

Redox-sensitive Pluronic F127-tocopherol micelles: synthesis, characterization, and cytotoxicity evaluation

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Abstract: Pluronic F127 (F127), an amphiphilic triblock copolymer, has been shown to have significant potential for drug delivery, as it is able to incorporate hydrophobic drugs and self-assemble into nanosize micelles. However, it suffers from dissociation upon dilution owing to the relatively high critical micelle concentration and lack of stimuli-responsive behavior. Here, we synthesized the α -tocopherol (TOC) modified F127 polymer (F127-SS-TOC) via a redox-sensitive disulfide bond between F127 and TOC, which formed stable micelles at relatively low critical micelle concentration and was sensitive to the intracellular redox environment. The particle size and zeta potential of the F127-SS-TOC micelles were 51.87 ± 6.39 nm and -8.43 ± 2.27 mV, respectively, and little changes in both particle size and zeta potential were observed within 7 days at room temperature. With 10 mM dithiothreitol stimulation, the F127-SS-TOC micelles rapidly dissociated followed by a significant change in size, which demonstrated a high reduction sensitivity of the micelles. In addition, the micelles showed a high hemocompatibility even at a high micelle concentration (1,000 $\mu\text{g/mL}$). Low cytotoxicity of the F127-SS-TOC micelles at concentrations ranging from 12.5 $\mu\text{g/mL}$ to 200 $\mu\text{g/mL}$ was also found on both Bel 7402 and L02 cells. Overall, our results demonstrated F127-SS-TOC micelles as a stable and safe aqueous formulation with a considerable potential for drug delivery.

Keywords: Pluronic F127, tocopherol, reduction sensitivity, micelles, hemolysis

Introduction

Copolymers, containing hydrophilic and hydrophobic portions, are promising multifunctional materials for clinical and biological applications. Owing to their amphiphilic chemical property, with a hydrophilic outer shell and a hydrophobic inner core, they are able to self-assemble into small micellar structures in aqueous solutions with a narrow size distribution of 10–200 nm.¹ Because of the small size, nanomicelles tend to sneak into cells, increasing cellular uptake efficiency. Furthermore, nanomicelles also facilitate high tumor accumulation on the basis of enhanced permeability and retention effect.^{1,2} To prove this concept, several polymeric micelles have been developed as drug carriers for cancer chemotherapy.^{3–5} Pluronic F127 (F127, BASF SE, Ludwigshafen, Germany) is one kind of poly(ethylene oxide) (PEO)-poly(propylene oxide) (PPO)-poly(ethylene oxide) (PEO) triblock copolymers, which is accepted as an excipient by the U.S. and British Pharmacopoeia and is widely used in clinical applications.⁶ The amphiphilic chemical structure, including a hydrophobic PPO segment and a hydrophilic PEO segment, allows it to incorporate hydrophobic drugs and self-assemble into micelles with a hydrophilic shell and a hydrophobic core in aqueous medium.^{7,8} Carlson et al⁹ developed the curcumin and resveratrol co-loaded

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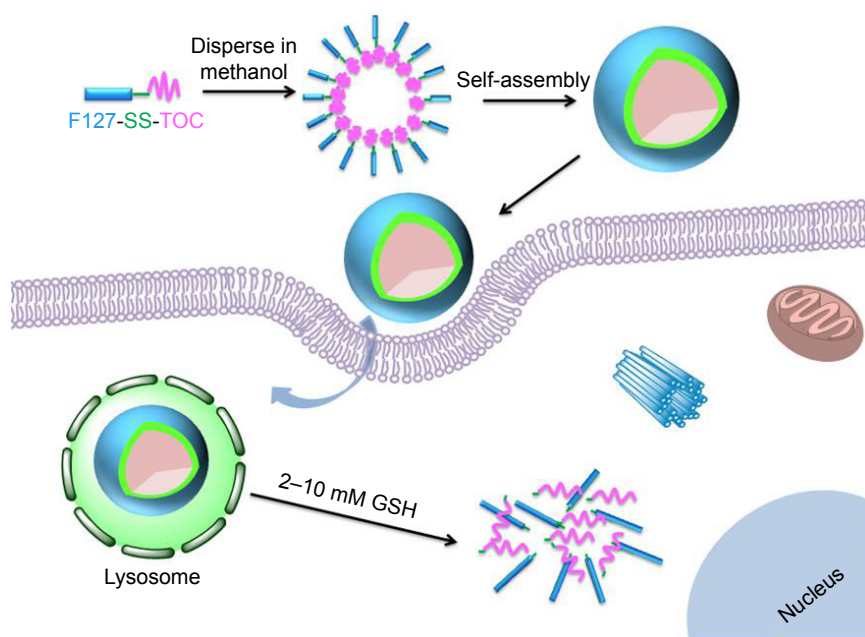


Figure 1 Schematic illustration of self-assembly of F127-SS-TOC micelles and hypothetical redox-sensitive micelle rupture in the presence of 2–10 mM GSH. **Abbreviations:** F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; GSH, glutathione; TOC, α -tocopherol.

micelles based on F127, simultaneously coadministered with doxorubicin hydrochloride, which showed cardioprotective effects and antiproliferative effect *in vitro*. Although promising, F127 micelles in the blood circulation have one limitation: their relatively high critical micelle concentration (CMC), which prevents them from keeping their structure upon dilution, thereby causing drugs to leak.^{6,10,11}

