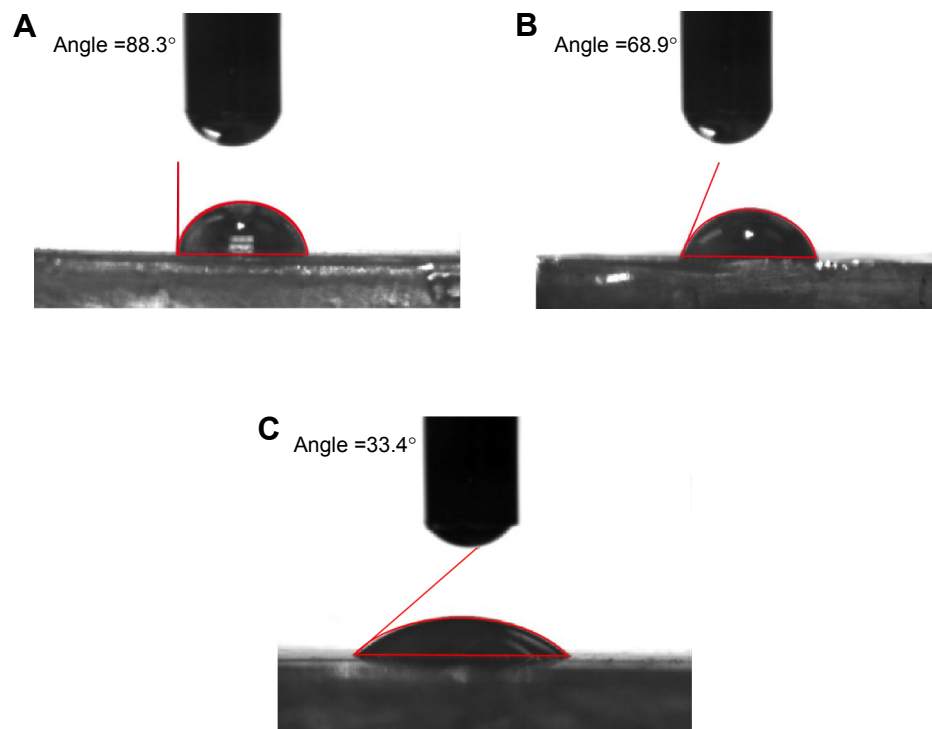


Figure 4 Representative contact angle images showing spreading of water droplet on control and coated mPE samples.

Notes: (A) Pristine mPE, (B) 12 hours extract-coated mPE, (C) 24 hours extract-coated mPE.

Abbreviation: mPE, metallocene polyethylene.

Therefore, the recorded micrographs evidently support the importance of microwave treatment in augmenting the adhesion and strength of extract coating. Meanwhile, the presence of a fibrous structure was noted from images of

A-12 h-mPE and A-24 h-mPE samples (1,500×), which once again confirms the existence of coating. In addition, the A-24 h-mPE surfaces exhibited dense sedimentation of fibers when compared with A-12 h-mPE, which corroborated

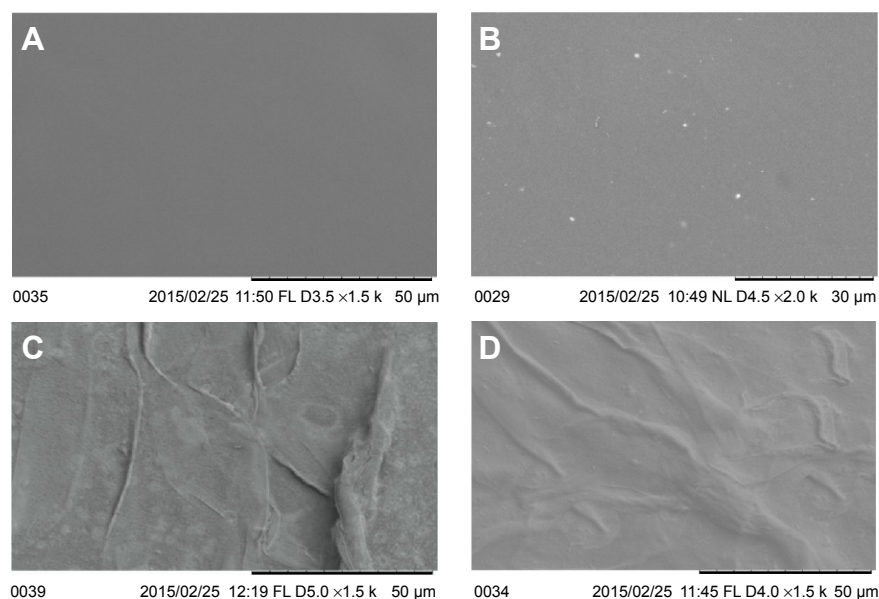


Figure 5 Representative SEM micrographs of control, microwave-treated, and extract-coated mPE samples.

