

# Linseed hydrogel-mediated green synthesis of silver nanoparticles for antimicrobial and wound-dressing applications

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**Abstract:** Polysaccharides are being extensively employed for the synthesis of silver nanoparticles (Ag NPs) having diverse morphology and applications. Herein, we present a novel and green synthesis of Ag NPs without using any physical reaction conditions. Linseed hydrogel (LSH) was used as a template to reduce  $\text{Ag}^+$  to  $\text{Ag}^0$ .  $\text{AgNO}_3$  (10, 20, and 30 mmol) solutions were mixed with LSH suspension in deionized water and exposed to diffused sunlight. Reaction was monitored by noting the change in the color of reaction mixture up to 10 h. Ag NPs showed characteristic ultraviolet-visible (UV/Vis) absorptions from 410 to 437 nm in the case of sunlight and 397–410 nm in the case of temperature study. Transmission electron microscopy images revealed the formation of spherical Ag NPs in the range of 10–35 nm. Face-centered cubic array of Ag NPs was confirmed by characteristic diffraction peaks in powder X-ray diffraction spectrum. Ag NPs were stored in LSH thin films, and UV/Vis spectra recorded after 6 months indicated that Ag NPs retained their texture over the storage period. Significant antimicrobial activity was observed when microbial cultures (bacteria and fungi) were exposed to the synthesized Ag NPs. Wound-healing studies revealed that Ag NP-impregnated LSH thin films could have potential applications as an antimicrobial dressing in wound management procedures.

**Keywords:** silver nanoparticles, green synthesis, antimicrobial studies, wound dressing, storage and stability

## Introduction

Metal-based nanoparticles (NPs) have intrigued material scientists in the fields of applied and fundamental research for the last two decades.<sup>1</sup> Unique electronic,<sup>2</sup> catalytic,<sup>3</sup> and optical<sup>4</sup> properties of such metal NPs have led to their potential applications in bioconjugation, biosensing,<sup>5,6</sup> and surface-enhanced Raman scattering.<sup>7</sup> Although NPs have been synthesized from a large variety of metals, silver nanoparticles (Ag NPs) have gained special importance over the last few years. Ag NPs have a wide range of potential applications in lithography,<sup>8</sup> microelectronics,<sup>9,10</sup> supermagnets,<sup>11</sup> and semiconductors.<sup>12</sup> Silver is also known to show antimicrobial properties, and several silver salts and their derivatives have been used as commercial antimicrobial agents.<sup>13,14</sup> Recently, Ag NPs have been employed as antimicrobial agents in water filters<sup>15</sup> and cotton fabrics.<sup>16</sup>

Owing to a large plethora of applications of Ag NPs, a number of methods have been devised for their synthesis. The more commonly used methods include chemical and photochemical reduction,<sup>17,18</sup> electrochemical method,<sup>19</sup> and laser ablation.<sup>20,21</sup>

Use of chemical reduction method for synthesis of Ag NPs involves environmentally hostile reducing agents,<sup>22,23</sup> toxic solvents,<sup>24</sup> and nonbiodegradable/synthetic

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stabilizing agents.<sup>25,26</sup> Such synthesis protocols can pose a serious threat to environment and also cause biological hazards. Further, high surface area-to-volume ratio of Ag NPs causes them to cling together with passage of time, thereby limiting their potential applications. Therefore, there has been keen interest to explore low-cost, biodegradable, nontoxic, and green agents<sup>27,28</sup> for synthesis of Ag NPs.<sup>29,30</sup>

Recently, polysaccharides are gaining importance as green reducing and self-capping agents for synthesis of Ag NPs.<sup>31,32</sup> The film-forming properties of polysaccharides can be additionally used for preventing the coagulation of Ag NPs over long-term storage.<sup>33</sup> Starch- and chitosan-based Ag NPs<sup>34,35</sup> have been synthesized but it was difficult to produce highly concentrated Ag NPs and store them for a longer period without getting agglomerated. Therefore, a need was felt to look for polysaccharides capable of producing highly concentrated and stable Ag NPs. In such an attempt, we have utilized linseed hydrogel (LSH) as a green reducing and self-capping agent for the formation of Ag NPs. LSH is composed of two major polysaccharidal fractions: rhamnogalacturonan and arabinoxylan.<sup>36,37</sup> The effect of AgNO<sub>3</sub> concentration, reaction time, and reaction temperature on the formation of Ag NPs is also studied. We are further focused to study antimicrobial properties of the pristine Ag NPs. Wound-healing capability of the synthesized Ag NPs will also be evaluated. The potential of LSH thin films as a storage medium for the Ag NPs is being reported for the first time.

## Experimental Materials

Linseed used for hydrogel extraction was purchased from local market. AgNO<sub>3</sub> was obtained from Sigma-Aldrich, St Louis, MO, USA. Analytical grade solvents and other reagents were procured from Fluka (Sigma-Aldrich) and used as received. Deionized water was used for preparation of all solutions and isolation of LSH.

## Isolation of LSH

LSH was conveniently isolated using hot water extraction.<sup>38</sup> Linseed (100 g) was thoroughly washed with deionized water to clean it from dust particles and soaked overnight in deionized water (1,000 mL). The swollen seeds were heated at 80°C for 30 min under stirring. Hydrogel extruded from these seeds was separated with the help of nylon gauze. LSH was washed with *n*-hexane to clean it from waxy or fatty material. LSH was further washed with deionized water and separated after centrifugation. Afterwards, it was dried under air for 24 h and then placed in vacuum oven at 60°C for the

next 24 h. Fine powder of LSH was collected after passing through stainless-steel mesh no 60 and stored in an air-tight container in a desiccator.

## Preparation of AgNO<sub>3</sub> solution

To prepare AgNO<sub>3</sub> solution (10 mmol), AgNO<sub>3</sub> (169.87 mg) was dissolved in deionized water (100 mL). Similarly, 20 and 30 mmol solutions of AgNO<sub>3</sub> were prepared by dissolving 339.74 and 509.61 mg of AgNO<sub>3</sub> in deionized water, respectively.

## Preparation of LSH suspension

LSH powder (0.2 g) was suspended in deionized water (10 mL) to prepare 2% (w/v) solution.

