

A sensitive and rapid chemiluminescence immunoassay for point-of-care testing (POCT) of copeptin in serum based on high-affinity monoclonal antibodies via cytokine-assisted immunization

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Purpose: Antibodies are key reagents in the development of immunoassay. We attempted to develop high-performance CPP immunoassays using high-affinity monoclonal antibodies prepared via cytokine-assisted immunization.

Methods: We used fetal liver tyrosine kinase 3 ligand (Flt3L), CC subtype chemokine ligand 20 (CCL20), and granulocyte-macrophage colony-stimulating factor (GM-CSF) to assist traditional subcutaneous immunization of preparing high-affinity monoclonal antibodies, and further to develop high-performance immunoassay methods for CPP.

Results: This novel immune strategy significantly enhanced immune response against CPP. Six anti-CPP monoclonal antibodies (mAbs) with high affinity were successfully screened and selected for application in a fully automated magnetic chemiluminescence immunoassay (CLIA). This robust and rapid assay can efficiently detect CPP in the range of 1.2–1250 pmol L⁻¹ with a detection limit of 6.25 pmol L⁻¹. Significantly, the whole incubation process can be completed in 30 min as compared to about 4.5 hr for the control ELISA kit. Furthermore, this assay exhibited high sensitivity and specificity, low intra-assay and inter-assay coefficients of variation (CVs < 15%). The developed assay was applied in the detection of CPP in 115 random serum samples and results showed a high correlation with data obtained using a commercially available ELISA kit (correlation coefficient, 0.9737).

Conclusion: Our assay could be applied in the point-of-care testing of CPP in the serum samples, and also the method developed in this study could be adopted to explore the detection and diagnosis of other biomarkers for various diseases.

Keywords: copeptin, cytokine-assisted immunization, high-affinity antibodies, chemiluminescence immunoassay, point-of-care tests, clinical application

Introduction

The need for a simple, rapid, and more accurate diagnosis and prognostic assessment of various diseases to assist with treatment decision has led to the investigation of new biomarkers. A previous study has indicated that CPP mirrors individual stress levels even more subtly than circulating cortisol.¹ This new biomarker can enable early decision making in clinical practice and also provide a reliable degree of correlation with various acute disease states, such as cerebrovascular event,²

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myocardial infarction³ or pneumonia,⁴ kidney disease and hypertension,⁵ osmotic alterations,⁶ human obesity,⁷ bipolar disorder,⁸ major depression,⁹ and childhood maltreatment.¹⁰ In triage of chest pain patients, determination of CPP, in addition to troponin, improves early diagnostic performance, especially after the onset of chest pain.¹¹ During the onset of an acute myocardial infarction (AMI), CPP is rapidly released from the pituitary gland and starts to return to normal levels within a few hours while troponin T concentrations are still normal.¹² In a research on CPP diagnostic cutoff of AMI among 5,000 individuals, the 99th percentile, which is usually suggested as the universal definition of a biomarker, was 18.9 pmol L⁻¹, the 97.5th percentile was 13 pmol L⁻¹, and the 95th percentile was only 9.8 pmol L⁻¹.¹¹ Apparently, the clinical value of CPP can be maximized only by shortening the detection cycle as much as possible while ensuring the sensitivity and specificity of the method. However, till date, only a few techniques have been applied in the determination of CPP concentration, among which are microtubes chemiluminescence immunoassay (tubes CLIA),¹³ radioimmunoassay (RIA),¹⁴ enzyme-linked immunoassay (ELISA),¹⁵ and electrochemical assay.¹⁶ Although their sensitivity levels could meet the required standards, assay durations are generally too long, making them unsuitable for use in clinical settings and hence their application only in scientific research. To the best of our knowledge, the point-of-care testing (POCT) for CPP has yet not been reported, hence it would be expedient to develop a POCT for the detection of CPP in clinical samples.

High-affinity antibodies are important for the establishment of a good performance immunoassay. Effectively improving the immune response may be a viable way to obtain high-affinity antibodies. Several studies have shown that cytokines can effectively improve immune response and successfully break immune tolerance. For example, Flt3L, an important cytokine, has been shown to promote the proliferation, differentiation, and maturation of pre-lymphocytes, dendritic cells (DCs), natural killer cells (NKs), and cytotoxic lymphocytes (CLs) both in vivo and in vitro, and plays an important role in the anti-tumor immune response.^{17,18} GM-CSF recruits, activates, and enhances the function of antigen presenting cells, making it a useful adjuvant in vaccines. Numerous studies in different animal models have clearly shown that plasmids expressing GM-CSF can enhance immune responses generated against DNA vaccines.^{19–23} Some researchers have also suggested the potential use of

CCL20 chemokine as a vaccine adjuvant to enhance Th1 mediated cellular and humoral immune responses in hepatitis C virus (HCV) core DNA immunization.²⁴ CCL20 chemokine attracts immature DCs, effector/memory T cells, and B cells, thereby increasing the ability of DCs in the capture and presentation of antigens. However, these studies are currently limited to DNA immunization, and the role of this cytokine in the preparation of monoclonal antibodies by improving the immune response of subcutaneous immunization has not been investigated.

Chemiluminescence immunoassays have been widely applied in the routine clinical analysis due to their high sensitivity, wide detection range, methodological reliability, and relative stability.^{25,26} Recently, magnetic-particles (MPs) have proven to be a suitable tool for highly sensitive detection of various biomarkers due to their higher specific surface area for immobilizing antibody, improved capture efficiency, reduced incubation time, and easy separation.^{27,28} More importantly, the efficient enrichment of MPs provides an increased amount of captured CPP at extremely low concentrations and thus afford high sensitivity and specificity.

In this work, cytokine-assisted immunization was applied to increase the immunity of the BALB/c mice, and several high-affinity antibodies were successfully screened and purified. For fully automated CLIA constructing, MPs were coated with a high-affinity antibody, alkaline phosphates (AP) labeled another anti-CPP antibody which identifies another site of the CPP and the concentration of the CPP was computed based on the observed luminescence intensity of the substrate. This fully automated CLIA exhibited an excellent specificity, stability, and reproducibility in human serum samples test and has an added advantage of obtaining results in a short period. Furthermore, the result of this CLIA compared favorably with a commercially available ELISA test kit and could serve as an excellent alternative for the detection of CPP in clinical samples.

