

# Usutu virus: current knowledge and future perspectives

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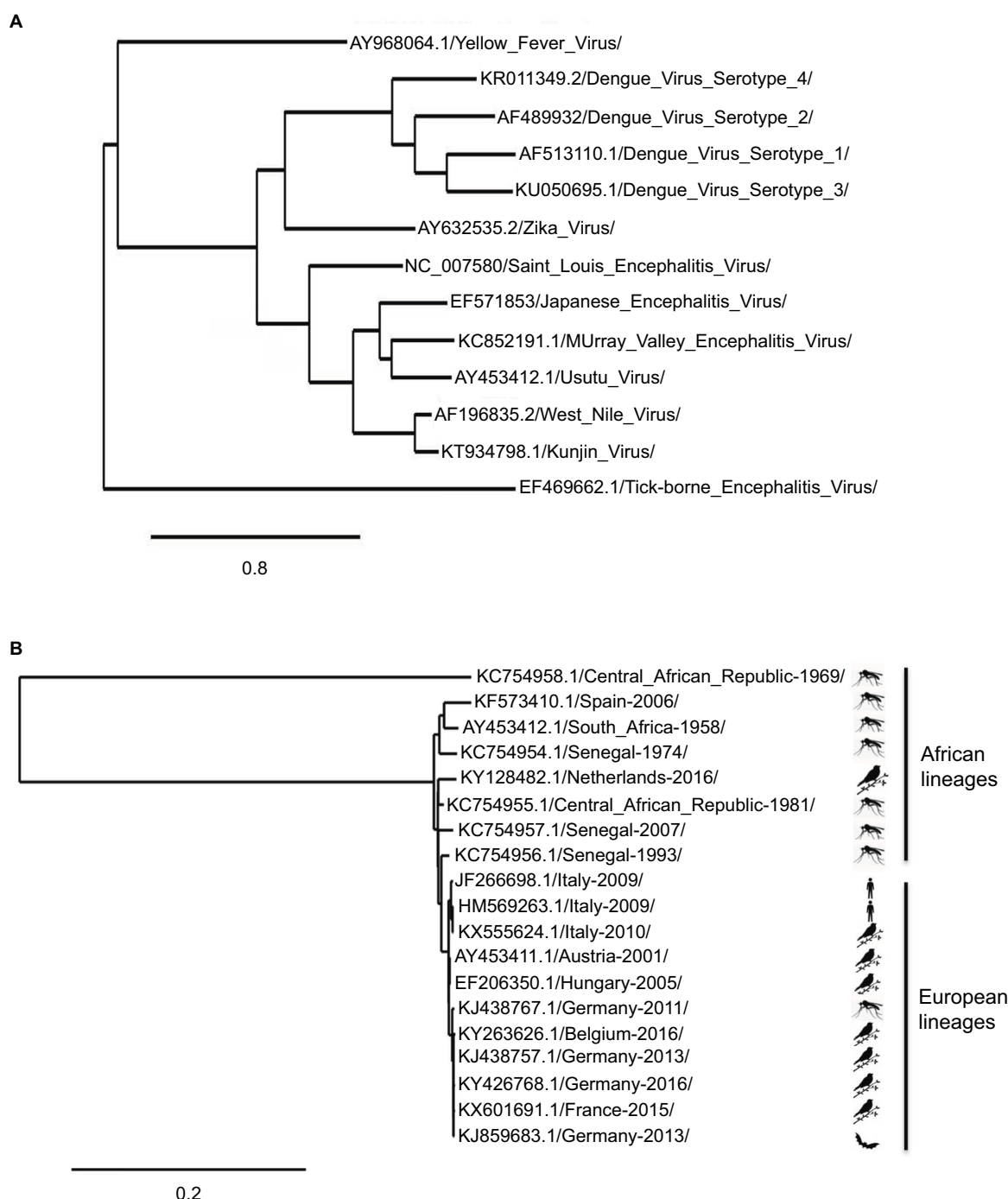
**Abstract:** The *Flavivirus* genus (Flaviviridae family) contains important pathogens such as yellow fever virus, Japanese encephalitis virus, St Louis encephalitis virus, West Nile virus, Usutu virus (USUV), Zika virus, and dengue virus, many of which constitute a worrisome threat to global human and animal health. USUV is transmitted by mosquitoes and, as any other flavivirus, is an enveloped plus-strand RNA virus. The virus was first isolated from *Culex neavei* mosquitoes in South Africa in 1959 near the Usutu River, from where it takes its name. Since then, the virus was confined to Africa until its first detection in Austria in 2001, although it was probably present in Europe since 1996 or even earlier. After that, USUV has spread throughout Europe, causing a considerable mortality among birds and a few neurologic cases in humans. The main USUV natural hosts are birds, but infection has also been reported in other vertebrate species, including humans. The fast spread of the virus through the continent, the relatively high mortality caused in birds, and the recent neuroinvasive human cases related to USUV infection reported in Europe have raised serious concerns about its possible consequences for public health. Here, an updated review of current knowledge about this emerging pathogen is presented.

**Keywords:** Usutu virus, flavivirus, host cell–virus interactions, surveillance, prophylaxis

## Biology of the virus Molecular classification and phylogeny

Usutu virus (USUV) is a member of the *Flavivirus* genus within the Flaviviridae family (Figure 1A). In 2004, the complete genomic sequences of two USUV strains, one African (South Africa-1959) and one European (Vienna-2001), were used to build a phylogenetic tree. These two strains showed a 97% and 99% homology at the nucleotide (nt) and amino acid (aa) level, respectively.<sup>1</sup> Both strains were closely related to Murray valley encephalitis virus (73% and 82% nt and aa similarity), Japanese encephalitis virus (71% and 81% nt and aa similarity), and to West Nile virus (WNV; 68% and 75% nt and aa similarity). Nowadays, dozens of complete sequences are available in the data banks. In a recent study, the sequence identity among seven USUV strains was 96%–99% and 99% at the nt and aa levels, respectively.<sup>2</sup> The only exception was a 1969 isolate from Central African Republic (CAR-1969) that presented homologies of 81% and 94% at the nt and aa levels with other African sequences (Figure 1B). Due to these differences, it has been suggested that CAR-1969 formed an USUV subtype strain.<sup>3</sup> A comparison with the original South African SAAR-1776 strain showed that the mutations found among different European and African strains were scattered throughout the genome, with some unique substitutions and some that were present

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**Figure 1** Usutu virus phylogram trees.