## LSH-mediated green synthesis of Ag NPs

To synthesize Ag NPs, freshly prepared solution of AgNO<sub>3</sub> (10 mmol, 10 mL) was added to the suspension of LSH (10 mL). The mixture was stirred in a dark area for 5 min at room temperature to ensure homogeneous mixing of AgNO<sub>3</sub> with LSH suspension. This mixture was then placed in sunlight for 10 h, and progress of reaction was monitored periodically by observing the change of color. Sample (1 mL) of this reaction mixture was also analyzed with the help of ultraviolet-visible (UV/Vis) spectrophotometry. Same protocol was adopted for the other AgNO<sub>3</sub> (20 and 30 mmol) solutions.

## Film formation of LSH containing embedded Ag NPs

A mixture of AgNO<sub>3</sub> (10, 20, and 30 mmol) and LSH was exposed to diffused sunlight for 10 h. Mixture was air dried in dark in a petri dish to produce LSH thin films containing the embedded Ag NPs and stored in an air-tight container. After 6 months, dry film was dissolved in deionized water, and the resultant suspension was characterized through UV/Vis spectrophotometry. Furthermore, Ag NPs were isolated from this suspension after centrifugation and then characterized by powder X-ray diffraction (PXRD) and transmission electron microscopy (TEM).

## UV/Vis spectrophotometry

Formation of Ag NPs with the passage of time was monitored by UV/Vis spectrophotometry. Sample (1.0 mL) of the reaction mixture was taken at selected time intervals (0.25, 0.5, 0.75, 1, 2, 4, 6, 8, and 10 h) and analyzed in a range 200–800 nm on UV/Vis spectrophotometer (Optizen POP, Mecasays, Korea). The effect of various reaction conditions

(concentration of  $\text{AgNO}_3$  solution and reaction time) on the progress of reaction was also studied. Moreover, effect of temperatures (50, 75, and  $100^\circ\text{C}$ ) on the synthesis of Ag NPs was also observed over a time period of 8 h. In a temperature-dependent study, the mixture of  $\text{AgNO}_3$  solution and LSH suspension was placed at different temperatures in a dark area to prevent any photochemical reaction.

## Powder X-ray diffraction

LSH suspension containing Ag NPs was freeze dried and characterized by PXRD. The PXRD spectra were recorded on a Bruker D8 Advance diffractometer (Bruker, Karlsruhe, Germany) (over a range of  $10^\circ\text{C}$ – $80^\circ\text{C}$ ,  $2\theta$ ), operated at 40 mA and 40 kV.

## Transmission electron microscopy

Morphology of Ag NPs was assessed by TEM. For this purpose, Ag NPs were isolated from solution by centrifugation and analyzed on a Philips 420 instrument. For recording TEM images, samples were drop casted on carbon-coated copper-wired TEM grid, and the instrument was operated at 120 kV. The LSH thin films were air dried in dark and stored for 6 months. After the specified storage period, Ag NPs were again analyzed by TEM. In a typical experiment, the stored LSH thin film impregnated with Ag NPs was dissolved in deionized water, and Ag NPs were isolated by centrifugation and viewed by TEM.

## Antimicrobial activity of Ag NPs

Aqueous solution of the synthesized Ag NPs (10 mg/mL) was used to study the antimicrobial activity against different bacterial (Gram positive and Gram negative) and fungal strains. The tested bacterial strains include *Streptococcus mutans* American-type culture collection (ATCC) 25175, *Staphylococcus epidermidis* ATCC 12228, *Pseudomonas aeruginosa* ATCC 27853, *Escherichia coli* ATCC 25922, *Staphylococcus aureus* ATCC 25923, *Bacillus subtilis* ATCC 6633, and *Actinomyces odontolyticus* ATCC 17929. The fungal strain used in the antimicrobial assay was *Aspergillus niger*. Mueller Hinton Agar Media (Thermo Fisher Scientific, Waltham, MA, USA) was used as a growth medium for bacterial species, while fungi were grown on Sabouraud Dextrose Agar (Hardy Diagnostics, Santa Maria, CA, USA). Inoculums (fungal and bacterial culture in respective media, 10 mL) were incubated at  $27^\circ\text{C}$ – $30^\circ\text{C}$  for 30–37 h for fungal strains and at  $37^\circ\text{C}$  for 24 h for bacteria. Fungal culture (7 days old) was washed, suspended in normal saline solution, and incubated at  $28^\circ\text{C}$  after being filtered through aseptic glass wool. The turbidity of inoculums was adjusted by 0.5 Mc Farland Standard.

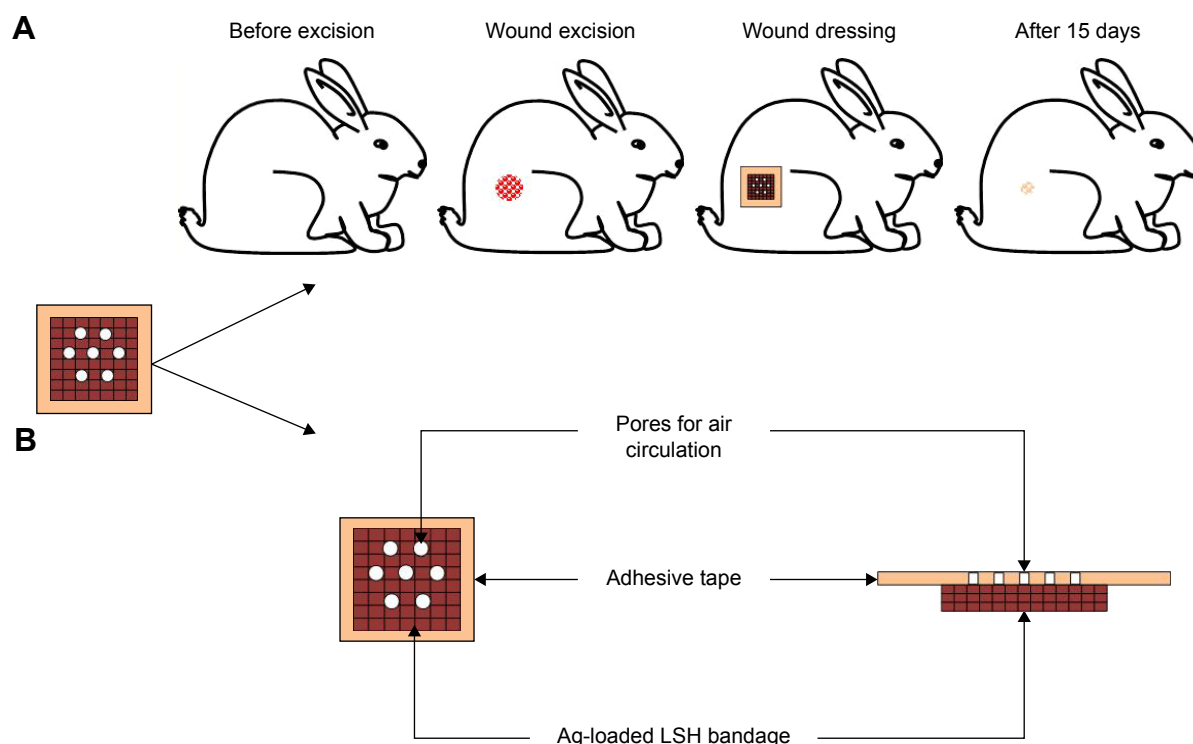
Disk diffusion method<sup>39</sup> was used to determine the antimicrobial activity of Ag NPs. In a typical experiment, aqueous solution of the Ag NPs (20  $\mu\text{L}$ ) was loaded/impregnated onto filter paper discs (Whatman filter paper no 1, 6 mm in diameter) and implanted on the surface of the microbial culture plates. These plates were then incubated at  $37^\circ\text{C}$  for 24 h for bacterial strains and at  $30^\circ\text{C}$  for 36 h for fungal strains. At the end of incubation period, zone of inhibition was measured in millimeter. Discs loaded with pure dimethyl sulfoxide and distilled water were used as negative control while  $\text{AgNO}_3$  solution (20 mmol) was used as positive control. All experiments were performed in triplicate and mean values are reported.