Notes: (A) Pristine mPE at 1,500×, (B) 60 seconds microwave-treated mPE at 2,000×, (C) 12 hours *Aloe vera*-coated mPE at 1,500×, and (D) 24 hours *Aloe vera*-coated mPE at 1,500×.

Abbreviations: mPE, metallocene polyethylene; SEM, scanning electron microscopy.

the results of contact angle measurement showing higher wettability for samples exposed for a long period. Thus, it can be ascertained that the properties of extract-glazed mPE surfaces enhance with increase in exposure time due to comprehensive settlement of active components present in *Aloe vera*.

Hirox-3D digital microscope studies

Inspired by the results of SEM analysis, the micrometric characterization of coated samples and the dispersal of fibrous coating were analyzed with the help of a Hirox 3D digital microscope. The main advantages of the Hirox microscope are its ability to capture real-time color images and to produce high-resolution two-dimensional (2D) and three-dimensional (3D) views, which meticulously explores the micro-topographical signatures present on the material. Unlike SEM, special preparation of samples, such as coating, is not required in Hirox analysis, which is remarkably suitable for sensitive surfaces. At first, 2D images of pristine-control mPE, A-12 h-mPE, and A-24 h-mPE samples were taken at 100 \times , 500 \times , and 1,000 \times , respectively, which are illustrated in Figure 6. From the images, we can clearly observe the presence of homogeneous *Aloe vera* coating in the form of dense hair-like structures. Like SEM images, the A-24 h-mPE sample exhibited an enriched number of fibers with dense morphology when compared with that of A-12 h-mPE substrates. It again confirmed the time-dependent deposition of active components on microwave-treated mPE. More interestingly, the fibrous coating formed resembles the structure of extra cellular matrix components. Extra cellular matrix is the building block of tissues which contains fibrous biomolecules like collagens, elastin, keratin, fibronectin, laminins, and vitronectin. These structures offer mechanical support to the tissue and regulate several features of cell behavior. These fibers have been reported to control proliferation, migration, and differentiation of cells; further, the orientation of these fibers are dependent on the tissues. Currently, several modern technologies such as electrospinning are mainly aimed at replicating this fibrous environment, thereby enhancing the recovery process by attracting and supporting new cells.⁴² Thus, from the images, we can determine that *Aloe vera*-coated surfaces not only possess essential components for tissue regeneration but also acquire the ability to mimic the biological environment.

Concurrently, the increase in roughness due to microwave treatment and its effect on coating characteristics were also analyzed. The 3D images of substrates recorded at 1,000 \times , shown in Figure 7, express the presence of crests and troughs. Briefly, number of crests and troughs was increased on

coated surfaces when compared with control. To provide a detailed insight, graphical illustration of crests and troughs were obtained for each sample utilizing built-in 3D profilometry software. Using this software, several sketches were randomly drawn on the images recorded at 1,000 \times , and the one which showed the maximum value was considered for further analysis. After choosing the desired profile, the built-in software generated an excel sheet containing values of coordinates (X, Y, and Z) obtained from 3D geometry, which was exploited to generate a graphical illustration.⁴³ Figure 8 compared the fingerprints of untreated and coated samples; the escalation in roughness is inferred due to the microwave treatment and the settlement of fibers on mPE surface. Apart from the surface roughness, the coating characteristics were also reported to depend on the number of peaks present on the material surface. Excitingly, the microwave-treated samples had more peaks when compared with untouched mPE, which motivated us to check the influence of the microwave-improved properties on the coating thickness achieved.

Hence to determine that, the value of coating thickness present on microwave-treated sample was compared with that of an untreated one. First, the glazed samples were cut into small pieces and vertically placed on the imaging stage using adhesives. When viewed in 100 \times , the presence of a thin coating was observed, and its thickness was measured by drawing a line. Interestingly, the microwave-treated sample had a coating thickness of 134.8 μm , while on the untreated mPE, the thickness of *Aloe vera* extract was only around 99 μm . So, the Hirox 3D analysis meticulously supports the fact of enhanced coating characteristics of rough surfaces formed by the microwave treatment and also demonstrates the existence of dense fibrous coating similar to SEM images.

AFM analysis

In addition to coating characteristics, the surface roughness also play a crucial role in determining the behavior of a material within a biological environment alongside surface wettability and energy. The phenomenon that makes the rougher surface highly suitable for cell adhesion and proliferation is the presence of grooves and ridges unlike its counterpart, that is, smooth surfaces.⁴⁴ Further, the range of roughness required for a promising biomaterial was also observed to vary depending on the cells and tissue it intended to assist. For instance, larger cells like osteoblasts were inferred to adhere and proliferate well on surfaces with micrometric roughness. On the other hand, irregularities in the range of the nanoscale were reported to be suitable for