Experimental Materials

Peptides related to CPP were chemically synthesized, purified, and quality-controlled by Jill Biochemical Co., Ltd. (Shanghai, China). The peptides were N-CPP (sequence CATQLDGPAGALLRLV, representing positions 132–147 of pre-provasopressin plus an N-terminal cysteine residue), C-CPP (sequence CLAGAPEPFEPAPDAY, representing positions 149–164 of pre-provasopressin plus an N-terminal cysteine residue), and CPP (sequence

ATQLDGPAGALLRLVQLAGAPEPFEPAPQDAY, representing positions 132–164 of pre-provasopressin). Peptides were conjugated via mmaleimidobenzoyl-N-hydroxysuccinimide ester to keyhole limpet hemocyanin (KLH) to act as the immunogen (KLH-C-CPP, KLH-N-CPP). Murine CCL20, Flt3L, GM-CSF coding sequence cloned into pCAGGS vector under the control of a CAG promoter using the EcoRI and EcoRI restriction sites were assisted by Atagenix Biotechnology Co., Ltd. (Wuhan, China). The final construct was confirmed by sequencing and transfection of HEK 293 T cells which purchased from the Cell Bank of Chinese Academy of Sciences. Female six-to-eight-week-old BALB/c mice were purchased from the Experimental Animal Center of Nanjing Medical University (Nanjing, China) and were allowed to acclimatize for 1 week prior to immunization. The carboxyl micromagnetic particles ($2.8 \pm 0.2 \mu\text{m}$), alkaline phosphates, and the chemiluminescent substrate (4-methoxy-4-(3-phosphatephenyl)-spiro-(1,2-dioxetane-3,2-adamantane) (AMPPD) were supplied by Guangzhou Wondfo Biotech Co., Ltd. (Guangzhou, China). All buffer reagents and other inorganic chemicals were purchased from Sigma-Aldrich (Shanghai, China). Random clinical serum samples were obtained from The Third Affiliated Hospital of Guangzhou University of Chinese Medicine (Guangzhou, China). All aqueous solutions were prepared using double distilled water.

Apparatus

Automatic chemiluminescence immunoassay instrument (Limray1200) was supplied by Rayto Life Science Co., Ltd. (Shenzhen, China). The concentration of antibody was detected by NanoDrop (2000c) spectrophotometer from Thermo fisher scientific (Shanghai, China). The affinity of antibodies was evaluated by the BLItz[®] System from PALL ForteBio LLC (Menlo Park, USA). Commercial enzyme-linked immunosorbent assay (ELISA) kits obtained from Cusabio Biotech Co. Ltd. (Wuhan, China) were used in the determination of CPP concentration in serum samples following the manufacturer's instructions.

Production and characterization of anti-CPP monoclonal antibody

Female BALB/c mice were injected subcutaneously with 100 μg immunogen emulsified with the identical volume of complete Freund's adjuvant for the first immunization.

Subsequent booster immunizations with 50 μg immunogen emulsified with an equal volume of incomplete Freund's adjuvant were injected at two-week intervals. Seven days after each immunization, cytokine plasmids (10 μg) were injected through the tail vein. Seven days after the second and third injection, antibody titers in mouse serum samples collected from the tail vein were checked by ELISA. The hybridoma development and monoclonal antibody characterization were carried out as previously reported.^{29,30} Meanwhile, the affinity of antibodies was evaluated according to the protocol provided by the manufacturer. All animal experiments and welfare of the animals were performed under ethical approval from and in agreement with the guidelines of the Institutional Animal Care and Use Ethics Committee of South China University of Technology and also in accordance with the policy of the National Ministry of Health.

Coating MPs with CPP antibody

High-affinity CPP monoclonal antibody was covalently coupled to MPs in the binding buffer via the terminal amine. Firstly, 20 mg mL^{-1} MPs were placed in a 2 mL Eppendorf tube. The MPs were then washed five times with binding buffer. During the washing, the tube was placed on a magnetic concentrator and the supernatant was removed. Afterward, the MPs were resuspended in 2 mL binding buffer. Antibody solution was added to the above suspension for the conjugation of CPP antibodies and MPs by incubation at 37°C with shaking overnight. After incubation, the tube was placed in a magnetic concentrator to separate the supernatant. The residual binding sites on the MPs were blocked with 3% bovine serum albumin (BSA) incubated at 37°C with slight shaking for 2 hrs. After washing 5 times, the coated MPs (mAb-MPs) were dispersed in 2 mL buffer and stored at 4°C.

Preparation of AP-conjugated CPP antibody

The AP and anti-CPP antibody were coupled with glutaraldehyde. First, the AP and anti-CPP antibody were suspended in ultrapure water and diluted to 4 and 8 mg mL^{-1} , respectively. A 250 μL aliquot of the 4 mg mL^{-1} AP solution was then transferred to a 1.5 mL Eppendorf tube and gently mixed with 250 μL of 8 mg mL^{-1} anti-CPP antibody solution. Second, 0.5 mL of 1% glutaraldehyde in 0.1 mol L^{-1} phosphate buffer (pH 7.4) was added to the solution. The resulting

mixture was incubated at 37°C with gentle shaking for 4 hrs in the dark. Third, 0.1 mL of 1 mol L⁻¹ mono-ethanolamine solution was added to the mixture, which was subsequently incubated with shaking for 2 hrs at room temperature. The mixture was dialyzed against PBS solution at 4°C overnight. After dialysis, CPP antibody-AP conjugate was then transferred to an Eppendorf tube and mixed with an equal volume of glycerin and 1% BSA. Finally, the AP-conjugated CPP antibody (AP-mAbs) was stored at -20°C for use in further experiments.

MPs-based fully automated CLIA

CPP was detected with the automatic chemiluminescence immunoassay instrument (Limiray1200) which is based on the sandwich immunoassay using the anti-CPP antibody coated MPs and AP-labeled anti-CPP antibody. These two monoclonal antibodies can recognize different epitopes of the CPP. The reaction procedure for the MPs-based CLIA was slightly different from the conventional microtubes CLIA. In the MPs-based CLIA, MPs coated antibodies served as the solid phase instead of the microtubes, and hence the whole immunoassay process can be performed using the automated chemiluminescence immunoassay instrument. A schematic diagram of the methodology is shown in Figure 1. About 50 µL of mAb-MPs and CPP samples (30 µL) of different concentrations were pipetted into the corresponding tubes and incubated at 37°C for 20 mins (capture time) with gentle shaking. The tubes then were passed through washing stations with magnets to wash with washing buffer (0.01 mol L⁻¹ PBS containing 0.05% Tween 20) for three times to remove non-specific bindings. Then, the AP-mAbs was added and incubated at 37°C for 10 min with gentle shaking. At this point, the sandwich immunocomplex MPs-CPP-AP had formed. The sandwich immunocomplex was magnetically separated, and the excess AP-mAbs were removed by washing. Subsequently, the solution containing AMPPD (200 µL) was added to the sandwich complex. The resulting mixture was placed in the immunoassay instrument, and the value of relative light units (RLU) was measured.

Optimization of immunoassay reagents

Serial dilutions of AP-mAbs (1:50, 1:100, 1:200, 1:500) and mAb-MPs (1:20, 1:50, 1:100, 1:200, 1:500) were allowed to react with a standard positive sample S3 and negative sample S0 of CPP as described earlier. The

optimal dilution was determined from the maximum RLU ratio of the positive vs negative samples (RLU_{S3}/RLU_{S0}). Other experimental parameters, such as the pH, incubation time, accuracy, precision, sensitivity, and stability were also investigated. To monitor the feasibility of the newly developed immunoassay, 115 clinical serum specimens were analyzed using our newly developed immunoassay test kit and a commercially available ELISA kit.