To overcome its limitations and improve the drug delivery functions of F127, several approaches, including chemical modifications or mixed micelles, have been proposed to overcome these disadvantages.^{12,13} Gao et al⁶ prepared amphiphilic copolymer stearic-coupled F127 nanoparticles to deliver doxorubicin (DOX) for cancer therapy, which had better efficacy compared with free DOX on MCF-7 cells *in vitro*. Saxena et al⁷ developed Gambogic acid-loaded Poloxamer 407/D- α -Tocopheryl polyethylene glycol 1000 succinate mixed micelles for the treatment of breast and multidrug-resistant cancer. α -Tocopherol (TOC), known as vitamin E, is toxic to various cancer cells but not to normal cells.¹⁴ It has a hydrophobic substance consisting of various methylated phenols, making it easy to penetrate into the phospholipid bilayer of the cell membrane.^{14,15} Studies demonstrated that designed functional polymeric micelles were able to stably encapsulate drugs and release them at tumor sites in response to external stimuli such as redox, pH, temperature, and enzyme conditions.^{16–19} In particular, redox-sensitive micelles were designed to trigger burst drug release owing to the high concentration of glutathione (GSH, 4 μ M/g) that is normally found

in tumor tissues.^{20,21} Disulfide bond is stable under normal physiological conditions yet responsive to the reductive environment (eg, GSH) of intracellular fluids in cancer cells, indicating that disulfide bonds can be used as the linker of hydrophilic and hydrophobic portions in the redox-sensitive polymeric micelles for controlling the delivery of intracellular drug.²²

Here, we synthesized a redox-sensitive F127-SS-TOC polymer using a disulfide bond to link F127 and TOC, which formed nanomicellar structures in aqueous solution and ruptured in the presence of glutathione (GSH, 2–10 mM) (Figure 1) owing to a redox reaction. F127-SS-TOC polymer was synthesized and confirmed with ¹H nuclear magnetic resonance spectra (¹H NMR) and Fourier transform infrared spectra (FTIR). The particle size, zeta potential, stability, as well as the reduction sensitivity of the micelles were subsequently evaluated. Then the *in vitro* hemolysis and cytotoxicity of the F127-SS-TOC micelles were investigated and found to have a high hemocompatibility at a high micelles concentration (1,000 μ g/mL) and a low cytotoxicity at concentrations ranging from 12.5 μ g/mL to 200 μ g/mL on both Bel 7402 and L02 cells. These results demonstrated the potential applications of F127-SS-TOC as stable and safe aqueous formulations for a drug delivery system.

Materials and methods

Materials

Pluronic F127 (F127, molecular weight =12,600) was purchased from BASF SE (Ludwigshafen, Germany). Tocopherol

and 3,3'-dithiodipropionic acid were purchased from Aladdin Industrial Corporation (Shanghai, People's Republic of China). Dicyclohexylcarbodiimide (DCC) and 4-dimethylaminopyridine (DMAP) were obtained from GL Biochem., Ltd. (Shanghai, People's Republic of China). Millipore water was prepared with a Milli-Q Plus System (Merck Millipore Co., Billerica, MA, USA). All chemicals were in analytical grade.

Human hepatoma cell line Bel 7402 and human hepatic cell line L02 were supplied by American Type Culture Collection (ATCC, Rockville, MD, USA). Dulbecco's Modified Eagle's Medium, fetal bovine serum, phosphate-buffered saline, penicillin-streptomycin, and 0.25% (w/v) trypsin/1 mM EDTA were obtained from Thermo Fisher Scientific (Waltham, MA, USA).

Synthesis and characterization of F127-SS-TOC polymer

The F127-SS-TOC polymer was synthesized with two-step esterification (Figure 2).²³ F127-SS was prepared by the

conjugation of F127 polymer with 3,3'-dithiodipropionic acid by the esterification reaction. Briefly, F127 polymer (1.0 mmol), 3,3'-dithiodipropionic acid (2.2 mmol), DCC (2.2 mmol), and DMAP (2.2 mmol) were dissolved in 20 mL anhydrous dimethyl sulphoxide (DMSO) and stirred well for 24 h at room temperature. Then the resulting solution was dialyzed against DMSO in a dialysis bag with molecular weight cutoff at 3,500 Da to remove unreacted materials, and the by-product *N,N'*-dicyclohexylurea was removed by filtration. Next, the dialysis bag was transferred into Millipore water to remove DMSO further. The intermediate product F127-SS was obtained by freeze-drying. Subsequently, TOC was conjugated with F127-SS via the esterification reaction between the carboxyl group of F127-SS and the hydroxy group of TOC. Similarly, F127-SS (1.0 mmol), TOC (2.2 mmol), DCC (2.2 mmol), and DMAP (2.2 mmol) were dissolved in 20 mL DMSO with stirring for 24 h at room temperature. The excess of TOC, DCC, and DMAP was removed by dialysis in a dialysis bag (molecular weight

