

Multifunctional gold nanorods and docetaxel-encapsulated liposomes for combined thermo- and chemotherapy

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Abstract: Personalized and precise nanomedicines are highly demanded for today's medical needs. Liposomes are ideal candidates for the construction of multifunctional drug delivery systems. In this study, a liposome was used to improve the clinical issues of docetaxel (Doc), a potent antimitotic chemotherapy for prostate cancer (PC). RLT, a low-density lipoprotein receptor (LDLR)-binding peptide, and PEG were conjugated to the liposomes, and gold nanorods (GNRs) were also incorporated into the liposomes. The GNRs/Doc-liposome-RLT (GNRs/DocL-R) was tested in PC-3 cells and in PC-3 tumor-bearing nude mice. Results showed that GNRs/DocL-R possessed a diameter approximately 163.15 ± 1.83 nm and a zeta potential approximately -32.8 ± 2.16 mV. GNRs/DocL-R showed enhanced intracellular entrance, increased accumulation in the implanted tumor region, and the highest tumor inhibition in vitro and in vivo. Therefore, the multifunctional GNRs/DocL-R was a potential cancer treatment via combined chemo- and chemotherapy.

Keywords: gold nanorods, docetaxel, liposomes, prostate cancer, LDL receptor targeting

Introduction

Cancers are still unmet medical issues with increasing incidence and high mortality. Cancer cells are difficult to control because of their fast proliferation, invasive migration, poor drug permeability, adaptive drug resistance, and multiple self-protective mechanisms. A combined therapy of two or more anticancer agents is an ideal strategy for enhanced efficacy. The combined therapy should possess minimum toxicity to healthy tissues and organs. Gold nanorods (GNRs) have been explored as anticancer agents¹⁻³ for photothermal therapy (PTT). GNRs possess a transverse absorption at the wavelength of 520 nm and a longitudinal peak at the near-infrared (NIR) region.^{4,5} GNRs have the maximum absorption and the highest heating efficiency around 800 nm.⁶ NIR light has deep tissue penetration and causes minimum damage to normal tissues.⁷ By applying NIR light, absorbed light by GNRs was converted into local heating⁸ that would cause cell injury/death via cell necrosis and/or apoptosis.^{2,9} Docetaxel (Doc) is a potent and broad-spectrum anticancer drug that acts on the synthesis, migration, and division of microtubules to inhibit the growth of cancer cells.¹⁰ However, GNRs possessed poor stability in biological environment and reduced cellular uptake.¹¹ Doc is a lipophilic compound that has poor specificity and high toxicity in vivo.¹⁰ These disadvantages limit the applications and anticancer efficacy of GNRs and Doc.

Liposomes have been the earliest and most well-developed drug delivery systems since 1965. Over the past 5 decades, many efforts have been performed to prolong

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the *in vivo* circulation of liposomes, control the release rate of drugs from liposomes, and improve the tumor targeting of liposomes; therefore, some liposomal formulations have entered the market.¹² The natural merits of liposomes make them to be ideal candidates for the construction of multi-functional drug delivery systems and advantageous tools to apply combined therapies.

Low-density lipoprotein receptors (LDLRs) were over-expressed on the membrane of cancer cells to take extra essential fatty acids for cell proliferation.¹³ Hayavi and Halbert¹⁴ first synthesized an LDLR-binding peptide RLT (CEKLKEAFRLTRKRLKLA, underlined sequence represented the binding region to LDLR) that preserved the binding sequence of native low-density lipoprotein (LDL). Lysine (Lys) and Arginine (Arg) are amino acids with positive charges and were added to RLT to assist endo-lysosome escape. Later studies proved that RLT could bind to LDLR and be taken up via LDLR-mediated endocytosis. RLT could also facilitate the delivery of paclitaxel to glioblastoma multiforme cells and improve the efficacy of paclitaxel.^{15,16} RLT-modified DNA nanocomplexes showed enhanced intracellular delivery of doxorubicin (Dox) and higher tumor cell inhibition.¹⁷ In addition, *in vivo* studies confirmed that RLT increased the tumor accumulation of submicron emulsion and GNRs in mice.^{3,18} In this study, liposomes are used to improve the application of GNRs and Doc by encapsulating lipophilic Doc into the bilayer of liposomes and loading GNRs into the hydrophilic core of liposomes. Polyethylenes glycol (PEG) and RLT were conjugated to the liposomes to increase the stability and tumor targeting of liposomes. The PEG- and RLT-modified liposomes, and GNRs and Doc-loaded liposomes (GNRs/Doc-liposome-RLT, GNRs/DocL-R), were then characterized and investigated for *in vitro* and *in vivo* antitumor efficacy and safety.

Materials and methods

Materials

Sodium borohydride (NaBH_4), chloroauric acid trihydrate ($\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$), cetyltrimethyl-ammonium bromide (CTAB), silver nitrate (AgNO_3), sodium sulfate (Na_2SO_4), and L-ascorbic acid (AA) were purchased from Alfa chemicals (Berkshire, UK). Ultrapure Milli-Q water (18 $\text{M}\Omega \text{ cm}$; Millipore-Q; EMD Millipore, Billerica, MA, USA) was used for all solution preparations. Sangon Biotech (Shanghai, China) provided cholesterol (chol). Aladdin (Shanghai, China) supplied soy phosphatidylcholine (SPC) and hydrogenated SPC (HSPC). 1,2-Distearoyl-sn-glycero-3-phosphoethanolamine-conjugated polyethylene glycol (DSPE-PEG (2000)-Mal) was

obtained from Nanocs (NY, USA). RLT(-SH) and fluorescein isothiocyanate (FITC)-labeled RLT(-SH) were synthesized by AC Scientific (Xi'an, China). Doc and Dox were purchased from Hisun (Zhejiang, China). HyClone (Logan, UT, USA) supplied penicillin-streptomycin solution (100 $\times$) and 0.25% trypsin. MTT kit was purchased from Sigma-Aldrich Co. (St Louis, MO, USA). Solarbio® Life Sciences (Beijing, China) provided dimethyl sulfoxide (DMSO), RPMI-1640 cell culture medium, apoptosis assay kit, cell cycle kit, and reactive oxygen species (ROS) probe (dichloro-dihydro-fluorescein diacetate [DCFH-DA]). 1,1-Dioctadecyl-3,3,3,3-tetramethylindotricarbocyanine iodide (DiR iodide) was purchased from AAT Bioquest (Sunnyvale, CA, USA). Prostate cancer cell line (PC-3) was obtained from Chinese Academy of Science Cell Bank for Type Culture Collection (Shanghai, China).

Preparation and characterization of GNRs/DocL-R

GNRs were synthesized using a modified seed-mediated growth method in aqueous solution as described previously (Supplementary materials).¹⁹⁻²¹ GNRs/DocL-R was prepared by using lipid-film hydration method.²² Briefly, chol (10 mg, 25.9 nM), SPC (150 mg, 193.4 nM), HSPC (5 mg, 6.4 nM), Doc (5.5 mg, 6.8 nM), and DSPE-PEG (2000)-Mal (2.5 mg, 0.88 nM) were dissolved in chloroform using bath sonication. Chloroform was then removed by rota-evaporation, and the lipid solution was dried to a thin and transparent film at the bottom of flask. GNR solution (0.4 nM, 4 mL) was then added to the flask to hydrate the film. The flask was sonicated in ice bath for 40 times (200 W) (every sonication lasted for 3 s; every sonication contained two 3 s sonication), and the obtained solution was centrifuged at 8,000 rpm for 10 min. Supernatant was obtained and mixed with RLT (1.2 μg , 0.44 nM) solution (24 μL). The mixture was vortexed for 1 min and shaken for 12 h at room temperature to obtain GNRs/DocL-R.

Size and zeta potential of GNRs/DocL-R were analyzed by Zetasizer Nano ZS90. The morphology of GNRs/DocL-R was observed under transmission electron microscopy (TEM; TecnaiG20; FEI, Hillsboro, OR, USA). Briefly, fresh samples were diluted in Millipore water and mixed well. Before visualization, the fresh samples were placed on copper grids with film and settled for 1 min. Excess liquid was absorbed by paper, and the samples were air dried before TEM examination.

Loading and release of Doc and thermal responses of GNRs/DocL-R

Loading efficiency (LE) and loading capacity (LC) were analyzed by high-performance liquid chromatography

(HPLC; Agilent 1100 [Beijing, China]; flow rate: 1.0 mL/min; mobile phase: acetonitrile/water 53/47; column: Diamonsil™ C18, 4.6×200 mm, 5 μm; detection wavelength: 230 nm, and temperature: 25°C). The *in vitro* release of Doc was carried out using a dialysis unit (molecular weight cutoff [MWCO] 20,000 Da; Thermo Fisher Scientific, Waltham, MA, USA) at 37°C. Briefly, 1 mL of GNRs/DocL-R (concentration of GNRs 0.4 nM) was sealed to a dialysis unit. The dialysis unit was submerged into 50 mL PBS that contained 0.5% (w/w) Tween 80 (pH 7.4 or pH 5.0). The dialysis unit with GNRs/DocL-R (concentration of GNRs 0.4 nM) + laser was exposed to laser radiation (808 nm) for 10 min. At set time points (0.5, 1, 2, 4, 8, 12, 24, 48, 72, 96, and 120 h), 1 mL of release media was withdrawn and 1 mL of fresh 0.5% (w/w) Tween 80 PBS solution was refilled. The obtained release samples were centrifuged at 10,000 rpm for 10 min. Supernatants were analyzed by HPLC to calculate the cumulative release (%) of Doc.

To test thermal response, GNRs/DocL-R was exposed to laser radiation at 808 nm for 10 min. Temperature increase was recorded at set time points. Curves of thermal response to laser radiation were plotted.

In vitro cell inhibition

PC-3 cells were in an exponential growth phase and cultured in RPMI-1640 medium supplemented with 10% (v/v) fetal bovine serum (FBS; 5% CO₂, 37°C) unless otherwise specified. Cells were incubated for 24 h prior to the experiment. Cell inhibition was evaluated by the standard MTT assay. Briefly, a serial dilutions of GNRs, GNRs + laser, Doc, GNRs/DocL, GNRs/DocL + laser, GNRs/DocL-R, and GNRs/DocL-R + laser (corresponding Doc concentrations 1, 10, 20, 40, 60, 80, 100, and 120 μg/mL) were added to cells in 96-well plates (cell density 1×10⁵ cells/well) and incubated for 12, 24, 48, and 72 h to determine cell viability. Groups with laser radiation (808 nm) were exposed to laser for 5 min after 4 h of treatment. Standard MTT assay was conducted after these treatments. The absorbance of samples was measured at 490 nm using a Synergy H1 plate reader (BioTek, Winooski, VT, USA). The cell inhibition rate was calculated, and the time- and concentration-dependent cell inhibition curves were plotted.

