

WISP-1 overexpression upregulates cell proliferation in human salivary gland carcinomas via regulating MMP-2 expression

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Background: WISP-1 is a member of the CCN family of growth factors and has been reported to play an important role in tumorigenesis by triggering downstream events via integrin signaling. However, little is known about the role of WISP-1 in proliferation of salivary gland carcinoma (SGC) cells.

Methods: In this study, we investigated the WISP-1 expression in SGC tissues via immunohistochemical staining, Western blotting assay, and real-time quantitative polymerase chain reaction method, and then evaluated the regulatory role of WISP-1 in the growth of SGC A-253 cells. In addition, the role of MMP-2 in the WISP-1-mediated growth regulation was also investigated.

Results: It was demonstrated that the WISP-1 expression was upregulated at both mRNA and protein levels in 15 of 21 SGC tumor tissues, compared to the non-tumor tissues (five of 21), associated with the lymph node dissection and bone invasion. The in vitro CCK-8 assay and colony-forming assay demonstrated that the exogenous WISP-1 treatment or the WISP-1 overexpression promoted the growth of A-253 cells. In addition, we confirmed that the WISP-1 overexpression upregulated the MMP-2 expression in A-253 cells with the gain-of-function and loss-of-function strategies, and that the MMP-2 knockdown attenuated the WISP-1-mediated growth promotion of A-253 cells.

Conclusion: We found that WISP-1 was overexpressed in the human SGCs, and the WISP-1 overexpression promoted the salivary gland cell proliferation via upregulating MMP-2 expression. Our study recognized the oncogenic role of WISP-1 in human SGCs, which could serve as a potential target for anticancer therapy.

Keywords: WISP-1, SGCs, proliferation, MMP-2

Background

The incidence of head and neck carcinomas, including salivary gland carcinomas (SGCs), is increasing worldwide, while the survival rate of patients with head and neck carcinoma has not substantially improved over the last 2 decades¹ due to the heterogeneity of the carcinoma.² Thus, novel therapies are urgently needed to supplement the treatment currently available for SGCs. SGCs account for ~3%–6% of all head and neck malignancies.^{3,4} In recent years, multiple molecular markers, such as PTTG, CDKN2A, TP53, and EGFR, have been found to be prognostic of SGCs.^{5–8} In particular, the EGFR positivity has been recognized in ~82%–91% of SGC samples.^{6,9}

The CCN family of growth factors derived their name from the acronym of the first three members of the family: *cyr61*, *CTGF*, and *NOV*.¹⁰ They share properties including stimulation of cell proliferation, migration, adhesion, and extracellular matrix (ECM)

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formation, as well as angiogenesis and tumorigenesis.^{11,12} WISP-1, also known as CCN4, is a cysteine-rich, secreted matricellular protein.¹³ WISP-1 is a downstream target of Wnt-1 and β -catenin.¹⁴ Recently, the deregulation of WISP-1 has been indicated to result in various pathologies including osteoarthritis,¹⁵ fibrosis,¹⁶ and cancers.^{17,18} However, the exact role of WISP-1 in tumorigenesis is not clear. Although the WISP-1 overexpression correlates with metastasis of these cancers and hence poor overall survival, it is not clear whether the oncogenic molecule contributes to tumorigenesis of SGCs.

In the present study, we evaluated the WISP-1 overexpression in the human SGC specimens by immunohistochemical analysis, Western blot analysis, and real-time quantitative polymerase chain reaction (RT-qPCR). To investigate the regulatory role of WISP-1 in the proliferation of SGC cells, we overexpressed WISP-1 in A-253 cells or treated A-253 cells with exogenous WISP-1, and then investigated the influence of WISP-1 on the proliferation of A-253 cells. In some specimens, we knocked down WISP-1 with WISP-1-specific siRNA, and reevaluated the proliferation of A-253 cells. In addition, the regulation by WISP-1 of the expression of MMP-2 in A-253 cells was investigated. It was implied that WISP-1 plays a regulatory role in the proliferation of salivary gland cells, and could serve as a potential target for anticancer therapy.

Materials and methods

SGC specimens, A-253 cell culture, and treatment

Our study was approved by the Medical Ethics Committee of the First Affiliated Hospital of Henan University of Science and Technology. Twenty-one human SGC specimens and 21 control salivary gland specimens (as control) were collected by surgical resection, before any radiotherapy or chemotherapy, from SGC patients or from salivary gland adenoma patients. Each patient provided written informed consent for the use of specimens. Clinicopathological data of patients were recorded prospectively, including age, sex, tumor size, and other items (Table 1). Human salivary gland A-253 cell line was purchased from American Type Culture Collection (ATCC) (Manassas, VA, USA) and was cultured in McCoy's 5a Medium Modified (Thermo Fisher Scientific, Waltham, MA, USA), which was supplemented with 10% fetal bovine serum (Thermo Fisher Scientific), at 37°C in a humidified incubator with 5% CO₂. Human recombinant WISP-1 (R&D Systems China, Shanghai, People's Republic of China) was dissolved in McCoy's 5a Medium Modified

Table 1 Association of WISP-1 mRNA level with the clinicopathologic characteristics of patients with salivary gland carcinoma