## Designing of a wound-dressing patch

LSH thin layer containing the embedded Ag NPs was evaluated as a wound-dressing material. Dressing patch consists of two layers. One layer is a cubic porous adhesive (United States Pharmacopeia, Rockville, MD, USA) backing membrane (4×4 cm) and the second layer is cubic sterile cotton patch (2×2 cm) immersed in LSH impregnated with Ag NPs (20 mmol, 0.5 mL). This moist antiseptic patch was implanted at the center of adhesive membrane as shown in Figure 1B. This adhesive membrane will help the dressing patch to stick to the body. Freshly prepared patch was used during wound-healing studies.

## Wound-healing studies

Nine healthy male rabbits ( $2.5\pm 0.2$  kg, 4 months old) were selected and divided into three equal groups: control group, standard group, and test group. All rabbits were caged under standard laboratory conditions for 48 h before experiment and provided standard food and water ad libitum throughout the study. Rabbits were given anesthesia and placed on surgical table in natural position. Hair from the rear leg of the rabbit was shaved and excised a circular wound of 1 cm in diameter using a biopsy forceps. Wound of control group rabbits was left open and untreated. A Band-aid® dressing was applied to the wound of standard group while the test group was applied with dressing patch made as described above. The healing attributes were studied by tracing the raw wound area on tracing paper till the wound was completely epithelialized.<sup>40</sup> An illustration of wound-healing procedure is shown in Figure 1. The study protocol was approved by Pharmacy Research Ethics Committee of University of Sargodha, Sargodha, Pakistan (Ref no 75–2014/PREC) and in accordance with the National Institute of Health's Guidelines for the Care and Use of Laboratory Animals.



**Figure 1** Schematic illustrations of wound treatment (A) and LSH wound-dressing patch containing embedded Ag nanoparticles (B).  
**Abbreviation:** LSH, linseed hydrogel.

## Collagen estimation

Collagen contents of the regenerated tissue were measured using a reported method.<sup>40</sup> For this purpose, regenerated tissue of the scratched wound was chopped and suspended into 0.5 M acetic acid (10% w/v) after being washed with 0.5 M sodium acetate. It was stirred for 48 h and then centrifuged for 2 h at  $5,000 \times g$ . Collagen was precipitated by adding sodium chloride (10% w/v) and filtered on a weighed Whatman filter paper. Amount of collagen was calculated from the difference in the weight of filter paper before and after filtration.

## Statistical analysis

The data of collagen estimation were expressed as the mean  $\pm$  standard deviation (SD) and analyzed using one-way analysis of variance. A  $P$ -value  $< 0.05$  was considered statistically significant and denoted by asterisk.

## Results and discussion

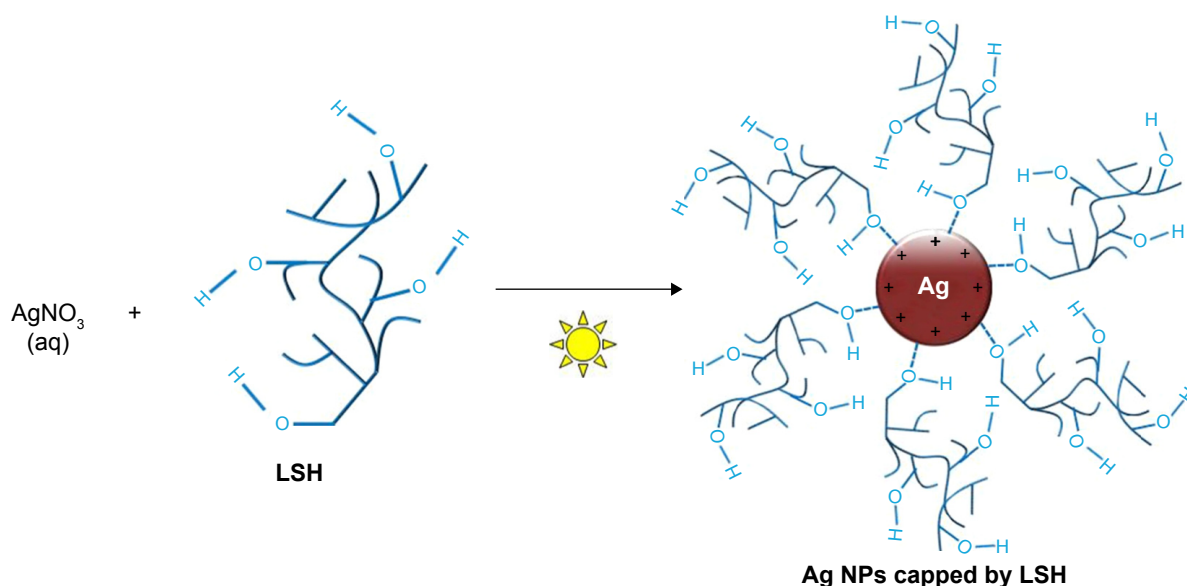
### Green synthesis of Ag NPs

On subjecting the  $\text{AgNO}_3$  solutions to diffused sunlight, hydrated electrons are generated in the system. These electrons can be used to reduce monovalent silver cations ( $\text{Ag}^+$ ) to zero-valent silver atom ( $\text{Ag}^0$ ). The  $\text{Ag}^0$  atoms so generated usually

