

Comparative study of in vitro effects of different nanoparticles at non-cytotoxic concentration on the adherens junction of human vascular endothelial cells

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Background: Effects of different nanoparticles (NPs) exposure at acutely non-cytotoxic concentrations are particularly worthy to figure out, compare, and elucidate.

Objective: To investigate and compare the effect of a small library of NPs at non-cytotoxic concentration on the adherens junction of human umbilical vein endothelial cells (HUVECs), obtaining new insights of NPs safety evaluation.

Materials and methods: The HUVECs layer was exposed to NPs including gold (Au), platinum (Pt), silica (SiO₂), titanium dioxide (TiO₂), ferric oxide (Fe₂O₃), oxidized multi-walled carbon nanotubes, with different surface chemistry and size distribution. Cellular uptake of NPs was observed by transmission electron microscopy. and the cytotoxicity was determined by Cell Counting Kit-8 assay. The NP-induced variation of intracellular reactive oxygen species (ROS) and catalase (CAT) activity was measured using the probe of 2',7'-dichlorodihydro fluorescein diacetate and a CAT analysis kit, respectively. The level of VE-cadherin of HUVECs was analyzed by Western blot, and the loss of adherens junction was observed with laser confocal microscopy.

Results: The acutely non-cytotoxic concentrations of different NPs were determined and applied to HUVECs. The NPs increased the level of intracellular ROS and the activity of CAT to different degrees, depending on the characteristics. At the same time, the HUVECs lost their adherens junction protein VE-cadherin and gaps were formed between the cells. The NP-induced oxidative stress and gap formation could be rescued by the supplementary N-acetylcysteine in the incubation.

Conclusion: The increase of intracellular ROS and CAT activity was one common effect of NPs, even at the non-cytotoxic concentration, and the degree was dependent on the composition, surface chemistry, and size distribution of the NP. The effect led to the gap formation between the cells, while could be rescued by the antioxidant. Therefore, the variation of adherens junction between endothelial cells was suggested to evaluate for NPs when used as therapeutics and diagnostics.

Keywords: reactive oxygen species (ROS), catalase, HUVEC, VE-cadherin

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Introduction

Engineered nanoparticles (NPs) have been attracting growing research and development interests as well as serious concerns for how to find out and properly evaluate their impacts on the biological system due to their unique physicochemical characteristics and uncertainty of biological effects.¹ In particular, NPs that are

intended for uses in therapy and diagnosis are likely to enter into the blood circulating system by various exposure ways. On the wall of blood vessels, there is an endothelial layer composed of endothelial cells connecting each other, which is the most important biological barrier between blood and tissues to maintain the vascular function and of homeostasis.²⁻⁴ The injury of the endothelial layer may result in acute inflammation,^{5,6} leading to various pathophysiological conditions, such as atherosclerosis, thrombosis, and myocardial infarction.^{7,8}

Interactions between NPs and endothelial cells have been investigated in previous reports. The multiplicity of NPs includes gold (Au), platinum (Pt), silica (SiO₂), titanium dioxide (TiO₂), iron(III) oxide (Fe₂O₃), iron(II,III) oxide (Fe₃O₄), Zinc oxide (ZnO), copper (II) oxide (CuO), magnesium oxide (MgO), aluminum oxide (Al₂O₃), and multi-walled carbon nanotubes (MWCNTs).⁹⁻¹⁹ The exposure dosage, size distribution, and surface modification of NPs employed in the previous literatures were largely various, even for the same kind of NPs, which lead to hard comparisons among their biological effects. Take Au NPs as an example, the size range is from 10 to 50 nm, and the concentration range employed with vascular endothelial cells was from 100 nM to 100 μ M in different labs.^{9,20} It has been recognized that in addition to inducing cells death, NPs are able to affect the integrity of vascular endothelial layer, for examples, titanium dioxide (23.5 nm, at 10 μ M) was demonstrated to cause endothelial cell leakiness through physical interactions.¹⁴ Similarly, effects of six kinds of metal oxide NPs on human cardiac microvascular endothelial cells are reported, the exposed concentration of the NPs ranges from 0.001 to 100 μ g/mL, and average diameter is about 40 nm. Among them, NPs of ZnO, CuO, and MgO increase lactate dehydrogenase of vascular endothelial cells, while NPs of Fe₂O₃, Fe₃O₄, and Al₂O₃ have no significant effects on the permeability.¹⁸ In addition, NPs of Fe₂O₃ (50 μ g/mL, with a mean diameter of 298 nm),¹⁷ MWCNTs (3.86 μ m long, 2.5 μ g/mL),¹¹ and silver NPs (110 nm, 1 μ g/mL)²¹ affect VE-cadherin expression by increasing intracellular reactive oxygen species (ROS). However, at acutely non-cytotoxic concentration, effects of NPs on endothelial cells have little touched yet, which are particularly worthy to note, because there may be some unseen biological effects.