Statistical analysis

Bland-Altman plot and Passing-Bablok regression analysis were performed with MedCalc Software (MedCalc, Mariakerke, Belgium). Statistical analysis on a completely randomized design was conducted using the one-way ANOVA procedure, with SPSS 19.0 software, at a level of significance set at $p=0.05$. All data were presented as mean±standard deviation (mean±SD). Differences were accepted as significant when $p<0.05$.

Results and discussion

Production and characterization of anti-CPP mAbs

In antibody affinity maturation, the average affinity of the antibodies produced in the secondary immune response is usually higher than the primary immune response.³¹ Longer duration of antigen presentation is likely to generate higher levels of immunity. For example, a simple extension of the interval between influenza vaccine doses resulted in higher levels of neutralizing antibodies.³² Likewise, a modeling study predicted that sequential immunogens would be more effective than mixed immunogens to target the HIV-1 Env CD4bs epitope.³³ Hence, antibodies preparation for research, diagnostic, and therapeutic applications frequently relied on a series of immunization of animals with antigen over the process of several weeks to enhance the proliferation of mature cells producing antigen-specific antibodies. Adjuvants are often used for promotion of affinity maturation through increases the persistence of antigen.³⁴ The use of CCL20 as a vaccine adjuvant could enhance Th1-mediated cellular and humoral immune responses.²⁴ CCL20 chemokine attracts immature antigen presenting cells, and Flt3L could promote the proliferation, differentiation, and maturation. Similarly, GM-CSF is also able to recruit, activate, and enhance the function of antigen presenting cells. Subcutaneous and cytokine-assisted immunization approaches were taken to produce monoclonal antibodies.

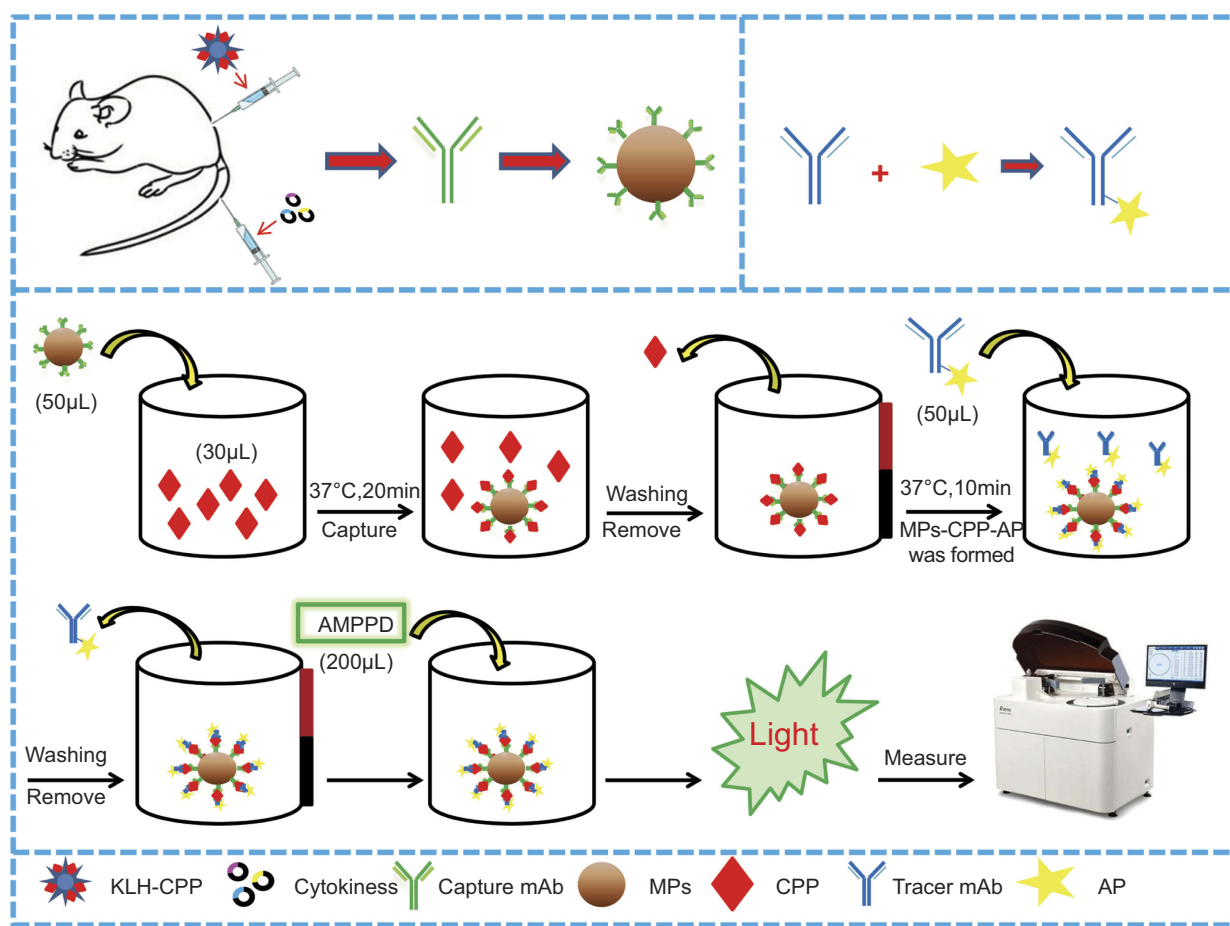


Figure 1 Schematic illustration of the MPs-based fully automated CLIA method for quantitative detection of coceptin.

Abbreviations: KLH-CPP, coceptin conjugated with keyhole limpet hemocyanin; mAb, monoclonal antibody; MPs, magnetic particles; CPP, coceptin; AP, alkaline phosphatase; AMPPD, 4-methoxy-4-(3-phosphatophenyl)-spiro-(1,2-dioxetane-3,2'-adamantane).

For the cytokine-assisted immunization, we first subcutaneously immunized mice and after a few days, cytokines were delivered via the tail vein to further enhance the presentation of the antigen. In the second and third vaccinations, we observed that serum titers were more increased with cytokine-assisted immunization. The rate of positive hybridoma and the amount of stable hybridoma was greatly increased following the application of this novel immunization strategy (Table S1). Eight hybridoma cell lines with a stable secretion of mAb against C-CPP were successfully selected after a series of screening. It is worth noting that the affinity of monoclonal antibodies prepared via cytokine-assisted immunization was significantly improved (Figure 2A and 2B). As shown in Figure 2C, the $1/k_D$ value of cytokine-assisted antibody (6-18G10) was approximately 16.7 times higher than that of antibodies produced through the normal subcutaneous immunization (4-30E2). The smaller the k_D value, the higher the affinity, hence Li et al recommended that K_D values in the range of 5.3–14.0 nM or less are high-

affinity antibodies.³⁵ Six of antibodies produced through the cytokine-assisted method had K_D values less than 5.3 nM with high-affinity for antigen detection (Table S2). The purity of all mAbs was above 90% as analyzed on SDS-PAGE (Figure S1). The titer values of the antibodies against CPP standards were up to $1:10^6$ using CPP concentration of $1 \mu\text{g mL}^{-1}$ and absorbance above 0.3 at 450 nm (Figure S2). These properties are basic experimental requirements for any good antibody and hence these antibodies could be used in further investigation.