**Notes:** (A) Representative phylogram showing the genetic relatedness between different flaviviruses. Accession numbers are displayed in the tree. The scale indicates 0.8 nucleotide substitutions/site. (B) Representative phylogram showing the genetic relatedness between geographically and temporally different Usutu virus strains. Accession numbers, country, and collection date are displayed in the tree. The scale indicates 0.2 nucleotide substitutions/site. The trees were based on complete genome sequence, built from a multiple alignment using Clustal omega and Phylogeny.fr.

in various strains.<sup>4</sup> Eleven aa changes were common to all strains when compared with SAAR-1776, whereas some unique substitutions, not conclusively identified, have been suggested to be implicated in specific viral features. For instance, aa changes at positions S595G and D3425E in the Bologna-2009 strain, which are found in other neurovirulent

flaviviruses (dengue virus, Japanese encephalitis virus, and WNV), have been suggested to be related with viral neuroinvasiveness.<sup>2</sup> Furthermore, variations (A1236V and L1549F) in some German strains have also been related to pathogenesis,<sup>5</sup> as similar aas are found in WNV strains that have been reported to inhibit the interferon signaling

pathway, thus playing a role in the host immune response.<sup>6</sup> In any case, availability of a manageable infectious clone and experiments in suitable animal model will be necessary to clarify these points. Lately, phylogenetic analysis of recently detected strains from bats in Germany has pointed to a new introduction of the virus in the continent and to bats as possible amplifying reservoir hosts.<sup>5,7</sup> Even more, a further study described new strains from two dead juvenile Great gray owls (*Strix nebulosa*) from the Zoological Garden in Berlin in 2015. These sequences were closely related to the mosquito strain reported in Catalonia (Spain) in 2006<sup>8</sup> and, thus, point to different and frequent introduction of the virus in Europe and to a probable higher bird mortality than initially assumed.<sup>9</sup>

Further phylogenetic studies have reported an important differentiation between Central European and African USUV sequences, resulting in phylogeographic clustering of eight distinct lineages.<sup>10</sup> Phylogenetic trees from USUV European sequences indicated a common ancestor for the majority of them, except for some sequences from Spain<sup>8</sup> and Germany,<sup>9</sup> grouped in a branch divided into four well-supported clades.<sup>11</sup> Notably, sequences collected in Spain from birds differed in their phylogenetic origin than those from mosquitoes.<sup>12</sup> On the other hand, as mentioned above, a putative novel USUV lineage in Germany, called Europe 5, has been recently proposed.<sup>5</sup> Two other lineages, Europe 3 and Africa 2, were most likely introduced into Germany and recently spread to other western European countries, since the viruses detected in Belgium, France, and the Netherlands clustered with those previously circulating in mosquito vectors, wild birds, and/or bats in Germany between 2011 and 2014.<sup>13</sup>

According to a recent report,<sup>11</sup> the estimated time of the most recent common ancestor of the European strains dates back to 20.9 or 22.8 years, depending on whether the analysis is based on the NS5 or the E genomic sequences, thus suggesting that the probable time of arrival of USUV in Europe

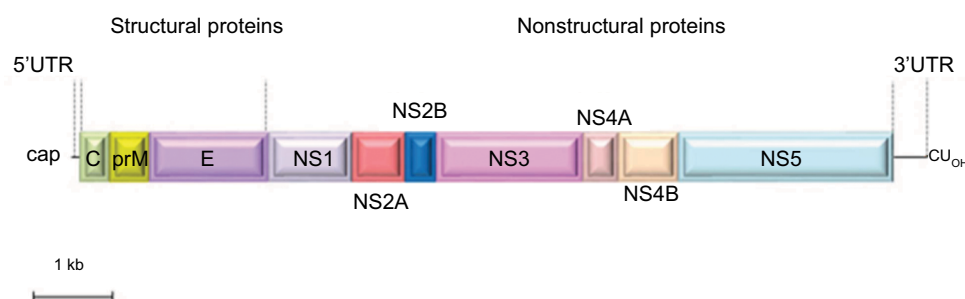
was in the middle of 1990s. This estimation is consistent with the retrospective identification of USUV in Italian dead bird samples from 1996.<sup>14</sup>

## Genome

Similar to other flaviviruses, USUV is a spherical, small, enveloped virus with a diameter of 40–60 nm. Its genome is constituted by a single-stranded RNA molecule of positive polarity. The sequence of USUV prototype strain, SAAR-1776, isolated from mosquitoes in South Africa in 1959, revealed a genome of 11,064 nt in length, containing a single open reading frame that encoded a polyprotein of 3,434 aa (Figure 2).<sup>1</sup> This polyprotein is flanked by two untranslated regions, a 96 nt region at the 5'-end and a 664 nt region at the 3'-end. The genomic RNA contains a 5'-cap structure, but lacks a polyadenylated tail. Untranslated region folding into stem loops present secondary structural elements that are necessary for the translation and replication of their genomes and are recognized and bind to different cellular proteins.<sup>15</sup> These regions have been suggested to interact with host-specific factors, since mutations in these regions led to limited host-range phenotypes.<sup>16</sup> These sequences also play a role in flaviviruses cytopathicity and pathogenicity.<sup>17</sup> In addition, it has been reported that USUV maintained the recognized conserved sequence motifs of the 3' stem loop present in yellow fever virus strains.<sup>18</sup>

## Proteins

As in other related flaviviruses,<sup>19,20</sup> the viral polyprotein is co- and posttranslationally processed by cellular and viral proteases into three structural proteins, capsid (C), pre-membrane/membrane (prM), and envelope (E), and seven nonstructural proteins (NS1, NS2A, NS2B, NS3, NS4A, NS4B, and NS5),<sup>2</sup> which promote virus replication through their protease (NS3), and polymerase and methyltransferase activities (NS5).<sup>1</sup>



**Figure 2** Schematic view of the genomic organization of USUV.

**Notes:** The single ORF (boxes) that encodes both structural (C-prM/M and E) and NS proteins (NS1, NS2A, NS2B, NS3, NS4A, NS4B, and NS5) is flanked by two UTRs.

**Abbreviations:** C, capsid; E, envelope; ORF, open reading frame; prM/M, pre-membrane/membrane; USUV, Usutu virus; UTRs, untranslated regions.