Clinicopathologic characteristics	Cases (N)	Expression of WISP-1	
		mRNA level	P-value
Age (years)			0.4365
≤55	12	1.7642±0.2104	
>55	9	1.6094±0.1836	
Sex			0.6217
Male	10	1.7856±0.2031	
Female	11	1.6184±0.1853	
Tumor size (cm)			0.1126
≤2.5	13	1.6178±0.1726	
>2.5	8	1.8275±0.2132	
Lymph node dissection			0.0213
No	14	1.5719±0.1705	
Yes	7	1.9476±0.2342	
Bone invasion			0.0182
No	16	1.5928±0.1823	
Yes	5	2.0336±0.2417	

and was utilized at a final concentration of 0, 10, or 30 ng/mL to treat A-253 cells. To overexpress WISP-1 in A-253 cells, the wild-type human WISP-1-coding sequence was amplified and cloned into the pcDNA3.1(+) vector. The expression of WISP-1 at both mRNA and protein levels in A-253 cells was examined at 0, 24, 48, or 72 hours post-transfection with the WISP-1-pcDNA3.1(+) or control Ctrl-pcDNA3.1(+) plasmids (which expressed enhanced green fluorescent protein). Lipofectamine 2000 (Thermo Fisher Scientific) was utilized to transfect plasmids into A-253 cells. To knock down WISP-1, siRNA-WISP-1 or control siRNA (siRNA-Ctrl) (GenePharma, Shanghai, People's Republic of China) was transfected with Lipofectamine RNAiMax (Thermo Fisher Scientific) into A-253 cells at a concentration of 30 or 60 nM.

Immunohistochemical staining for WISP-1 in SGC tissues

Paraffin-embedded slides of SGC tissues were successively deparaffinized by heating at 55°C, rehydrated serially with different concentrations of ethanol, and then were prepared for the antigen retrieval by heating at 98°C in 10 mM sodium citrate (pH 6.0). The endogenous peroxidase activity was blocked with 0.3% hydrogen peroxide for 20 minutes. The rabbit polyclonal antibody to human WISP-1 (Cat PA5-29150; Thermo Fisher Scientific) was utilized to stain the WISP-1. After the incubation with the primary WISP-1 antibody at room temperature for 2 hours, the slides were incubated with goat anti-rabbit IgG-HRP (Cat sc-2004; Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA) at room

temperature for 1 hour. Then, the slides were incubated with Pierce™ DAB Substrate Kit (Thermo Fisher Scientific) for several minutes to stain the WISP-1. Each slide was dehydrated and sealed with coverslips.

RT-qPCR analysis

mRNA from SGC tissues or from A-253 cells was isolated with the PolyAttract® mRNA Isolation Kit (Promega Corporation, Fitchburg, WI, USA) under the guidance of the kit's manual, and each mRNA sample was treated with 1 μ L RNasin® Plus RNase Inhibitor (Promega Corporation). mRNA samples were stored at -80°C before use. RT-qPCR was performed using an iTaq™ Universal one-step RT-qPCR Kit (Bio-Rad Laboratories Inc., Hercules, CA, USA), with β -actin as an internal control. The primers for WISP-1 were 5'-CTG CAT CAG CAC ACG CTC CT-3' (forward) and 5'-TGT AGG GGA TGC AGC ACC TAT-3' (reverse), and primers for β -actin were 5'-GCT GCT GAC CGA GGC CCC CCT-3' (forward) and 5'-GAA GGT CTC AAA CAT GAT CT-3' (reverse). All primers were synthesized by Shanghai Sangon company (Shanghai, People's Republic of China). The WISP-1 mRNA level was calculated as the fold change compared to the internal β -actin control with the $\Delta\Delta\text{Ct}$ method.¹⁹

Western blot analysis

WISP-1 protein level in tumor specimens or in A-253 cells was relatively quantified by Western blotting. The homogenized SGC specimens or cultured cells were isolated with the NE-PER™ Nuclear and Cytoplasmic Extraction Reagents (78833; Thermo Fisher Scientific), under the guidance of the kit's manual. Each protein sample was treated with a protease inhibitor cocktail (Sigma-Aldrich, St Louis, MO, USA) and was stored at -80°C before use. Electrophoresis with 12% sodium dodecyl sulfate-polyacrylamide gel was performed to separate the protein bands, which were then transferred to a polyvinylidene fluoride hydrophobic membrane (EMD Millipore, Billerica, MA, USA). The band of WISP-1 or β -actin was specifically bound by the polyclone rabbit anti-WISP-1 (Abcam, Cambridge, UK) or anti- β -actin antibody (Abcam), and was subjected to binding with the peroxidase-conjugated secondary antibody (Bio-Rad Laboratories Inc.), and detected via coloration with the electrochemiluminescence detection system (Thermo Fisher Scientific).

Cell proliferation and cell colony formation

CCK-8 assay (Dojindo, Kumamoto, Japan) was performed to examine the proliferation of A-253 cells. Briefly, A-253

cells that were ~85% confluent after the WISP-1 treatment or the siRNA (siRNA-Ctrl or siRNA-MMP-2) transfection were cultured at 37°C in a humidified incubator with 5% CO_2 for 12, 24, or 48 hours. Then, cells were incubated with CCK-8 until visual color appeared, and their absorbance at 450 nm was calculated. Colony formation assay for A-253 cells was performed as follows: A-253 cells (500 per well) were incubated in 12-well plates at 37°C in the presence of 5% CO_2 , and were treated with 0, 10, or 30 ng/mL WISP-1, or were transfected with WISP-1-pcDNA3.1(+) or Ctrl-pcDNA3.1(+) plasmid (500 ng per well), or siRNA-Ctrl or siRNA-MMP-2 (50 nM). Then, cells were incubated for another 48 hours, and were stained with crystal violet (0.005%) for 20 minutes to record the colony numbers.