have a size ranging in nanometers. Therefore, UV irradiation can be exploited as a green method for the synthesis of Ag NPs. This method can also be used to avoid environmentally toxic and hazardous reducing agents. In the present work, we have carried out the green synthesis of Ag NPs using LSH as a green reducing agent under diffused sunlight. The LSH was also evaluated as a storage medium for the synthesized Ag NPs. LSH-mediated synthesis of Ag NPs was carried out using different concentrations of  $\text{AgNO}_3$  solutions (10, 20, and 30 mmol). On mixing the reactants,  $\text{Ag}^+$  in the  $\text{AgNO}_3$  solution reacted with LSH to give  $\text{Ag}(\text{LSH})^+$  complex. When the mixture was subjected to diffused sunlight,  $\text{Ag}^+$  in the complex was reduced by LSH to produce Ag (LSH) precursor. The positive charge on the surface of Ag NPs was electrostatically stabilized and capped by the negative charge on polysaccharide hydroxyls, which resulted in colloidal stabilization of  $(\text{Ag}[\text{LSH}])$ .

The progress of formation of Ag NPs by irradiation of  $(\text{Ag}[\text{LSH}])^+$  complex was followed by noting the color change of reaction mixture with passage of time. The color of LSH and  $\text{AgNO}_3$  mixture changed from colorless to reddish brown and finally dark brown within 10 h indicating the complete reduction of  $\text{Ag}^+$ . A schematic illustration of synthesis of LSH-mediated synthesis of Ag NPs is shown in Figure 2.





**Figure 2** Schematic illustration showing synthesis of LSH-mediated Ag NPs.  
**Abbreviations:** LSH, linseed hydrogel; NPs, nanoparticles.

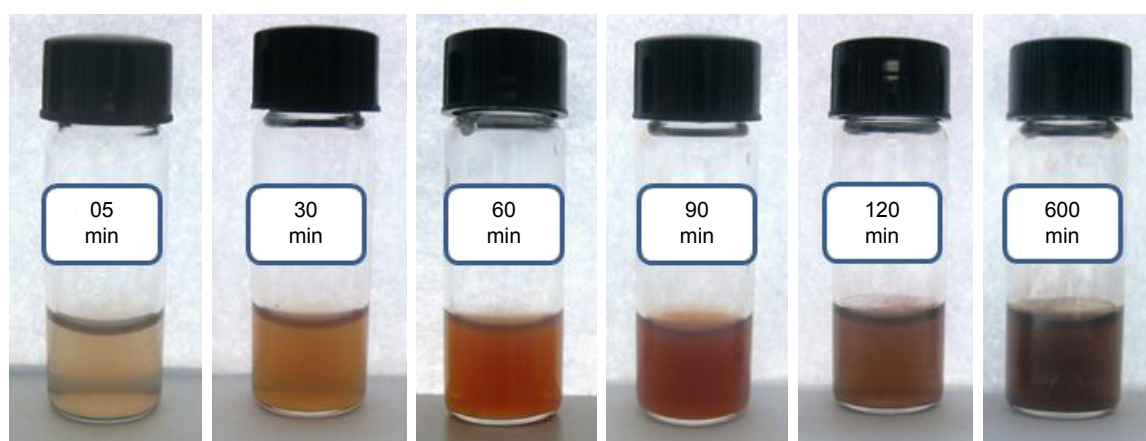
## Characterization of Ag NPs

### UV/Vis spectrophotometry

LSH suspension (2 mL) was mixed with different concentrations of  $\text{AgNO}_3$  (10, 20, and 30 mmol, 2 mL aliquot each) in dark and then placed in diffused sunlight. Reduction of  $\text{Ag}^+$  to  $\text{Ag}^0$  was indicated by change in color of the reaction mixture and monitored by UV/Vis spectrophotometry. The conduction electrons of Ag NPs exhibit surface plasmon resonance (SPR) because of their collective oscillation, and this SPR results in strong absorption in visible region of spectrum.<sup>41</sup> The wavelengths absorbed for SPR transitions of Ag NPs are changed by change in reaction time and size of Ag NPs, so reaction mixture showed a color change as the reaction progressed. The color changed from colorless to reddish

brown and finally dark brown with passage of time from 0.25 to 10 h. Photographs showing the color change of reaction mixture with progress of reaction are shown in Figure 3.

The LSH- $\text{Ag}^+$  solutions showed UV/Vis absorptions at 410, 415, 419, 425, 426, 428, 430, 431, and 436 nm for 10 mmol, whereas 412, 416, 422, 426, 428, 430, 431, 434, and 436 nm absorptions were recorded for 20 mmol LSH- $\text{Ag}^+$  solution. Likewise, 30 mmol solution showed UV absorptions at 413, 417, 424, 427, 429, 432, 433, 435, and 437 nm. These absorptions were recorded at reaction times of 0.25, 0.5, 0.75, 1, 2, 4, 6, 8, and 10 h, respectively. Similar results have been reported in the literature where Ag NPs were synthesized using exogenous reducing agents.<sup>42</sup> The absorption spectra of Ag NPs showed red shift indicating increase in size of



**Figure 3** Photographs of linseed hydrogel- $\text{Ag}^+$  mixture (20 mmol  $\text{AgNO}_3$ ) showing color change with passage of time.  
**Abbreviation:** min, minute.