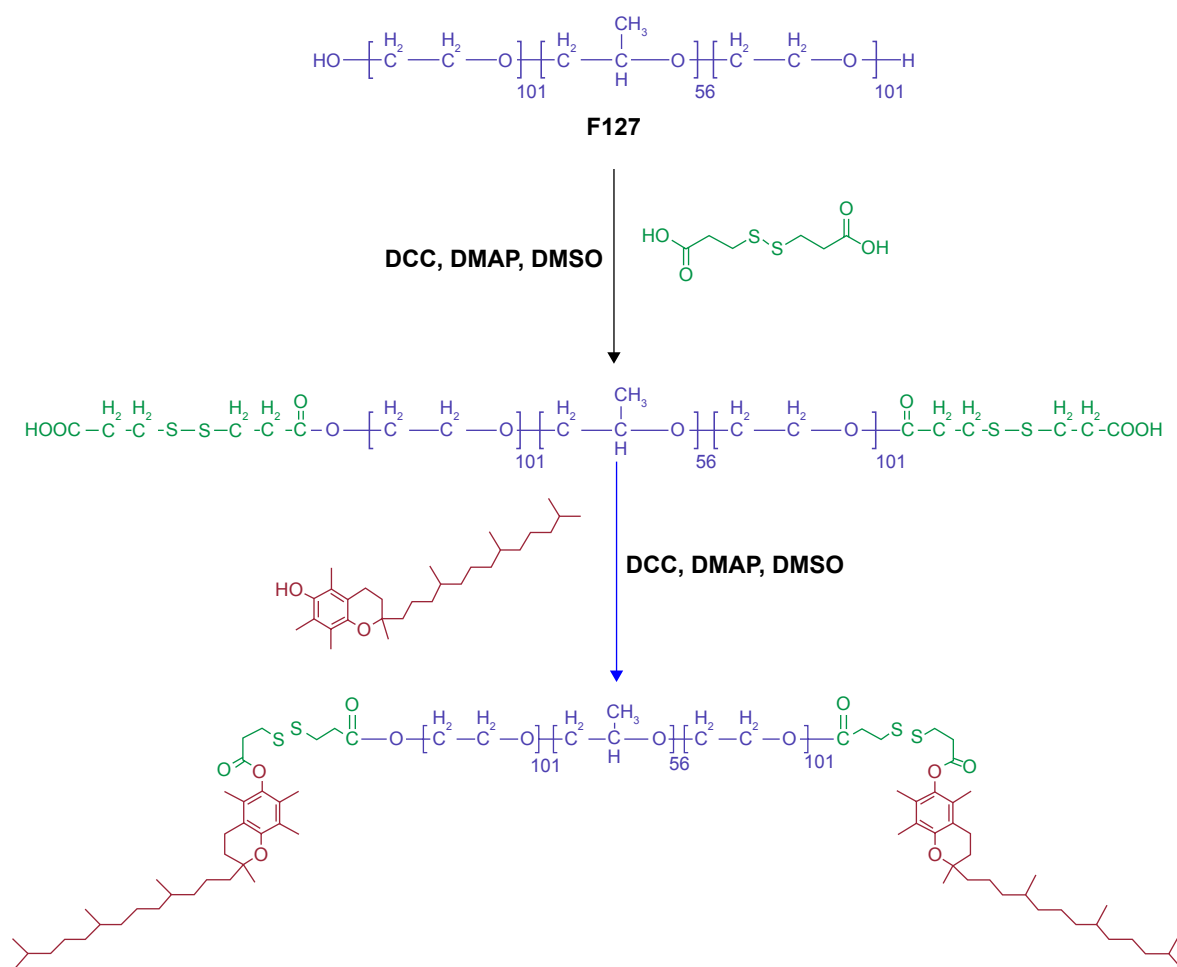


Figure 2 Synthesis route of F127-SS-TOC polymer.

Abbreviations: DCC, dicyclohexylcarbodiimide; DMAP, 4-dimethylaminopyridine; DMSO, dimethyl sulphoxide; F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; TOC, α -tocopherol.

cutoff = 3,500), and by-product *N, N'*-dicyclohexylurea was removed by filtration. F127-SS-TOC polymer was obtained by lyophilization and stored at 4°C for use.

The structure of F127, F127-SS, TOC, and F127-SS-TOC polymer was measured and confirmed by ¹H NMR (Bruker, Karlsruhe, Germany) at 400 MHz in deuterated dimethyl sulfoxide-d₆ (DMSO-d₆) solvent and FTIR (NEXUS 670, Nicolet, Hudson, NH, USA) within a spectrum range of 400–4,000 cm⁻¹.

Preparation and characterization of F127-SS-TOC micelles

F127-SS-TOC micelles were prepared by self-assembly using the solvent evaporation method.²⁴ 15 mg F127-SS-TOC polymer was completely dissolved in 1 mL methanol and added dropwise into distilled water and stirred well for 6 h at room temperature. The residual organic solvent was then removed by vacuum evaporation. The final F127-SS-TOC micelles were obtained by filtering with 0.45 μm microfiltration membrane and preserved at 4°C for use.

Dynamic light scattering (DLS) was employed to measure the average particle size and zeta potential of F127-SS-TOC micelles at room temperature with a Zetasizer Nano ZSP system (Malvern Instruments, Malvern, UK). Micelle morphology was observed and photographed with transmission electron microscopy (TEM, Tecnai G20, FEI Company, Hillsboro, OR, USA). A droplet of micelle sample was placed on the carbon-coated copper grid, followed by staining with phosphotungstic acid solution (2%, w/v). TEM images of the micelles were taken under 200 kV.

The stability of F127-SS-TOC micelles

The stability of F127-SS-TOC micelles was characterized by measuring the particle size and zeta potential for 7 days. During the whole testing period, the F127-SS-TOC micelles were stored and measured at room temperature with DLS.