Statistical analysis

All data are expressed as mean \pm standard error of the mean. The difference in WISP-1 positivity between the two groups was evaluated by chi-square test. The expression of WISP-1 or MMP-2 at mRNA or protein level, or the cell number of the two groups was analyzed by Student's *t*-test. The difference in the CCK-8 assay results was analyzed using analysis of variance. A *P*-value of ≤ 0.05 was considered statistically significant.

Results

Overexpression of WISP-1 in SGC tissues

To recognize the role of WISP-1 in SGC, we examined the WISP-1 expression in SGC tissues. First, immunohistochemical staining was performed to analyze the WISP-1 expression in situ in SGC (N=21) or control salivary gland tissues (N=21). Figure 1A and B demonstrates the positive staining for WISP-1 in SGC tissues (positivity =15/21), which was significantly higher than in the normal salivary gland tissues (positivity =5/21) ($P=0.002$). Then, Western blotting was performed to quantify the protein level of WISP-1 in the 15 WISP-1-positive SGC specimens. As indicated in Figure 1C, the relative protein level of WISP-1 compared to β -actin was also significantly higher in the SGC group ($P<0.001$, paired-samples *t*-test; Figure 1D). Finally, we performed the RT-qPCR assay to calculate the WISP-1 mRNA level in both groups. Figure 2A demonstrates that the relative mRNA level of WISP-1 in SGC specimens was 1.667 ± 0.315 (N=21), which was significantly higher than 1.000 ± 0.146 in control group (N=21) ($P<0.001$). Also, the relative mRNA level of WISP-1 was higher in five of six SGC specimens, compared to the peri-SGC specimens ($P<0.05$ or $P<0.01$; Figure 2B).

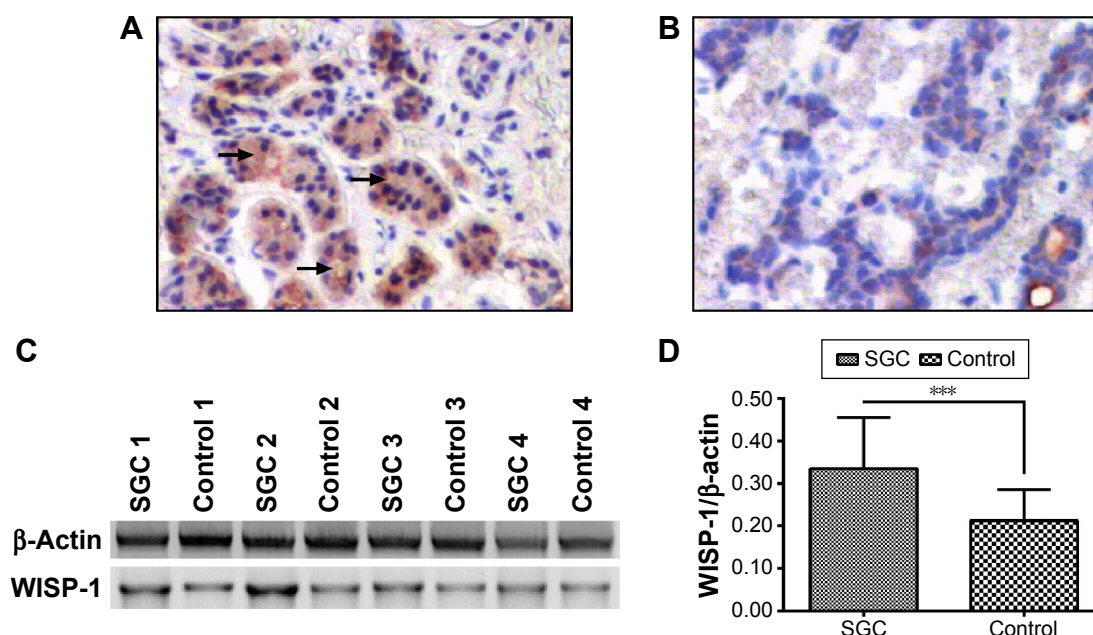


Figure 1 Immunohistochemical staining for WISP-1 expression in human salivary gland carcinoma tissues.

Notes: Representative WISP-1 staining in human salivary gland carcinoma (A, N=21) or control salivary gland tissues (B, N=21). (C) Representative Western blotting assay of WISP-1 protein level in control or human salivary gland carcinoma tissues. (D) Relative protein level of WISP-1 compared to β-actin in the control (N=15) or human salivary gland carcinoma tissues (N=15). (A) and (B) were ×400 under light microscope. (A) Positive staining was indicated by the black arrows. ***P<0.001.

Abbreviation: SGC, salivary gland carcinoma.

In addition, we associated the WISP-1 mRNA level with the clinicopathologic characteristics of SGC patients. As indicated in Table 1, there was a marked association of the WISP-1 overexpression with the lymph node dissection or bone invasion ($P=0.0213$ or $P=0.0182$). Taken together, we found that WISP-1 was overexpressed in SGC specimens.