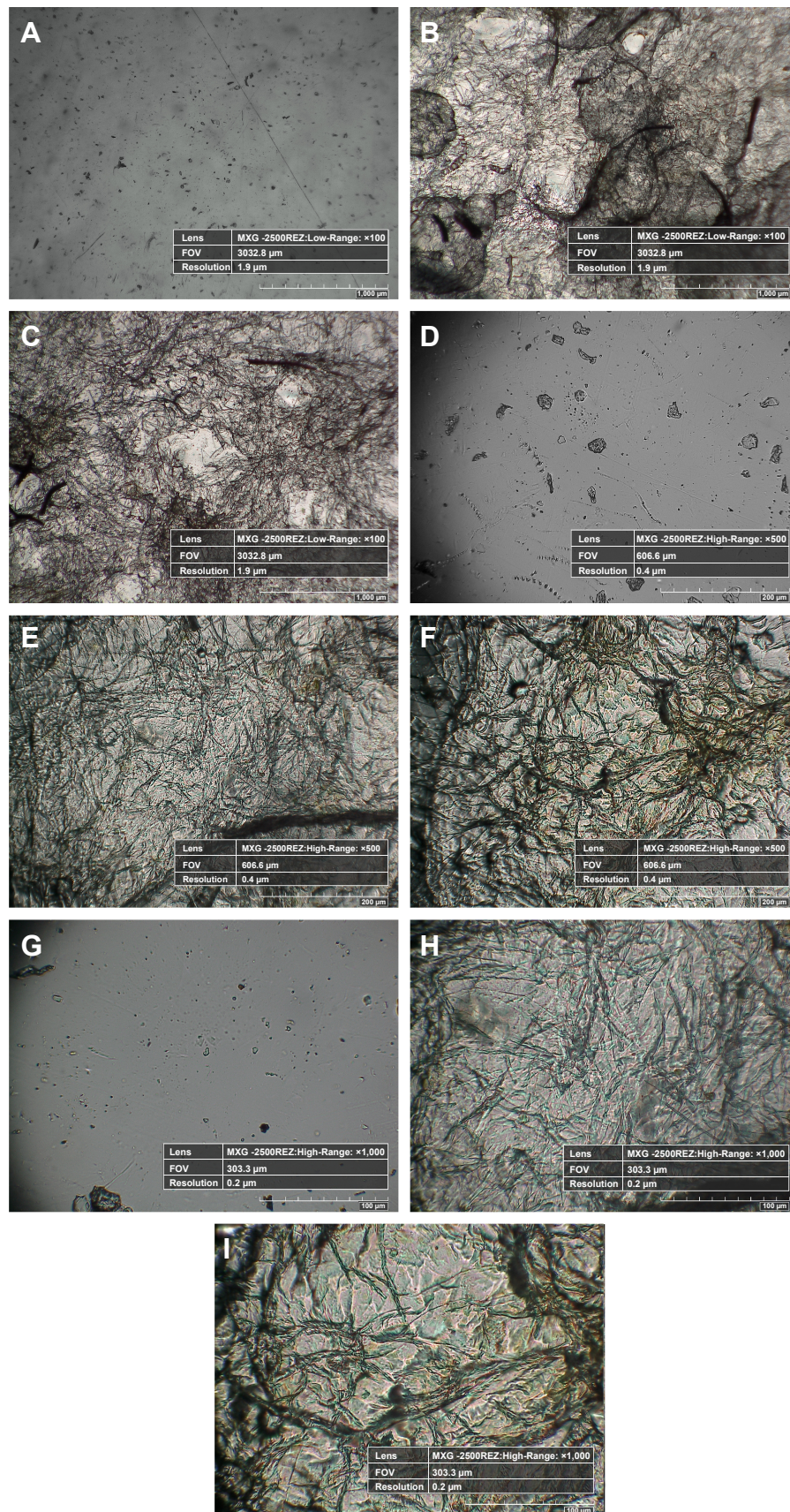


Figure 6 Hirox 2D images of control, 12 hours extract-coated mPE samples and 24 hours extract-coated mPE samples.

Notes: (A), (B), (C) Images taken at 100× (mPE, A-12 h-mPE and A-24 h-mPE, respectively), (D), (E), (F) Images taken at 500×, (G), (H), (I) Images taken at 1,000×.

Abbreviations: mPE, metallocene polyethylene; D, dimensional; A-12 h-mPE, 12 hours Aloe vera-coated; A-24 h-mPE, 24 hours Aloe vera-coated.