Fluorescence spectrometer (Lumina, Thermo Scientific) was applied to measure the CMC of F127-SS-TOC micelles using pyrene as a fluorescence probe.²⁵ A series of F127-SS-TOC micellar solutions with various concentrations ranging from 1.45×10⁻⁶ mg/mL to 0.76 mg/mL were prepared in the presence of pyrene at 6×10⁻⁷ M. The pyrene fluorescence of different micellar solutions was measured with an excitation wavelength of 338 nm and an emission wavelength of 380 nm. On formation of micelles, pyrene moved inside the micelles from the aqueous phase, followed by an alteration in the intensity ratio I_{338}/I_{333} in a certain micelle concentration range. In addition, a study based on a

well-known Tyndall phenomenon was used to further evaluate the stability of the F127-SS-TOC micelles.²⁶ In brief, a series of micellar solutions in concentrations ranging from 1.052×10⁻⁴ mg/mL to 1.052 mg/mL were prepared at room temperature and then illuminated with a laser pointer to observe whether the micellar solution showed a visible light beam when irradiated.

The reduction sensitivity of F127-SS-TOC micelles

The redox-sensitive behavior of F127-SS-TOC polymer was associated with the size change of micelles in response to dithiothreitol (DTT) (10 mM). 1 mL DTT (20 mmol/L) was added to 1 mL F127-SS-TOC micelles to form 10 mM DTT concentration. The solution was incubated in a shaker with a rotation speed of 100 rpm at 37°C. The reduction-triggered size change of the F127-SS-TOC micelles was traced by DLS at predetermined intervals.

Hemolysis assay

The hemolytic activity of F127-SS-TOC micelles was carried out in red blood cells from rats following the previous method.²⁷ In order to isolate and collect red blood cells (RBCs), fresh rat blood was diluted by physiological saline and centrifuged at 3,000 rpm for 10 min. RBCs were washed several times until the supernatant was clear. After that, the RBCs were diluted with physiological saline to obtain 2% (v/v) erythrocyte suspension before use. F127-SS-TOC micelles were dissolved in physiological saline at different concentrations, and a suspension of RBCs was added. The mixtures were maintained at 37°C for 3 h in a thermostatic tank, after which they were centrifuged and the supernatant of each sample was collected. The absorbance of supernatant was measured by a microplate reader (Spectra-Max M5, Molecular Devices LLC, Sunnyvale, CA, USA) at 540 nm. Distilled water and physiological saline were used as the positive and negative controls, respectively. The hemolysis rate was calculated by the following equation:

$$\text{Hemolysis rate (\%)} = \frac{A_{\text{sample}} - A_{\text{negative}}}{A_{\text{positive}} - A_{\text{negative}}} \times 100, \quad (1)$$

where A_{sample} , A_{negative} , and A_{positive} represented the absorbance of test samples, negative control, and positive control at 540 nm, respectively. All hemolysis experiments were operated in triplicate.

In vitro cytotoxicity study

The in vitro cytotoxicities of F127-SS-TOC micelles were evaluated on Bel 7402 cells and L02 cells by MTT assay.

The cells were seeded in 96-well plates at a density of 8,000 cells/well in 100 μ L Dulbecco's Modified Eagle's Medium containing 10% fetal bovine serum and incubated at 37°C for 24 h. The cells were incubated with F127-SS-TOC micelles at concentrations ranging from 12.5 μ g/mL to 200 μ g/mL. After 24–48 h incubation at 37°C, 20 μ L of MTT solution (5 mg/mL in phosphate-buffered saline) was added to each well, and the plates were returned to the incubator. The MTT solution was then removed from each well after 4 h. Formazan crystals were dissolved in DMSO. The absorbance of the solution was recorded by a microplate reader at 570 nm. The cell viability (%) was calculated according to the following equation:

$$\text{Cell viability (\%)} = \frac{A_{\text{sample}}}{A_{\text{control}}} \times 100. \quad (2)$$

In equation (2), A_{sample} and A_{control} represented the absorbances of the sample and control wells, respectively, which were measured in triplicate.

Statistical analysis

The data were presented as mean \pm standard deviation. Each value was the mean of at least three repetitive experiments. Statistical analysis was performed using GraphPad Prism 5 software (GraphPad Software, San Diego, CA, USA).

Results and discussion

Synthesis and characterization of F127-SS-TOC polymer

F127-SS-TOC polymer was synthesized and confirmed by ^1H NMR spectra. As shown in Figure 3, typical signals for F127 polymer were observed, including peaks a ($\delta \sim 1.02$ ppm for CH_3 of PPO), b ($\delta \sim 3.36$ ppm for CH of PPO), and c ($\delta = 3.41\text{--}3.54$ ppm for CH_2 of PPO and PEO), which appeared in the F127-SS-TOC polymer. In addition, the typical signals of TOC were also found in F127-SS-TOC polymer, including peaks d ($\delta = 1.95\text{--}2.05$ ppm for CH_3 on the benzene ring), e ($\delta = 1.01\text{--}1.15$ ppm for CH_2 belonged to the repeating units of side chain on TOC polymer), and f ($\delta = 0.75\text{--}0.87$ ppm for CH_3 on the side chain on TOC polymer). The characteristic peaks for CH_2 close to the disulfide bond in the F127-SS and F127-SS-TOC polymer were at 2.87 ppm (peak g) and 2.64 ppm (peak h). Moreover, FTIR spectra were used to further validate the structure of intermediate products and F127-SS-TOC polymer. Figure 4 showed the FTIR spectra of F127, F127-SS, TOC, and F127-SS-TOC polymer. In the spectra of TOC and F127-SS-TOC, sharp bands at $1,258\text{ cm}^{-1}$ were attributed to stretching vibration of --C--O group. In addition, the bands at $3,485$ and $3,475\text{ cm}^{-1}$ were responsible for stretching vibration of --OH in F127 and TOC, respectively. However, the signature peak of hydroxyl did not obviously exist in the F127-SS-TOC polymer, which indicated that this signature peak had disappeared owing to the esterification reaction.