WISP-1 promotes the proliferation of human salivary gland A-253 cells

To investigate the regulatory role of WISP-1 in SGC cells, we treated A-253 cells with 0, 10, or 30 ng/mL recombinant human WISP-1, and then examined the proliferation of A-253

cells. As indicated in Figure 3A, there was a significant difference in the growth curve of the A-253 cells after the treatment with 0, 10, or 30 ng/mL WISP-1 ($P<0.05$ or $P<0.01$ for 24 or 48 hours posttreatment, respectively). The colony-forming assay was also performed to examine the difference in the proliferation of WISP-1-treated or WISP-1-untreated A-253 cells. Results demonstrated that there were more colonies in the group of A-253 cells, which were treated with 10 or 30 ng/mL WISP-1, than in the A-253 cells not treated with WISP-1 ($P<0.01$; Figure 3B and C). Then, we overexpressed WISP-1 in A-253 cells via transfecting with the recombinant WISP-1-pcDNA3.1(+) plasmid. Figure 4A indicates that the

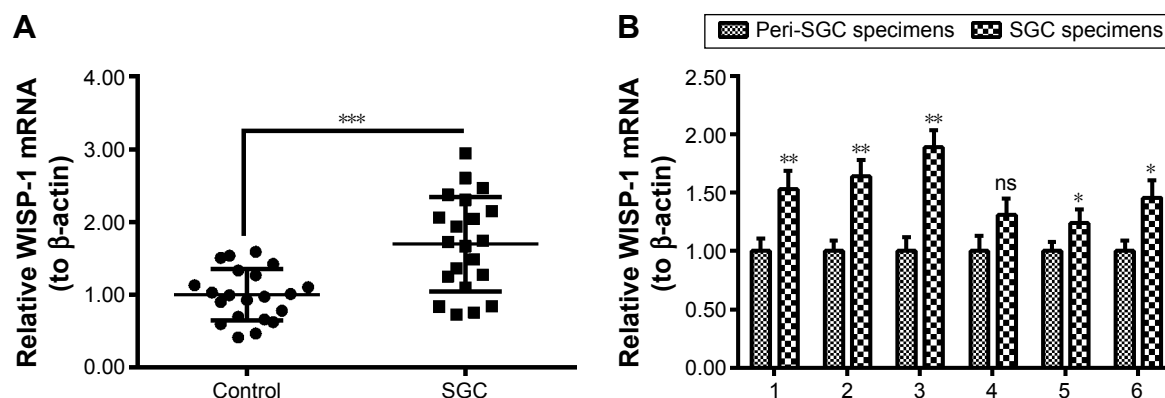


Figure 2 mRNA level of WISP-1 in human salivary gland carcinoma tissues.

Notes: (A) RT-qPCR analysis of the WISP-1 mRNA level in the control (N=21) or human salivary gland carcinoma (N=21) tissues. (B) Relative WISP-1 mRNA level in the human salivary gland carcinoma (N=6) or peri-tumor (N=6) tissues. β-Actin was taken as an internal control. *P<0.05, **P<0.01, and ***P<0.001.

Abbreviations: RT-qPCR, real-time quantitative polymerase chain reaction; SGC, salivary gland carcinoma; ns, no significance.

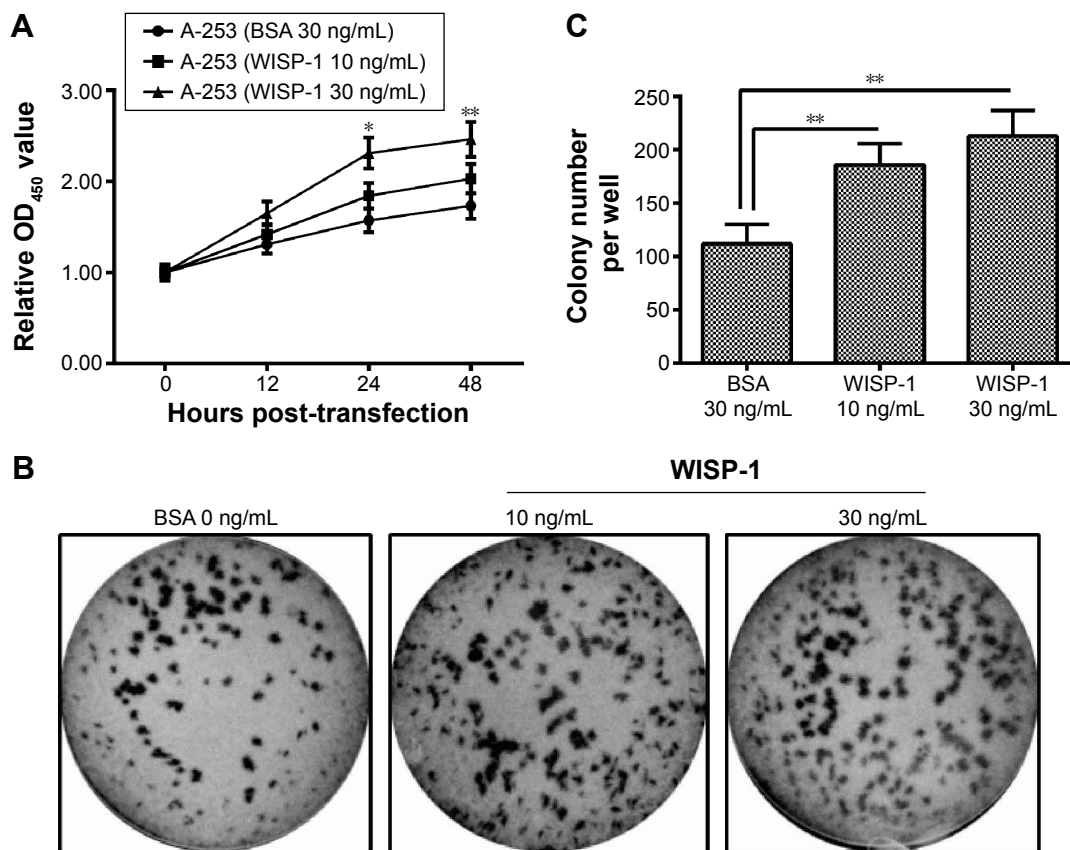


Figure 3 Exogenous recombinant human WISP-1 promotes the growth of A-253 cells.