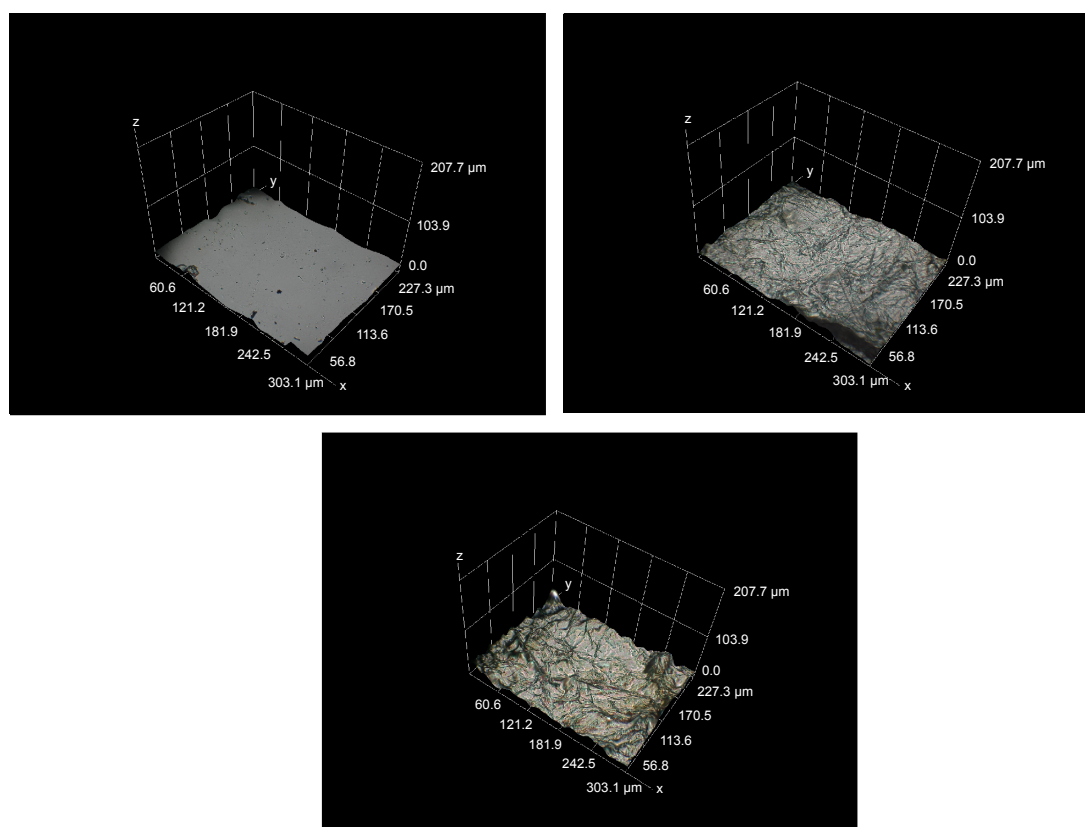


Figure 7 Hirox 3D images of control and extract-coated mPE samples taken at 1,000 \times .

Notes: (A) Pristine mPE, (B) A-12 h-mPE, (C) A-24 h-mPE.

Abbreviations: mPE, metallocene polyethylene; D, dimensional; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

better adhesion, proliferation, and differentiation of fibroblasts, endothelial, and neural cells.^{37,45} Moreover, it was also documented that surface roughness is a vital factor for determining the blood compatible property of a biomaterial, since it has the ability to delay the clot formation. Initially the role of surface roughness in improving coating characteristics was demonstrated using SEM micrographs and Hirox 3D images. Then, AFM is utilized to provide high resolution and 3D nanometric insights of the surface anomalies present on control and *Aloe vera*-coated samples.

The images were captured on different zones at an area of 10 \times 10 μm , and the mean value of Ra was statistically calculated using one-way ANOVA. Figure 9 shows the AFM images of pristine and A-24 h-mPE samples. Unlike pristine mPE, the extract-coated substrates emphasized the presence of small pits because of microwave treatment prior to coating, which was also demonstrated in SEM micrographs. In the interim, the mean Ra value for the control was calculated to be 2.069 nm. Conversely, the A-24 h-mPE sample showed a mean Ra of 7.796 nm; this significant improvement confirms the formation of pits and the existence of fibrous coating. Further, the recorded roughness value of coated samples

greatly corroborated with the optimum range prerequisite for successful tissue regeneration reported previously.³⁷ Thus, from the surface characterization studies, the *Aloe vera*-coated samples depicted to possess commendable signatures required for a promising biomaterial, which was further established by performing several blood compatibility studies.

In vitro blood compatibility tests

Coagulation assay

Hemocompatibility is one of the crucial aspects of medical implants; as soon as a prosthetic material is placed in our body, the first and foremost biological environment it supposed to encounter is what we call the river of life, “blood”. It is a complex mixture of RBCs, white blood cells, platelets, plasma proteins, and other essential biomolecules. Whenever a biomaterial comes in contact with the blood stream, it attracts the circulating plasma protein, which results in toping of the material surface with undesired biomolecules. Subsequently, the protein-doped surface activates several clotting factors and platelets moving in the blood stream. Activated platelets then adhere and aggregate on the material surface, which is followed by stimulation of several