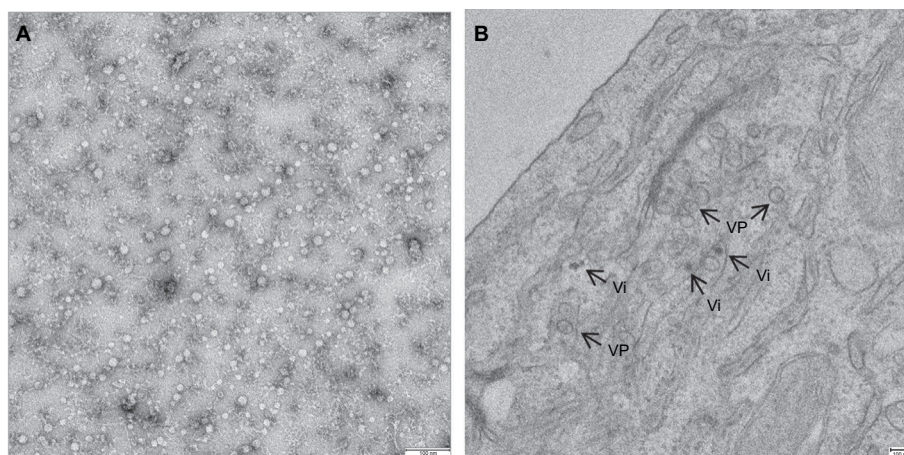
Cysteine residues within the polyprotein are well conserved among USUV strains with only slight differences.<sup>21</sup> Whether these changes affect the virus assembly or RNA replication is still unknown. Putative N-glycosylation sites (Asn-Xaa-Ser/Thr) have been identified at aa positions 118 and 154 of the E protein<sup>3</sup> and seem to be conserved among all USUV strains. In this sense, as in other related flaviviruses, functional significance of N-glycosylation sites has not been elucidated, as deglycosylated flaviviruses can keep the same antigenicity,<sup>22</sup> and their role in flavivirus life cycle still need to be clarified.<sup>23–25</sup> Flaviviral structural proteins are essential in viral RNA replication.<sup>26</sup> Thereby, the C protein associates with the genomic RNA to form the core of the virions.<sup>20</sup> The coexpression of the flavivirus prM and E glycoproteins induces formation of virus-like particles (Figure 3A) commonly referred to as recombinant subviral particles (RSPs) which, despite their reduced size, share both antigenic and immunogenic properties with whole virions.<sup>27</sup> The E protein, as reported for other flaviviruses, plays a role in cellular tropism, helping the virus to bind to specific receptors and initiate an infection.<sup>28</sup> The antigenic characteristics of the flaviviruses are mainly determined by their E protein structures.<sup>29</sup> In this regard, using computational tools, the presence of potential B-cell and T-cell epitopes on the E protein has been predicted,<sup>30</sup> showing a sequence identity of ~77.26% and a sequence similarity of ~85.02% with the E protein of WNV.

On the other hand, the specific functions of NS proteins of USUV are mainly unknown. Thus, the NS3 of viruses from the Flaviviridae family contains both a protease and a helicase coding region. A theoretical model of the three-dimensional structure of the helicase domain of USUV using as a template

the crystal structure of the Murray valley encephalitis virus helicase, the most phylogenetically related virus, showed similar structural features to those of other known Flaviviridae helicases.<sup>31</sup> The NS5, highly conserved among the USUV strains, is the viral RNA-dependent RNA polymerase<sup>32</sup> and, like in other flaviviruses,<sup>19</sup> it bears a methyltransferase domain necessary for capping the 5' end of the viral genomic RNA.<sup>3</sup>

## Host cell–virus interactions

USUV infectivity has been tested in several human and animal species cell lines. First reference of USUV growth in a micro-cultured method was performed in pig kidney cells (PS) in 1969.<sup>33</sup> Later on, the susceptibility to USUV infection was assessed in various cell lines and cultures, including HeLa (human), Vero (simian), ED (equine), PK-15 (porcine), RK-13 (rabbit), MDBK (bovine), MDCK (canine), DK (canine), CR (feline), BHK-21 (hamster), BF (hamster), C6 (rat), TH1 (turtle), primary goose embryo fibroblasts, and horse kidney cells.<sup>34</sup> Although replication was detected in all mammalian cell types, only Vero, PK-15, and goose embryo fibroblast cells developed cytopathic effects. Rodent cell lines derived from the bank vole were also susceptible to infection with USUV.<sup>35</sup> More recently, 13 additional human and animal cell lines have been shown to be susceptible either to USUV or Zika virus.<sup>36</sup> These cell lines (Tb 1 Lu, DF-1, Sf 1 Ep, EA.hy.926, FoLu, PI 1 Ut, OHH1.K, OK, DN1.Tr, LLC-MK2, BT, WCH-17, and Mv 1 Lu) included some domestic and peridomestic representative species. Even though both viruses replicate to significant titers in almost all cell lines, only 8 out of these 17 showed cytopathic effect. Susceptibility of ex vivo human neural cells to USUV infection has also been tested, showing



**Figure 3** Transmission electron microscopy of USUV RSPs and virions.

**Notes:** (A) Negative stained TEM image of purified Usutu RSPs. (B) Negative stained TEM image of Vero cells infected with USUV showing VP and electron-dense Vi.

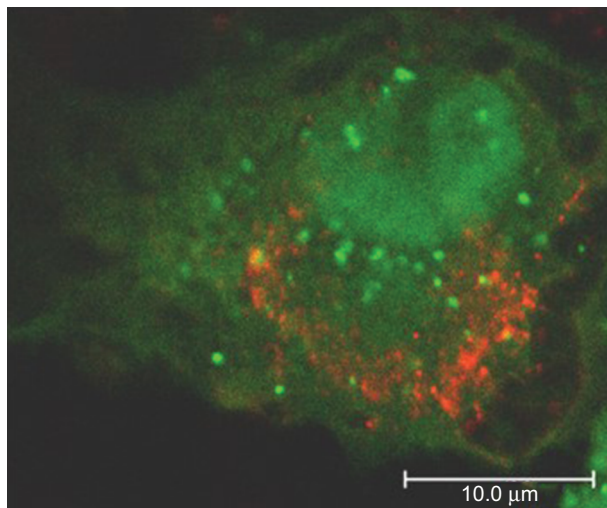
**Abbreviations:** RSPs, recombinant subviral particles; TEM, transmission electron microscopy; USUV, Usutu virus; VP, vesicle packets; Vi, virions.



that neurons, astrocytes, microglia, and induced pluripotent stem cells (iPSC)-derived human neuronal stem cells display strong cytopathic effect after viral infection.<sup>37</sup> The ability of USUV to replicate in cells of such different origin could be related to its transmission cycle, which includes replication in mosquito vectors and in a wide variety of mammalian hosts, such as birds, humans, and other vertebrates.