Notes: (A) CCK-8 assay for the growth curve of the A-253 cells, which were treated with 0, 10, or 30 ng/mL for 0, 12, 24, or 48 hours. (B) Colony-forming assay and (C) colony counting for the A-253 cells, which were treated with 0, 10, or 30 ng/mL for 48 hours. Results were averaged for triple independent experiments. * $P < 0.05$ and ** $P < 0.01$.

Abbreviations: OD, optical density; BSA, bovine serum albumin.

WISP-1 mRNA level was significantly upregulated in A-253 cells after the WISP-1-pcDNA3.1(+) transfection at 24, 48, or 72 hours ($P < 0.001$). The Western blotting result (Figure 4B) also indicated that the protein level of WISP-1 was upregulated in the WISP-1-pcDNA3.1(+)-transfected A-253 cells at 24, 48, or 72 hours ($P < 0.001$; Figure 4C). Moreover, the colony-forming assay also demonstrated that more colonies were formed by the A-253 cells, which were transfected with WISP-1-pcDNA3.1(+) (A-253 WISP-1 Up), than those which were transfected with the control plasmid (A-253 Ctrl Up) ($P < 0.01$; Figure 4D and E). Thus, our results found the promotive role of WISP-1 in the proliferation of A-253 cells.

WISP-1 upregulates MMP-2 expression in A-253 cells

WISP-1 has been indicated to promote the MMP-2 expression in various types of tumor cells.^{20,21} In the present study, we investigated the regulation by WISP-1 of the MMP-2 expression in A-253 (WISP-1 Up) cells. Figure 5A demonstrates that the MMP-2 mRNA was markedly higher in the A-253 cells transfected with the WISP-1-pcDNA3.1(+)

at 24, 48, or 72 hours ($P < 0.05$ or $P < 0.01$). The Western blotting assay also indicated that the protein level of MMP-2 was significantly higher in the A-253 (WISP-1 Up) cells than in the A-253 (Ctrl Up) cells, at 24, 48, or 72 hours post-transfection ($P < 0.01$; Figure 5B). Then, we knocked down WISP-1 with siRNA transfection in A-253 (WISP-1 Up) cells. Figure 5C indicates the marked downregulation of WISP-1 mRNA ($P < 0.01$). Moreover, the WISP-1 knockdown also reduced the MMP-2 mRNA level in A-253 (WISP-1 Up) cells ($P < 0.05$; Figure 5D). The WISP-1 knockdown and the MMP-2 downregulation were also confirmed at protein levels by the Western blotting assay ($P < 0.05$, $P < 0.01$, or $P < 0.001$; Figure 5E and F). Taken together, we found that WISP-1 promoted the MMP-2 expression in salivary gland A-253 cells.

MMP-2 knockdown inhibits the WISP-1-mediated growth promotion of A-253 cells

In order to examine the role of MMP-2 in the WISP-1-mediated growth promotion of A-253 cells, we knocked