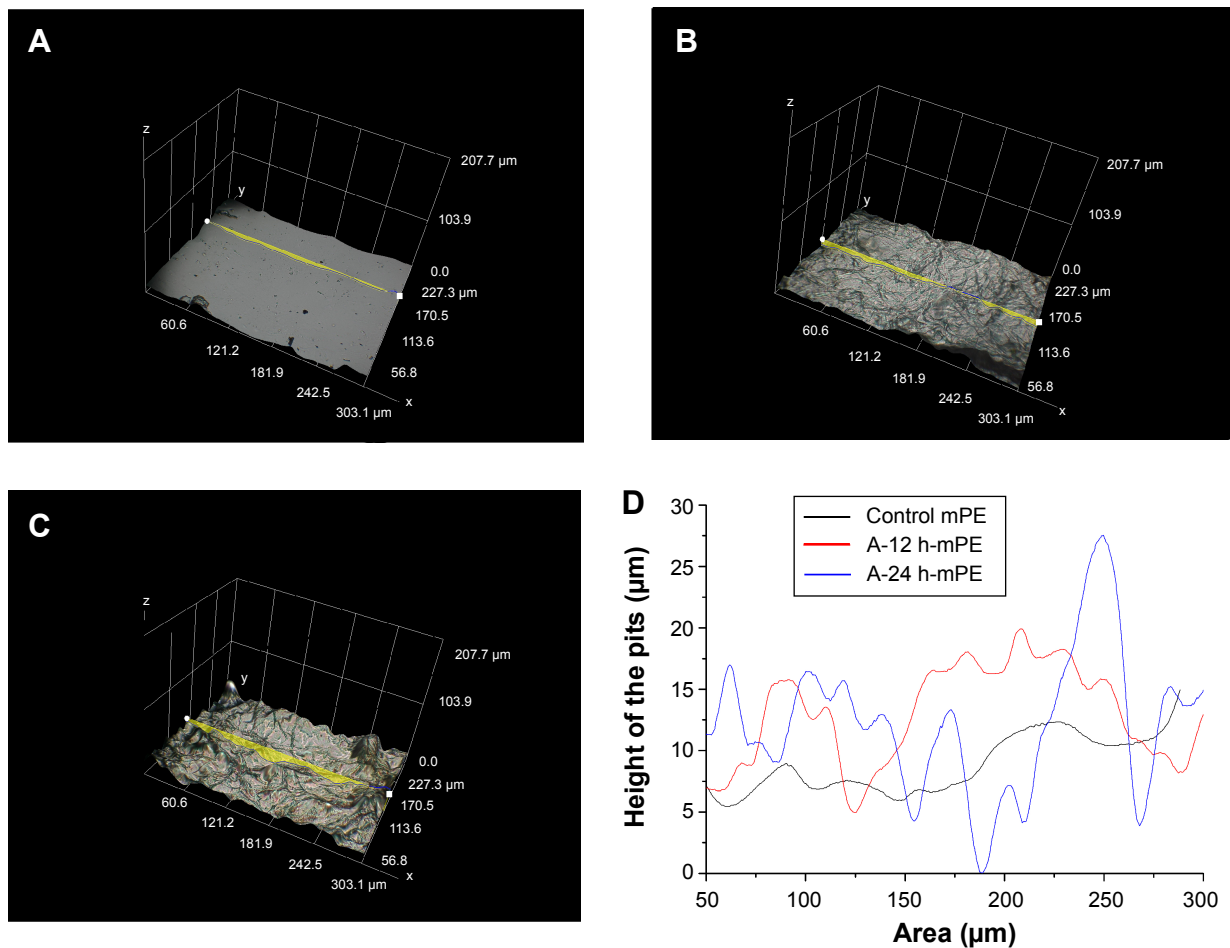


Figure 8 Hirox 3D images of control and extract-coated mPE samples taken at 1,000 \times with profiles.

Notes: (A) Pristine mPE, (B) A-12 h-mPE, (C) A-24 h-mPE, (D) outline of crests and troughs in figure (8A, B and C).

Abbreviations: mPE, metallocene polyethylene; D, dimensional; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