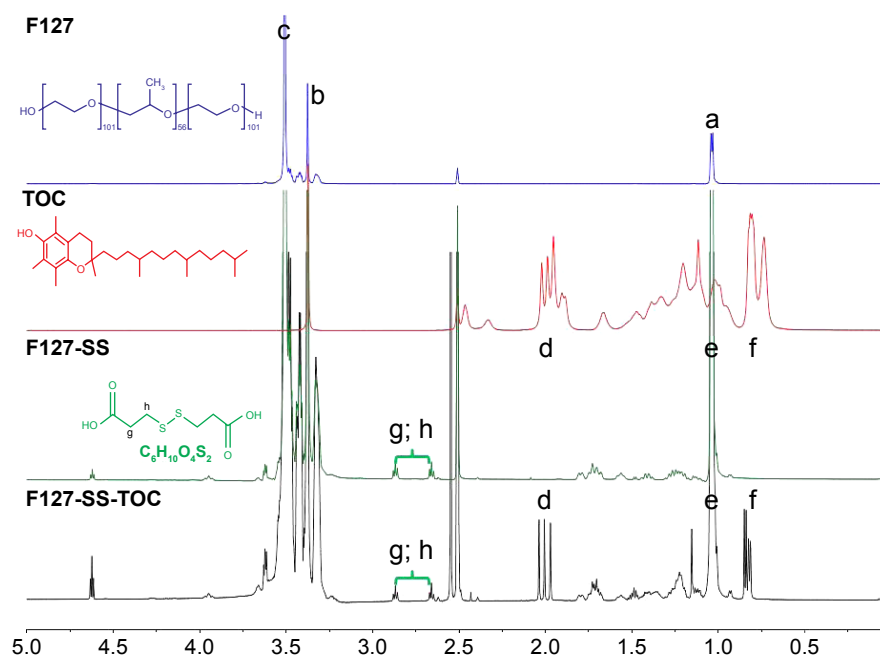


Figure 3 ^1H NMR spectra of F127, TOC, F127-SS, and F127-SS-TOC polymer.

Note: The letters a–h represent the typical signals in F127, TOC, F127-SS, or F127-SS-TOC polymers.

Abbreviations: F127, pluronic F127; F127-SS, 3,3'-dithiodipropionic acid modified F127 polymer; F127-SS-TOC, TOC modified F127 polymer; ^1H NMR, ^1H nuclear magnetic resonance spectra; TOC, α -tocopherol.

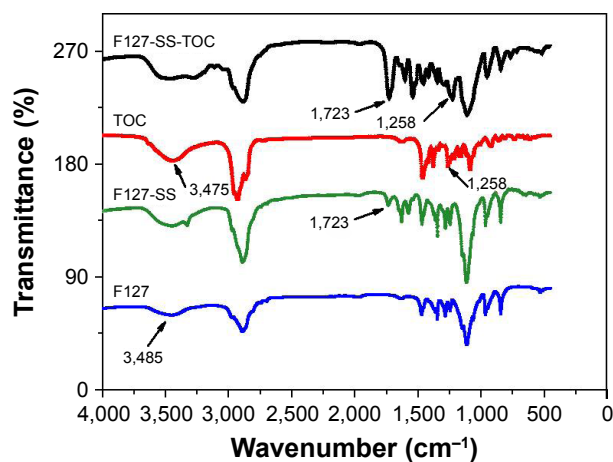


Figure 4 FTIR spectra of F127, F127-SS, TOC, and F127-SS-TOC polymer.
Abbreviations: F127, pluronic F127; F127-SS, 3,3'-dithiodipropionic acid modified F127 polymer; F127-SS-TOC, TOC modified F127 polymer; FTIR, Fourier transform infrared spectra; TOC, α -tocopherol.

Besides, the vibrational peak at $1,723\text{ cm}^{-1}$ represented the peak of C=O in the F127-SS and F127-SS-TOC polymer. Herein, the successful synthesis of the F127-SS-TOC polymer was proved by the ^1H NMR and FTIR spectra.

Preparation and characterization of F127-SS-TOC micelles

In order to better understand the characteristics of F127-SS-TOC micelles, their particle size, zeta potential, and

morphology were evaluated. The synthesized F127-SS-TOC polymer was prepared into micelles with a small particle size of $51.87 \pm 6.39\text{ nm}$ and narrow distribution (PDI: 0.184 ± 0.015) (Figure 5A), measured by DLS. Furthermore, F127-SS-TOC micelles exhibited negative zeta potential of $-8.43 \pm 2.27\text{ mV}$ (Figure 5B), which might be of benefit to the colloidal stability in aqueous solution.²⁸ The morphology of F127-SS-TOC micelles (Figure 5C) was observed with TEM, which showed homogeneous spherical shapes and smooth surfaces. However, the particle sizes shown in TEM images were smaller than the results measured by DLS, which was mainly due to the dehydration during the preparation of the TEM samples.²⁹ These results indicated that the small particle size of F127-SS-TOC micelles might benefit for tumor-specific accumulation by the enhanced permeability and retention effect.