Intracellular uptake

PC-3 cells were seeded at a density of 2×10⁵ cells/well in six-well plates over glass cover slips. Dox was selected as the fluorescent probe, and RLT was labeled with green FITC. Cells were incubated with Dox (1 μg/mL), GNRs/

DoxL (Dox concentration of 1 μg/mL), and GNRs/DoxL-R (Dox concentration of 1 μg/mL) for 1, 2, or 4 h at 37°C in the dark. Subsequently, the culture medium was removed, and the cells were washed thrice with PBS (pH 7.4). About 70% cold ethanol was used to fix the cell samples for 15 min at room temperature and gently washed twice with PBS. The prepared samples were observed using confocal laser scanning microscopy (Olympus FV1000; Olympus Corporation, Tokyo, Japan). Intracellular uptake was quantified using flow cytometry. After incubation with treatments for 1, 2, or 4 h at 37°C, trypsin was added to PC-3 cells, and the detached cells were collected into centrifuge tubes. The cells were washed with PBS thrice. The purified cell samples were collected into fluorescence activated cell sorting tubes to quantify fluorescence signals.

Flow cytometric analysis

Cell apoptosis analysis

Doc (50 μg/mL), GNRs + laser, and GNRs/DocL-R (+laser) (Doc concentration of 50 μg/mL) were added to PC-3 cells (cell density of 2.5×10⁵ cells/well) in six-well plates and incubated with the cells for 24 h. Subsequently, the cells were washed with PBS three times and incubated with trypsin (no EDTA). The obtained cell suspensions were centrifuged at 1,000 rpm for 10 min, and the cell pellets were suspended with 1 mL of cold PBS. The cells were washed twice using centrifugation. The purified cell pellets were suspended in 200 μL of binding buffer and then mixed with Annexin V-FITC (10 μL) and propidium iodide (PI; 10 μL). The cell solutions were incubated for 15 min in the dark. The additional 300 μL of binding buffer was gently added. The prepared samples were tested immediately using flow cytometry.

Cell cycle analysis

Doc (50 μg/mL), GNRs/DocL-R (Doc concentration of 50 μg/mL), and GNRs/DocL-R (+laser) (Doc concentration of 50 μg/mL) were added to PC-3 cells (cell density of 2.5×10⁵ cells/well) in six-well plates and incubated with the cells for 24 h. Subsequently, the cells were washed with PBS three times and incubated with trypsin (no EDTA). The cell suspensions were centrifuged at 1,000 rpm for 10 min. The cell pellets were suspended with 500 μL of cold ethanol and kept at 4°C overnight in the dark. The cell samples were washed with PBS before staining. RNase A (100 μL) was used to suspend the cells, and the cell suspensions were incubated for 30 min at 37°C. About 400 μL of PI was then added, and the cell samples were incubated at 4°C for 30 min

in the dark. The prepared samples were examined using a flow cytometer and analyzed by FlowJo software.

ROS analysis

Doc (50 $\mu\text{g/mL}$), GNRs, GNRs + laser, GNRs/DocL-R (Doc concentration of 50 $\mu\text{g/mL}$), and GNRs/DocL-R (+laser) (Doc concentration of 50 $\mu\text{g/mL}$) were added to PC-3 cells (cell density of 2.5×10^5 cells/well) in six-well plates and incubated with the cells for 24 h. Subsequently, cells were subsequently washed with PBS three times and incubated with trypsin (no EDTA). The cell suspensions were centrifuged at 1,000 rpm for 10 min. ROS probe (DCFH-DA) was diluted with culture medium (no serum) to 10 μM , and 1 mL of DCFH-DA solution was mixed with cells. The cell suspensions were incubated for 20 min at 37°C with gentle mixing every 5 min. Next, the cells were washed with culture medium (no serum) three times and suspended with 500 μL of culture medium (no serum) for ROS quantification using flow cytometry.

In vivo studies

BALB/c-nu male nude mice were purchased from the HuNan Experimental Animal Center of the Chinese Academy of Science. Animal care and experiments were performed with the approval of the animal ethical committee of Zhengzhou University (Zhengzhou, China), according to the requirements of the National Act on the Use of Experimental Animals (China). All animals were kept in a favorable environment and were acclimated at 25°C and 55% of humidity under natural light/dark conditions, with free access to a rodent diet and water. The experimental animals were acclimated for 1 week before the beginning of the study. The in vivo studies were conducted on 5-week-old nude mice with body weight around 18–20 g. PC-3 cells were collected from cell culture by trypsinization and subcutaneously implanted (1×10^7 cells in 200 μL of culture medium) in the upper-right flank of anesthetized nude mice. When PC-3 xenograft volume reached 100 mm^3 , the prostate cancer (PC) models were established successfully, and the nude mice were randomly divided into experimental groups. All procedures followed the guidelines of the institutional animal ethical committee of Zhengzhou University (Zhengzhou, China).

In vivo biodistribution

DiR iodide emits weak fluorescence in water but strong fluorescence in cell membrane or lipophilic biomacromolecules. The dark red fluorescence from DiR iodide can efficiently penetrate cells and tissues, which makes DiR iodide

an ideal fluorescent dye for in vivo imaging. GNRs/DiR liposomes and GNRs/DiR-RLT liposomes were prepared as described in Supplementary materials. The PC-3 xenograft-implanted nude mice were divided into three groups. DiR solution (100 $\mu\text{g/mL}$; dose 1 mg DiR/kg mice), GNRs/DiR liposomes (dose 1 mg DiR/kg mice), and GNRs/DiR-RLT liposomes (dose 1 mg DiR/kg mice) were given to the nude mice via tail vein injection. At set time points (1, 2, 4, 8, 12, 24, 48), the nude mice were anesthetized, and the dynamic fluorescent distribution in the nude mice was recorded by In Vivo FX PRO (excitation wavelength 748 nm, emission wavelength 780 nm, X-ray exposure time 30 s) (Bruker; Billerica, MA, USA).

Antitumor efficacy

The nude mice were randomly assigned into six treatment groups: saline, Doc solution (concentration 10 mg/kg mice/3 days), GNRs/DocL (concentration of Doc: 10 mg/kg mice/3 days), GNRs/DocL (+laser) (concentration of Doc: 10 mg/kg mice/3 days), GNRs/DocL-R (concentration of Doc 10 mg/kg mice/3 days), and GNRs/DocL-R (+laser) (concentration of Doc 10 mg/kg mice/3 days). The treatments were given to nude mice via tail vein injection at set days (day 1, day 4, day 7, day 10, day 13). The PC xenograft size and body weights of the mice were measured the day before injections. At day 15, the mice were sacrificed, and the PC xenografts were excised and weighed after euthanizing the mice.

Statistical analysis

All experiments in this study were conducted at least in triplicate. Statistical analyses were performed using SPSS 18.0 (SPSS Inc., Chicago, IL, USA), Origin 8.0, and GraphPad Prism 6 software (GraphPad Software, Inc., La Jolla, CA, USA). Quantified data were presented as mean $\pm$ SD. Student's *t*-test was applied to compare between two groups, and one-way analysis of variance (ANOVA) was performed to compare three or more groups (*p*-values < 0.05 indicated statistical significance).

Results

Preparation and characterization of GNRs/DocL-R

Doc is a potent antimitotic drug for PC, but the application of Doc is limited by the side effects and high tendency of cellular drug resistance of Doc. A multifunctional liposome that contained GNRs and Doc was designed and constructed in this study to improve Doc for PC treatment. The structure and compositions of the liposome are illustrated in Figure 1A.

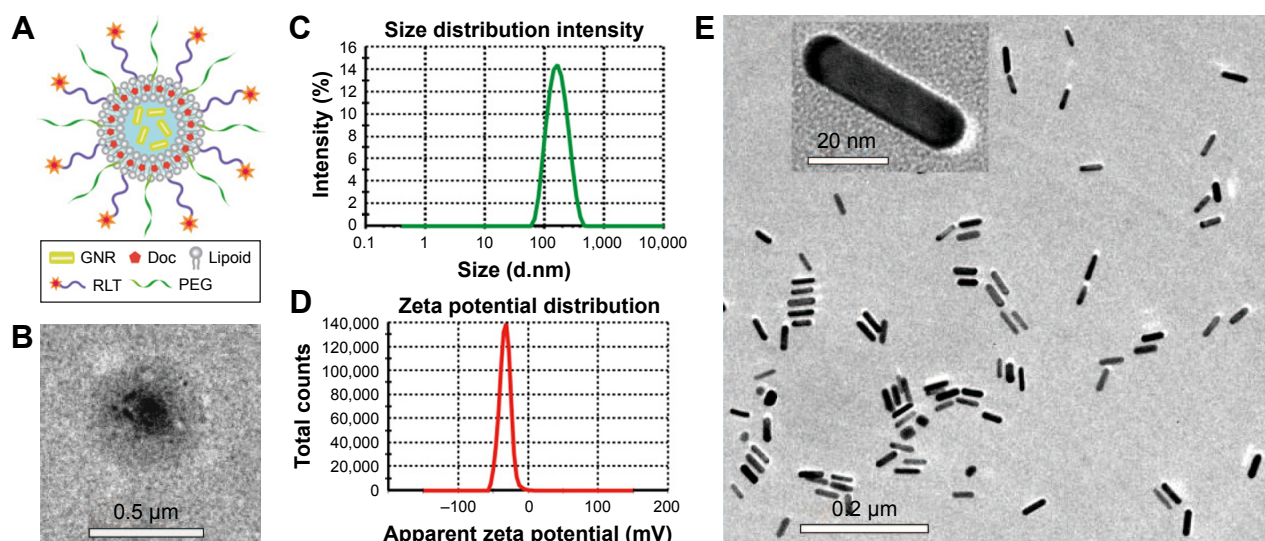


Figure 1 Characterization of GNRs and GNRs/DocL-R.

Notes: (A) The compositions and structure of GNRs/DocL-R. (B) A TEM image of GNRs/DocL-R. (C) Particle size distribution of GNRs/DocL-R by dynamic light scattering. (D) Zeta potential distribution of GNRs/DocL-R by Zetasizer. (E) A TEM image of GNRs.