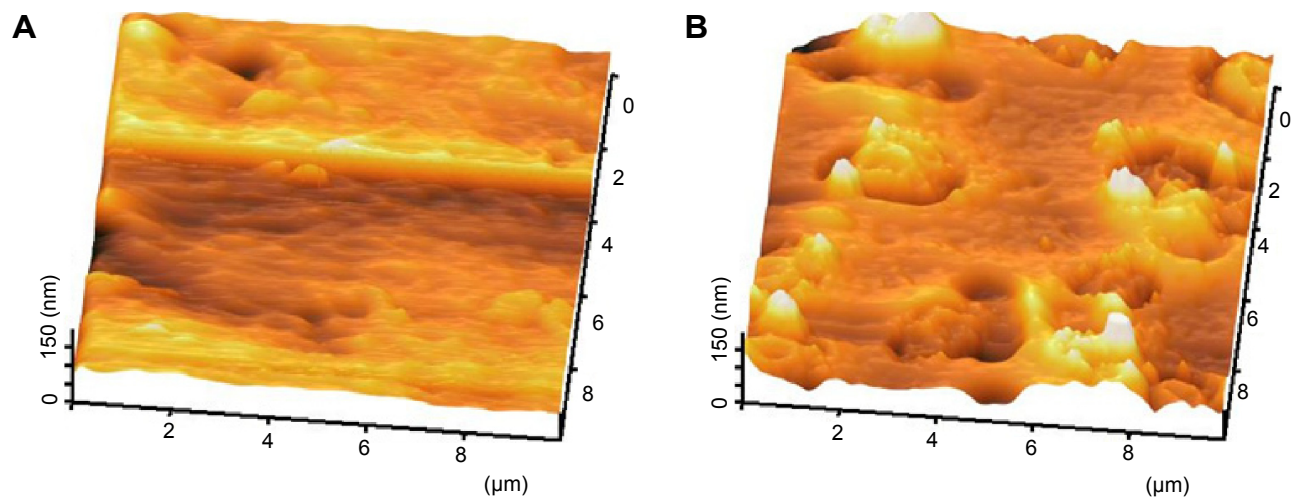


Figure 9 Representative 3D topographical view of control and 24 hours coated sample obtained by AFM.

Notes: (A) Pristine mPE, (B) A-24 h-mPE.

Abbreviations: mPE, metallocene polyethylene; AFM, atomic force microscopy; D, dimensional; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

coagulation pathways. The intrinsic and extrinsic pathways are the two main mechanisms greatly engaged in subsequent clot formation. These chain reactions alert our defense system, and paves the way for the occurrence of inflammation and foreign body responses which ultimately ends in rejection of the implant.^{16,17}

However, a biomaterial with commendable surface features has been reported to avert these complications either by halting protein adsorption or by allowing the displacement of undesired proteins adhered with cellular compatible biomolecules, as mentioned previously.^{37,38} This property not only saves the implant from rejection but also boosts its job of assisting diseased tissue. The characterization studies carried out so far reveal the improvement of mPE surface properties after extract coating. *Aloe vera*, used in this work, consists of various natural anti-inflammatory agents in the form of sugars, amino acids, lipids, active enzymes, and salicylic acid. And the extract has been exploited as a powerful wound healing agent for several centuries. Thus, in order to disclose the ability of *Aloe vera*-improved mPE surfaces in combating the expected blood mediated complications, in vitro coagulation assays were performed.

Activated partial thromboplastin time

APTT assay was performed on both control and coated mPE samples. The experiment was repeated three times ($n=3$) to check the reproducibility of outcomes observed. Statistical analysis was performed through one-way ANOVA, and the mean values are illustrated in Figure 10. The control mPE had

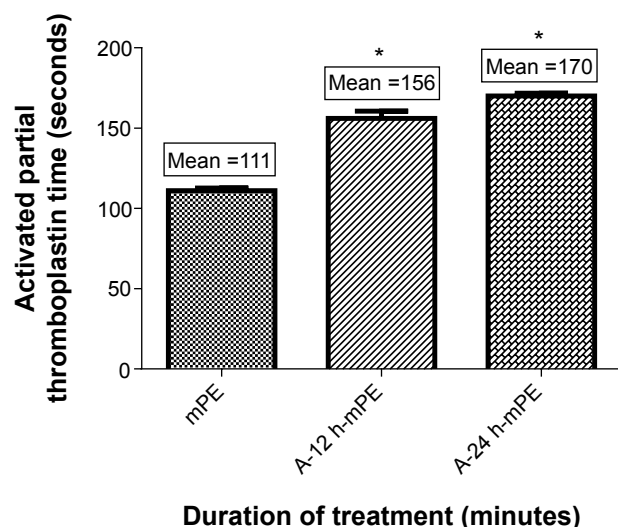


Figure 10 Comparison of activated partial thromboplastin time of control and coated mPE ($n=3$).

Notes: Values shown are mean \pm SD and the difference in mean is significant with $P<0.05$. *Differences between the mean values is significant with respect to control ($P<0.05$).

Abbreviations: mPE, metallocene polyethylene; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

a mean clotting time of 111 seconds, whereas the *Aloe vera*-coated sample demonstrated a notable delay in clot formation. The A-12 h-mPE surfaces were able to avert clot formation for a period of 156 seconds, and the A-24 h-mPE samples exhibited an increased clot formation time of 170 seconds. The statistical analysis showed a significant difference ($P<0.05$) between control and coated samples.

Prothrombin time

Prothrombin time was calculated to illustrate the ability of *Aloe vera*-coated mPE surfaces to delay the formation of clots through the extrinsic pathway. Like APTT, the experiment was repeated three times, and the mean value obtained for each sample was considered as its PT. As expected, the coated samples showed a better delay in clot formation when compared with controls. The A-12 h-mPE samples had a mean clotting time of 36.1 seconds compared with 39 seconds for A-24 h-mPE substrates, while for controls, the mean PT was 27.23 seconds. Moreover, the statistical analysis carried out using one-way ANOVA showed a significant difference ($P<0.05$) between control and extract-coated samples, which is represented in Figure 11. The above outcomes illustrate the ability of *Aloe vera* coating to delay the development of clots through contact activation, that is, both intrinsic and extrinsic pathways.