Ag NPs with increase in reaction time. Absorption intensity also showed an increase when reaction time was increased from 0.25 to 10 h, which indicated continuous reduction of  $\text{Ag}^+$  by LSH with passage of time. Similarly, an increase in absorption intensity was also observed as the concentration of  $\text{AgNO}_3$  solution was increased from 10 to 30 mmol as evident in Figure 4A–C. This increase in absorption intensity is due to increase in size of the synthesized Ag NPs as shown in TEM images in Figure 5. Results of UV/Vis analyses are summarized in Figure 4.

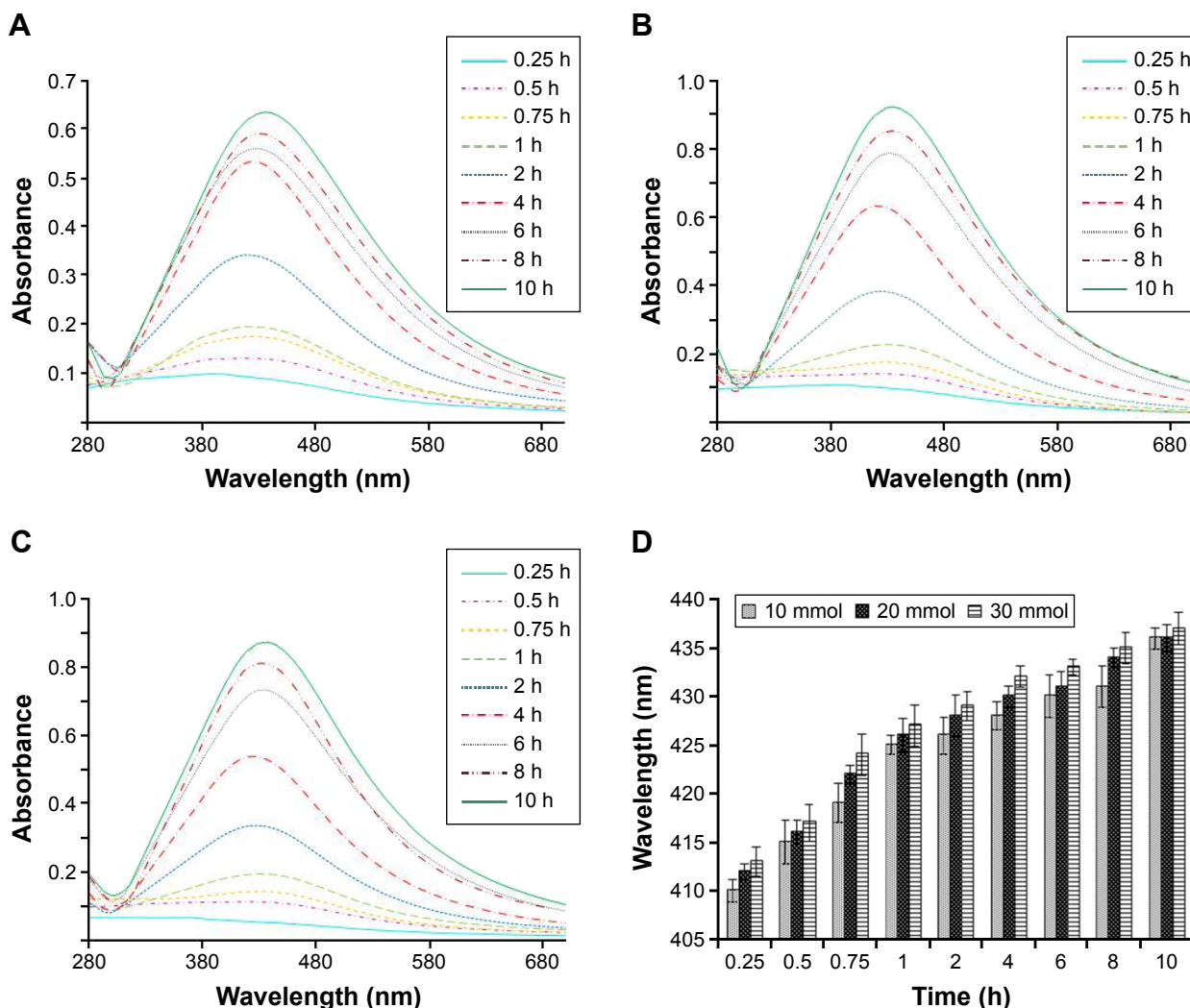
## Effect of temperature on formation of Ag NPs

Effect of temperature on rate of formation and size of LSH-mediated Ag NPs was also noted.  $\text{AgNO}_3$  (10, 20, and 30 mmol) solution was mixed with LSH solution and

progress of reaction was monitored at different temperatures (50, 75, and 100°C). This experiment was performed in dark to avoid any photochemical reaction. Comparison of absorption bands showed that Ag NPs exhibit higher absorption with increase in temperature (Figure 6). The increase in absorption wavelength for a given reaction time at higher temperature indicated that rate of formation of Ag NPs and size of synthesized NPs are dependent on the temperature. These findings are in line with the observations by other scientists.<sup>43</sup>

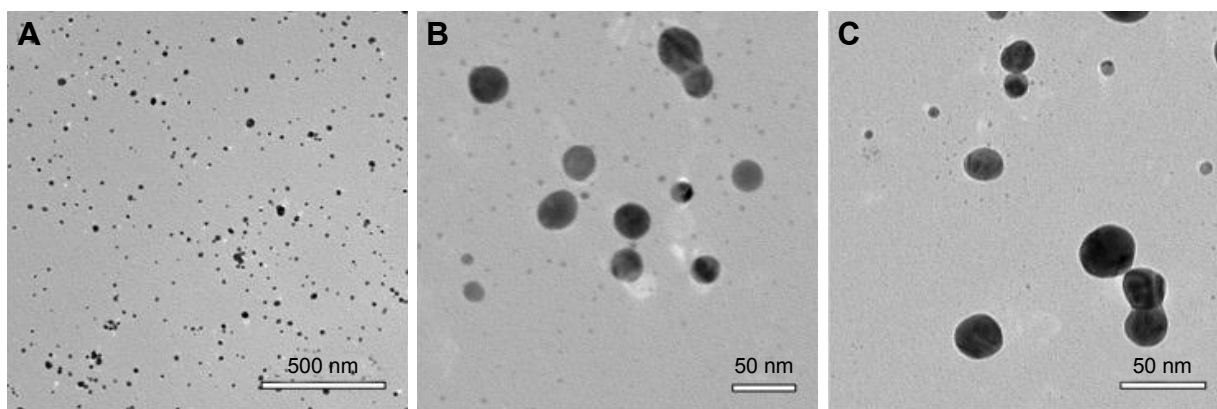
## TEM of isolated Ag NPs

Size distribution and morphology of Ag NPs were assessed by TEM. Ag NPs were separated from LSH and  $\text{AgNO}_3$  (10, 20, and 30 mmol) solutions by centrifugation, and TEM image was recorded. TEM images confirmed the formation