There is not much information available on how USUV enters the host cell. As reported for other flaviviruses, the mechanism of penetration into the cytoplasm must be initiated by the fusion of the viral E with the membranes of the cellular endosomes from the host cell, a process triggered by the acidic pH inside cellular endosomes.<sup>38</sup> The implication of cellular cofactors in membrane rearrangements induced by USUV has been analyzed by transmission electron microscopy after USUV infection, showing, as previously reported for other flaviviruses,<sup>39</sup> the presence of vesicle packets with electron-dense virions<sup>40</sup> (Figure 3B). Lipid requirement analyses for USUV replication have confirmed fatty acid and other cellular lipids as essentials for replication, since treatment with acetyl coenzyme A carboxylase inhibitors impaired USUV multiplication.<sup>40,41</sup> Likewise, the involvement of neutral sphingomyelinase in USUV infection has unveiled a connection between sphingomyelin metabolism and USUV biogenesis.<sup>42,43</sup>

Moreover, infection with USUV triggered an autophagic response in the host cell (Figure 4).<sup>44</sup> Autophagy is an important cellular pathway that plays important roles in viral infections and pathogenesis, and the upregulation of the autophagic pathway has been well documented for various flaviviruses,<sup>45</sup> although the involvement of autophagy in WNV infection is still controversial.<sup>46</sup>



**Figure 4** Confocal image of Vero cell transfected with the autophagosome marker GFP-LC3 plasmid and infected with USUV (red).

**Note:** Green LC3 aggregates indicate the induction of autophagy in the cell.

**Abbreviation:** USUV, Usutu virus.

Infection with USUV induces endoplasmic reticulum stress, which results in activation of a cellular signaling response termed unfolded protein response,<sup>44</sup> consistent with that observed for other members of the *Flavivirus* genus.<sup>45</sup>

## Vectors Mosquitoes

USUV, as other related flaviviruses such as WNV,<sup>19</sup> is mainly transmitted by ornithophilic *Culex* mosquitoes. In fact, USUV was isolated for the first time in 1959 from a female adult *Culex neavei* mosquito in South Africa.<sup>47</sup> Later on, the virus has been isolated from several mosquito species throughout the African continent, mainly in countries where surveillance programs are implemented, such as Senegal, CAR, Burkina Faso, Cote d'Ivoire, Nigeria, and Uganda,<sup>48</sup> and more recently in Kenya.<sup>49</sup> Experimental oral infection of *C. neavei* has been demonstrated and, although virus transmission to bitten mice was not initially observed, USUV infection, dissemination, and transmission rates were later on reported, showing the importance of high virus titers in the blood meal for virus infection.<sup>50</sup> In addition to *C. neavei*, USUV has been also isolated from other *Culex* species, including *Culex quinquefasciatus*, an anthropophilic vector that was directly implicated in the emergence of the virus in Europe,<sup>48</sup> and from *Aedes* mosquitoes.

After the emergence of USUV in Europe, viral genome amplification was achieved from *Culex pipiens* collected in Catalonia (Spain) in 2006,<sup>8</sup> and in the south of the country in pools from *Culex perexiguus* collected in 2009.<sup>51</sup> During a regional WNV surveillance program based on reverse transcription polymerase chain reaction (RT-PCR) detection conducted in Italy in the same year, 56 pools (54 of *C. pipiens* and 2 of *Aedes albopictus*) out of 1,789 mosquito pools were USUV positive by RT-PCR,<sup>52</sup> and similar results have been reported since then in different regions of Italy.<sup>53–57</sup> These findings confirm the role of *Culex* species as the main vector in the virus cycle, although *Aedes* mosquitoes also play a role on it, and point to an association between the presence of USUV-positive mosquitoes and drought conditions and a wide temperature range. USUV was also isolated from a pool of *C. pipiens* mosquitoes trapped in summer of 2010 in Germany<sup>58</sup> and from *Culex modestus* in Czech Republic the same year.<sup>59</sup>

## Hosts Birds

Nowadays, few data are available regarding USUV circulation in birds in Africa, where the virus was first isolated.

The virus was detected in Nigeria in 1972 in piping hornbill (*Bycanistes sharpei*), little greenbull (*Andropadus virens*), and Kurrichane thrush (*Turdus libonyana*) ([www.ncid.cdc.gov/arbocat/](http://www.ncid.cdc.gov/arbocat/)), and, more recently, one blackbird (*Turdus merula*) resulted positive against USUV in a serosurvey conducted in Morocco in 2008<sup>60</sup> and 9 out of 226 laughing doves (*Spilopelia senegalensis*) were USUV seropositive in Tunisia in 2015.<sup>61</sup>

In 2001, USUV was detected for the first time out of Africa in dead birds from Austria;<sup>62</sup> however, retrospective analysis of preserved tissue paraffin blocks from Italian wild birds found dead in 1996 resulted in the amplification of USUV RNA, thus indicating that the virus was already circulating in Europe at that time.<sup>14</sup> Later on, as part of WNV serosurveys conducted in different countries, USUV seropositivity has been found among different birds species (Table 1), mainly among blackbirds (*T. merula*).