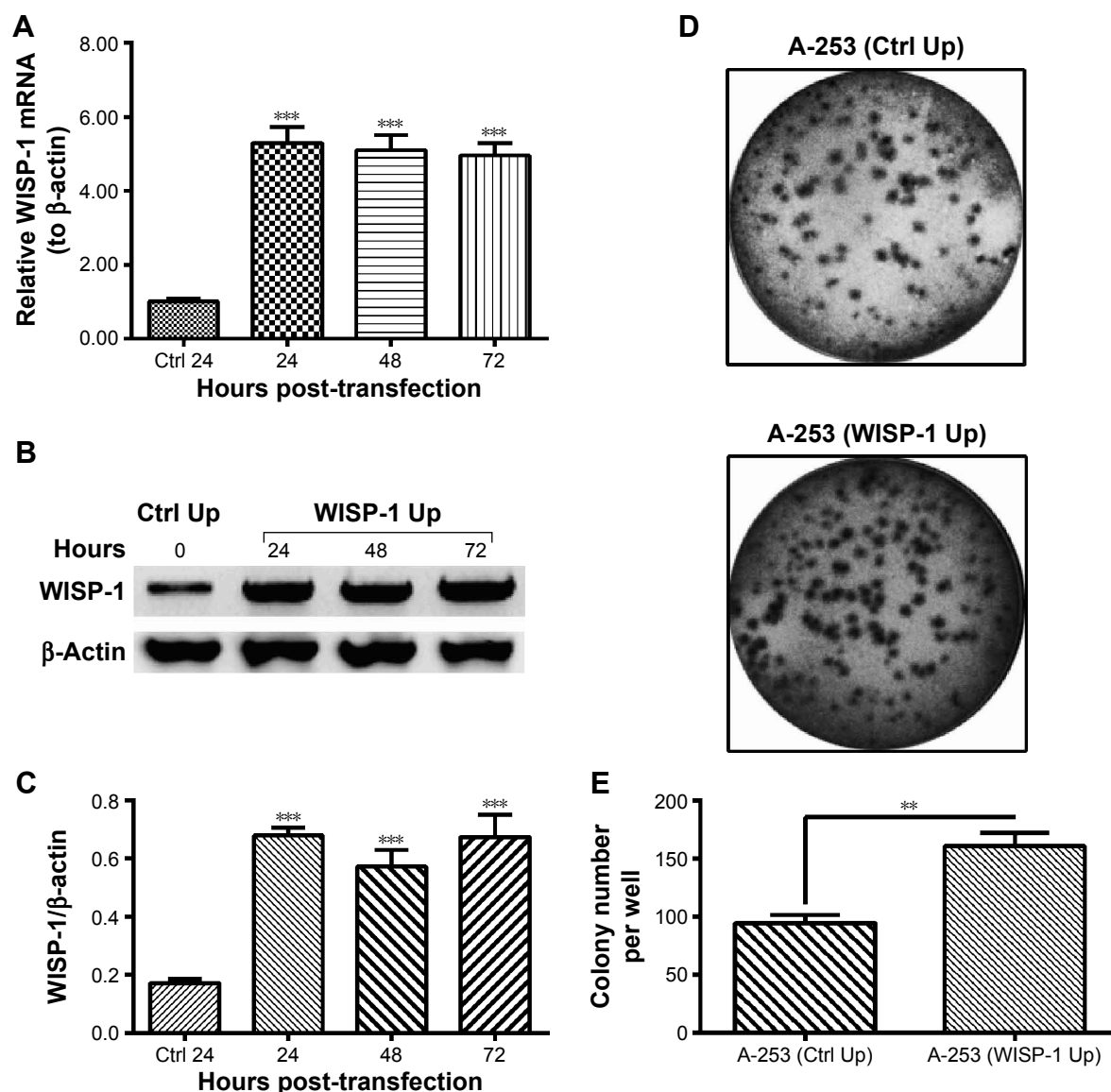


Figure 4 Overexpressed WISP-1 promotes the growth of A-253 cells.

Notes: (A) RT-qPCR analysis of the WISP-1 mRNA in the A-253 cells, which were transfected with Ctrl-pcDNA3.1(+) plasmid for 24, 48, or 72 hours. (B) Western blotting assay of WISP-1 in the Ctrl-pcDNA3.1(+) or WISP-1-pcDNA3.1(+) transfected A-253 cells for 24, 48, or 72 hours. (C) WISP-1 was presented as a relative level to β-actin. Colony-forming assay (D) and colony counting (E) for the Ctrl-pcDNA3.1(+) or WISP-1-pcDNA3.1(+) transfected A-253 cells for an inoculation of 48 hours. Experiments were performed independently in triplicate. Statistical significance is shown as ** $P < 0.01$ and *** $P < 0.001$.

Abbreviations: RT-qPCR, real-time quantitative polymerase chain reaction; Ctrl, control.

down MMP-2 and reevaluated the growth of A-253 (WISP-1 Up) cells. Figure 6A confirms the knockdown of MMP-2 at mRNA level in the A-253 (WISP-1 Up) cells after the transfection with 30 or 60 nM siRNA-MMP-2 ($P < 0.01$ or $P < 0.001$). Then, the growth curve and the colony formation of A-253 (WISP-1 Up) cells after MMP-2 knockdown were reexamined. Figure 6B indicates a reduced growth of the A-253 (WISP-1 Up) cells after the transfection with 60 nM siRNA-WISP-1. There were less colonies in the group of A-253 (WISP-1 Up) cells after MMP-2 knockdown (60 nM) ($P < 0.05$; Figure 6C and D). All results indicated that MMP-2

might regulate the WISP-1-mediated growth promotion of A-253 cells.

Discussion

SGCs represent a group of tumors with morphologically remarkable heterogeneity²² and are characterized by loco-regional invasion and metastasis.²³ The heterogeneous cellular make-up poses challenges for the awareness of the tumorigenesis of SGCs, and for the diagnosis and treatment of the carcinoma. Challenge for SGC diagnosis hinders the improvement in SGC prognosis. Several deregulated