In this study, we selected six kinds of NPs, including Au, Pt, SiO₂, TiO₂, Fe₂O₃, and MWCNTs, with different size, surface chemistry, and composition to set up a small library. By screening the cytotoxicity of each kind of the

NPs, we had endothelial cells exposed to the different NPs at acutely non-cytotoxic concentrations, investigating the effects of the NPs on the adherent conjunction of vascular endothelial cells and underlying reasons.

Materials and methods

Materials

Dextran-Fe₂O₃, DMSA-Fe₂O₃ NPs were provided by the Nanjing Nanoeast Biotech Co., Ltd. TiO₂ NPs (25, 30, 80, 100 nm) were provided by Prof. Lingyu Piao, National Center for Nanoscience and Technology, China. SiO₂ (20, 50, 100 nm) and platinum NPs (5, 30, 70 nm) were purchased from the nanocomposix company. The product of original multi-walled carbon nanotube (50 μ m) was purchased from Chengdu Organic Chemicals Co., Ltd., Chinese Academy of Sciences. Oxidized multi-walled carbon nanotubes and gold NPs (10, 30, 50 nm) were synthesized according to protocols of references.²²⁻²⁴ Milli-Q water (18 M Ω cm) was used for the preparation of all solutions.

Size and charge

Dynamic light scattering analysis (Nano ZS90, Malvern Instruments Ltd, UK) was used to measure the hydrodynamic diameter and Zeta potential of particles. In short, the solutions of NPs were dispersed in 1 mL Milli-Q water or serum containing medium. Each measurement was repeated three times.

Cell culture

Primary human umbilical vein endothelial cells (HUVECs), endothelial cell medium, endothelial cell growth supplements, and Poly-L-Lysine were all purchased from ScienCell Research Laboratories (San Diego, CA). Cells were grown on a cell culture plates coated with 1 μ g cm⁻² poly-L-lysine, and cultured at 37°C in a wet incubator with 5% CO₂.

Cell viability assay

HUVECs were seeded into 96-well cell culture plates at a density of 1.5×10^4 cells/well and cultured 24 hrs for adherence. The cells were incubated with 100 μ L NPs at 37°C for 24 hrs, followed by washing with PBS and incubated with fresh medium containing 10 μ L Cell Counting Kit-8 (CCK-8, Dojindo Molecular Technologies, Inc) for 2 hrs. The absorbance of the medium was measured at 450 nm using the Synergy H1 hybrid

multifunction microplate reader (BioTek Instruments, Winooski, VT, USA). The measurements were carried out in three parallel lines, and the relative cell viability was expressed as a percentage of the control.

Cell uptake assay through transmission electron microscopy (TEM)

HUVECs were seeded at a density of 1.5×10^6 cells/dish in a 100 mm dish and incubated overnight for cell attachment. The cells were treated with NPs for 24 hrs. Then, cells were washed twice with PBS, scraped gently from the dishes and centrifuged, then fixed with 2.5% glutaraldehyde (Fluka) and dehydrated gradually by alcohol and embedded in epon. Sections were then cut and placed on copper grids for observation (TEM-1400 plus, JEOL Ltd., Tokyo, Japan). In the process, NPs were dyed with Uranium acetate and lead citrate dyeing except Dex- Fe_2O_3 .

Intracellular ROS measurement

HUVECs were seeded at a density of 1×10^5 cells/well in the 24-well cell culture plate and kept overnight to adhere. For the rescue experiment, the cells were treated with 3 mM N-acetylcysteine (NAC; Sigma-Aldrich, USA) for 1 hr before exposure to NPs. The cells were treated with NPs for 1 hr at 37°C and then washed with PBS. After that, PBS containing 10 μM DCFH-DA (Sigma-Aldrich, USA) was added at 37°C for 30 mins and then washed with PBS twice. Finally, the cells were detached from the plates with 100 μL well⁻¹ trypsin, and collected by centrifugation. Flow cytometric analysis was conducted on the various cell treatment groups, using an excitation wavelength of 488 nm and emission wavelength of 525 nm (AccuriTM C6 flow cytometer, BD Biosciences, San Jose, CA). All groups had three replicate wells.

Catalase (CAT) activity

HUVECs were grown at a density of 7×10^5 cells well⁻¹ on six well plates overnight and exposed to NPs for 1 hr at 37°C . Subsequently, cells were washed twice with PBS and scraped using 100 μL of RIPA buffer (PMSF: RIPA=1:100). The samples were centrifuged at 2,500 rpm for 10 mins at 4°C . Then, cell extracts were collected for measurement of CAT activity using a CAT analysis kit (Nanjing Jiancheng Biology Engineering Institute, China). Then, the absorbance of the medium was measured at 405 nm using the Synergy H1 hybrid multifunction microplate reader. All groups had three replicate wells.