Abbreviations: Doc, docetaxel; GNRs, gold nanorods; GNRs/DocL-R, GNRs/Doc-liposome-RLT; TEM, transmission electron microscopy.

The liposomes were composed of GNRs, Doc, lipid (chol, SPC, and HSPC), PEG, and RLT. The TEM image showed a round shape of the multifunctional liposomes and a central location of GNRs (Figure 1B). Diameter and zeta potential of the liposomes were around 163.2 nm and -32.8 mV, respectively (Figures 1B, C and D). The TEM images of GNRs are shown in Figure 1E. The size of GNRs was 40 nm (length) $\times$ 10 nm (diameter). At aspect ratio (L:D) 4, GNRs have strong absorbance at a wavelength around 810 nm. Three different GNR concentrations were used to prepare liposomes. There were no significant differences between the size and polydispersity index (PDI) of the GNRs/DocL-R prepared using GNRs with different concentrations (Figure 2A). However, negative charge of the liposomes reduced with increased GNR concentration (Figure 2B). At a higher concentration of GNRs (0.635 nM), the aggregation of GNRs was observed after a few days, indicating a low stability (Table S1). Figure 3A shows a lower temperature increase after laser radiation using low GNR concentration. Therefore, to ensure the heating efficiency and stability of GNRs *in vitro* and *in vivo*, 0.4 nM of GNRs was selected. Lipid contained chol, SPC, and HSPC. HSPC was reduced from SPC by the addition of hydrogen and has significantly increased stability and phase transition temperature compared to SPC. The addition of HSPC can increase the stability of the liposomes. Varied ratios between SPC and HSPC were used to prepare the liposomes. Results showed that at SPC:HSPC (1:30; Table S2), optimized liposomes were obtained. Chol can regulate the mobility of the double-layer

membrane of lipid and reduce the permeability of the liposomes. The addition of Chol could increase the stability and reduce the drug leakage of the liposomes. At the ratio of (HSPC + SPC):Chol (8:1), the liposomes showed a high stability and a proper particle size (Table S3). RLT was conjugated to the liposomes via DSPE-PEG (2000)-Mal. DSPE-PEG (2000)-Mal was added during the preparation of liposomes. Partial DSPE-PEG (2000)-Mal might distribute into the inner layer of the double-layer membrane of the liposomes and could not react with RLT. HPLC was used to quantify conjugated RLT (Supplementary materials). Results showed that a higher DSPE-PEG (2000)-Mal:RLT ratio (3:1) resulted in a higher conjugation percentage of RLT (%) to DSPE-PEG (2000)-Mal ($\sim 90\%$; Table S4, Figure 2C). Because of the low concentration of DSPE-PEG (2000)-Mal to maintain a stable liposome system, higher DSPE-PEG (2000)-Mal:RLT ratio (3:1) led to a lower RLT concentration (0.1125%). Other studies reported the trend that nanoparticles with a higher percentage of targeting agents showed more internalization by the targeted cells.^{23,24} A low RLT modification rate may reduce the targeting ability of the liposomes. Therefore, molar ratio of DSPE-PEG (2000)-Mal:RLT (2:1) was selected in this study. In sum, the ratio of SPC:HSPC:Chol (1:30:3.75) was used to prepare a stable liposome. Molar modification rates of DSPE-PEG (2000)-Mal and RLT to the liposomes were 0.375% and 0.133%, respectively, and 200 W and 240 s were selected as the optimized power and time of sonication, respectively (Table S5, Figure 2D).

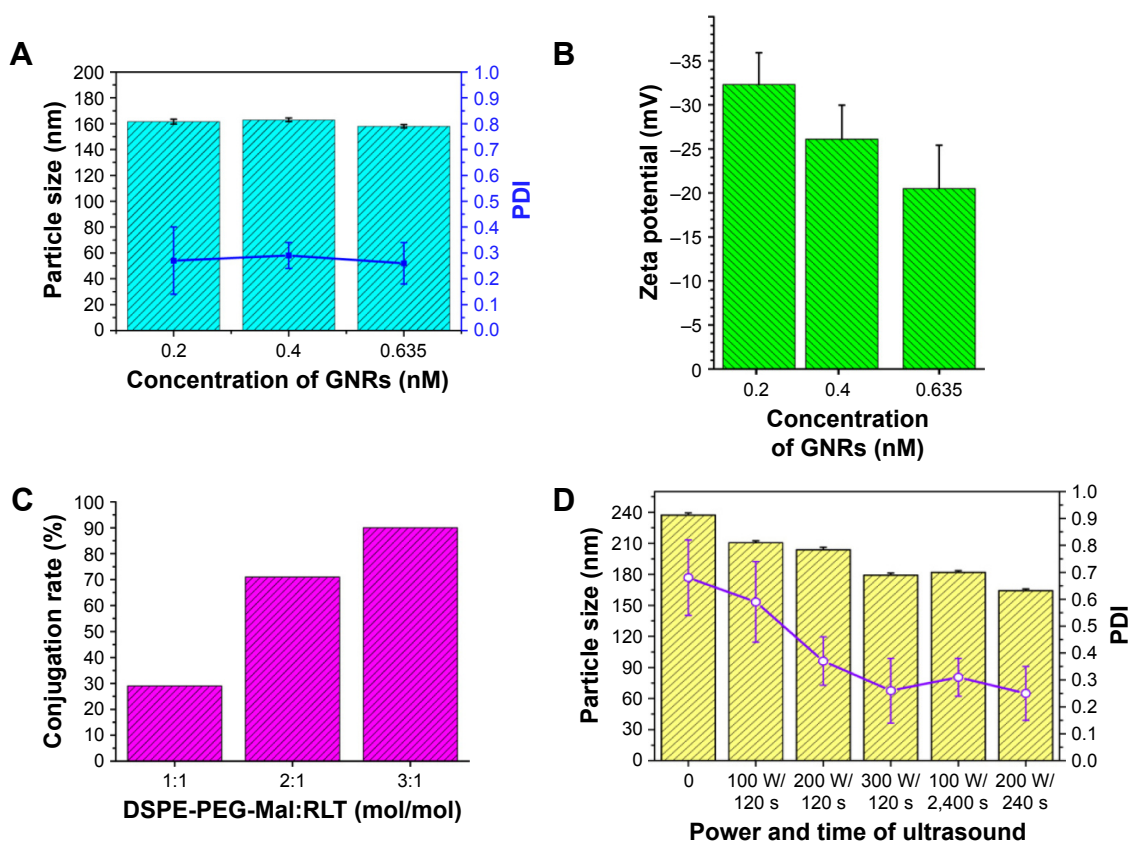


Figure 2 The optimization of compositions and preparation of GNRs/DocL-R.

Notes: (A) Influence of GNR concentration on the size and PDI of GNRs/DocL-R. (B) Influence of GNR concentration on the zeta potential of GNRs/DocL-R. (C) Influence of DSPE-PEG-Mal:RLT molar ratio on the conjugation rate of RLT. (D) Influence of power and time of ultrasound on the size and PDI of GNRs/DocL-R.

Abbreviations: Doc, docetaxel; GNRs, gold nanorods; GNRs/DocL-R, GNRs/Doc-liposome-RLT; PDI, polydispersity index.

Release of Doc and thermal responses of GNRs/DocL-R

The LE and LC of Doc were $98.45\% \pm 0.37\%$ and 1.41 ± 0.11 mg/mL, respectively. About 0.5% (w/w) Tween 80 was added to PBS to increase the solubility of Doc in PBS. The results showed that laser radiation and low pH accelerated Doc release (Figure 3B and C). Temperature increase caused by laser radiation possibly reduced the stability of liposomes and resulted in Doc release. Figure 3C shows the relationship between the concentration of GNRs and laser radiation-induced temperature increase. The results indicated that a higher GNR concentration could produce a higher temperature for thermal therapy.

PC-3 cell inhibition and intracellular uptake by PC-3 cells

Concentration- and time-dependent PC-3 cell inhibition was examined. All the listed treatments showed a higher cell inhibition rate than free Doc (Figure 4A). The addition of GNRs to the liposomes (GNRs/DocL) did not cause

more cell inhibition, whereas laser radiation induced a higher cell inhibition rate. The conjugation of RLT also enhanced PC-3 cell inhibition. At Doc of 80 μ g/mL, time-dependent cell inhibition (Figures 4B and S1) showed a similar trend with concentration-dependent cell inhibition. Free Doc caused a low cell inhibition rate, whereas Doc-loaded liposomes caused significant cell inhibition. The multifunctional liposomes (GNRs/DocL-R with(out) laser radiation) result in the cell inhibition rates to around 100%. With 24 h incubation, the cell inhibition rate caused by Doc plus GNRs + laser was approximately equal to that caused by GNRs/DocL-R + laser (additive effect); however, the cell inhibition rate using GNRs/DocL-R + laser treatment was higher than that using Doc plus GNRs + laser (synergistic effect) with longer incubation (48 h and 72 h), indicating a shift from the additive effect of Doc and GNRs + laser to a synergistic effect on PC-3 cell inhibition with longer incubation time (Figure S1).