Platelet adhesion studies

Adhesion of platelets is another important phenomenon, which is actively involved in stimulation of undesired

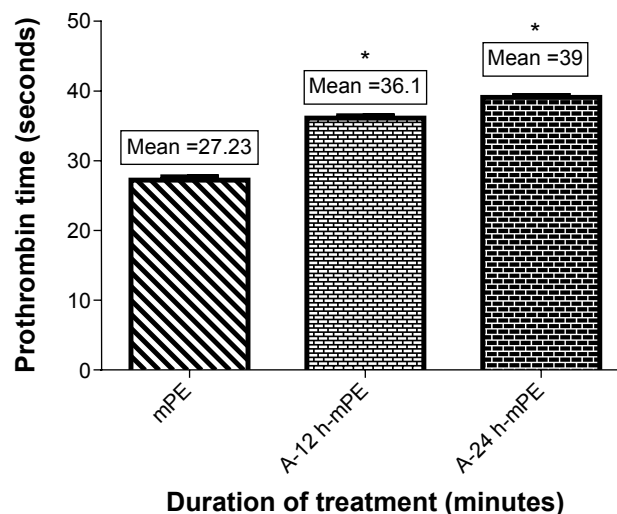


Figure 11 Comparison of prothrombin time of pristine and extract-coated metallocene polyethylene ($n=3$).

Note: Values shown are mean \pm SD and the difference in mean is significant with $P<0.05$. *Differences between the mean values is significant with respect to control ($P<0.05$).

Abbreviations: mPE, metallocene polyethylene; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

hemo-complications. In this work, the platelet-repellent property of *Aloe vera*-coated substrates were explored using PRP. Images of all samples taken using a light microscope at 40× were compared in Figure 12. The platelets were observed to readily adhere and aggregate extensively on pristine mPE, whereas on A-12 h-mPE samples, the adhesion was very minimal, and the aggregation also reduced significantly. On the other hand, the A-24 h-mPE samples exhibited an excellent ability to combat platelet attachment by reducing the adhesion percentage less than one (approximately). This significant improvement may indicate the presence of antioxidants and platelet-repellent biomolecules available in *Aloe vera* on coated samples. The same was reported in the in vivo studies performed by Singh and Fahim.⁴⁶ In the case of pristine samples, the absence of coating and lack of required topographical characteristics pave the way for deposition of unspecific plasma proteins. It in turn facilitates the activation, adhesion, and aggregation of platelets. In addition to *Aloe vera* extract, the presence of nano-roughness is also suspected to influence the platelet-repellent property of the modified

mPE surface, which is highly desirable in temporary and permanent blood-contacting medical devices.

Hemolysis assay

Apart from deterring clot formation, a material should also demonstrate good compatibility with blood cells. RBC is the abundant cell type present in the blood, and a commendable relationship with these cells will serve as a marker for better cellular compatibility of an appropriate biomaterial. For determining this, HA was performed, and the absorbance value recorded at 542 nm related the damage caused to RBCs upon biomaterial interaction. The mean absorbance elicited by the control was 0.018. Interestingly, for A-12 h-mPE, the mean absorbance was 0.015, and for A-24 h-mPE samples, it was 0.0065 (Figure 13), which is only a quarter percentage of uncoated samples. This reduction in absorbance with increasing time of *Aloe vera* exposure reveals the enhancement in compatible properties of mPE surfaces after coating. Hence, from the above interpretation, we can conclude that the *Aloe vera*-glazed mPE substrates