Optimization of immunoassay

Selection of the optimal antibody combination

Two C-CPP antibodies (6-20G5 and 6-18G10) with high affinity and one N-CPP antibody (4-30E2) were used for coating and labeling, respectively. In pairs 1 and 2, 4-30E2 was used as the coating antibody with MPs while 6-18G10 and 6-20G5 were labeled with AP. Conversely, in pairs 3 and 4, 6-20G5 and 6-18G10 were used for coating MPs and

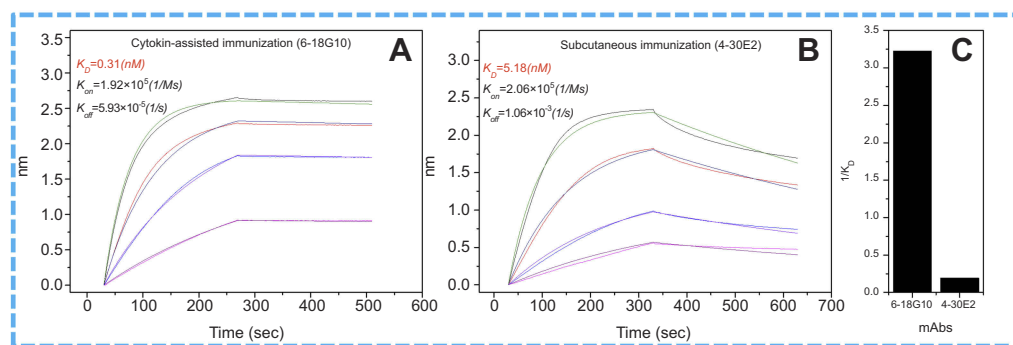


Figure 2 Affinity of the paired antibodies. **(A)** The affinity of capture antibody (6-18G10) from cytokine-assisted immunization. **(B)** The affinity of labeled antibody (4-30E2) from subcutaneous immunization. **(C)** Comparison of both antibodies by the value of $1/K_D$.

Abbreviations: K_D , equilibrium dissociation constant; K_{on} , association rate constant; K_{off} , dissociation rate constant.

4-30E2 for labeling. Since there were four possible combinations, the first development step was the selection of the optimal antibody combination. A standard positive sample S3 and negative sample S0 of CPP were analyzed under all combinations. Then, the signal ratio of RLU_{S3}/RLU_{S0} was calculated and compared for the determination of the best antibody combinations. As shown in Figure 3, the RLU_{S3}/RLU_{S0} of pairs 1 and 2 were lower than the other two pairs because of the lower affinity for 4-30E2 (only 5.18 nM), showing relatively weak capture capacity for the antigen. It can be observed that the high-affinity antibodies were more suitable for use as capture antibodies. Ultimately, pair 3 was selected as the most suitable antibody pair with the highest RLU_{S3}/RLU_{S0} ratio of all ($p < 0.05$).

Optimization of AP-mAbs and mAb-MPs concentration

The amount of AP-mAbs antibody and mAb-MPs are key parameters that affect the sensitivity and accuracy of immunoassay, especially in a sandwich immunoassay. In this study, the dilution ratios of AP-mAbs and mAb-MPs were optimized by comparing with serial dilutions of standard positive sample (S3) and negative sample (S0). Similarly, the signal ratio of RLU_{S3}/RLU_{S0} served as the parameter for the determination of the best assay. As shown in Figure 4, the RLU_{S3}/RLU_{S0} increased when the dilution ratios of AP-mAbs increased from 1:500 to 1:50 at all dilution ratios of the examined mAb-MPs ($p < 0.05$). When the dilution ratios of AP-mAbs were 1:50 and 1:100, the difference in RLU_{S3}/RLU_{S0} was insignificant. In

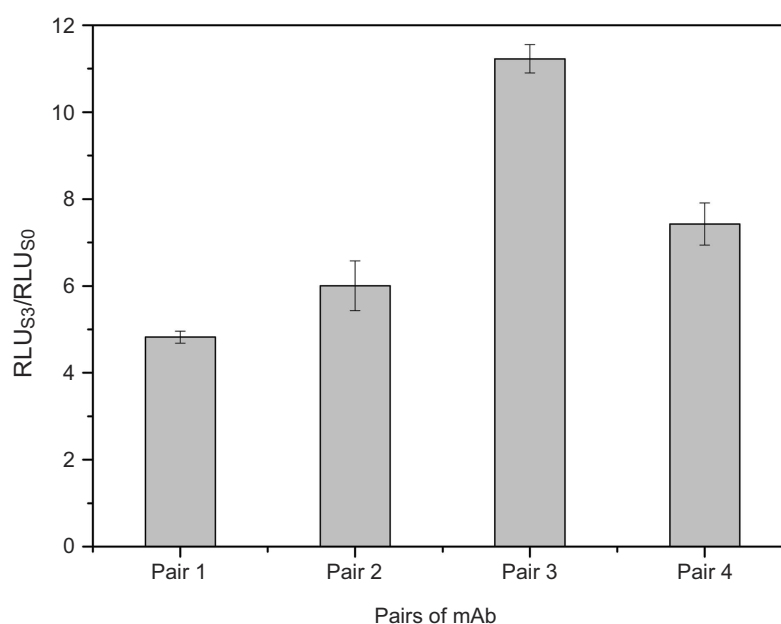


Figure 3 Selection of the optimal antibody combination. Detection methods: RLU_{S3}/RLU_{S0} ratio obtained for all antibody pairs, with 0 and 12.5 pmol L^{-1} of the CPP standards. Pair 1: 6-20G5-labeling with 4-30E2-coating, Pair 2, 6-18G10-labeling with 4-30E2-coating, Pair 3: 4-30E2-labeling with 6-18G10-coating, Pair 4: 4-30E2-labeling with 6-20G5-coating.

Abbreviations: mAb, monoclonal antibody; RLU, relative light units.

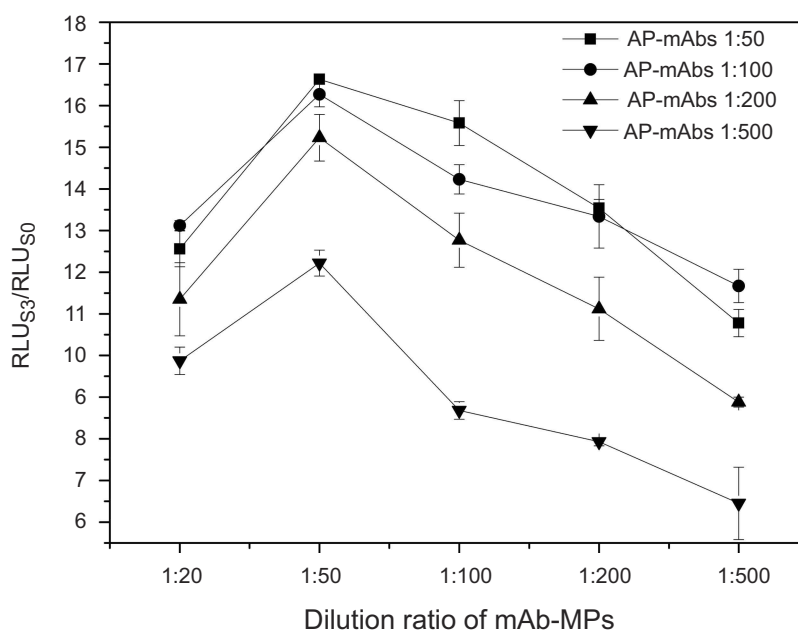


Figure 4 Optimization of AP-mAbs and mAb-MPs concentration. Detection methods: RLU_{S3}/RLU_{S0} ratio obtained for all tests, with 0 and 12.5 pmol L^{-1} of the CPP standards.