Dox is a potent chemotherapy that has similar hydrophobicity and molecular weight with Doc. Dox was selected as

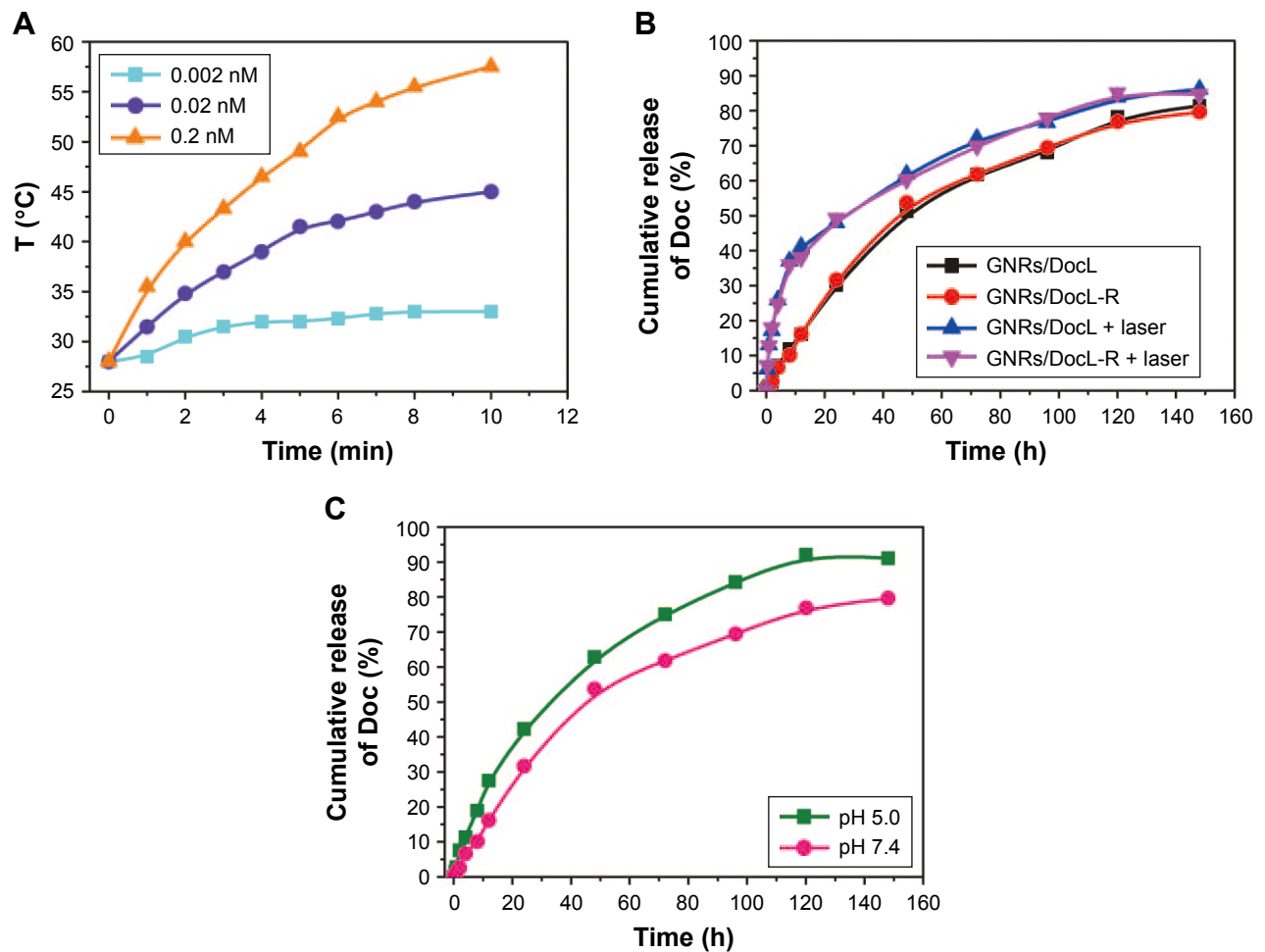


Figure 3 Doc release and thermal responses of GNRs/DocL-R.

Notes: (A) Thermal responses of GNRs/DocL-R to laser radiation (concentrations represented the GNR concentration in GNRs/DocL-R). (B) Doc release from the listed formulation with(out) laser radiation in pH7.4 PBS plus 0.5% Tween 80. (C) Doc release from GNRs/DocL-R in pH5.0 PBS plus 0.5% Tween 80 and pH7.4 PBS plus 0.5% Tween 80.

Abbreviations: Doc, docetaxel; GNRs, gold nanorods; GNRs/DocL-R, GNRs/Doc-liposome-RLT.

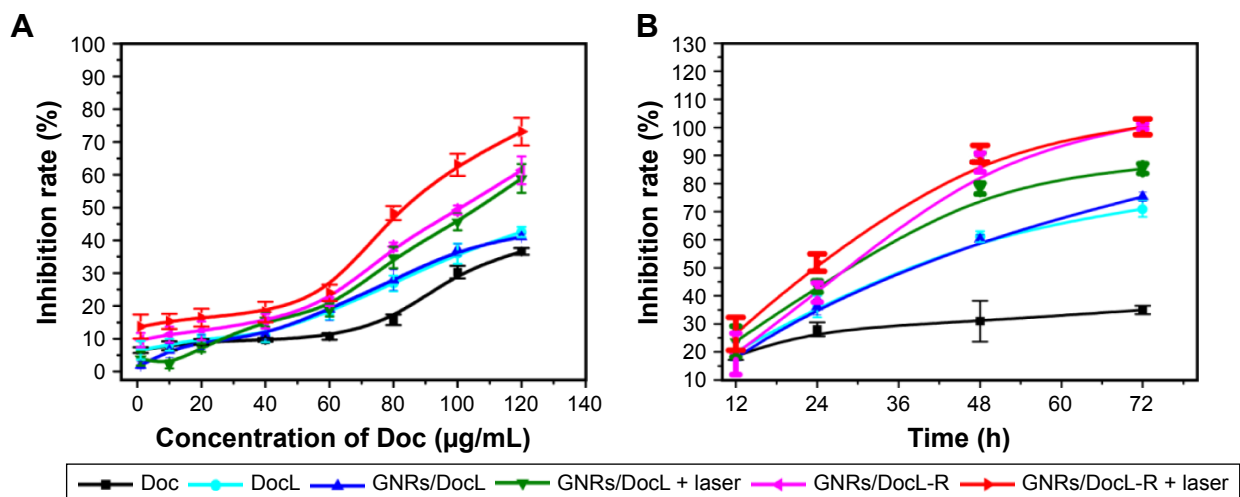


Figure 4 (Continued)

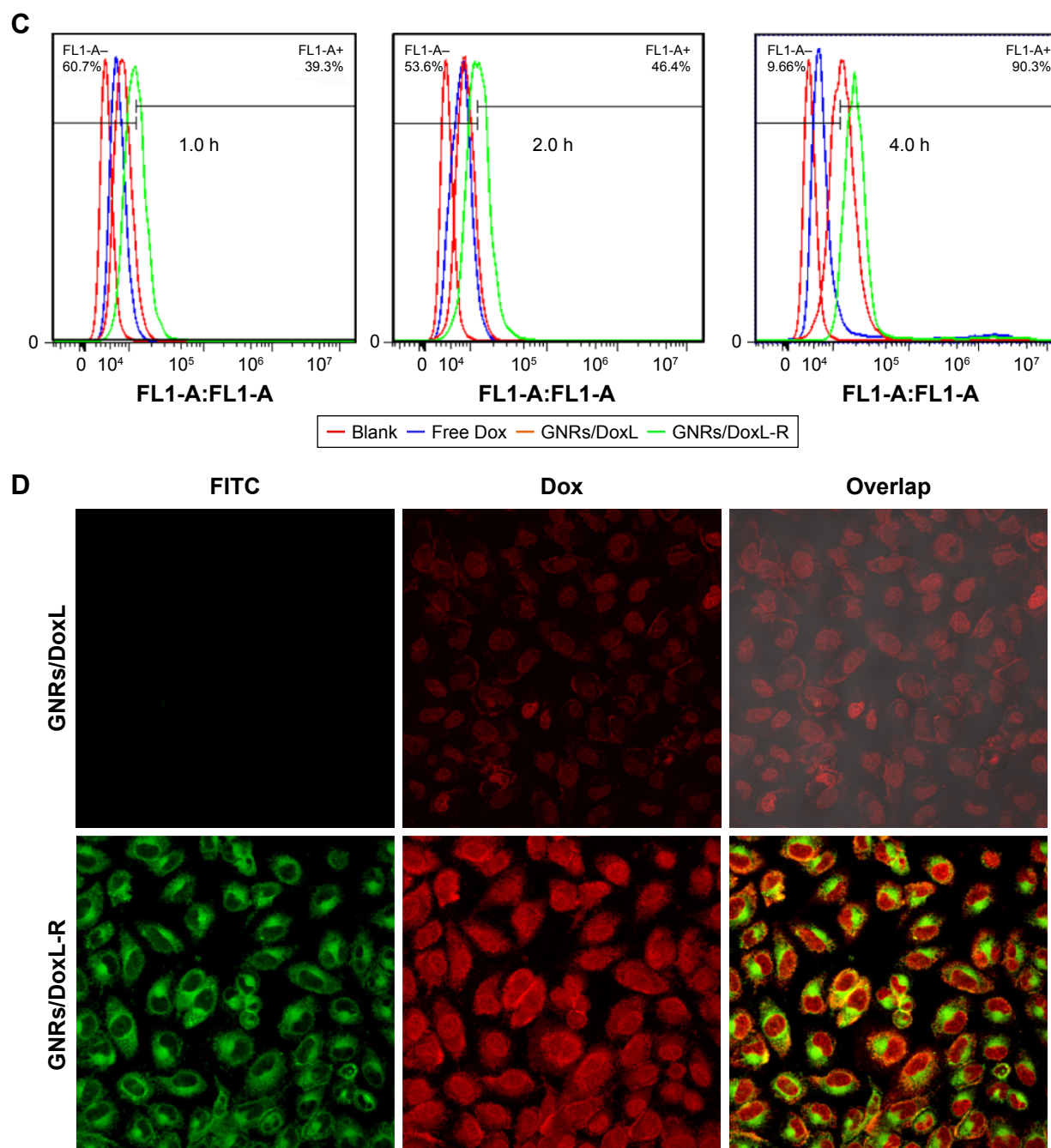


Figure 4 Evaluation of PC-3 cell inhibition by Doc and intracellular uptake of Dox.

Notes: (A) PC-3 cell inhibition rate by the listed treatments with increasing Doc concentration. (B) PC-3 cell inhibition rate by the listed treatments with increasing incubation period (the concentration of Doc for all treatments was 80 $\mu\text{g/mL}$). (C) Flow cytometric histograms of intracellular uptake of Dox by PC-3 cells. (D) Confocal microscopy images of PC-3 cell after 4 h incubation with GNRs/DoxL and GNRs/DoxL-R. Red fluorescence and green fluorescence represented the location of Dox and GNRs/L-R, respectively. Orange fluorescence indicated the colocalization of Dox and GNRs/L-R.

Abbreviations: Doc, docetaxel; Dox, doxorubicin; FITC, fluorescein isothiocyanate; GNRs, gold nanorods; PC-3, prostate cancer cell line.

fluorescent agent to investigate the intracellular uptake of the liposomes. Figure S2 shows that GNRs/DoxL-R-treated cells had a much stronger intracellular red fluorescence than free Dox and GNRs/DoxL, indicating a higher concentration of intracellular Dox using GNRs/DoxL-R. Flow cytometry was used to quantify the intracellular uptake of Dox.

The results showed that intracellular uptake rates of Dox using free Dox, GNRs/DoxL, and GNRs/DoxL-R were 2.5%, 9.9%, and 50.6%, respectively, with 1 h incubation. After 4 h incubation, intracellular uptake rates of Dox using free Dox, GNRs/Dox liposomes, and GNRs/Dox-RLT liposomes were increased to 14.3%, 61.1%, and 96.2%, respectively.