Immunostaining and laser confocal microscopy assay

HUVECs were seeded at a density of 1×10^5 cells/well in 24-well cell culture plates (pre-loaded with cell climbing glass slices) and incubated overnight for adhesion. To carry out the rescue experiment, the cells were treated with 3 mM NAC for 1 hr before exposure to the NPs. The cells were treated with medium containing NPs for 1 hr and fixed with 4% polyformaldehyde at 37°C for 30 mins, permeabilized using 0.3% TritonX-100 for 15 mins. After rinsing with PBS and incubated with 2% BSA for 1 hr, the cells were labeled with VE-cadherin XPTM Rabbit antibody (1:200 dilution, Cell Signaling Technology, USA) in 2% BSA/PBS overnight at 4°C followed by washed with PBS and incubated with Goat anti-rabbit antibody labeled with Alexa Fluor488 (1:400 dilution, Cell Signaling Technology) and mounted with an aqueous mounting medium containing DAPI (Zhongshan Goldenbridge biotechnology Co., China). The microphotographs were taken by using the FluoView FV1000 confocal microscope (Olympus). Each group was repeated three times.

Western blot

HUVECs were seeded at a density of 6×10^5 cells/well in six-well cell culture plates and incubated overnight and then incubated with the medium containing NPs for 1 hr. After washed with PBS and scraped using RIPA buffer (PMSF: RIPA=1:100), the cells were centrifuged at 12,000 rpm for 15 mins at 4°C , and the supernatants were collected. Protein concentrations were determined using PierceTM BCA Protein Assay Kit (Thermo Fisher Scientific, USA). Proteins (30 μg lane⁻¹) were resolved by means of SDS-PAGE and electrotransferred onto PVDF membranes (Millipore, Bedford, Massachusetts). The membranes were blocked using TBST containing 5% skim milk, and then incubated overnight at 4°C with primary antibodies (VE-cadherin XPTM Rabbit antibody, Cell Signaling Technology). The membranes were next incubated for 2 hrs at room temperature with horseradish peroxidase conjugated secondary IgGs (1:10,000). Finally, the immunoreactive bands were visualized with the Immobilon[®] Western HRP Substrate luminal reagent and peroxide solution (Millipore Corporation, Billerica).

Measurement of endothelial permeability

HUVECs were seeded in 24-well cell culture plates containing cell climbing glass slices that had been pre-loaded

with collagen and incubated for two days. The cells were treated with NPs for 1 hr at 37°C, which was then washed before being immersed in 18 μ M 40 kDa FITC-dextran (Sigma-Aldrich) for 5 mins. Thereafter, follow the procedure in laser confocal microscopy assay. Confocal Y and Z-slice images were taken by a confocal microscope. For data analysis, a rectangle box of about 510×70 pixels in each cell layer was calculated as the boundary with ImageJ. The upper rectangle frame represented the average pixel intensity of 18 μ M FITC-dextran strength. A second box of width 510 pixels was drawn below the cell layer, containing the whole height of collagen, which represents the average pixel intensity of FITC-dextran that has entered the collagen. The accumulation of dextran was calculated by dividing the average pixel intensity within the collagen by that above the collagen.

Statistical analysis

Statistical significance was ascertained through Student's two-tailed *t*-test with SPSS software (SPSS17.0). **P*<0.05, ***P*<0.01, respectively.

Results

Characterization of NPs

The characteristics of employed NPs, including composition, particle size, and coating are summarized in Table 1. Images from the transmission electron microscope (TEM) showed morphologies of the NPs dispersed in aqueous solution (Figure 1). Most of the selected particles are spherical except for MWCNTs having a tubular structure. For the particles with the same composition, the smaller one is likely to aggregate in aqueous solution. From the TEM images of HUVECs incubated with NPs for 24 hrs, most of the NPs were included in the vesicles and especially in the lysosomes

structures. While, Dex-Fe₂O₃ NPs were more located in the junction area of the cells (Figure 2).