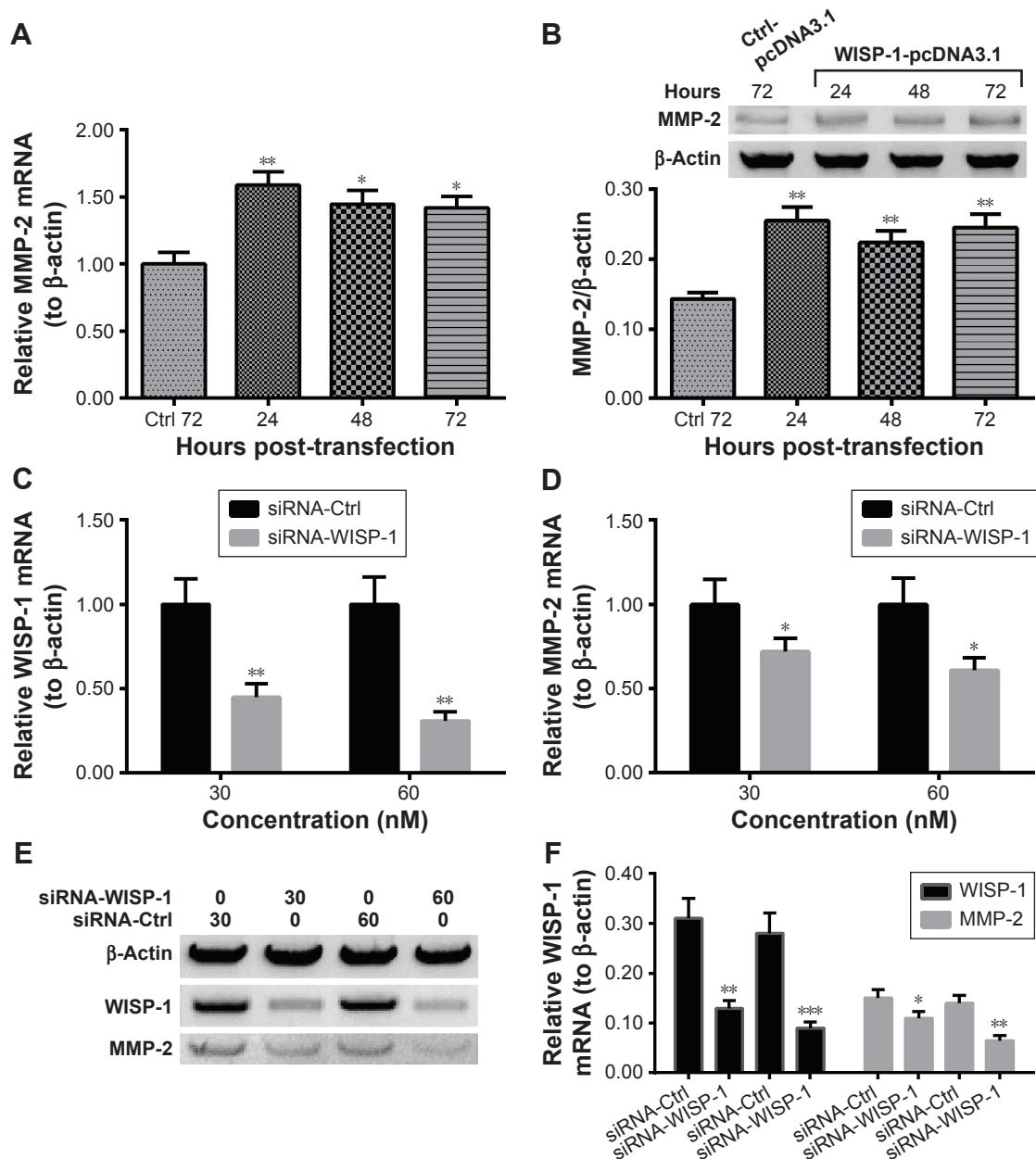


Figure 5 Overexpressed WISP-1 upregulates the expression of MMP-2 in A-253 cells.

Notes: (A) RT-qPCR analysis of the MMP-2 mRNA in the A-253 cells, which were transfected with Ctrl-pcDNA3.1(+) plasmid for 24 hours, or with WISP-1-pcDNA3.1(+) plasmid for 24, 48, or 72 hours. (B) Western blotting assay of MMP-2 in the Ctrl-pcDNA3.1(+) or WISP-1-pcDNA3.1(+) transfected A-253 cells, at 24, 48, or 72 hours. RT-qPCR analysis of the (C) WISP-1 and (D) MMP-2 mRNA in the A-253 cells, which were transfected with 30 or 60 nM siRNA-Ctrl or siRNA-WISP-1 for 24 hours. (E) Western blotting assay and (F) quantification of WISP-1 and MMP-1 in the siRNA-Ctrl- or siRNA-WISP-1-transfected (60 nM) A-253 cells, at 24, 48, or 72 hours. The quantitative data were averaged for triple independent results. Statistical significance is shown as * $P < 0.05$, ** $P < 0.01$, or *** $P < 0.001$.

Abbreviations: RT-qPCR, real-time quantitative polymerase chain reaction; Ctrl, control.

biomarkers have been recognized in SGCs, such as PTTG, TP53, and EGFR,⁵⁻⁸ in association with their malignancy. In the present study, we found the WISP-1 overexpression at both mRNA and protein levels in SGC tissues, via various methods, including immunohistochemical staining, Western blotting assay, and RT-qPCR. The WISP-1 overexpression was associated with the lymph node dissection or the bone invasion of SGCs.

Deregulation of WISP-1 has been indicated to correlate with various pathologies including osteoarthritis, fibrosis, and cancer.²⁴ Particularly, the impact of WISP-1 has been identified in various types of tumors in processes such as tumor cell proliferation, migration, and survival.²⁵ Recombinant WISP-1 has been confirmed to induce the proliferation of alveolar epithelial cells and adenocarcinomic alveolar basal epithelial cells.²⁶ Overexpressed WISP-1 has been found to