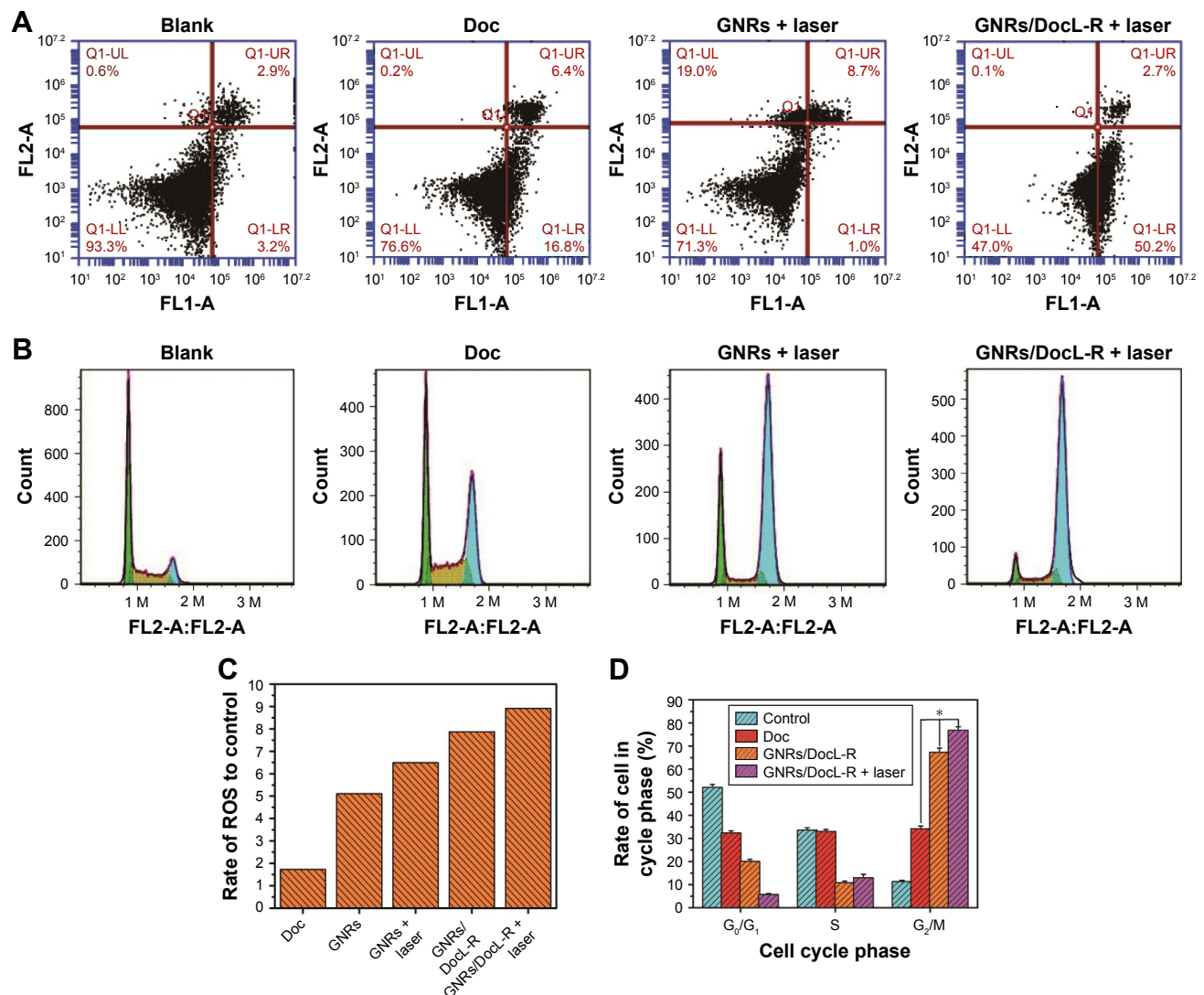


Figure 5 Flow cytometric analysis of anticancer mechanisms of listed treatments in vitro.

Notes: (A) Apoptotic statistics of PC-3 cells by the listed treatments. (B) Flow cytometry histograms of PC-3 cell cycle by the listed treatments. (C) Quantification of ROS in PC-3 cells that incubated with the listed treatments. (D) Cell cycle analysis of PC-3 cells after the listed treatments. Data are presented as mean $\pm$ SD (N=3). *Doc vs GNRs/DocL-R vs GNRs/DocL-R + laser, $p < 0.05$.

Abbreviations: Doc, docetaxel; GNRs, gold nanorods; GNRs/DocL-R, GNRs/Doc-liposome-RLT; PC-3, prostate cancer cell line; ROS, reactive oxygen species.

(Figure 4C). RLT was labeled with FITC, and thus the location of GNRs/DocL-R in cells could be tracked by green fluorescence. As shown in Figure 4D, red fluorescence was observed in both cytoplasm and nucleus area of the cells, whereas green fluorescence was observed in the cytoplasm area of the cells. The overlapped cell image demonstrated the red nucleus and the yellowish cytoplasm.

Flow cytometry analysis

After 24 h of incubation with Doc, GNRs + laser, and GNRs/DocL-R + laser, cell apoptosis was analyzed by flow cytometry. As shown in Figure 5A, Doc caused 0.2% cell necrosis, 16.8% early apoptosis, and 6.4% late apoptosis;

GNRs + laser induced 19.0% cell necrosis, 1.0% early apoptosis, and 8.7% late apoptosis; GNRs/DocL-R + laser resulted in 0.1% cell necrosis, 50.2% early apoptosis, and 2.7% late apoptosis. GNRs/DocL-R + laser was significantly more potent to PC-3 cells than Doc plus GNRs + laser. The results of cell cycle analysis showed that blank PC-3 cells had around the half of cells in G₀/G₁ phase, preparing for DNA duplication and subsequent cell division. The PC-3 cells in G₂/M phase were around 11.3% $\pm$ 0.6%. Incubation of the cells with Doc resulted in an increase in the cells in G₂/M phase to 34.2% $\pm$ 1.2%. GNRs/DocL-R increased G₂/M phase arrest to 67.4% $\pm$ 1.8%, whereas G₀/G₁ phase arrest and S phase arrest of PC-3 cells reduced to 20.0% $\pm$ 0.9% and

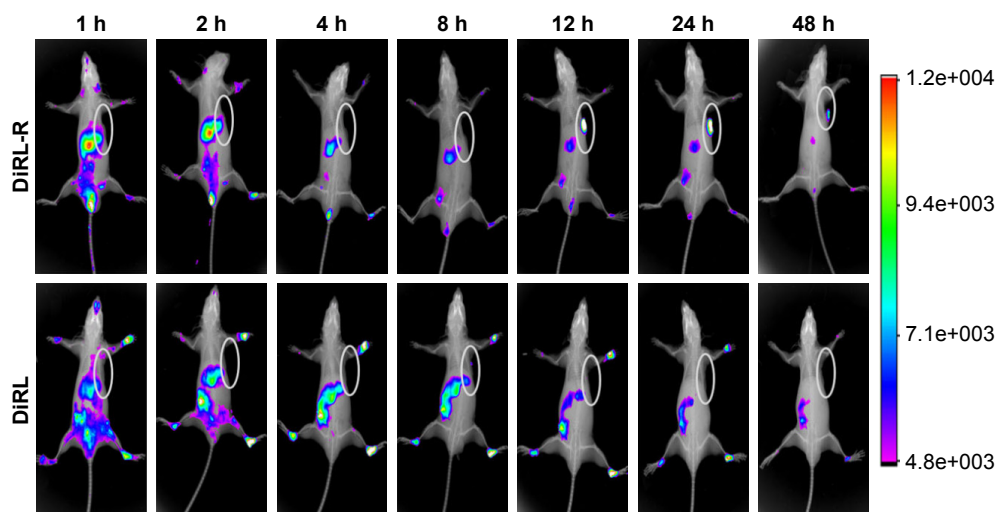


Figure 6 In vivo distribution of GNRs/DiRL-R and GNRs/DiRL after tail vein injection to nude mice.

Notes: Observed fluorescence indicated the location of DiR-encapsulated liposomes. The ellipses indicated the location of implanted PC-3 tumors.

Abbreviations: DiR iodide, 1,1-diiododecyl-3,3,3,3-tetramethylindotricarbocyanine iodide; DiRL, DiR liposomes; DiRL-R, DiR-RLT liposomes; GNRs, gold nanorods; PC-3, prostate cancer cell line.

10.8%±0.7%, respectively. GNRs/DocL-R + laser reduced G_0/G_1 phase arrest to 5.7%±0.4% and increased G_2/M phase arrest to 76.8%±1.6% (Figure 5B and D). Doc is a potent antimitotic chemotherapy and inhibits cell growth by G_2/M phase arrest as shown in this study. GNRs/DocL-R + laser significantly enhanced the G_2/M phase arrest of PC-3 cells compared to Doc. The existence of ROS is necessary to maintain the characteristics of cells. However, excess ROS will cause damage to intracellular macromolecules and lead to cell apoptosis. Doc could cause increased intracellular ROS. GNRs/DocL-R caused a much higher intracellular ROS because of its higher intracellular delivery of Doc. Laser radiation to cells also produced more intracellular ROS because of the heat generation by GNRs (Figure 5C).

In vivo study

RLT-modified lipid submicron emulsion increased the delivery of Doc to the tumor site of mice.¹⁸ To elucidate the dynamic distribution of the liposomes and the RLT-modified liposomes, DiR liposomes and DiR-RLT liposomes were tracked after tail vein injection to mice using In Vivo FX PRO. The fluorescence signal indicated the location of DiR-encapsulated liposomes in vivo. DiR liposomes showed strong fluorescence in the liver and kidney and no fluorescence in PC-3 tumor-implanted site. RLT-modified DiR liposomes demonstrated strong fluorescence in PC-3 tumor-implanted site within 12 h, and the fluorescent signal was maintained until 48 h (Figure 6). The results indicated that RLT-modified DiR liposomes could penetrate and accumulate in tumor

sites. In vivo antitumor efficacy was evaluated by measuring the volume and weight of PC-3 tumor and calculating the inhibition rate of PC-3 tumor. As shown in Figure 7A–C, all the listed treatments showed the reduced volume and weight of PC-3 tumor, and the increased inhibition rate of PC-3 tumor compared with Doc treatment. GNRs/DocL-R + laser resulted in the lowest volume and weight of PC-3 tumor, and the highest inhibition rate of PC-3 tumor. Body weight can be an indicator of mice health. As shown in Figure 7D, mice treated with Doc lost the most body weight at the end of treatment, whereas GNRs/Doc-RLT liposomes + laser treatment maintained the highest body weight of mice.