Abbreviations: RLU, relative light units; AP-mAbs, alkaline phosphates-conjugated antibodies; mAb-MPs, monoclonal antibody-coated magnetic-particles.

the case of the mAb-MPs, the RLU_{S3}/RLU_{S0} showed a peak corresponding to the dilution ratio of 1:50 ($p < 0.05$), the RLU_{S3}/RLU_{S0} increased when the concentration of mAb-MPs decreased from 1:50 to 1:500, indicating that not all of the available CPP was captured. When 1:50 of mAb-MPs were added, there was sufficient binding of the antigen to mAb-MPs resulting in the highest RLU_{S3}/RLU_{S0} ratio. By increasing the concentration of antibodies above 1:50 of mAb-MPs, that is, 1:20 dilution, the excess mAb-MPs led to lower sensitivity because of the close association between mAb-MPs, thereby absorbing the emitted light and thereby blocking luminescence.³⁶ Hence, considering both the sensitivity and the assay cost, we selected the dilution ratios of 1:100 and 1:50 for AP-mAbs and mAb-MPs, respectively.

Optimization of substrate buffer pH

The pH value and incubation time are also two important parameters since the acidity of the solution greatly affects the activity of the immobilized protein and extended incubation time probably results in dissociation of the antigen-antibody complex. These two parameters were assessed by detecting higher maximal RLU values for the immunoassay detection of CPP standard sample S9. AP-mAbs solution (1:100, 50 μL), mAb-MPs solution (1:50, 50 μL), and standard sample (S9, 30 μL) were incubated at 37°C for 0–60 mins. The MPs-CPP-AP complex was washed at 5-min interval, and the

chemiluminescent substrates (200 μL) with different pH values were added to measure the RLU. Substrate buffers of different pH were prepared by adding 1 mol L^{-1} NaOH solution. The influence of pH and incubation time on the overall kinetics of light emission is shown in Figure 5. The RLU intensity increased gradually until it reached a steady state. The observed biphasic behavior may be due to the overall dynamics of light emission, which was a two-step process that causes emission delays before steady state chemiluminescence.³⁷ When the pH was 7.0, the RLU value increased from 0 to 20 mins and reached an unstable plateau. Furthermore, the maximum RLU at this pH was lower than others. The RLU apparently reached a maximum at pH of 9.0, thus making it the best suitable for diagnostic testing, similar to the results reported in a literature.³⁶ Concerning the incubation time of the assay, the maximal RLU increased with incubation time from 0 to 30 mins. However, there was no change from 30 to 50 mins, indicating that the antigen-antibody complex formation had reached equilibrium. Hence, 30 mins were selected as the most suitable incubation time period.

Method evaluation

Calibration curve for the determination of CPP

Under the optimal condition, a series of concentrations of standard CPP solutions (Table 1) were measured. The

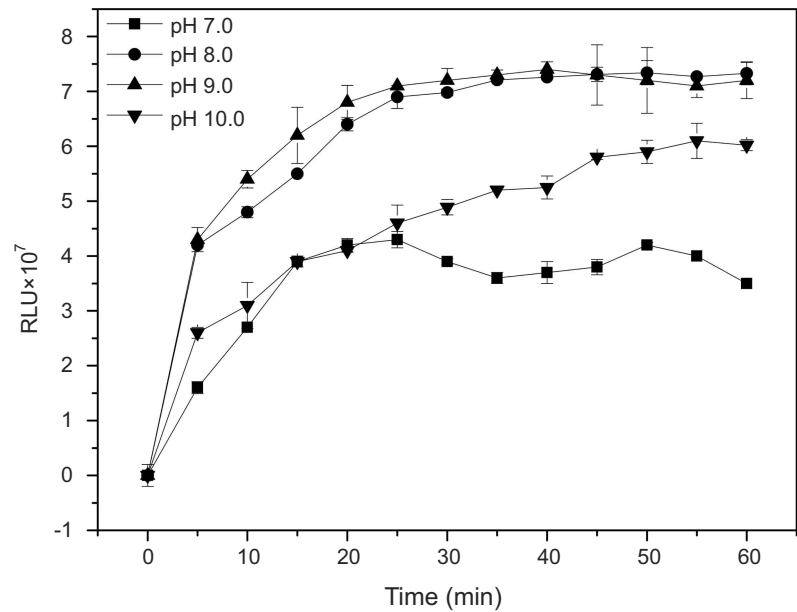


Figure 5 Optimization of substrate buffer pH and incubation time. Detection methods: RLU_{S9} obtained for all tests, which 1,250 pmol L⁻¹ of the CPP standards. **Abbreviation:** RLU, relative light units.

Table 1 The concentration of standard CPP solutions (pmol L⁻¹)

| S0 | S1 | S2 | S3 | S4 | S5 | S6 | S7 | S8 | S9 |
|----|-----|------|------|----|------|-----|-----|-----|------|
| 0 | 2.5 | 6.25 | 12.5 | 25 | 62.5 | 125 | 250 | 625 | 1250 |

Note: S0-S9 indicate samples of standard CPP with a series of concentrations. **Abbreviation:** CPP, copeptin.

calibration curve for CPP standards was constructed, as shown in Figure 6. The limit of detection (LOD) was determined from the average absorbance by adding twice the mean of the standard deviation from 10 S0 wells. The standard curve exhibited a detection range of 1.2–1250 pmol L⁻¹ with a LOD of 6.25 pmol L⁻¹ and a correlation coefficient of 0.9993. These features of our newly developed assay are within the required parameters for use in clinical diagnostic (below 18.9 pmol L⁻¹).¹¹ We also compared this assay with previously reported assays used in the detection of CPP to evaluate the competitive advantage of this assay. As shown in Table 2, the incubation time of our assay was much lower than that of previously reported assays, including Tubes CLIA, RIA, and ELISA. Although the assay time for the electrochemical assay method was only 5 min, the LOD was 37 pmol L⁻¹, which is far beyond the clinical requirement limit for use in clinical applications. Taken together, these results indicated that our fully automated MPs based-CLIA developed displayed better performance.