Stability of F127-SS-TOC micelles

Stability is a vital indicator for better understanding of the properties of micelles. As shown in Figure 5D, F127-SS-TOC micelles showed no drastic increases in particle size or zeta potential for 7 days. The average particle sizes were maintained below 100 nm , and the changes in zeta potential were about $\pm 2\text{ mV}$ at room temperature, indicating F127-SS-TOC micelles could remain stable during the storage.

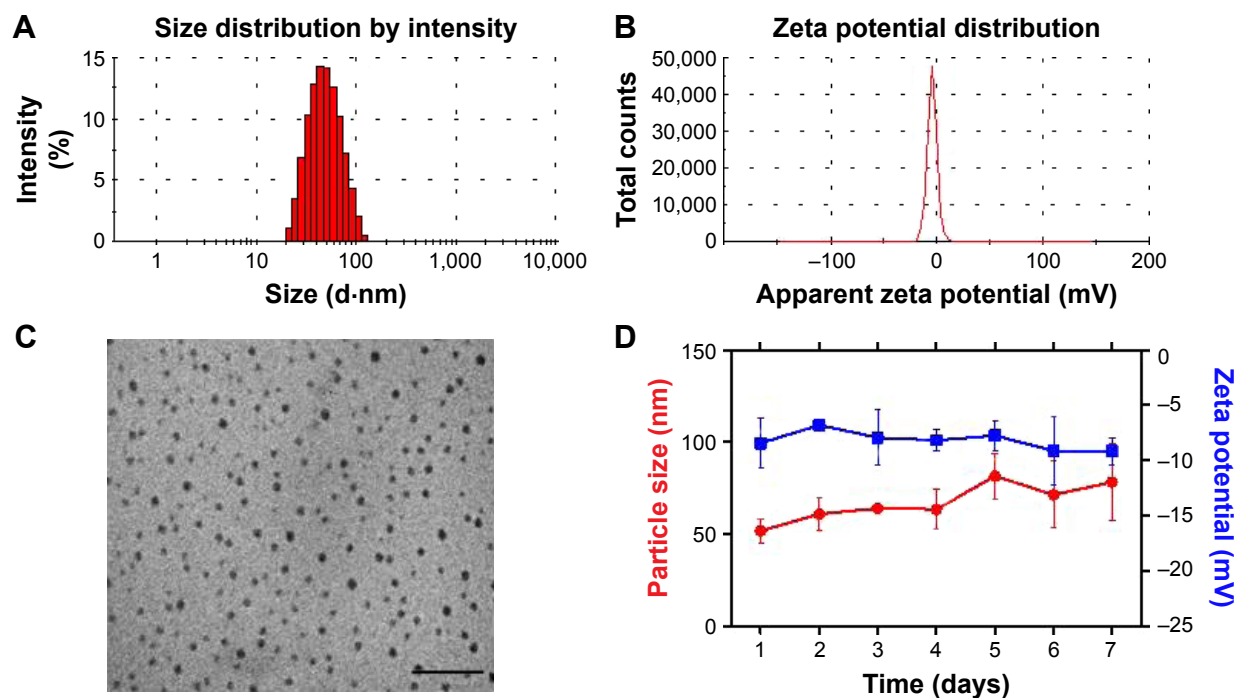


Figure 5 Characterization of F127-SS-TOC micelles.

Notes: (A) Size distribution of F127-SS-TOC micelles. (B) Zeta potential of F127-SS-TOC micelles. (C) TEM images of F127-SS-TOC micelles. The scale bar corresponds to $200\text{ }\mu\text{m}$ in the images. (D) The changes in micellar particle size and zeta potential during 7 days' storage. The red circle represents particle size, and the blue square represents zeta potential. The data were expressed as mean \pm SD of three experiments.

Abbreviations: F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; SD, standard deviation; TEM, transmission electron microscope; TOC, α -tocopherol.

CMC is an important characteristic to evaluate the aggregation properties and reflect the self-assembly stability of F127-SS-TOC micelles. It was determined by pyrene as the fluorescent probe.^{26,30} The CMC of the F127-SS-TOC micelles was ~ 7.58 mg/L ($\sim 5.4 \times 10^{-7}$ M), which was much lower than the CMC (6.9×10^{-5} M) of Pluronic F127 owing to the conjugation of the hydrophobic TOC chain,¹¹ suggesting that F127-SS-TOC micelles could maintain micellization at low concentrations and might have the ability to resist dilution in blood circulation (Figure 6A). Meanwhile, the Tyndall phenomenon of a series of micellar solutions was observed at different concentrations, which were irradiated by a laser beam.²⁶ The light beam could penetrate the micellar solutions by irradiation, and the light beam faded with the dilution of the F127-SS-TOC micelles (Figure 6B). The light intensity decreased significantly between the sample S5 (10.52 mg/L) and S6 (5.26 mg/L), and the brightness of sample S6 was similar with water. The low CMC of the F127-SS-TOC micelles could maintain the structural integrity under diluted conditions, which matched the results of the Tyndall phenomenon.

Reduction sensitivity of F127-SS-TOC micelles

The reduction-sensitive behavior of the micelles was related to the particle size changes in response to the reducing agent

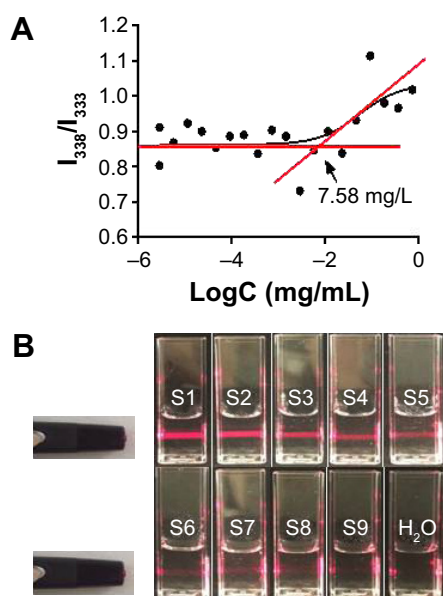


Figure 6 The determination of CMC and Tyndall phenomenon.