Discussion

Personalized and precise medicines are potential solutions for unmet medical needs. Nanomedicines that employ the latest drug delivery technologies are ideal candidates for personalized and precise medicines. The nanomedicines should be well designed and precisely constructed. The combination of gold nanoparticles and chemotherapy has been previously reported. For example, Yu et al have injected the physical mixture of a high thermosensitive liposome with GNRs to nude mice, and the results showed that GNRs not only facilitated co-accumulation of liposomes into tumor site but also triggered drug release from the liposomes.³⁹ However, in Yu et al's work, the interaction between GNRs and liposomes was not clear. GNRs possibly interacted with the liposomes to achieve co-accumulation in tumor. Factors that affected the integrity of the physical mixture of GNRs and

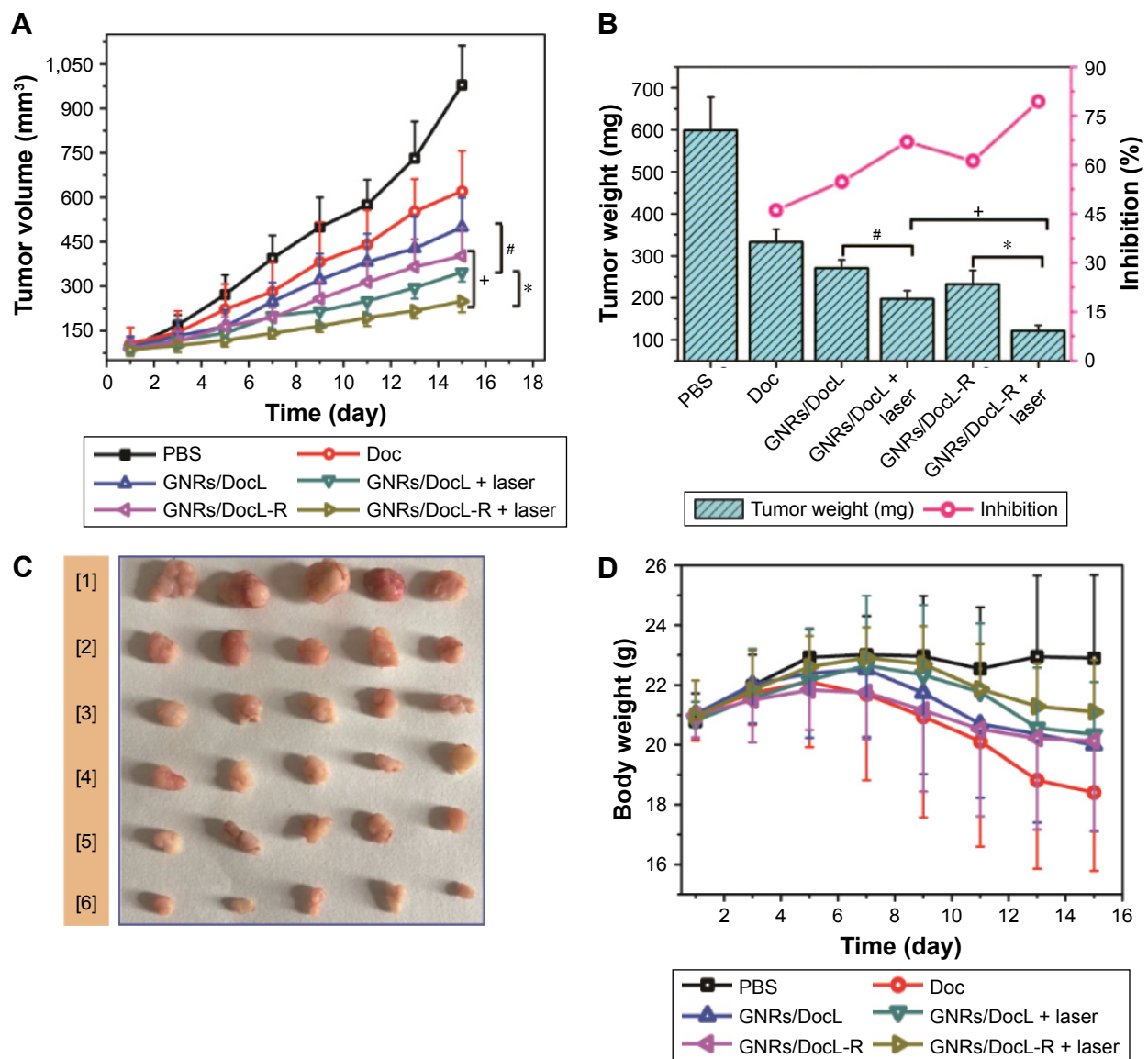


Figure 7 In vivo evaluation of the antitumor efficacy of the listed treatments.

Notes: (A) Tumor volume data during 15 days of treatment with the listed formulations. (B) Tumor weight and calculated tumor inhibition rate after 15 days of treatment with the listed formulations. (C) Tumor images after 15 days of treatment with the listed formulations (1–6: saline; Doc solution; GNRs/DocL; GNRs/DocL + laser; GNRs/DocL-R; GNRs/DocL-R + laser, respectively). (D) Body weight data during 15 days of treatment with the listed formulations. Data are presented as mean $\pm$ SD (N=5). *GNRs/DocL-R vs GNRs/DocL-R + laser, $p < 0.05$. *GNRs/DocL + laser vs GNRs/DocL-R + laser, $p < 0.01$. #GNRs/DocL vs GNRs/DocL + laser, $p < 0.05$.

Abbreviations: Doc, docetaxel; GNRs, gold nanorods; GNRs/DocL-R, GNRs/Doc-liposome-RLT.

liposomes might reduce antitumor efficacy. Kang and Ko²⁶ also developed lipid-covered gold nanoparticles and GNRs that contained Doc in the lipids. Compared to liposomes, monolayered lipids had limited drug LC. Both studies showed enhanced tumor inhibition via the combined use of gold nanoparticles and chemotherapy. However, drug release from the two systems was too fast for clinical application. In sum, the composition and interaction of a precise nanomedicine should be well characterized. Precise nanomedicines should be stable and reliable, can load sufficient drugs, can achieve sustained/controlled release based on the treatment demands, etc.

In this study, Doc was loaded into the lipophilic bilayer of the liposomes, and GNRs were encapsulated into the core of the liposomes. Compared with the physical mixture of GNRs/gold nanoparticles and liposomes,^{25,27} and the liposome–GNR hybrids,^{28–30} GNRs/DocL-R could protect GNRs and Doc from physical environment, ensure simultaneous in vivo behavior of the liposomes and GNRs, and enhance GNR-triggered Doc release from the liposomes. The issue of drug leakage from the liposomes was improved compared to previous studies. Around 15% of drug was released within 12 h, at which significant accumulation of DiRL-R had been observed at tumor

site (Figure 6). Once GNRs/DocL-R arrived tumor site, drugs needed to be released to kill tumor. Therefore, the drug release profile could meet the need for this study. For the long-term storage, GNRs/DocL-R can be lyophilized and stored in dry place. Laser radiation increased local temperature and might trigger Doc release. Mathiyazhakan et al³¹ proposed that gold nanoparticles triggered drug release by inducing phase transition of the liposomes as well as the generation of microbubble cavitation. The phase transition temperature of HSPC and SPC were 50°C and below 41°C, respectively.³² In this study, laser radiation to GNRs generated a local temperature higher than 50°C (Figure 3C), which might increase membrane fluidity of the liposomes and accelerate Doc release. Increased Doc release induced the enhanced in vitro and in vivo anticancer efficacy (Figures 4, 5, and 7).

Chol is an essential component for the synthesis of new cell membrane. LDLRs that response for the transportation of chol are thus highly active in tumor cells. Liposomes are generally believed to be taken via two steps: adsorption onto the cell surface and endocytosis. Schaefer et al³³ reported a clathrin-dependent internalization of unmodified liposome–polyethyleneimine complexes by SKOV-3 cells. In this study, DSPE-PEG (2000)-Mal was modified at a molar ratio of 0.375% of total lipids in the liposomes. The low modification rate was consistent with the previous report from Li and Takeoka.³⁴ The purpose was to maintain an original physical characteristics, such as size, lipid compositions, drug-to-lipid ratio, drug encapsulation efficiency, stability, and zeta potential, of the liposomes. Li and Takeoka³⁵ showed the enhanced cellular uptake of liposomes via thiol-mediated transport with the addition of low concentrated DSPE-PEG (2000)-Mal, which was consistent with our results that the low-level modification of RLT to the liposomes increased intracellular uptake. PEGylation prolongs the in vivo circulation of liposomes but attenuates their endocytosis by cells.³⁶ RLT was used to enhance LDLR-mediated endocytosis and assist endo-lysosomal escape in cells.^{14–16}

Doc is an anti-microtubule agent that induces cell apoptosis and cell cycle arrest via the disorganization of microtubule structure.³⁷ Cell responses to PTT are mainly cell injury/death via necrosis.³⁸ In this study, Doc mainly caused PC-3 cell apoptosis, whereas GNRs + laser resulted in necrosis (Figure 5A). However, only significant apoptosis was observed using GNRs/DocL-R + laser. The result was different from our previous study that the addition of laser radiation to Dox/DNA complexed GNR system (Dox/DNA-GNRs-PEG/R) mainly led to enhanced cell necrosis.³ The contrary results might be caused by the different structures of

the two drug delivery systems. Because of the central location of GNRs, heating generated by GNRs/DocL-R + laser might induce the disruption of liposomes and the release of Doc, whereas laser-radiated Dox/DNA-GNRs-PEG/R released the heating directly to the environment. Doc increased M phase arrest by destroying microtubules that were essential for cell division. GNRs/DocL-R could significantly increase M phase arrest of PC-3 cells due to the enhanced and sustained intracellular delivery of Doc. The cell apoptosis and cell cycle arrest results explained the significantly reduced in vivo tumor weight and volume (Figure 7C).