Accuracy, precision, and stability analysis

The recovery rate of dilution is often used to assess the accuracy of assays.¹³ In this study, five serum samples were tested after linear dilutions (up to 1:32) (Figure 7). The obtained values were multiplied by the dilution factor and compared with the original undiluted concentrations. None of the 5 samples showed a deviation >15% of the original concentrations. To confirm the precision of this method, intra-assay and inter-assay calibrations were performed using three different concentrations of CPP standards with six duplicates for both intra- and inter-assays conducted in 7 days. As shown in Table 3, both CVs were below 15%. Furthermore, the stability of the fully automated CLIA was also investigated. Upon storage of the AP-mAbs at 4°C, and CPP standards and AP-mAbs at -20°C for 30 days, there was no significant change in the readings obtained (Table S3). These results point to the establishment of a high performance and accurate CPP detection method in clinical samples.

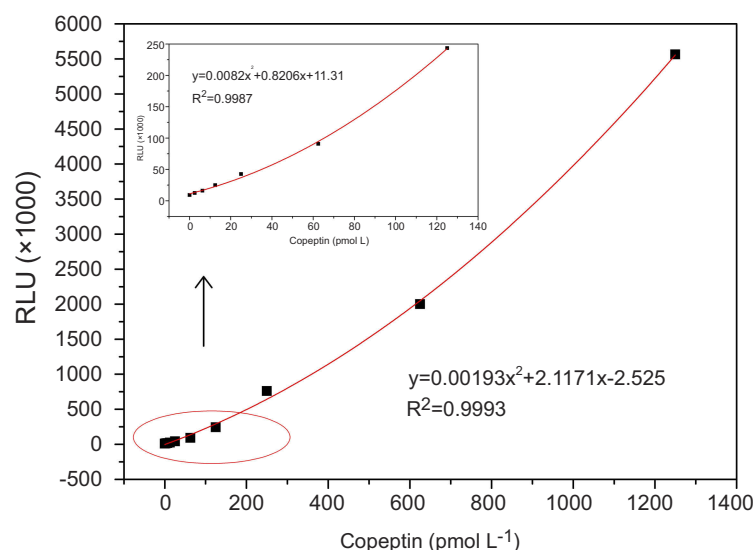


Figure 6 CPP dose-response calibration curve obtained from fully automated CLIA. A series of CPP standard samples (0, 2.5, 6.5, 12.5, 25, 62.5, 125, 250, 625 and 1,250 pmol L⁻¹) were used (n=3).

Abbreviations: CPP, copeptin; RLU, relative light units.

Table 2 Comparison of performance CPP detection methods

| Method | LOD | Detection range | Incubation time | Reference |
|-----------------------|---------------------------|--------------------------------|-----------------|-----------|
| Tubes CLIA | 2.25 pmol L ⁻¹ | 2.25–1215 pmol L ⁻¹ | 2 hrs | 13 |
| RIA | — | 0–2500 pmol L ⁻¹ | >24 hrs | 14 |
| ELISA | 19.5 pmol L ⁻¹ | 19.5–1250 pmol L ⁻¹ | 4.5 hrs | 15,38 |
| Electrochemical assay | 37 pmol L ⁻¹ | 25–125 pmol L ⁻¹ | 5 mins | 16 |
| Fully automated CLIA | 6.25 pmol L ⁻¹ | 1.2–1250 pmol L ⁻¹ | 30 mins | This work |

Note: This would be a highlight that the incubation process was only 30 min for the proposed assay.

Abbreviations: CPP, copeptin; LOD, limit of detection; CLIA, Chemiluminescence immunoassay; RIA, radioimmunoassay; ELISA, enzyme-linked immunoassay.

Cross-reactivity studies

The specificity of the CLIA for CPP was evaluated using other structural or functional analogs such as B-type natriuretic peptides (BNP), N-terminal pro-B-type natriuretic peptide (NT-proBNP), troponin T (cTnT), myoglobin, creatine kinase-myocardial band (CK-MB) that are often detected in combination with CPP in the diagnosis of heart failure or AMI.^{11,39} The RLU value was measured for each analyte and the cross-reactivity (CR) for each analog was calculated according to the following equation:

$$CR = \frac{\text{RLU of analyte}}{\text{RLU of CPP}} \times 100\%$$

As indicated in Table S4, none of these analytes showed significant reactivity with CPP antibody (<0.2%)

even at a higher concentration than that of CPP. These observations demonstrated that the antibody obtained by cytokine-assisted immunization were of high affinity and excellent sensitivity for the detection of CPP.

Method comparison

To further assess the effectiveness of this analytical method, 115 serum specimens consisting of 53 low-level concentration samples (<18.9 pmol L⁻¹), 46 median value samples (18.9–50 pmol L⁻¹), and 16 high-value samples (>50 pmol L⁻¹) were analyzed and compared with a commercially available ELISA kit. It is worth mentioning that the incubation process of the ELISA method for the diagnosis of CPP takes over 4.5 hrs, while this proposed method was 30 mins. The results of both methods were compared based on the Passing-Bablok regression

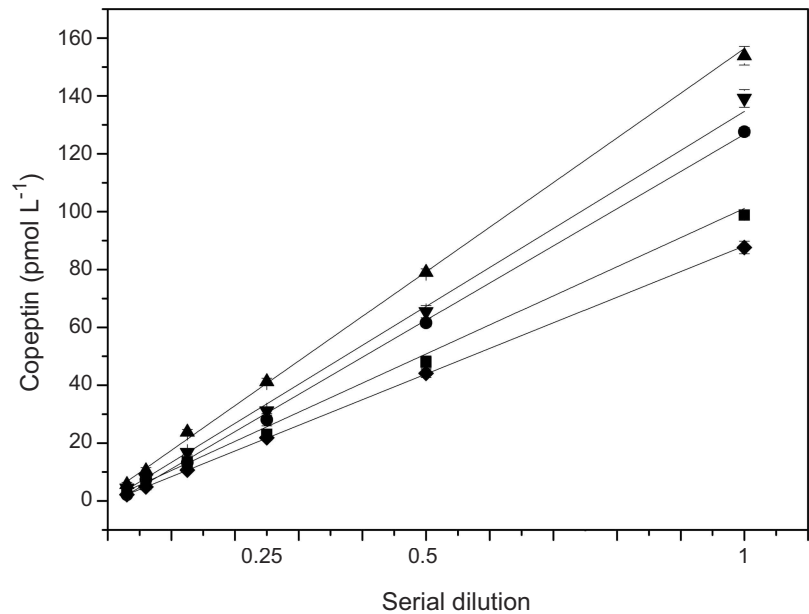


Figure 7 Linearity on dilution (up to 1:32) of 6 representatives of 5 tested samples.