Cytotoxicity of NPs on HUVECs

In order to determine the acutely non-cytotoxic concentration for each kind of the NPs, the viability of HUVECs incubated with the NPs for 24 hrs was measured (Figure 3). Different concentration ranges of NPs were employed firstly to obtain the acutely non-cytotoxic concentration for each NP. The cell viability of control group was set as a 100%. In general, the effect of NPs on the cell viability was closely associated with the NPs composition (Au, Pt, MWCNT), coating (Dex-/DMSA-Fe₂O₃), and size (TiO₂, SiO₂). Au NPs did not affect the viability of HUVEC in the range of testing concentration, possibly due to the sodium citrate on the surface,^{25,26} while Pt NPs modified with the same sodium citrate reduced the viability of HUVECs in a concentration-dependent manner, which was attributed to the chemical property of Pt. Pt-5 showed greater cytotoxicity than Pt-30 and Pt-70 due to the larger specific surface area of the smaller particles. Similar to Pt NPs, TiO₂ NPs affected the viability of HUVEC. Among the four TiO₂ NPs, TiO₂-25 was more cytotoxic than the pure anatase type in a dose-dependent manner. The cytotoxicity of SiO₂-100 was weaker than that of SiO₂-20 and SiO₂-50, while SiO₂-50 was more toxic than SiO₂-20, this is possibly due to the aggregation of SiO₂-20, evidenced by that the average dynamic diameter of SiO₂-20 in the culture medium was 202.37±0.25 nm, which led to the decrease of specific surface area to a certain degree and lower cytotoxicity. It was also observed the cytotoxic effect of SiO₂-20 and SiO₂-50 was associated with the exposure concentration. The two types of magnetic iron oxide particles displayed different cytotoxicity. The Dex-Fe₂O₃ NPs did not affect the cell viability until the content of Fe was higher than 240 μ g mL⁻¹, however, the DMSA-Fe₂O₃ NPs showed significant cytotoxic and statistical

Table 1 The small library of NPs used in this work

Category	Composition	Coating	Size (nm)	Abbreviation
Metal	Au Pt	Sodium citrate Sodium citrate	10/30/50 ^a 5/30/70 ^b	Au-10/30/50 Pt-5/30/70
Oxide	SiO ₂ TiO ₂ Fe ₂ O ₃	None None Dex DMSA	20/50/100 ^b 25/30/80/100 ^c 30.15 ^d 45.3±11.2 ^d	SiO ₂ -20/50/100 TiO ₂ -25/30/80/100 Dex-Fe ₂ O ₃ DMSA-Fe ₂ O ₃
Carbonaceous	Carbon	None	128.5±1.3 ^a	MWCNTs

Note: ^aMeasured from DLS measurement, ^bindicated on the label of products, ^cmeasured from TEM images, ^dcame from the reference.

Abbreviations: Dex, polydextrose sorbitol carboxymethyl ether; DMSA, 2, 3-dimercaptosuccinic acid; DLS, Dynamic light scattering.