The current study developed a promising therapy for combating PC. To elucidate in vivo behavior of GNRs/DocL-R, pharmacokinetic (PK) study was under investigation in our laboratory. Preliminary data were consistent with in vivo distribution of GNRs/DocL-R (Figure 6). Mice treated with GNRs/DocL-R showed a significantly higher concentration and longer circulation of Doc in the blood plasma compared with Doc and GNRs/DocL (data unpublished). Despite high targeting to tumor, significant liver accumulation of DiRL-R was observed within 0–2 h after injection. Most liposomes have the issue of high accumulation in the liver, which may disrupt the liposomes and cause drug leakage from the liposomes. Further PK study will quantify Doc concentration in the liver, and the liposomes will be modified to avoid the metabolism by liver.

Conclusion

In this study, a multifunctional liposome was developed for improved anticancer therapy. Doc and GNRs were loaded into the liposomes as chemotherapy and thermotherapy agents. The liposomes achieved a prolonged and sustained release of Doc. Laser radiation to GNRs led to Doc release from the liposomes in vitro and in vivo. Tumor targeting was increased by modifying the liposomes with RLT. The developed GNRs/DocL-R showed a promising anticancer efficacy and low toxicity in in vitro and in vivo evaluation. The results suggested that the stable and easily prepared GNRs/DocL-R could be an efficient and sustained therapy for PC. Thermal therapy using laser radiation further enhanced anticancer efficacy via increased release and intracellular delivery of Doc to PC-3 cells. Future work will focus on the PK study of GNRs/DocL-R to quantify the distribution and elimination of Doc and confirm the potential of GNRs/DocL-R for clinical use.

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Disclosure

The authors report no conflicts of interest in this work.

References

- Huang XH, El-Sayed IH, Qian W, El-Sayed MA. Cancer cell imaging and photothermal therapy in the near-infrared region by using gold nanorods. *J Am Chem Soc*. 2006;128(6):2115–2120.
- Tong L, Zhao Y, Huff TB, Hansen MN, Wei A, Cheng J-X. Gold nanorods mediate tumor cell death by compromising membrane integrity. *Adv Mater*. 2007;19(20):3136.
- Zhang N, Li S, Hua H, et al. Low density lipoprotein receptor targeted doxorubicin/DNA-gold nanorods as a chemo- and thermo-dual therapy for prostate cancer. *Int J Pharm*. 2016;513(1–2):376–386.
- Huang HC, Barua S, Kay DB, Rege K. Simultaneous enhancement of photothermal stability and gene delivery efficacy of gold nanorods using polyelectrolytes. *ACS Nano*. 2009;3(10):2941–2952.
- Huang HC, Rege K, Heys JJ. Spatiotemporal temperature distribution and cancer cell death in response to extracellular hyperthermia induced by gold nanorods. *ACS Nano*. 2010;4(5):2892–2900.
- Choi WI, Kim JY, Kang C, Byeon CC, Kim YH, Tae G. Tumor regression in vivo by photothermal therapy based on gold-nanorod-loaded, functional nanocarriers. *ACS Nano*. 2011;5(3):1995–2003.
- Helmchen F, Denk W. Deep tissue two-photon microscopy. *Nat Methods*. 2005;2(12):932–940.
- Kuo WS, Chang CN, Chang YT, et al. Gold nanorods in photodynamic therapy, as hyperthermia agents, and in near-infrared optical imaging. *Angew Chem Int Edit*. 2010;49(15):2711–2715.
- He XM, Wolkers WF, Crowe JH, Swanlund DJ, Bischof JC. In situ thermal denaturation of proteins in dunning AT-1 prostate cancer cells: implication for hyperthermic cell injury. *Ann Biomed Eng*. 2004;32(10):1384–1398.
- Cortes JE, Pazdur R. Docetaxel. *J Clin Oncol*. 1995;13(10):2643–2655.
- Shen S, Tang H, Zhang X, et al. Targeting mesoporous silica-encapsulated gold nanorods for chemo-photothermal therapy with near-infrared radiation. *Biomaterials*. 2013;34(12):3150–3158.
- Allen TM, Cullis PR. Liposomal drug delivery systems: from concept to clinical applications. *Adv Drug Deliv Rev*. 2013;65(1):36–48.
- Chen Y, Hughes-Fulford M. Human prostate cancer cells lack feedback regulation of low-density lipoprotein receptor and its regulator, SREBP2. *Int J Cancer*. 2001;91(1):41–45.
- Hayavi S, Halbert GW. Synthetic low-density lipoprotein, a novel biomimetic lipid supplement for serum-free tissue culture. *Biotechnol Prog*. 2005;21(4):1262–1268.
- Nikanjam M, Blakely EA, Bjornstad KA, Shu X, Budinger TF, Forte TM. Synthetic nano-low density lipoprotein as targeted drug delivery vehicle for glioblastoma multiforme. *Int J Pharm*. 2007;328(1):86–94.
- Nikanjam M, Gibbs AR, Hunt A, Budinger TF, Forte TM. Synthetic nano-LDL with paclitaxel oleate as a targeted drug delivery vehicle for glioblastoma multiforme. *J Control Release*. 2007;124(3):163–171.
- Zhang N, Tao J, Hua HY, Sun PC, Zhao YX. Low-density lipoprotein peptide-combined DNA nanocomplex as an efficient anticancer drug delivery vehicle. *Eur J Pharm Biopharm*. 2015;94:20–29.
- Zhang N, Miao J, Sun P, et al. Pharmacokinetics, tissue distribution and anti-tumor effect of low density lipoprotein peptide conjugated submicron emulsions. *Biomed Pharmacother*. 2016;82:614–619.
- Orendorff CJ, Murphy CJ. Quantitation of metal content in the silver-assisted growth of gold nanorods. *J Phys Chem B*. 2006;110(9):3990–3994.
- Nikoobakht B, El-Sayed MA. Preparation and growth mechanism of gold nanorods (NRs) using seed-mediated growth method. *Chem Mater*. 2003;15(10):1957–1962.
- Sau TK, Murphy CJ. Seeded high yield synthesis of short Au nanorods in aqueous solution. *Langmuir*. 2004;20(15):6414–6420.
- Shariat S, Badiie A, Jalali SA, et al. P5 HER2/neu-derived peptide conjugated to liposomes containing MPL adjuvant as an effective prophylactic vaccine formulation for breast cancer. *Cancer Lett*. 2014;355(1):54–60.
- Chen Z, He N, Chen M, Zhao L, Li X. Tunable conjugation densities of camptothecin on hyaluronic acid for tumor targeting and reduction-triggered release. *Acta Biomater*. 2016;43:195–207.
- Chu S, Tang C, Yin C. Effects of mannose density on in vitro and in vivo cellular uptake and RNAi efficiency of polymeric nanoparticles. *Biomaterials*. 2015;52:229–239.
- Chen P, Huang N-T, Chung M-T, Cornell TT, Kurabayashi K. Label-free cytokine micro- and nano-biosensing towards personalized medicine of systemic inflammatory disorders. *Adv Drug Deliv Rev*. 2015;95:90–103.
- Kang JH, Ko YT. Lipid-coated gold nanocomposites for enhanced cancer therapy. *Int J Nanomedicine*. 2015;10(Spec Iss):33–45.
- Guo HY, Kim JC. Photothermally induced release from liposome suspended in mixture solution of gold nanoparticle and thermo-sensitive polymer. *Colloid Surf A*. 2015;469:73–82.
- Reyes-Ortega F, Cifuentes A, Rodriguez G, et al. Bioactive bilayered dressing for compromised epidermal tissue regeneration with sequential activity of complementary agents. *Acta Biomater*. 2015;23:103–115.
- Taruttis A, Lozano N, Nunes A, et al. siRNA liposome-gold nanorod vectors for multispectral optoacoustic tomography theranostics. *Nanoscale*. 2014;6(22):13451–13456.
- Lozano N, Al-Jamal WT, Taruttis A, et al. Liposome-gold nanorod hybrids for high-resolution visualization deep in tissues. *J Am Chem Soc*. 2012;134(32):13256–13258.
- Mathiyazhakan M, Yang Y, Liu Y, et al. Non-invasive controlled release from gold nanoparticle integrated photo-responsive liposomes through pulse laser induced microbubble cavitation. *Colloids Surf B Biointerfaces*. 2015;126:569–574.
- Chen J, Yan GJ, Hu RR, et al. Improved pharmacokinetics and reduced toxicity of brucine after encapsulation into stealth liposomes: role of phosphatidylcholine. *Int J Nanomedicine*. 2012;7:3567–3577.
- Schaefer J, Hoeberl S, Bakowsky U, Aigner A. Liposome-polyethylenimine complexes for enhanced DNA and siRNA delivery. *Biomaterials*. 2010;31(26):6892–6900.
- Li T, Takeoka S. A novel application of maleimide for advanced drug delivery: in vitro and in vivo evaluation of maleimide-modified pH-sensitive liposomes. *Int J Nanomedicine*. 2013;8:3855–3866.
- Li T, Takeoka S. Enhanced cellular uptake of maleimide-modified liposomes via thiol-mediated transport. *Int J Nanomedicine*. 2014;9:2849–2861.
- Miller CR, Bondurant B, McLean SD, McGovern KA, O'Brien DF. Liposome-cell interactions in vitro: effect of liposome surface charge on the binding and endocytosis of conventional and sterically stabilized liposomes. *Biochemistry*. 1998;37(37):12875–12883.
- Wang LG, Liu XM, Kreis W, Budman DR. The effect of antimicrotubule agents on signal transduction pathways of apoptosis: a review. *Cancer Chemother Pharmacol*. 1999;44(5):355–361.
- Fernandez Cabada T, Sanchez Lopez de Pablo C, Martinez Serrano A, del Pozo Guerrero F, Serrano Olmedo JJ, Ramos Gomez M. Induction of cell death in a glioblastoma line by hyperthermic therapy based on gold nanorods. *Int J Nanomedicine*. 2012;7:1511–1523.
- Yu M, Guo F, Tan F, Li N. Dual-targeting nanocarrier system based on thermosensitive liposomes and gold nanorods for cancer thermo-chemotherapy. *J Control Release*. 2015;215:91–100.