Thereby, the presence of USUV-specific neutralizing antibodies was reported in the sera from healthy resident and migrant birds from different species, carrion crows (*Corvus corone*), magpies (*Pica pica*), and turkeys (*Meleagris gallopavo*) sampled between 2001 and 2002 in the UK, although

in many cases, the specificity of the antibodies could not be determined as cross-reactivity with WNV was observed.<sup>63</sup> Afterward, USUV seroconversion in chickens (*Gallus gallus*) sampled was described in 2004 in the UK.<sup>64</sup> However, further studies conducted in the UK did not confirm USUV circulation in the country.<sup>65,66</sup> Seroconversion was also observed in Italy in the summer of 2005<sup>67</sup> and in 2007–2008,<sup>68</sup> although no virus could be rescued from these animals, and in France in 2009–2010, even though cross-reactivity with WNV could not be discarded.<sup>69</sup>

A specific USUV surveillance program was conducted in Austria and Hungary between 2003 and 2006 in 322 dead birds from 52 species.<sup>70</sup> While no USUV was detected in 2003 or 2004, a blackbird resulted positive by RT-PCR, immunohistochemistry (IHC), and in situ hybridization in 2005, and six more birds were positive by next summer, from one of which the virus could be isolated and sequenced, displaying a very high similitude to the original Austrian strain found in 2001. Additional studies showed that 92/177 birds collected in 2003 in Austria were positive, most of them being blackbirds. The exceptions were two great tits (*Parus major*), one nuthatch (*Sitta europaea*), one song thrush (*Turdus*

**Table 1** Circulation of USUV among avian species in Europe

Order	Common name	Scientific name	Country	References
Passeriformes	Blackbird	<i>Turdus merula</i>	Austria	62, 71, 72
			Hungary	70
			Switzerland	76
			Italy	14, 54, 79, 80
			Germany	82, 83
			Czech Republic	86
	Barn swallow	<i>Hirundo rustica</i>	Austria	72
	Black redstart	<i>Phoenicurus ochruros</i>	Austria	72
	Blue tit	<i>Parus caeruleus</i>	Switzerland	76
			Germany	82
	Canary	<i>Serinus canaria domestica</i>	Germany	82
	Common reed bunting	<i>Emberiza schoeniclus</i>	Austria	72
	Common starling	<i>Sturnus vulgaris</i>	Italy	14
			Germany	82
	Eurasian blackcap	<i>Sylvia atricapilla</i>	Austria	72
	Eurasian bullfinch	<i>Pyrrhula pyrrhula</i>	Belgium	85
	Eurasian jay	<i>Garrulus glandarius</i>	Austria	72
			Italy	54, 71
	Eurasian nuthatch	<i>Sitta europaea</i>	Austria	71, 72
	European greenfinch	<i>Carduelis chloris</i>	Switzerland	76
	European pied flycatcher	<i>Ficedula hypoleuca</i>	Austria	72
	European robin	<i>Erithacus rubecula</i>	Austria	71, 72
			Switzerland	76

(Continued)

Table 1 (Continued)

Order	Common name	Scientific name	Country	References
Strigiformes	Garden warbler	<i>Sylvia borin</i>	Austria	72
	Great tit	<i>Parus major</i>	Austria	71, 72
	Hooded crow	<i>Corvus cornix</i>	Austria	72
	House martin	<i>Delichon urbica</i>	Austria	72
	House sparrow	<i>Passer domesticus</i>	Switzerland	76
			Germany	82
	Jackdaw	<i>Corvus monedula</i>	Austria	72
	Lesser whitethroat	<i>Sylvia curruca</i>	Austria	72
	Magpie	<i>Pica pica</i>	France	69
			Italy	54
	Redstart	<i>Phoenicurus phoenicurus</i>	Germany	73
	Reed warbler	<i>Acrocephalus scirpaceus</i>	Austria	72
	Song thrushes	<i>Turdus philomelos</i>	Spain	12
			Germany	83
			Austria	71, 72
	Whitethroat	<i>Sylvia communis</i>	Austria	72
	Barn owl	<i>Tyto alba</i>	Austria	72
	Eagle owl	<i>Bubo bubo</i>	Austria	72
	Hawk owl	<i>Surnia ulala ulala</i>	Switzerland	76
	Great gray owl	<i>Strix nebulosa lapponica</i>	Switzerland	76
			Germany	82 83
	Long-eared owl	<i>Asio otus</i>	Austria	72
			Italy	54 79, 80
			Germany	83
Galliformes	Northern hawk-owl	<i>Surnia ulula</i>	Germany	83
	Pygmy owl	<i>Glaucidium passerinum</i>	Switzerland	76
	Snowy owl	<i>Nyctea scandiaca</i>	Switzerland	76
	Tawny owl	<i>Strix aluco</i>	Austria	72
	Tengmalm's owl	<i>Aegolius funereus</i>	Switzerland	76
	Ural owl	<i>Stix uralensis</i>	Austria	72
	Chickens	<i>Gallus gallus domesticus</i>	Italy	67, 68
	Common pheasants	<i>Phasianus colchicus</i>	Spain	81
			Austria	72
	Indian peafowl	<i>Pavo cristatus</i>	Austria	72
	Red-legged partridges	<i>Alectoris rufa</i>	Spain	81
			Italy	54
Accipitriformes	Osprey	<i>Pandion haliaetus</i>	Germany	73
	Bearded vulture	<i>Gypaetus barbatus</i>	Austria	72
	Marsh harrier	<i>Circus aeruginosus</i>	Austria	72
Anseriformes	Mute swan	<i>Cygnus olor</i>	Serbia	84
	Mallard	<i>Anas platyrhynchos</i>	Italy	80
Charadriiformes	Black-headed gull	<i>Larus ridibundus</i>	Poland	74
	Inca tern	<i>Larosterna inca</i>	Germany	83
Ciconiiformes	White stork	<i>Ciconia ciconia</i>	Germany	73
Columbiformes	Eurasian collared dove	<i>Streptopelia decaocto</i>	Austria	72
			Italy	54
Caprimulgiformes	Rock pigeon	<i>Columba livia</i>	Italy	80
	Nightjar	<i>Caprimulgus europaeus</i>	Italy	54
Coraciiformes	Common kingfisher	<i>Alcedo atthis</i>	Germany	82
Falconiformes	Krestel	<i>Falco tinnunculus</i>	Austria	72
Gruiformes	Common coot	<i>Fulica atra</i>	Czech Republic	75
Piciformes	European green woodpecker	<i>Picus viridis</i>	Germany	83
	Great spotted woodpecker	<i>Dendrocopos major</i>	Austria	72
			Belgium	85

Abbreviation: USUV, Usutu virus.