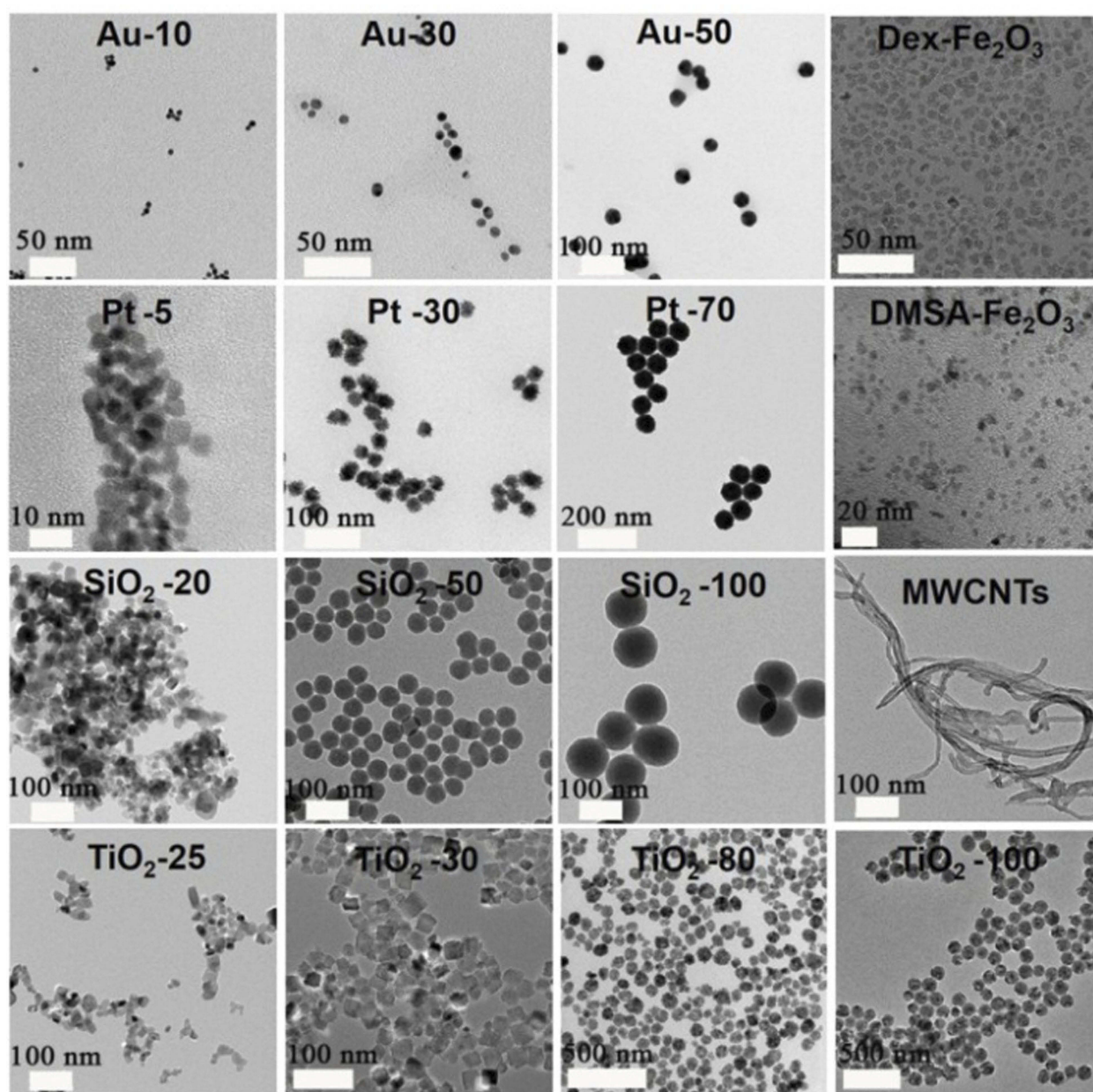


Figure 1 TEM images of various NPs employed in this work.

Abbreviations: Au, gold; Pt, platinum; SiO₂, silica; TiO₂, titanium dioxide; Fe₂O₃, iron(III) oxide; MWCNTs, multi-walled carbon nanotubes; Dex, polydextrose sorbitol carboxymethyl ether; DMSA, 2, 3-dimercaptosuccinic acid.

difference among the different concentrations when the content of Fe was higher than 30 $\mu\text{g mL}^{-1}$. As for the MWCNTs, the cell viability was less affected when the concentration was not higher than 120 $\mu\text{g mL}^{-1}$. Hence, we selected an acutely non-cytotoxic concentration for each kind of the NPs, under which the cell viability was no less than 80% (rectangle in Figure 3).

Oxidative stress induced by NPs

As shown in Figure 4A, comparing with the untreated group, 1 hr of NPs exposure increased the intracellular ROS level,

including Pt NPs, SiO₂ NPs, Fe₂O₃ NPs, and MWCNTs. However, it was not detectable ROS variation for cells incubated with TiO₂ NPs or Au NPs. It is well documented that when the extra ROS was produced, antioxidants in the cells would be increased to maintain the redox balance.²⁷ To provide a side evidence of the other NPs increasing intracellular ROS, we examined the intracellular level of CAT because the increase of CAT is recognized as one crucial marker of ROS upregulation.^{28–30} Here, we examined the CAT variation in the cells incubated with the largest size of Au NPs and TiO₂ NPs at the non-cytotoxic concentration reference to the untreated