**Figure 4** Ultraviolet-visible spectra of linseed hydrogel-mediated Ag NPs: 10 mmol (A), 20 mmol (B), and 30 mmol solution of  $\text{AgNO}_3$  (C) at different reaction times and cumulative graphical representation (D) showing increase in absorption of Ag NP solutions with increase in concentration and reaction time.

**Abbreviations:** h, hour; NPs, nanoparticles.



**Figure 5** Transmission electron microscopy images of Ag nanoparticles isolated from (A) 10 (10–25 nm), (B) 20 (10–30 nm), and (C) 30 mmol (10–35 nm) linseed hydrogel–Ag<sup>+</sup> solution. Magnification 7,500 $\times$ , 75,000 $\times$  and 55,000 $\times$ , respectively.

of spherical Ag NPs in the size regimen of 10–25, 10–30, and 10–35 nm for 10, 20, and 30 mmol AgNO<sub>3</sub> solution (Figure 5), respectively. For film formation, a mixture of LSH and AgNO<sub>3</sub> (20 mmol) solution was air dried in dark and stored for 6 months. After selected storage period, the film was dissolved in deionized water and Ag NPs were isolated by centrifugation. TEM image of isolated Ag NPs showed that the NPs retained their morphology and size over the storage period. Therefore, it was inferred that the LSH thin film could be used as storage media for Ag NPs.

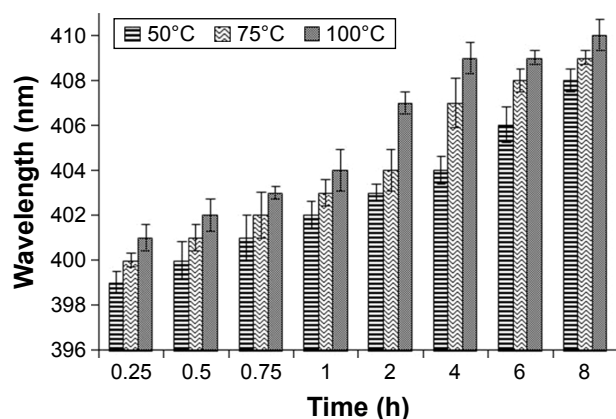
### Powder X-ray diffraction

PXRD (Figure 7) was used to confirm the crystal phase of the isolated Ag NPs in the range of 10°C–80°C, 2 $\theta$ . The diffraction peaks centered at (111), (200), (220), and (311) indicated that Ag NPs had face-centered cubic lattice in all samples. Diffraction pattern was also recorded for the Ag NPs isolated from stored thin films (6 months). It was revealed that there was no change in diffraction peak of samples after

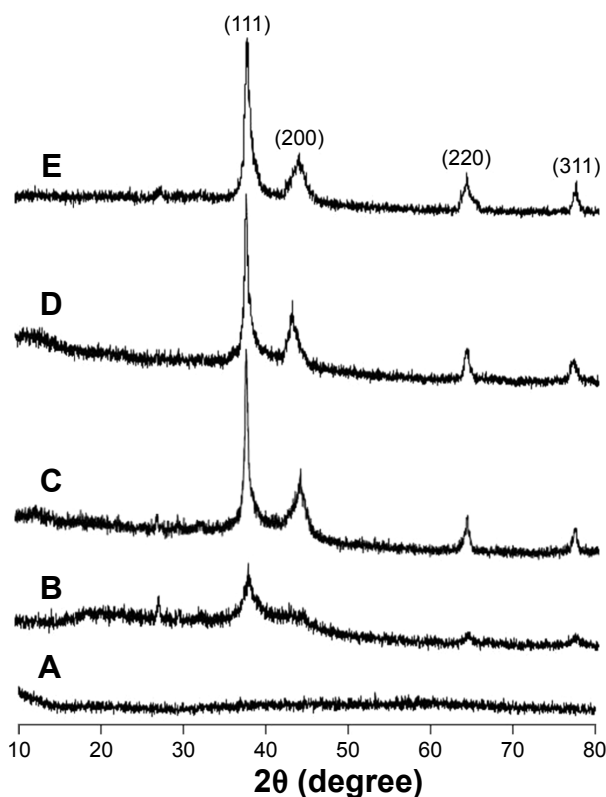
storage. Therefore, it was concluded that LSH could be used for the long-term storage of Ag NPs without affecting their morphology.

### Storage of Ag NPs in LSH thin films

Potential of LSH for the storage of Ag NPs in solution and in the form of a thin film was evaluated. For this purpose, Ag NPs were synthesized by reducing the AgNO<sub>3</sub> (20 mmol) solution with LSH over 10 h. UV/Vis spectra were recorded



**Figure 6** Ultraviolet-visible spectroscopy absorptions of Ag nanoparticles obtained from AgNO<sub>3</sub> solution (10 mmol) at different temperatures and reaction times. **Abbreviation:** h, hour.



**Figure 7** Powder X-ray diffraction spectra: LSH (A), Ag NPs-embedded LSH film (B), and isolated Ag NPs, 10 mmol (C), 20 mmol (D), and 30 mmol (E). **Abbreviations:** LSH, linseed hydrogel; NPs, nanoparticles.

after 10 and 24 h, and the sample was stored as dry thin film in dark. Absorption spectra of the isolated Ag NPs were acquired by dissolving the films in deionized water. Comparable absorption spectra were obtained after the storage period (15 days, 30 days and 6 months). No significant change in the absorption wavelength and intensity was observed for the stored samples. Therefore, it was inferred that the Ag NPs did not undergo agglomeration on drying and storage in the form of thin films. TEM images of isolated Ag NPs from stored thin film (6 months) were recorded and observed uniform size and shape as of freshly prepared Ag NPs. Results of storage experiments are illustrated in Figure 8.