*philomelos*), and one European robin (*Erithacus rubecula*); however, only 11/224 birds in 2004 and 4/103 in 2005 proved positive, all of which were blackbirds.<sup>71</sup> Although the drop in screened positive birds in the region during the course of the sampled years may be due to herd immunity acquired in previous seasons, the study showed that the virus spread from the original area of emergence to surrounding areas of the country. In agreement with these results, a serosurvey conducted in eastern Austria in the same years showed that the number of positive birds increased each year,<sup>72</sup> and reported the presence of positive serology in birds from different orders (Strigiformes, Galliformes, Accipitriformes, Falconiformes, Piciformes, Columbiformes), with Passeriformes, mainly blackbirds, being again the most affected birds. This study also suggested the occurrence of maternal transfer of antibodies in Ural owls.<sup>72</sup>

Analysis of over 3,000 bird samples for the presence of WNV antibodies in Germany (2002–2005) reported the presence of USUV neutralizing antibodies only in three birds:<sup>73</sup> one white stork (*Ciconia ciconia*), one redstart (*Phoenicurus phoenicurus*), and one osprey (*Pandion haliaetus*). One year later, positive serology was also reported in one black-headed gull (*Larus ridibundus*) in Poland<sup>74</sup> and in one common coot (*Fulica atra*) in the Czech Republic.<sup>75</sup>

Along the summer of 2006, dead birds were found around the Zurich zoo in Switzerland and further investigations point to USUV as the causative agent. In the following year, several birds from the zoo died of USUV infection.<sup>76</sup> Consequently, a study was conducted in sera from 372 birds collected in 2006 and 2007 in the zoos of Vienna, Basel, Zurich, and Budapest. A seroprevalence of 8.75%, 6.59%, 5.3%, and 0% specific neutralizing antibodies was found in the samples from these zoos, respectively,<sup>77</sup> with the highest USUV seroprevalence found among owls (*S. nebulosa*).

In 2008 and 2009, bird-based surveillance studies conducted in Italy found 12 and 11 USUV-positive samples out of over 1,000 samples tested per year.<sup>52,54</sup> An additional study conducted in the same years in the Po valley also detected USUV in dead birds,<sup>78</sup> and USUV infection was diagnosed in the country at that time in two free-living blackbirds and three captive owls.<sup>79</sup> Afterward (2012–2013), a serosurvey conducted in 47 different avian species found 1.35% (4/297) samples with neutralizing antibodies against USUV.<sup>80</sup>

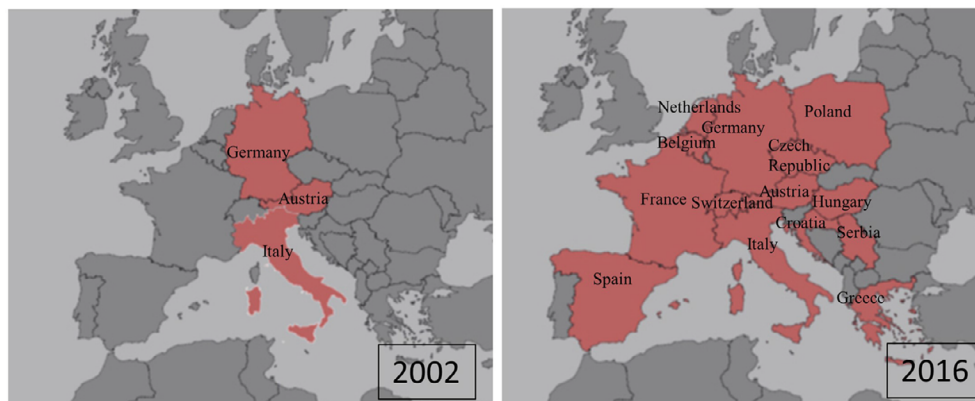
A flavivirus surveillance study carried out in southern Spain in 2011–2012 in hunted red-legged partridges (*Alectoris rufa*) and common pheasants (*Phasianus colchicus*) reported a 13% prevalence of anti-USUV neutralizing antibodies.<sup>81</sup> In the same area and year, USUV detection in

two song thrushes (*T. philomelos*) confirmed the virus as the cause of the diagnosed encephalitis in these birds.<sup>12</sup> At that time, USUV was also detected among several resident and migratory bird species in Germany,<sup>82</sup> and since then, the virus has been frequently found in dead birds in the southwest of the country.<sup>83</sup> USUV RNA-positive samples were amplified from blackbirds, thrush (*Turdus* sp.), long-eared owl (*Asio otus*), European green woodpecker (*Picus viridis*), inca tern (*Larosterna inca*), great gray owl (*S. nebulosa*), and northern hawk owl (*Surnia ulula*) along 2012 and 2013. Neutralizing specific antibodies were also found in a mute swan (*Cygnus olor*) collected in Serbia in 2012.<sup>84</sup> The same year, USUV RNA was detected in a Eurasian bullfinch (*Pyrrhula pyrrhula*) held in captivity for its entire lifespan and in a wild great spotted woodpecker (*Dendrocopos major*), both with neurologic signs, in Belgium.<sup>85</sup> The virus was also isolated from a blackbird found dead in Brno (Czech Republic) in 2011, and viral RNA amplification was achieved from two other blackbirds 1 year later in the same area.<sup>86</sup> In 2014, during a WNV surveillance conducted in Greece, USUV seropositivity was reported among pigeons (*Columba livia domestica*).<sup>87</sup> One year later, a French surveillance network reported increased fatalities in blackbirds in eastern France, and molecular analysis detected USUV in five bird tissues.<sup>88</sup> The spread of USUV throughout the continent has been further supported by its recent emergence in the Netherlands, where an outbreak, with a considerable bird mortality in blackbirds and great gray owls, has been recently reported.<sup>89</sup> In addition, widespread USUV activity was also described in 2016 in Belgium, France, and Germany, with 17 live and 147 dead USUV-positive birds in these countries.<sup>13</sup> Phylogenetic analysis indicates that the virus was most probably introduced from Germany by infected semi-resident birds, although the role of migrating birds or mosquitoes cannot be excluded. All these data clearly demonstrated a spread of USUV activity throughout the continent (Figure 5).