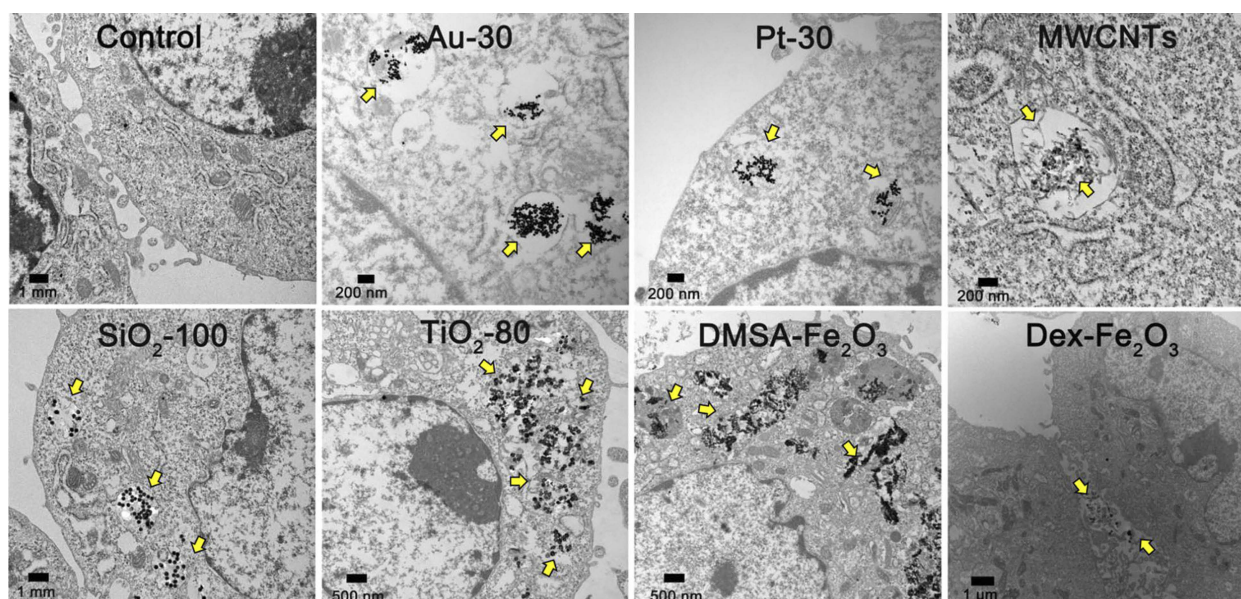


Figure 2 TEM images of cell uptake of various NPs. The NPs were distributed in/beside the cells as the arrow indicated.

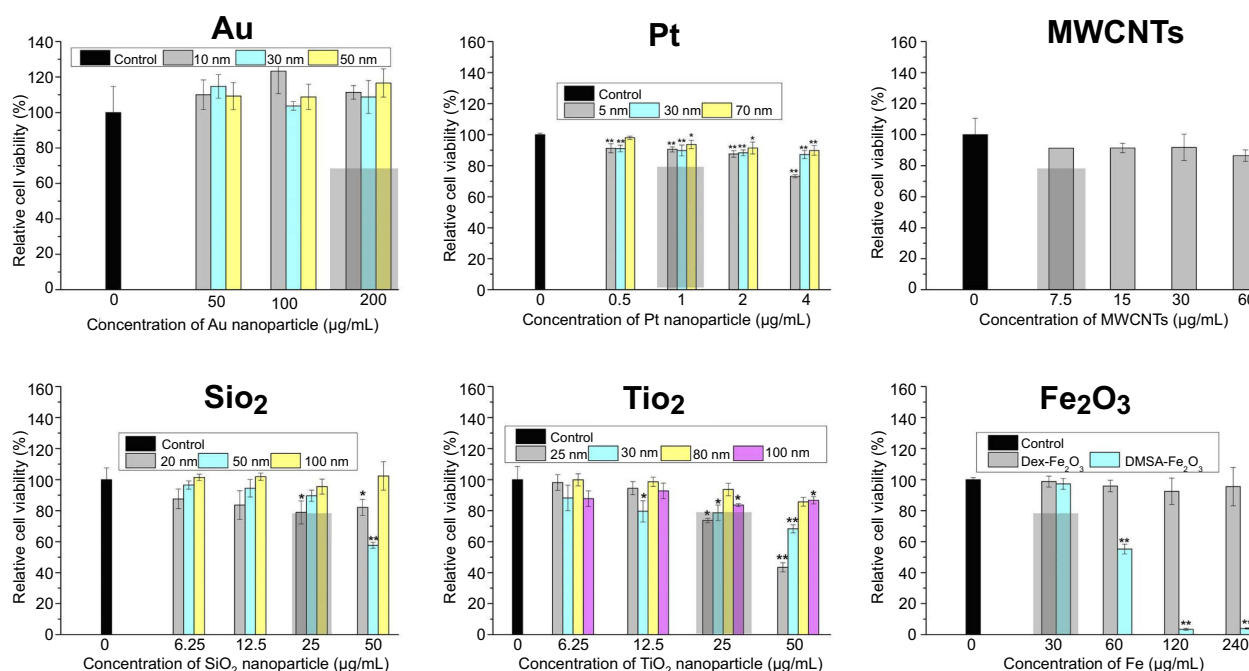


Figure 3 Viability of HUVECs exposed with NPs from CCK-8 assay. The * indicates significant differences between the control group and the nanoparticles treatment group (*: $P < 0.05$, **: $P < 0.01$).

group, at the same time, the effect of the largest size of Pt NPs and SiO₂ NPs, and the other NPs on the CAT variation were examined as well. Results showed that each kind of NPs induced upregulation of CAT, the variation degree was associated with the chemistry of the NPs (Figure 4B). Taken above together, all the NPs were capable of inducing oxidative stress in the HUVECs to different degrees, and the Au NPs induced the weakest oxidative stress.