Table 3 Intra-assay and inter-assay tests

| Times | Added concentration of CPP(pmol L ⁻¹) | | | | | | | | |
|-------|---|------|--------|------------|------|--------|-------------|------|--------|
| | 10.0 (n=6) | | | 50.0 (n=6) | | | 100.0 (n=6) | | |
| | Average | SD | CV (%) | Average | SD | CV (%) | Average | SD | CV (%) |
| 1 | 9.86 | 0.36 | 3.70 | 50.14 | 0.71 | 1.42 | 100.55 | 3.01 | 2.99 |
| 2 | 9.92 | 0.28 | 2.85 | 49.94 | 1.39 | 2.80 | 99.51 | 3.24 | 3.26 |
| 3 | 9.94 | 0.39 | 4.01 | 49.69 | 1.66 | 3.34 | 99.50 | 2.07 | 2.08 |
| 4 | 10.25 | 0.65 | 6.29 | 49.93 | 2.07 | 4.14 | 100.43 | 2.95 | 2.94 |
| 5 | 10.38 | 1.28 | 12.30 | 51.08 | 2.83 | 5.53 | 99.61 | 2.98 | 2.99 |
| 6 | 10.03 | 0.59 | 5.92 | 47.51 | 2.62 | 5.52 | 96.46 | 2.39 | 2.49 |

Note: Intra-assay and inter-assay tests were performed using three different concentrations of CPP standards.

Abbreviations: CPP, copeptin; CV, coefficient of variation; SD, standard deviation.

analysis (Figure 8A) and Bland–Altman plot (Figure 8B). The correlation coefficient (R^2) of the regression line of the Passing–Bablok regression analysis was 0.9737, indicating a good linear relationship between the two measurements. The Bland–Altman plot between the two methods was performed with serum samples, and the mean relative difference (95% limits of agreement) was 3.5% of the Bland–Altman plot results, revealing that there was no significant bias between these two methods.

Conclusions

The production of high-quality antibodies and the application of advanced analytical technologies would guarantee

the development of high-performance immunoassay methods. In this study, we have successfully produced ultra-high affinity antibodies against the CPP and have applied them in a chemiluminescence immunoassay for the rapid and sensitive determination of CPP in human serum. The affinity of the monoclonal antibody prepared via cytokine-assisted immunization was significantly improved approximately 16.7 times that of the traditional immunization method. Using these high-affinity antibodies for the detection of CPP, the sensitivity and specificity of our novel assay were significantly improved. Ultra-high affinity antibody weakens the dissociation of the antigen-antibody complex and MPs provide a large surface-to-area ratio,

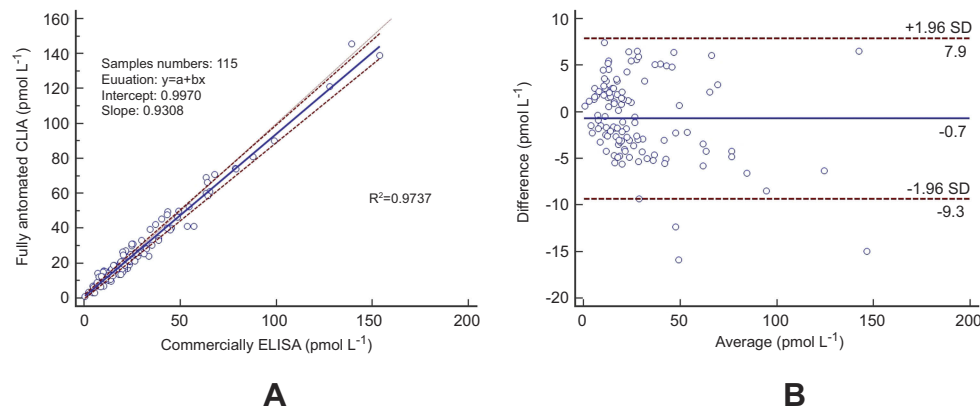


Figure 8 Comparison between fully automated CLIA and commercially available ELISA test kits. The correlation (A) and the average difference (B) between the two methods.

Abbreviations: CLIA, Chemiluminescence immunoassay; ELISA, enzyme-linked immunoassay.

hence the enhanced detection of CPP at low concentration. The utilization of these antibodies in the detection of CPP provides a suitable detection range between 1.2–1250 pmol L⁻¹ and a detection limit of 6.25 pmol L⁻¹ which is far below the standard requirement for the detection of CPP (<18.9 pmol L⁻¹). In addition, the application of the automatic chemiluminescence immunoassay instrument greatly shortened the incubation time (within 30 mins), whereas a minimum of 2-hr incubation time is required for other commercially available methods. This method was also applied in the detection of CPP in clinical serum specimens, and the results were significantly consistent as compared with other commercially available CPP detection ELISA kit. Hence, our assay could provide an alternative platform for the rapid and accurate detection of CPP in clinical samples. The method developed in this study could be adopted to explore the detection and diagnosis of other biomarkers for various diseases.

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Disclosure

The authors report no conflicts of interest in this work.

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Supplementary materials

Table S1 The comparison of subcutaneous and cytokine-assisted immunization

| Immunization strategy | Number of mice | Serum titer of 2nd vaccinations | Serum titer of 3rd vaccinations | Rate of positive hybridoma | Amount of stable hybridoma |
|-----------------------|----------------|---------------------------------|---------------------------------|----------------------------|----------------------------|
| Subcutaneous | 1 | <1: 10 ³ | 1: 10 ⁴ | - | - |
| | 2 | 1: 10 ⁴ | 1: 10 ⁵ | 9/146 (0.062) | 2 |
| | 3 | <1: 10 ³ | <1: 10 ³ | - | - |
| | 4 | 1: 10 ⁴ | 1: 10 ⁶ | 12/136 (0.089) | 2 |
| | 5 | <1: 10 ³ | <1: 10 ³ | - | - |
| Cytokine-assisted | 1 | 1: 10 ⁴ | 1: 10 ⁶ | - | - |
| | 2 | 1: 10 ⁵ | 1: 10 ⁶ | 104/337 (0.309) | 8 |
| | 3 | <1: 10 ³ | 1: 10 ⁴ | - | - |
| | 4 | <1: 10 ³ | 1: 10 ⁵ | - | - |
| | 5 | <1: 10 ³ | <1: 10 ³ | - | - |

Note: Serum titer was evaluated via CPP concentration of 1 $\mu\text{g mL}^{-1}$ and absorbance above 0.3 at 450 nm. “-” represents mice that were not used in hybridoma preparation.