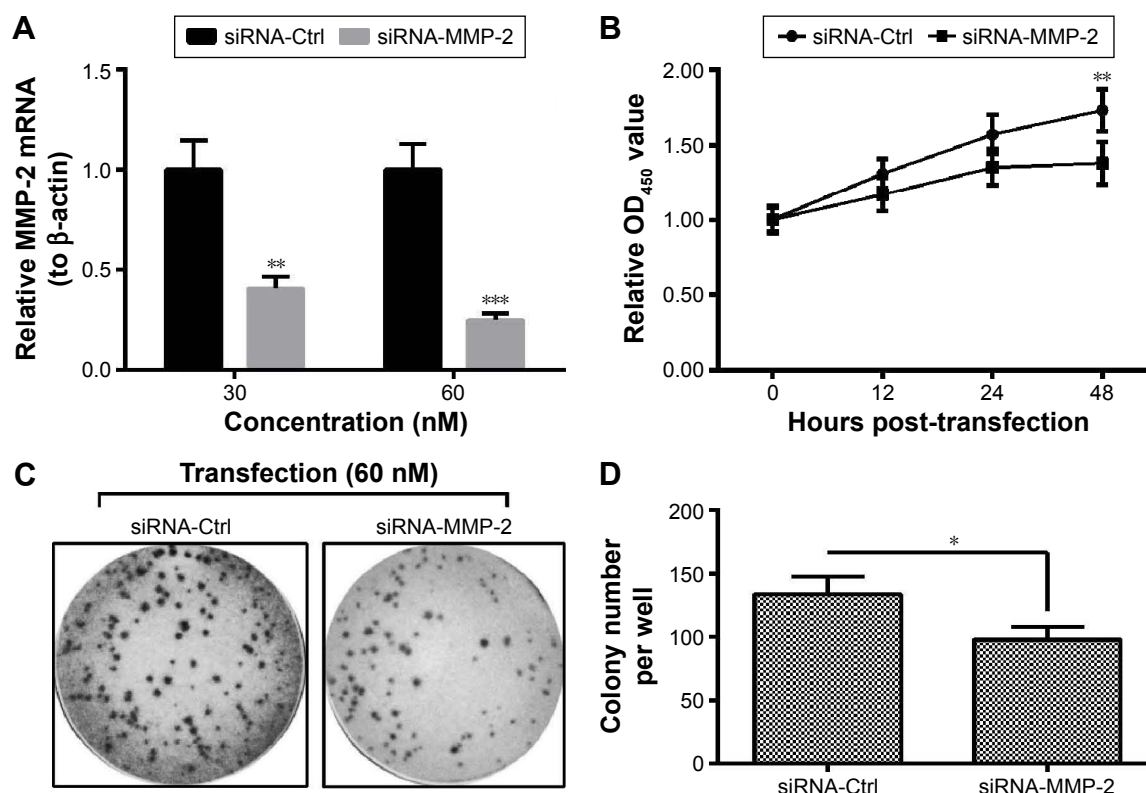


Figure 6 Influence of the MMP-2 knockdown on the growth of A-253 cells.

Notes: (A) RT-qPCR analysis of the MMP-2 mRNA in the A-253 cells, which were transfected with 30 or 60 nM siRNA-Ctrl or siRNA-MMP-2 for 24 hours. (B) CCK-8 assay for the growth curve of the A-253 cells, which were transfected with 60 nM siRNA-Ctrl or siRNA-MMP-2 for 0, 12, 24, or 48 hours. (C) Colony-forming assay and (D) colony counting for the siRNA-Ctrl- or siRNA-MMP-2-transfected (60 nM) A-253 cells for an inoculation of 48 hours. Experiments were performed independently in triplicate. Statistical significance is shown as * $P < 0.05$, ** $P < 0.01$, or *** $P < 0.001$.

Abbreviations: RT-qPCR, real-time quantitative polymerase chain reaction; CCK-8, Cell Counting Kit; Ctrl, control.

increase the growth rate of mouse embryonic fibroblasts²⁷ and human esophageal cancer cells.²⁸ On the other side, WISP-1 has been indicated to be inhibitory to the growth of melanoma cells.^{29,30} The migration and invasion of cells have also been regulated by WISP-1 in gastric carcinoma,²⁷ cholangiocarcinoma,²¹ and prostate cancer.³¹ Our study demonstrated that the overexpressed WISP-1 promoted the growth of salivary gland A-253 cells by results of both growth curve and colony-forming assay. Both exogenous WISP-1 treatment and intracellular overexpression of WISP-1 exerted such promotion on the A-253 cell proliferation.

MMPs are a family (>20 members) of zinc-containing enzymes,³² which play an important role in remodeling of the ECM under physiological situations and in the morphogenesis of salivary glands.³³ Overexpression of MMPs, particularly MMP-2 or MMP-9, promotes the breakdown of ECM which occurs in many pathological conditions such as proliferation, invasion, and metastasis of cancers.³⁴ MMP-2 and MMP-9 have been implicated in the progression, invasion, and metastasis of various types of cancers.^{35–37} MMP-2 expression in particular seems to be related to the invasive properties and

the malignant potential of SGCs.^{33,38,39} Moreover, it was indicated that the MMP-2 expression was regulated by WISP-1.²¹ Therefore, we speculated that the WISP-1 overexpression and its promotion of SGC cell proliferation might be mediated by MMP-2. Our results confirmed the upregulation by WISP-1 of the MMP-2 expression in salivary gland A-253 cells. On the other side, the knockdown of WISP-1 reduced the MMP-2 expression in the WISP-1-overexpressed A-253 cells. In addition, the MMP-2 knockdown inhibited the WISP-1-promoted A-253 cell proliferation. However, the association of WISP-1 with other types of MMPs in SGC is not clear. Also, the regulatory role of such association in the SGC progression is not known. This should be explained by further investigations.

Conclusion

We found that WISP-1 was overexpressed in the human SGCs, and the WISP-1 overexpression promoted the salivary gland cell proliferation and upregulated MMP-2 expression. Moreover, MMP-2 knockdown inhibited the WISP-1-mediated growth promotion of A-253 cells. Our study recognized the

oncogenic role of WISP-1 in human SGCs, which could serve as a potential target for anticancer therapy.

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Disclosure

The authors report no conflicts of interest in this work.

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