Effects of NPs on adherens junction of HUVECs

The upregulation of intracellular ROS level implied possible changes in endothelial cells functions.⁴ One of the most important proteins for vascular endothelial cells is VE-cadherin that is essential for the adhesion junction between HUVECs, reflecting the integrity of vascular endothelial

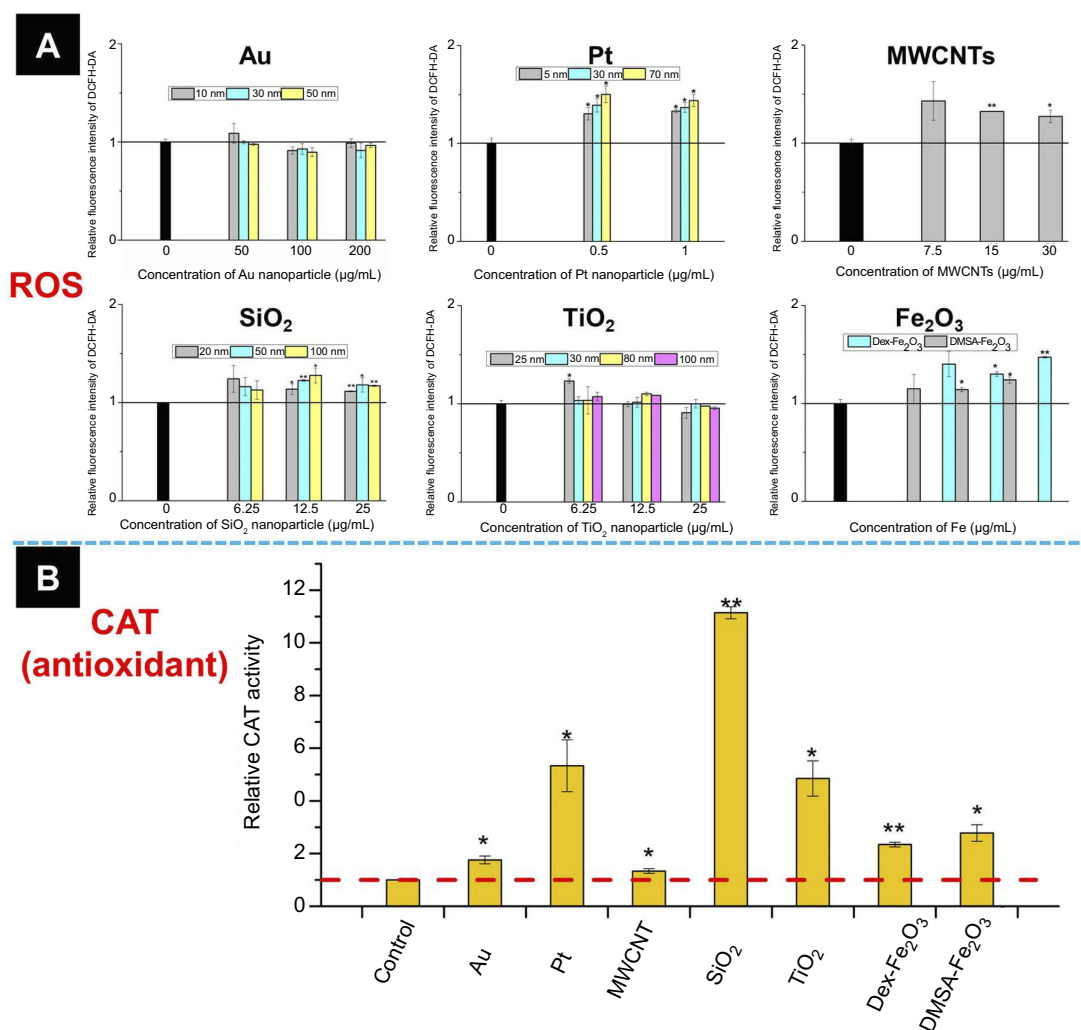


Figure 4 (A) The intracellular ROS level and **(B)** catalase activity of HUVECs incubated with various NPs. The concentration of NPs in **(B)** was listed below: Au (50 nm): 200 $\mu\text{g mL}^{-1}$, Pt (70 nm): 1 $\mu\text{g mL}^{-1}$, MWCNTs: 7.5 $\mu\text{g mL}^{-1}$, SiO₂ (100 nm): 25 $\mu\text{g mL}^{-1}$, TiO₂ (100 nm): 25 $\mu\text{g mL}^{-1}$, Dex-Fe₂O₃: 30 $\mu\text{g Fe mL}^{-1}$, and DMSA-Fe₂O₃: 30 $\mu\text{g Fe mL}^{-1}$. *, $P < 0.05$, **, $P < 0.01$ compared with the control group.