Notes: (A) The CMC of the F127-SS-TOC micelles. The arrow shows that the CMC value of the F127-SS-TOC micelles was 7.58 mg/mL. (B) Photos of F127-SS-TOC micellar solutions with the irradiation of red laser at various concentrations. (S1: 1.052 mg/mL; S2: 5.260×10^{-1} mg/mL; S3: 1.052×10^{-1} mg/mL; S4: 5.260×10^{-2} mg/mL; S5: 1.052×10^{-2} mg/mL; S6: 5.260×10^{-3} mg/mL; S7: 1.052×10^{-3} mg/mL; S8: 5.260×10^{-4} mg/mL, and S9: 1.052×10^{-4} mg/mL).

Abbreviations: F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; CMC, critical micelle concentration; $\text{Log}C$, log concentration; TOC, α -tocopherol.

DTT (10 mM), measured by DLS. There was no obvious change in the size of the F127-SS-TOC micelles within 3 h compared with the 0 mM DTT (Figure 7A). Notably, the intensity of the micelles had multiple peaks starting from 5 h, which suggested that the intermediate disulfide bonds between F127 and TOC were cleaved after interaction with 10 mM DTT, resulting in the size changes or eventually inducing aggregation or breakdown of the micelles. In addition, the changes in particle sizes were also verified by TEM. As shown in Figure 7B, 7 h later, the micelles had irregular surfaces and began to disassemble in the presence of 10 mM DTT, indicating that the F127-SS-TOC micelles were sensitive to reductive conditions.

Hemolysis assay

Hemolysis is considered to be an essential and reliable measurement to evaluate hemocompatibility of materials. The hemolysis assay was performed according to the previous report, employing rat red blood cells.²⁷ The hemolytic activity in vitro of the F127-SS-TOC micelles was evaluated at concentrations of 50, 100, 500, and 1,000 $\mu\text{g}/\text{mL}$. The F127-SS-TOC micelles did not exhibit hemolysis even at a high concentration of 1,000 $\mu\text{g}/\text{mL}$ (Figure 8A). The percentage of hemolysis of the F127-SS-TOC micelles at

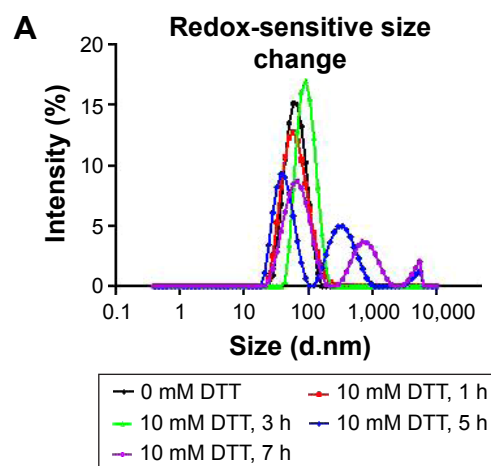


Figure 7 Reduction sensitivity of F127-SS-TOC micelles in vitro.

Notes: (A) Size change with the addition of 10 mM DTT during 7 h determined by DLS measurement. (B) TEM images of the F127-SS-TOC micelles with the addition of 10 mM DTT after 7 h. The scale bar corresponds to 100 μm in the images.

Abbreviations: F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; DLS, dynamic light scattering; DTT, dithiothreitol; TEM, transmission electron microscope; TOC, α -tocopherol.

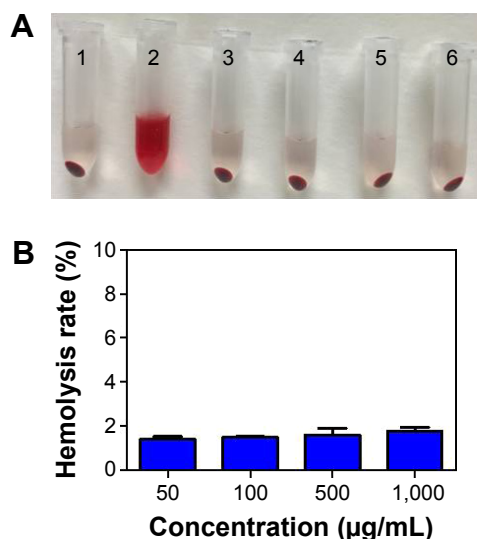


Figure 8 The hemolysis assay of F127-SS-TOC micelles.

Notes: (A) RBC suspensions were centrifuged after incubation with F127-SS-TOC micelles of various concentrations for 3 h. Sample 1 represented the negative control (RBCs suspended in physiological saline, 0% lysis); Sample 2 represented positive control (RBCs suspended in distilled water, 100% lysis); Samples 3, 4, 5, and 6 represented the RBCs suspended in 50, 100, 500, and 1,000 µg/mL concentrations of F127-SS-TOC micelles, respectively. (B) The hemolysis rate (%) of RBCs incubated with F127-SS-TOC micelles at various concentrations. The data were expressed as mean \pm SD of three experiments.