Table S2 The affinity of the antibodies prepared via cytokine-assisted immunization

| | K _D (nM) | K _{on} (1/ Ms) | k _{on} Error | K _{off} (1/s) | K _{off} Error | Rmax | Rmax error | Requilibrium | X ² | R ² |
|---------|---------------------|-------------------------|-----------------------|------------------------|------------------------|-------|------------|--------------|----------------|----------------|
| 6-20G5 | 0.14 | 2.08E+05 | 1.41E+03 | 2.93E-05 | 6.94E-06 | 2.592 | 0.005201 | 2.586 | 1.843 | 0.9985 |
| 6-8D2 | 2.09 | 4.35E+04 | 1.48E+02 | 9.08E-05 | 2.46E-06 | 1.685 | 0.004778 | 1.659 | 0.03397 | 0.9999 |
| 6-18H3 | 5.07 | 1.14E+05 | 4.07E+02 | 5.75E-05 | 4.70E-06 | 2.539 | 0.003137 | 2.529 | 0.8136 | 0.9994 |
| 6-5G8 | 0.7 | 9.57E+04 | 3.89E+02 | 6.67E-05 | 4.28E-06 | 2.607 | 0.00354 | 2.594 | 0.8124 | 0.9995 |
| 6-2A9 | 3.12 | 8.72E+04 | 3.62E+02 | 2.72E-04 | 5.17E-06 | 2.522 | 0.005042 | 2.465 | 0.7758 | 0.9994 |
| 6-3C10 | 19.34 | 6.42E+04 | 1.06E+03 | 1.24E-03 | 1.68E-05 | 2.504 | 0.03084 | 2.186 | 1.361 | 0.9977 |
| 6-18G10 | 0.31 | 1.92E+05 | 4.25E+02 | 5.93E-05 | 2.86E-06 | 2.413 | 0.002477 | 2.402 | 0.8382 | 0.9995 |
| 6-11C3 | 15.18 | 3.39E+05 | 6.16E+03 | 5.14E-03 | 6.85E-05 | 2.191 | 0.01945 | 1.784 | 6.697 | 0.9857 |

Note: The antibodies that K_D value were less than 5.3 nM could be regared as high-affinity antibodies.

Abbreviations: K_D, equilibrium dissociation constant; k_{on}, association rate constant; k_{off}, dissociation rate constant.

Table S3 Stability of the reagents at store (n=3)

| CPP (pmol L ⁻¹) | Days 10 | | Days 20 | | Days 30 | |
|-----------------------------|-------------|--------|-------------|--------|-------------|--------|
| | Value | RD (%) | Value | RD (%) | Value | RD (%) |
| 5.6 | 6.21±0.55 | 10.89 | 5.68±0.58 | 1.43 | 5.54±0.75 | -1.01 |
| 15.8 | 14.24±1.81 | -9.87 | 16.25±1.59 | 2.84 | 14.66±1.63 | -7.21 |
| 30.5 | 29.87±3.33 | -2.08 | 30.1±3.69 | -1.31 | 28.81±3.08 | -5.53 |
| 50.4 | 50.66±3.9 | 0.52 | 51.89±4.43 | 2.97 | 52.01±4.54 | 3.2 |
| 132.5 | 131.31±3.18 | -0.9 | 127.67±6.09 | -3.64 | 130.82±2.01 | -1.26 |

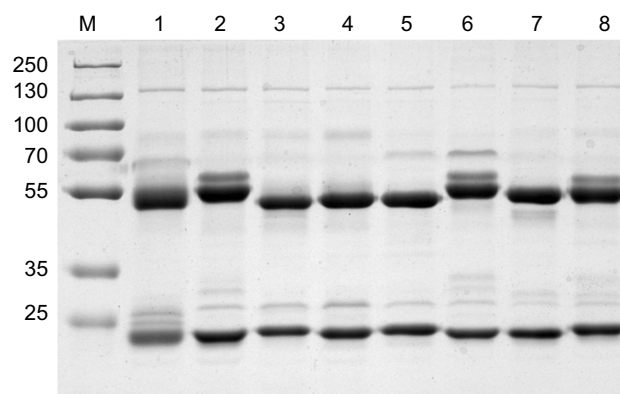
Note: RD=(Test values after storage - Original values)/Original values×100%; All of the RD values were less than ±15%, indicated that the kit has good stability.

Abbreviations: CPP, copeptin; RD, relative recovery deviation.

Table S4 Cross-reactivity of CLIA to related compounds

| Analyte | Added concentration | RLU×1000 | Cross-reactivity |
|-----------|---------------------------|----------|------------------|
| CPP | 100 pmol L ⁻¹ | 227.81 | 100% |
| BNP | 1000 pmol L ⁻¹ | 0.2 | 0.09% |
| NT-proBNP | 1000 pmol L ⁻¹ | 0.33 | 0.14% |
| cTnT | 0.3 ng mL ⁻¹ | 0.46 | 0.2% |
| Myoglobin | 200 ng mL ⁻¹ | 0.12 | 0.05% |
| CK-MB | 30 ng mL ⁻¹ | 0.14 | 0.06% |

Abbreviations: CLIA, chemiluminescence immunoassay; CPP, copeptin; BNP, B-type natriuretic peptides; NT-proBNP, N-terminal pronatriuretic peptides; cTnT, troponin T; CK-MB, creatine kinase-myocardial band.

**Figure S1** Purification results of the mAb against CPP by SDS-PAGE.

Notes: Lane 1: 6-20G5; lane 2: 6-8D2; lane 3: 6-18H3; lane 4: 6-5G8; lane 5: 6-2A9; lane 6: 6-3C10; lane 7: 6-18G10; lane 8: 6-11C3. There were only two straps after the mAb was purified, and these were heavy chain and light chain, respectively, CV. The results indicate that the purity of all mAbs was above 90% as analyzed on SDS-PAGE.

Abbreviations: mAb, monoclonal antibody; CV, coefficient of variation.

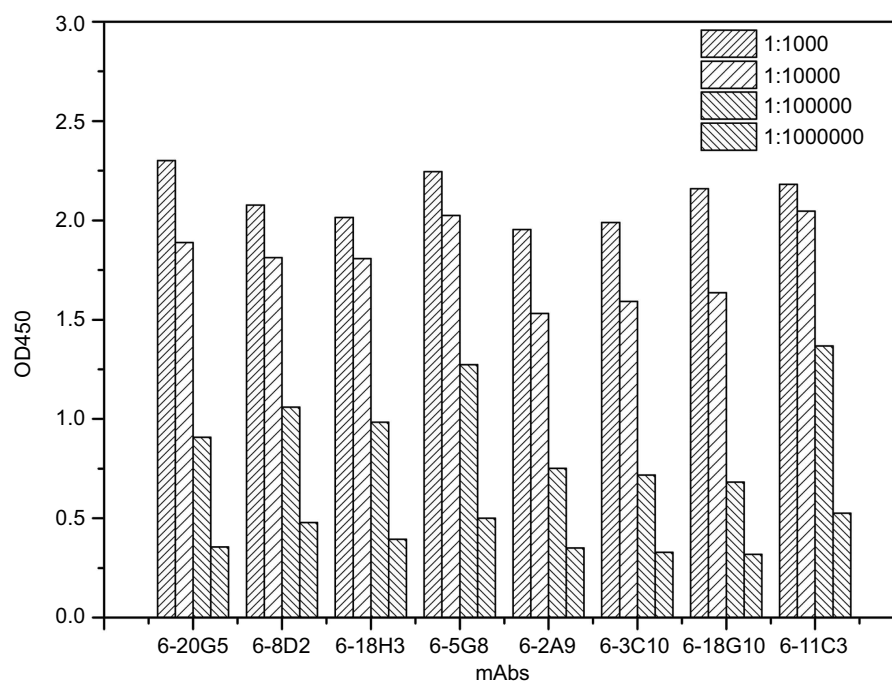


Figure S2 The titer of the mAbs against CPP.

Abbreviation: mAbs, monoclonal antibodies.

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