Supplementary materials

Materials and methods

Synthesis and characterization of gold nanorods (GNRs)

GNRs were synthesized by using a modified seed-mediated growth method in aqueous solution. The solutions were prepared and stored in the dark. To prepare “gold seed solution,” 5 mL of 5×10^{-4} M HAuCl_4 was added to 5 mL 0.2 M cetyltrimethyl-ammonium bromide (CTAB) at 25°C and mixed rapidly until the color of the solution turned golden brown. About 600 μL of ice-cold 0.01 M NaBH_4 was added to the golden brown solution under vigorous stirring for at least 2 min. The color of the solution turned dark brown, and the seed solution was kept at 28°C for 3 h before later usage. To prepare “gold growth solution,” 200 μL of freshly prepared AgNO_3 solution (4 mM) was added to 5 mL 0.2 M CTAB at 25°C. About 5 mL of 1×10^{-3} M HAuCl_4 was then added to the mixture, and the solution changed to golden brown after mixing, and 70 μL of 0.0788 M ascorbic acid was added under vigorous stirring until the color of the solution became transparent. Gold seed solution (1.1 mL) was added and mixed well with the gold growth solution (10.27 mL). The mixture was settled for 5 min, and the solution color turned dark red. GNRs were synthesized by incubating the mixture at 30°C overnight. GNR mixture was then processed with centrifugation at 7,000 rpm for 15 min. Supernatant was collected and centrifuged at 14,000 rpm for 15 min. GNR pellet was obtained and suspended with purified water. One more centrifugation was conducted (14,000 rpm for 15 min), and the final gold pellet was collected for later studies.

Size and zeta potential of GNRs were analyzed by Zetasizer Nano ZS90 (Malvern Instruments, Malvern, UK). Morphology of GNRs was observed by transmission electron microscopy (TEM; TecnaiG20; FEI, Hillsboro, OR, USA). Briefly, fresh samples were diluted in Millipore water and mixed well. The samples were then transferred into cuvette for GNR size and zeta potential measurements. To visualize GNR morphology, a fresh sample was placed on copper grids with film and settled for 1 min. Excess liquid was

absorbed by paper, and the sample was air dried for TEM examination.

Preparation of 1,1-dioctadecyl-3,3,3,3-tetramethylindotricarbocyanine (DiR) solution

DiR (2 mg) was weighed and placed in 1.5 mL eppendorf tube. Ethanol (600 μL) was added to the EP tube. DiR solution was vortexed before usage.

Preparation of GNRs/DiR liposomes and GNRs/DiR-RLT liposomes

Cholesterol (chol; 20 mg), soy phosphatidylcholine (SPC; 150 mg), hydrogenated SPC (HSPC; 5 mg), and 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-conjugated polyethylene glycol (DSPE-PEG (2000)-Mal; 2.5 mg) were dissolved in 15 mL chloroform. DiR ethanol solution (120 μL) was added to the mixture under sonication. Sonication was continued for 15 min in the dark. Chloroform was then removed by rota-evaporation, and the lipid solution was dried to a thin and transparent film at the bottom of the flask. GNR solution (0.4 nM, 4 mL) was then added to the flask to hydrate and dissolve the film by sonication. The flask was then sonicated in ice bath for 40 times (200 W) (3 s, 3 s) and centrifuged at 8,000 rpm for 10 min. Supernatant was obtained as GNRs/DiR liposomes and kept at 4°C. To prepare GNRs/DiR-RLT liposomes, the supernatant was mixed with RLT solution. The mixture was vortexed for 1 min and shaken for 12 h at room temperature. The obtained GNRs/DiR-RLT liposomes were kept at 4°C.

High-performance liquid chromatography (HPLC) for the quantification of reacted RLT

Liposomes prepared with different DSPE-PEG (2000)-Mal:RLT ratios were injected to and analyzed by HPLC (Agilent 1100, flow rate: 1.0 mL/min, mobile phase: acetonitrile: water 20.8%: 79.2% (0.05% trifluoroacetic acid), column: Diamonsil™ C18, 4.6×200 mm, 5 μm , detection wavelength: 215 nm and temperature: 25°C). Unreacted RLT was calculated based on the area under the peak.

Table S1 The influence of GNR concentration on particle size, PDI, zeta potential, and appearance (N=3)

GNRs (nM)	Particle size (nm)	PDI	Zeta potential (mV)	Appearance
0.635	158.2±1.29	0.26±0.08	-20.5±4.92	Aggregation of GNRs after a few days
0.4	163.1±1.47	0.29±0.05	-26.1±3.86	Little aggregation of GNRs after a few days
0.2	161.6±1.9	0.27±0.13	-32.3±3.61	No aggregation

Abbreviations: GNRs, gold nanorods; PDI, polydispersity index.

Table S2 The influence of SPC:HSPC on particle size, PDI, zeta potential, densification of film, and stability of liposomes (N=3)

SPC:HSPC (mol/mol)	Particle size (nm)	PDI	Zeta potential (mV)	Densification of film	Stability
50:1	136.1±2.72	0.78±0.10	-21.5±1.64	Loose	Unstable
40:1	140±1.73	0.49±0.08	-26.1±0.97	Loose	Semi-stable
1:30	169.4±1.62	0.250±0.05	-32.8±1.04	Compact	Stable
1:40	204±2.18	0.28±0.09	-31.4±2.09	Compact	Stable
1:50	214±1.83	0.41±0.13	-24.8±0.57	Compact	Semi-stable

Abbreviations: HSPC, hydrogenated SPC; PDI, polydispersity index; SPC, soy phosphatidylcholine.

Table S3 The influence of chol addition on particle size, PDI, zeta potential, and stability of liposomes (N=3)

(H)SPC:chol (mol/mol)	Particle size (nm)	PDI	Zeta potential (mV)	Stability
10:1	145.1±2.06	0.37±0.09	-37.2±0.69	Unstable
8:1	152.8±1.47	0.21±0.07	-34.0±1.19	Stable
6:1	190.4±1.62	0.32±0.05	-30.8±1.03	Semi-stable

Note: (H)SPC = SPC + HSPC.

Abbreviations: chol, cholesterol; HSPC, hydrogenated SPC; PDI, polydispersity index; SPC, soy phosphatidylcholine.

Table S4 The influence of molar ratio between DSPE-PEG (2000)-Mal and RLT on the reaction rate of RLT (N=3)

DSPE-PEG (2000)-Mal:RLT	Conjugation percentage of RLT (%) to DSPE-PEG (2000)-Mal
1:1	29
2:1	71
3:1	90

Note: DSPE-PEG (2000)-Mal, 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-conjugated polyethylene glycol.

Table S5 The influence of ultrasonic condition on particle size, PDI, zeta potential, and appearance (N=3)

Power and time of sonication	Particle size (nm)	PDI	Zeta potential (mV)	Appearance
No sonication	237.2±2.15	0.68±0.14	-30.2±2.61	Translucent viscous liquid
100 W, 120 s	210.5±1.93	0.59±0.15	-29.6±1.94	Good transparency and dispersity
200 W, 120 s	203.8±2.37	0.37±0.09	-30.9±2.08	Good transparency and dispersity
300 W, 120 s	179.2±2.09	0.26±0.12	-32.1±3.07	Generation of bubbles during sonication
100 W, 240 s	181.9±1.72	0.31±0.07	-31.7±1.82	Good transparency and dispersity
200 W, 240 s	164.2±1.80	0.25±0.1	-34.9±2.15	Good transparency and dispersity

Abbreviation: PDI, polydispersity index.

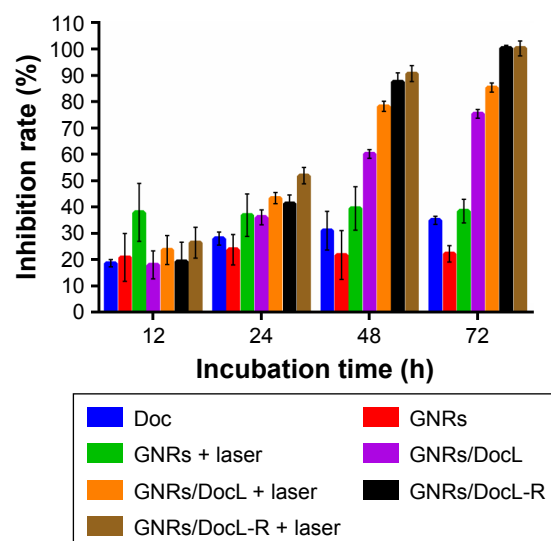


Figure S1 PC-3 cell inhibition caused by the listed treatments.

Notes: Data are presented as mean $\pm$ SD (n=3). Concentration of Doc, 80 μ g/mL; concentration of GNRs, 0.023 nM.

Abbreviations: Doc, docetaxel; GNRs, gold nanorods; GNRs/DocL-R, GNRs/Doc-liposome-RLT; PC-3, prostate cancer cell line.

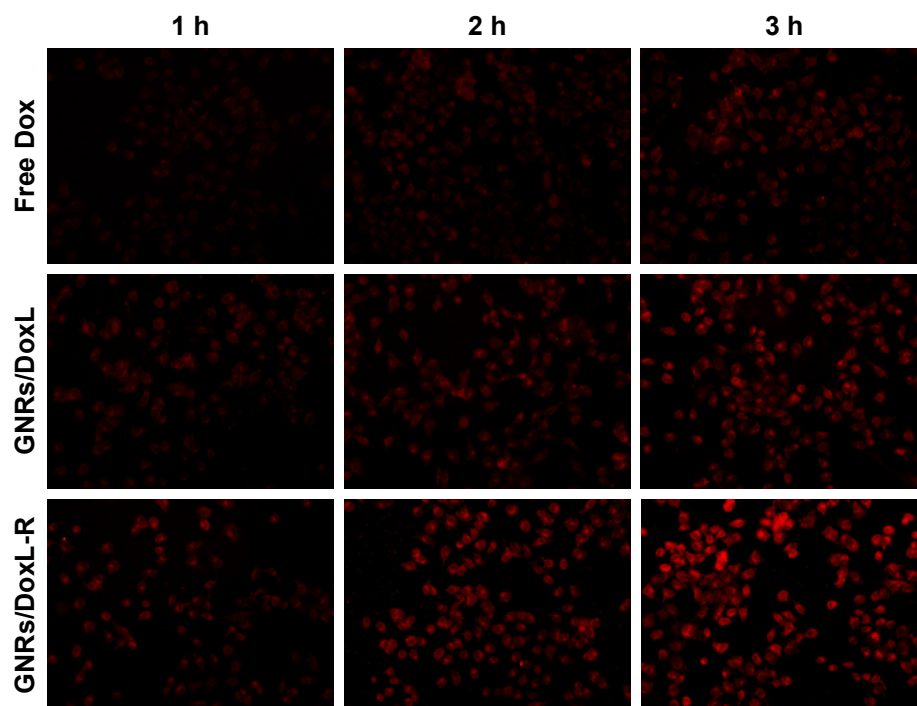


Figure S2 Cellular uptake of Dox by PC-3 cells.

Abbreviations: Dox, doxorubicin; PC-3, prostate cancer cell line.

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