Abbreviations: F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; RBC, red blood cell; SD, standard deviation; TOC, α -tocopherol.

different concentrations was calculated using equation (1). As shown in Figure 8B and Table 1, hemolysis rates of the F127-SS-TOC micelles at concentrations of 50, 100, 500, and 1,000 µg/mL were 1.387% \pm 0.122%, 1.438% \pm 0.042%, 1.589% \pm 0.297%, and 1.745% \pm 0.205%, respectively, which showed similar hemolysis rates with no statistically significant difference among different concentrations. In accordance with the American Society for Testing and Materials (ASTM F756-00, 2000), the materials were classified, according to the degree of hemolysis, into three kinds: hemolytic (>5% of hemolysis), slightly hemolytic (2%–5% of hemolysis), and nonhemolytic (0%–2% of hemolysis).³¹ The hemolysis rates for all four concentrations were <2%, which demonstrated the excellent blood compatibility of the F127-SS-TOC micelles as well as their potential application as safe carriers for intravenous injection.

Table 1 Quantification of hemolysis test of F127-SS-TOC micelles with different concentrations (mean \pm SD, n=3)

F127-SS-TOC concentration (µg/mL)	1,000	500	100	50
Hemolysis rate (%)	1.745 \pm 0.205	1.589 \pm 0.297	1.483 \pm 0.042	1.387 \pm 0.122

Abbreviations: F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; n, number; SD, standard deviation; TOC, α -tocopherol.

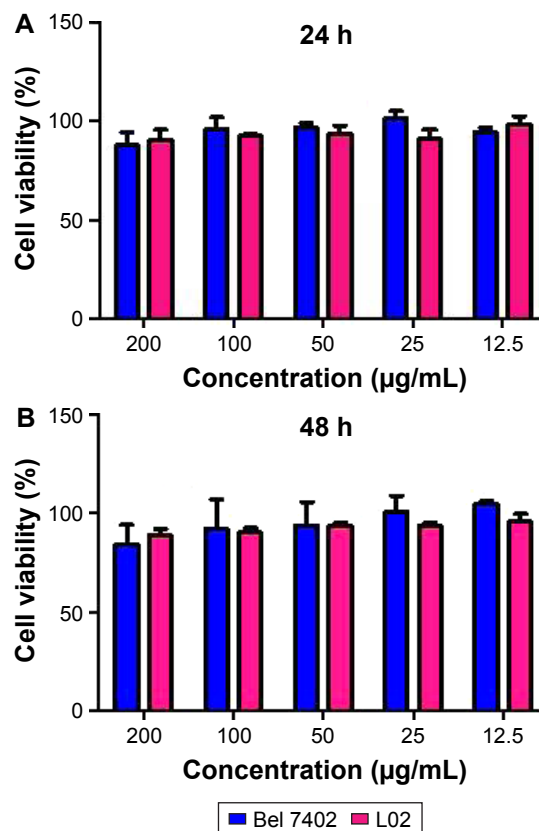


Figure 9 Cell viability of Bel 7402 and L02 cells treated with different concentrations of F127-SS-TOC micelles for (A) 24 h and (B) 48 h.

Note: The data were expressed as mean \pm SD of three experiments.

Abbreviations: F127, pluronic F127; F127-SS-TOC, TOC modified F127 polymer; SD, standard deviation; TOC, α -tocopherol.

In vitro cytotoxicity

The cytotoxicity of polymeric micelles is a crucial parameter for medical application. The cytotoxicity of the F127-SS-TOC against the Bel 7402 and L02 cells was evaluated by the MTT assay in terms of cell viability.¹⁵ The Bel 7402 and L02 cells were treated with various concentrations of F127-SS-TOC micelles for 24 h and 48 h, respectively. As shown in Figure 9, the F127-SS-TOC micelles did not show obvious cytotoxicity, with the micellar concentrations ranging from 12.5 to 200 µg/mL. After incubation for 24 h, the cell viability of both cells was >85%. Even after incubation for 48 h, the F127-SS-TOC micelles showed low cytotoxicity with a cell viability of about 85% in vitro. These results demonstrated the low biological toxicity of the F127-SS-TOC micelles at certain concentrations and its potential in the biomedical field.

Conclusion

In summary, we developed a biodegradable disulfide-containing polymer F127-SS-TOC, which proved to have

redox-sensitive properties in vitro. The micelles based on the F127-SS-TOC polymer had homogeneous spherical shapes and smooth surfaces, with a particle size as small as 51.87 ± 6.39 nm. The zeta potential of the micelles was -8.43 ± 2.27 mV, indicating that it might be beneficial for the colloidal stability in aqueous solution. Besides, the F127-SS-TOC micelles were proved to be stable at room temperature within 7 days, and the low CMC could ensure that the micelles maintained self-assembly under diluted conditions. Furthermore, the obvious reduction sensitivity of the F127-SS-TOC micelles was verified by measuring the size change in the presence of 10 mM DTT. The F127-SS-TOC micelles possessed favorable blood compatibility even at a micelle concentration of 1,000 $\mu\text{g/mL}$. In vitro study showed low cytotoxicity of the F127-SS-TOC micelles, indicating that the F127-SS-TOC polymer is safe to use. Thus, the F127-SS-TOC polymer exhibits excellent properties as micelles and has the potential to become a redox-sensitive carrier for delivering hydrophobic agents in cancer therapy. Besides, we will fully elucidate the application of drug-loaded micelles based on the F127-SS-TOC polymer in further work.

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Disclosure

The authors report no conflicts of interest in this work.

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