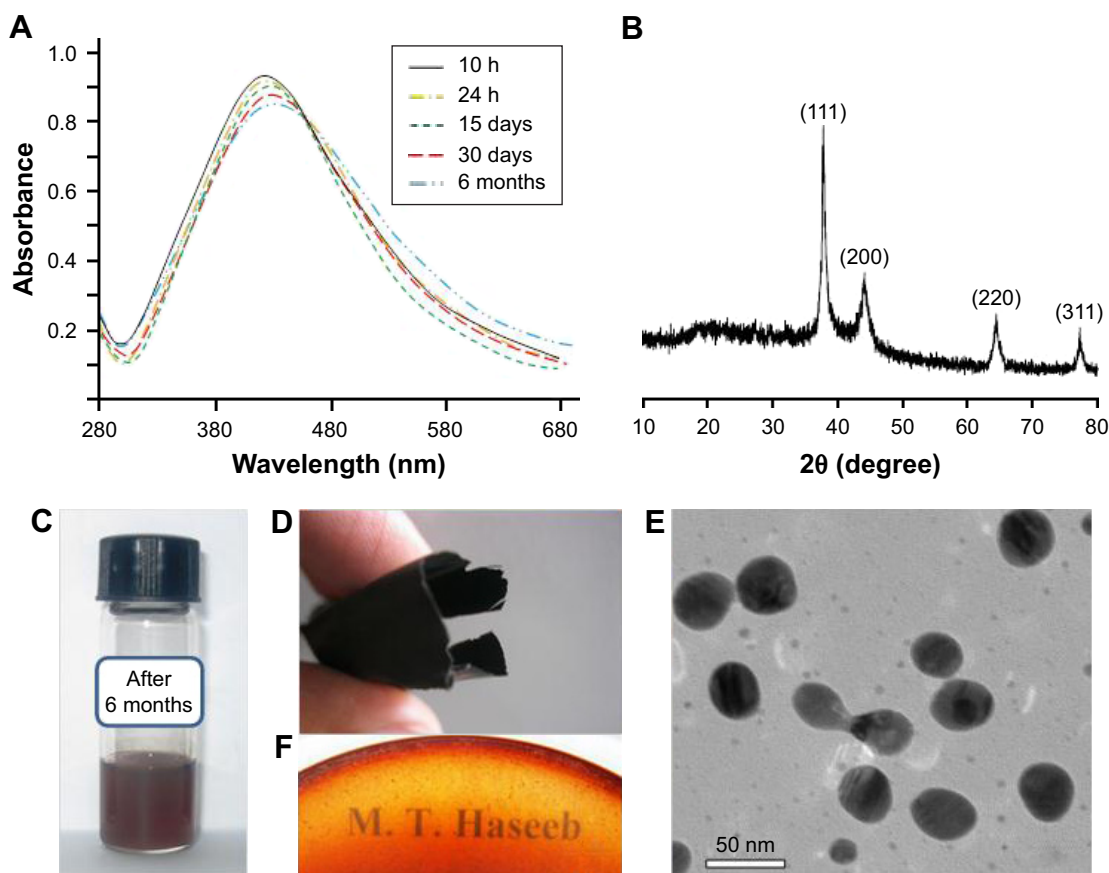
### Antimicrobial activity of Ag NPs

LSH-based Ag NPs were tested for antimicrobial properties. Significant antibacterial and antifungal activity was observed against *S. mutans*, *S. epidermidis*, *P. aeruginosa*, *E. coli*, *S. aureus*, *B. subtilis*, *A. odontolyticus*, and *A. niger* strains and showed inhibition zones of 22, 13, 18, 09, 20, 15, 16, and 10 mm, respectively, on being exposed to the aqueous

solution of Ag NPs synthesized with  $\text{AgNO}_3$  (20 mmol) solution. No antimicrobial activity was noticed for control experiments performed by using LSH solution and distilled water; however,  $\text{AgNO}_3$  (20 mmol) solution was found active against the above-mentioned strains as shown in Figure 9. Antimicrobial properties of Ag NPs prepared from 10 to 30 mmol  $\text{AgNO}_3$  solution were also tested. Results of antimicrobial activity of Ag NPs (20 mmol) are summarized in a bar graph in Figure 9, and the average of three readings is reported. As a typical example, antimicrobial activity of Ag NPs (20 mmol) against *P. aeruginosa* is shown in Figure 9. It is revealed that LSH-based Ag NPs had a clear zone of inhibition against the above-mentioned strains, while LSH alone did not show any effect on the microbial cultures.

### Wound-healing studies

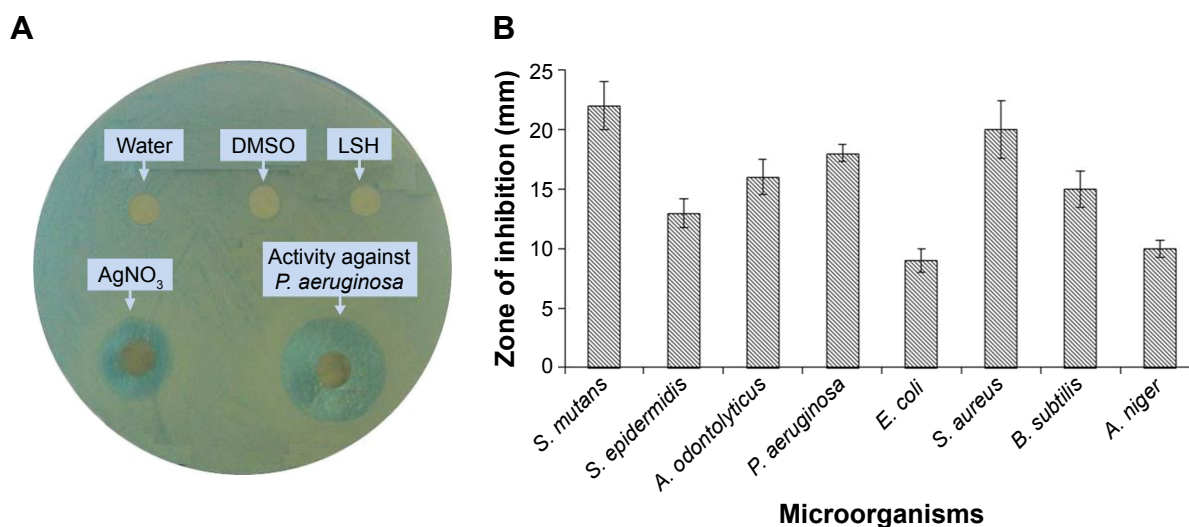
To evaluate wound-healing properties of LSH films impregnated with Ag NPs, excision wounds were created on rear leg of the rabbit. Average wound diameter was calculated for different groups of rabbits. Wound healing was estimated