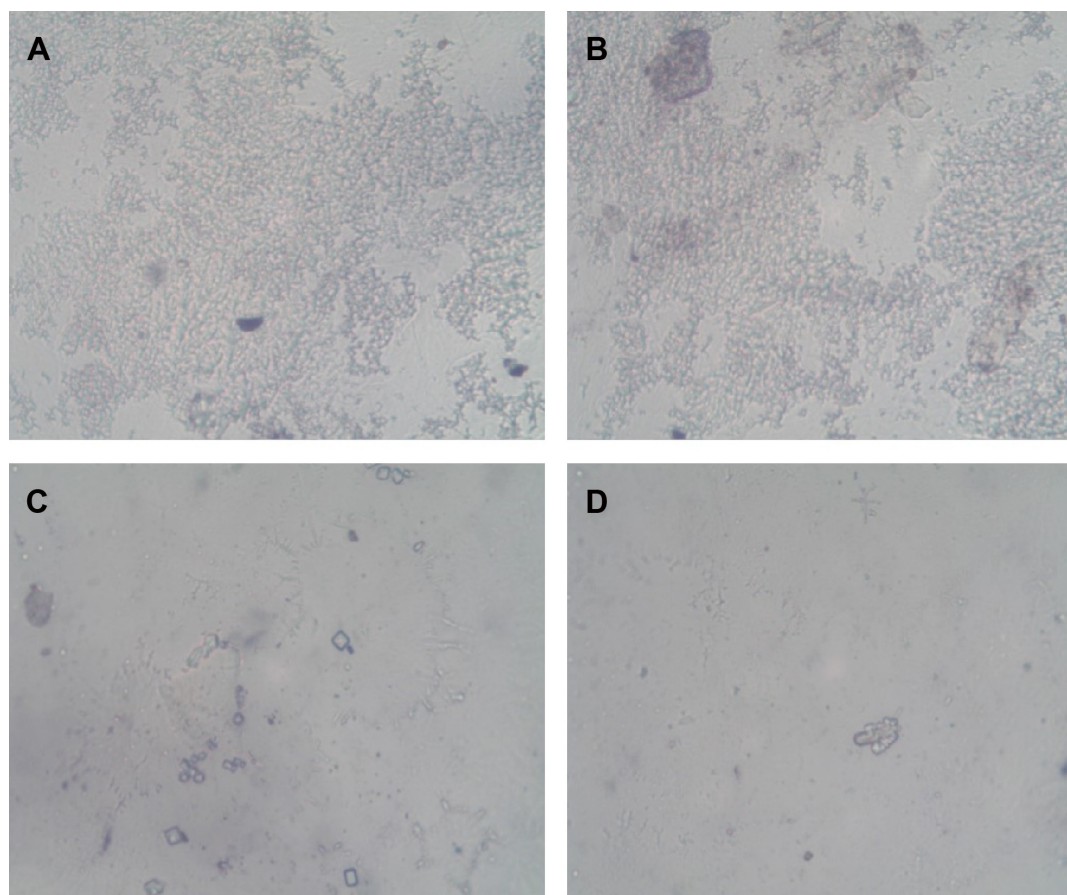


Figure 12 Light microscopic images showing adhered platelets on control and extract-coated mPE.

Notes: (A), (B) Pristine mPE, (C) A-12 h-mPE, (D) A-24 h-mPE.

Abbreviations: mPE, metallocene polyethylene; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

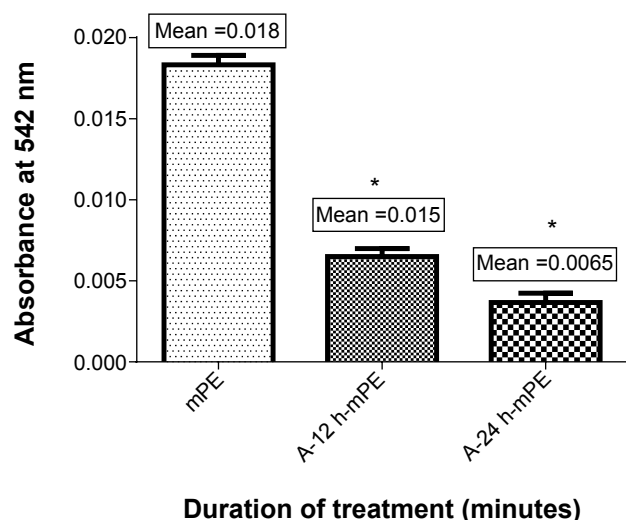


Figure 13 Comparison of absorbance of mPE and *Aloe vera*-coated metalocene polyethylene (n=3).

Notes: Values are shown in mean \pm SD, and differences in the mean are significant ($P < 0.05$). *Differences between the mean values is significant with respect to control ($P < 0.05$).

Abbreviations: mPE, metalocene polyethylene; A-12 h-mPE, 12 hours *Aloe vera*-coated; A-24 h-mPE, 24 hours *Aloe vera*-coated.

acquire the ability to avert expected complications in the biological environment.

Conclusion

Research on exploring novel materials and surface modification techniques in the field of biomaterials has been going on for decades. In this study, the biocompatibility of mPE, a polymer in clinical utilization, is improved by coating with *Aloe vera* extract with the help of microwaves. The results of surface characterization studies such as FTIR, contact angle, SEM, Hirox 3D, and AFM analysis demonstrate the achievement of optimum wettability, surface energy, and roughness followed by the coating of dense fibrous *Aloe vera* extract. Further, the importance of the microwave treatment in influencing the surface morphology and improving the coating thickness is also demonstrated using SEM micrographs and Hirox 3D images. Ultimately, the active biomolecules of *Aloe vera* present on the coated surface has enriched the blood compatibility of mPE as demonstrated through APTT, PT, platelet adhesion, and HA studies. Hence, from the results obtained in this work, it is concluded that *Aloe vera*-glazed mPE surfaces have great potential to support and guide the tissue rehabilitation process. Further, this cost-effective system can be plausibly employed for multifaceted tissue engineering and biomedical applications, especially in the field of maxillofacial, orthopedic, and cardiovascular implants.

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Disclosure

The authors declare that they have no conflict of interest in this work.

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