It is noteworthy that, even though information on the distribution and pathogenicity of USUV in Africa is scarce, apparently, it is a nonpathogenic virus for local bird populations, which could be due to the immunity that originated in the bird population as a result of exposure to USUV and other antigenically related flaviviruses over a long period of time.<sup>1</sup>

Histopathologic and IHC studies in birds have frequently described hepatomegaly and splenomegaly.<sup>12,54,62,76,82,89</sup> Histologic lesions have also been reported in heart, liver, kidney, and spleen. Slightly swollen liver and hyperemic lungs, as well as discrete histopathologic lesions in heart and liver, neuronal necrosis, and leukocytolysis in brain blood vessels are





**Figure 5** Map showing the spread of Usutu across Europe from 2002 to 2016.

less frequently observed. A more detailed study<sup>70</sup> described unspecific gross lesions, such as general organ congestion, and the presence of acute hepatitis with multiple inflammatory and necrotic foci, focal necrosis in the spleen, perivascular infiltrations in the kidney and heart, focal myocardial degeneration, vacuolar degeneration of tubular epithelial cells in the kidney, and perivascular and perineuronal edema in the brain. Likewise, immunoreactive accumulation of glial cells in the brain was detected by IHC, and multiple IHC-positive foci were present in the cerebral cortex. USUV antigens were also observed, to different extents, in the heart, lungs, spleen, pancreas, and intestines. Another study conducted in two thrushes with severe encephalitis showed congestion and Purkinje cell necrosis, gliosis, satellitosis, neuronophagia, and endothelial cell swelling and vasculitis, along with some lesions in the kidney, liver, and spleen.<sup>12</sup> All these findings indicating USUV replication in a large number of organs and tissues point to multiorgan failure as the cause of death.

## Humans

USUV has been isolated from human sera in Africa only twice: in 1981 in the CAR from a patient with fever and rash, and in 2004 in Burkina Faso from a 10-year-old patient with fever and jaundice.<sup>48</sup>

In 2009, the first case of USUV infection in humans was reported in Europe. A patient with a neuroinvasive infection with diffuse large B-cell lymphoma who presented with fever and neurologic symptoms was diagnosed with meningoencephalitis in Italy. USUV was amplified by RT-PCR from the cerebrospinal fluid (CSF), serum, and plasma samples of the patient.<sup>90</sup> The same year, a case of USUV-related illness was reported in Italy in a woman who received an orthotopic liver transplant as a final consequence of a thrombotic thrombocytopenic purpura during an acute episode of USUV infection.<sup>91</sup>

Later on, the only USUV neuroinvasive infection in humans outside Italy was described in Croatia in 2013 during a WNV outbreak.<sup>92</sup> The main clinical features in all patients were headache, fever, nuchal rigidity, hand tremor, and hyperreflexia. USUV neutralizing antibodies were detected in all patients, and seroconversion was documented by enzyme-linked immunosorbent assay (ELISA) in two of them.

The first screening using an in-house ELISA detected the presence of specific IgG against USUV in 1.1% (4/359) blood donors tested in northern Italy without history of other flavivirus infections.<sup>93</sup> A further study on the presence of neutralizing antibodies against USUV in 6,000 sera collected in 2010–2011 from healthy blood donors in the northeastern part of the country found 14 (0.23%) samples positive for USUV.<sup>94</sup> A survey conducted in Germany in 4,200 serum samples from healthy blood donors collected in January 2012 detected only one USUV IgG- and IgM-positive sample.<sup>95</sup> However, analysis of 13,023 blood donations for the presence of WNV antibodies revealed an acute USUV infection in one of the donors.<sup>96</sup> Evidence of USUV circulation has also been reported in Croatia in 2013 after detecting USUV neutralizing antibodies in 3 out of 95 patients with clinically suspected WNV infection.<sup>97</sup>

Reports of viremia or USUV RNA-positive samples in humans are scarce. Molecular analysis conducted in 2010 on the plasma/serum and/or CSF sampled from 30 human patients suffering from acute meningoencephalitis did not give any USUV-positive result;<sup>54</sup> neither positive amplification was obtained by RT-PCR in 96 CSF samples from patients with encephalitis of unknown etiology diagnosed in 2010–2013 in Tuscany, Italy.<sup>98</sup> However, USUV RNA was detected by quantitative real-time polymerase chain reaction in the CSF specimens of three immunocompetent patients suffering from meningoencephalitis.<sup>99</sup> Likewise, very recently,

a retrospective analysis reported the presence of USUV RNA in 1.1% and USUV antibodies in 6.57% of the CSF and serum samples collected between 2008 and 2011 from 915 patients with or without neurologic impairments in the area of Modena, Italy.<sup>100</sup>

## Other vertebrates

Information about USUV circulation in vertebrate species other than birds and humans is scarce. A flavivirus isolate obtained from a furred rat (*Praomys* sp.) in CAR, initially described as USUV, turned out to be a related flavivirus, the Yaounde virus.<sup>48</sup> Even though no isolation from mammals other than humans has been reported, guinea pigs and newborn and weaning mice have been shown to be sensitive to intracerebral inoculation with USUV.<sup>101</sup> Although it has been described that a rodent species (*Arvicanthis abyssinicus*) showed a trace of viremia after intraperitoneal inoculation,<sup>102</sup> the susceptibility of USUV in adult mice is limited.<sup>103</sup>

Circulation of USUV in horses was reported in Europe during a 2-year study (2008–2009) carried out in Italy,<sup>104</sup> during which time a high neutralizing antibody titer against the virus was also detected in a horse in Serbia.<sup>105</sup> Likewise, during 2011, 2/69 WNV ELISA-reactive horse sera collected in two different locations in Croatia resulted USUV positive by virus neutralization test.<sup>106</sup> Later on, a study carried out in 172 horses sampled between 2011 and 2012 in the island of Mallorca (Spain) found a 1.2% seroprevalence of antibodies against USUV and reported seroconversion in some of the animals included in the study.<sup>107</sup> Similarly, seroneutralization tests conducted in military working horses and dogs in Morocco in 2012 suggested exposure of these animals to USUV infection,<sup>108</sup> and in 2014, another study reported the presence of antibodies against USUV in 10 equids from southwestern Tunisia.<sup>109</sup> In 2013, two dead *Pipistrellus* bats found in southwest Germany resulted USUV positive and the full genome sequences were directly amplified from their brains.<sup>7</sup> Presence of USUV-specific neutralizing antibodies has also been reported in four wild boars hunted in Serbia in 2011–2012,<sup>110</sup> and a retrospective serosurvey conducted in 4,693 wild ruminants, including red deer (*Cervus elaphus*), fallow deer (*Dama dama*), mouflon (*Ovis aries musimon*), and roe deer (*Capreolus capreolus*), collected between 2003 and 2014 from the hunting states across Spain reported a prevalence of USUV-specific antibodies of 0.1%–0.2%.<sup>111</sup>