**Figure 8** Ultraviolet-visible spectra of  $\text{AgNO}_3$  (20 mmol) solution in LSH recorded after 10 h, 24 h, 15 days, 30 days, and 6 months storage (A), powder X-ray diffraction spectrum of Ag NPs isolated after 6 months storage of LSH–Ag NP film (B), vial containing solution of stored film in water (C), foldable and see-through Ag NPs-embedded LSH thin film (D and F), respectively, and transmission electron microscopy image of isolated Ag NPs (10–30 nm) from thin films after 6-month storage in dark (E). Magnification 75,000 $\times$ .

**Abbreviations:** LSH, linseed hydrogel; NPs, nanoparticles.





**Figure 9** Photograph indicating the antimicrobial activity of Ag nanoparticles (20 mmol) against *P. aeruginosa* along with negative and positive control (A) and measurements of zone of inhibition of different bacterial and fungal strains (B).

**Abbreviations:** *A. niger*, *Aspergillus niger*; *A. odontolyticus*, *Actinomyces odontolyticus*; *B. subtilis*, *Bacillus subtilis*; DMSO, dimethyl sulfoxide; *E. coli*, *Escherichia coli*; LSH, linseed hydrogel; *P. aeruginosa*, *Pseudomonas aeruginosa*; *S. aureus*, *Staphylococcus aureus*; *S. epidermidis*, *Staphylococcus epidermidis*; *S. mutans*, *Streptococcus mutans*.

from percentage closure of wound area (mm<sup>2</sup>) noted at different times after wound induction. Control group showed a wound closure of 0.25, 1.11, 20.32, 38.76, 62.55, and 83.27% on 1st, 3rd, 6th, 9th, 12th, and 15th day, respectively. The animals treated with standard Band aid dressing showed closure area of 2.17, 19.45, 63.67, 87.27, 98.49, and 100%, while the test group showed 1.41, 16.73, 54.62, 93.21, 99.22, and 100% wound closure on 1st, 3rd, 6th, 9th, 12th, and 15th day, respectively. Results indicated that tissue regeneration and percentage wound closure of LSH-based wound dressing were comparable to the standard Band aid dressing. Results of wound-healing study for various groups are summarized in Table 1.

To understand the mechanism of wound healing, collagen content and tensile strength of the epithelialized wound tissue were measured. The control group showed a collagen content of 38 mg/kg while standard and test groups showed a collagen content of 69 and 65 mg/kg, respectively. Therefore, it was noticed that the standard and test groups had higher collagen content than the control group, which resulted in rapid wound healing in the said groups in contrast to the control. Tensile strength was also found higher for test and standard

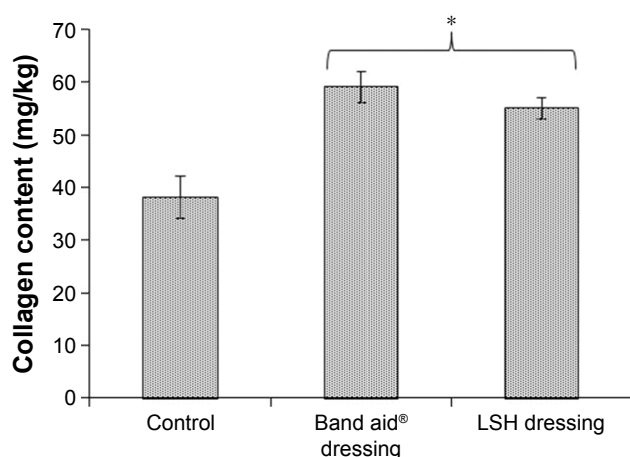
groups as compared to the control. Results of collagen content of regenerated tissue for various groups are depicted in Figure 10. It was inferred that collagen synthesis is initiated at the wound area through formation of a polypeptide precursor. Inter- and intramolecular cross-linking of this collagen then results in rapid tissue regeneration and wound closure.

## Conclusions

Herein, we report on a cost-effective and eco-friendly method for the synthesis of Ag NPs using biodegradable and nontoxic LSH as a template nanoreactor and reducing agent. Effect of AgNO<sub>3</sub> concentration, reaction time, and change in temperature on the size and distribution of Ag NPs has been studied. Narrow size distribution and uniform dispersion of Ag NPs were achieved in the presence of LSH as a green reducing agent. LSH thin films proved to be an excellent storage medium to prevent agglomeration of Ag NPs with passage of time. Ag NPs-embedded LSH films can be exploited for potential biomedical and antimicrobial applications. Exploration of catalytic properties and surface-enhanced Raman scattering of Ag NPs are future prospects of the study.

**Table 1** Wound diameter and wound closure % after selected day intervals

	Wound diameter in mm ± standard deviation (% of wound closure) at day					
	1st	3rd	6th	9th	12th	15th
Control	5.98±0.06 (0.25)	5.93±0.05 (1.11)	4.78±0.03 (20.32)	3.67±0.10 (38.76)	2.25±0.06 (62.55)	1.00±0.02 (83.27)
Band aid® dressing	5.87±0.04 (2.17)	4.83±0.08 (19.45)	2.18±0.09 (63.67)	0.76±0.05 (87.27)	0.09±0.08 (98.49)	(100)
Linseed hydrogel dressing	5.91±0.04 (1.41)	4.94±0.07 (17.73)	2.72±0.08 (54.62)	0.41±0.06 (93.21)	0.05±0.08 (99.22)	(100)



**Figure 10** Collagen contents of epithelialized wound tissue of various groups after 15th day.

**Note:** Statistical significance from control group and is shown by \* $P < 0.05$ .

**Abbreviation:** LSH, linseed hydrogel.

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## Disclosure

The authors report no conflicts of interest in this work.

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