## Diagnosis

Virus isolation in susceptible cell culture is the gold standard for virus detection, but it is usually hampered by the

short duration and low levels of viremia, and thus, it is not routinely applied for USUV diagnosis. Earlier, the diagnosis was based on complement fixation and agar gel precipitation technique,<sup>112,113</sup> and then, viral neutralization tests and immunofluorescence assays were implemented.<sup>114</sup> However, as with any flavivirus, cross-reactivity between viral antigens is the greatest drawback for proper serologic diagnosis,<sup>19</sup> and thus, sera have to be tested against different related viruses and results have to be subsequently confirmed by different assays, such as immunofluorescence assay, hemagglutination inhibition, or, preferably, plaque reduction neutralization test. A 4-fold increase in plaque reduction neutralization test titers between two sequential serum samples collected 2–3 weeks apart usually confirms an acute infection, and USUV neutralizing titers 4-fold higher than titers to other related flavivirus can usually be taken as a specificity probe. In any case, diagnosis has been mainly based on in-house serologic and molecular tests, since only very recently, a commercial USUV IgG ELISA (EUROIMMUN, Lübeck, Germany) as well as a recombinant NS1 NS protein (The Native Antigen company, Oxford, UK) have been made available.

On the contrary, no molecular kit has yet been commercialized for USUV diagnosis. The first USUV-specific real-time RT-PCR targeting the NS5 gene and based on European strains was reported in 2011.<sup>99</sup> The assay can be used in human blood and CSF samples, and showed a sensitivity of 50 genomic copies per reaction (corresponding to 2,200 copies/mL) and 1 plaque forming units (PFU)/mL of USUV isolate. Later on, a similar assay based on sequences from Europe and Africa,<sup>115</sup> and a one-step real-time multiplex RT-PCR, which efficiently detects and differentiates in a single reaction WNV strains from lineages 1 and 2 and USUV, have been reported.<sup>116</sup>

More recently, a NS1-antigen protein microarray for the serologic differential diagnosis of flavivirus infections in horses, including USUV, has been developed and validated.<sup>117</sup> In any case, the lately widespread occurrence of USUV throughout Europe, the relatively high avian mortality recorded, and the reported human cases make necessary the development of accurate, fast, and economically affordable specific diagnostic tools.

## Prevention

Nowadays, there are neither specific licensed drugs nor vaccines to fight USUV infection, and thus, the only efficient prophylaxis is avoiding mosquito bites. Thereby, since the number of human cases reported until now is too low to have experience with therapeutic interventions, as with other flaviviruses, current treatment against USUV is based only

on supportive care measures, such as intravenous antipyretics, fluids, respiratory support, and prevention of secondary infections. Despite this fact, many different drugs are being tested against other related flaviviruses, such as WNV<sup>118</sup> and ZIKV,<sup>119</sup> which, in many instances, will more probably be useful against USUV. Indeed, some investigations have already reported the activity of several drugs against USUV multiplication in cell culture that may help to develop future antiviral strategies. So far, the acetyl-CoA carboxylase inhibitors TOFA and MEDICA 16<sup>120</sup> and valproic acid<sup>121</sup> have been shown to inhibit USUV production in vitro. Treatment with inhibitors of autophagy 3-methyladenine or wortmannin also showed a significant reduction in the virus yield in USUV infection, since this virus takes advantage of autophagy for replication.<sup>44</sup>

Even fewer data have been published regarding vaccines against USUV, as vaccine development against some flaviviruses present several drawbacks: relatively low incidence, cost-effectiveness, implementation for wild life vaccination, availability of animal models, and so on. For instance, in the case of USUV, only laboratory suckling mice, but not adult mice, exhibit susceptibility to infection, thus hampering the analysis of the immune responses and complicating the evaluation of vaccine candidates against this pathogen.<sup>103</sup> In any case, vaccination of adult mice with WNV RSPs induced low, although detectable, levels of circulating IgG cross-reactive with USUV, and a boost in these levels was observed upon infection with USUV; however, protection studies of mice immunized with WNV RSPs against USUV challenge were not conducted, as no lethal challenge model for USUV was available.<sup>27</sup> Later on, it was shown that mice deficient in the alpha/beta interferon receptor (IFNAR [−/−] mice) were highly susceptible to USUV infection, thus providing a lethal challenge model for vaccine testing. In fact, a single intramuscular immunization of these mice with a plasmid DNA vaccine candidate encoding the precursor of membrane (prM) and E proteins of USUV was sufficient to elicit a significant level of protection against challenge with USUV and to prime the production of anti-USUV neutralizing antibodies.<sup>122</sup>

In any case, as mentioned above, nowadays, the main preventive measures against USUV and other related flaviviral infections are prevention of mosquito bites responsible for disease transmission. These measures can be summarized as: implementation of mosquito control programs, use of insect repellents, minimization of skin surface exposed to mosquito bites, elimination of standing water where mosquitoes can lay eggs, installation of window and door screens, and

minimization of outdoor activities coincident with the maximum activity of mosquitoes. An early detection of enzootic circulation based on mosquito and avian surveillance should be performed to support prevention and control measures.

## Conclusion

Recent expansion of different flaviviruses, such as WNV and ZIKV, to new geographic regions has caused social alarm and demands health attention. In this sense, USUV is an obvious example for the introduction, spread, and establishment of tropical arboviruses in moderate climate. In fact, since its first detection in Europe in 2001, the wide and quick spread of USUV throughout Europe, the relatively high avian mortality recorded, and the reported human cases have made USUV a potential public health concern. Based on the previous behavior of other related arboviruses, at this moment, a risk that further large outbreaks will occur in regions already colonized by the virus, or in new regions where USUV is not yet established, cannot be dismissed. Thus, there is a need for the development of USUV-specific diagnosis tools to help clinicians to discriminate encephalitis cases of unknown etiology. In addition, further USUV circulation surveillance and control measures should be established around Europe. Likewise, although the current knowledge based on epidemiologic data and clinical manifestations of the infection does not urge to routinely test blood donations, we should be aware that additional measures may have to be implemented. Therefore, as with any other infectious emerging pathogen, multidisciplinary interventions and research that includes virologists, ornithologists, entomologists, climatologists, veterinarian, and physicians should be conducted to increase our current knowledge about USUV, and to do so, policymakers should provide and implement the adequate guidelines and funding to coordinate them between European countries.

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## Disclosure

The authors report no conflicts of interest in this work.

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