Abbreviation: ROS, reactive oxygen species.

cells.^{31,32} In the normal cultivation (control group), HUVECs exhibited cobblestone phenotype, and the intercellular junction between the cells was clear and side-by-side with laser confocal microscopy observation (Figure S1). After 1 hr of incubation with the NPs, the VE-cadherin on the membrane became blur (Figure 5A), and at the same time, the expression of VE-cadherin in the whole cell was decreased in reference to that in the control (Figure 5B), both of which indicated that the adherens junction was not so tightly together between cells. Images from the confocal observation for cells incubated with the NPs with various sizes and coatings were given in Figure S2. We further measured the gap-width of intercellular junctions, and the measurement approach was illustrated in Figure S3 according to the reference.³³ Strikingly, after incubated with NPs at their non-

cytotoxic concentrations for 1 hr, intercellular gaps were detected in all the groups except for cells incubation with Au NPs (Figure 5C), though no statistical difference was shown among the size, coating, and component. The permeability of endothelial monolayer pre-treated with different NPs was also evaluated using the assay of 40-kD FITC-dextran movement across the monolayer,¹⁴ results were consistent with the statistical result of gap formation (Figure S4).

Rescue on adherens junction of HUVECs

We next investigated whether the NPs-induced gap formation in HUVECs could be rescued by antioxidant NAC. It was seen that the pre-incubation with NAC has no effect on the control group. However, in the NPs treated group, pre-incubation with NAC could decrease the level of NPs

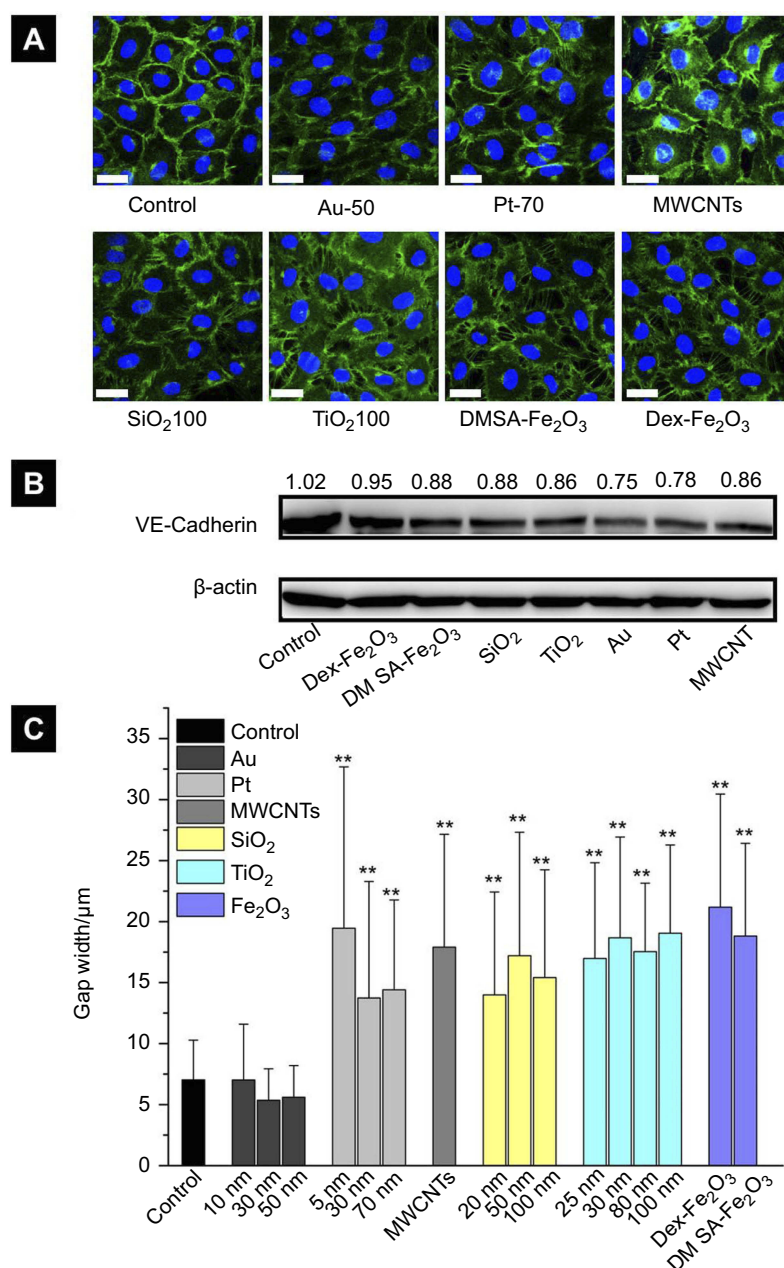


Figure 5 (A) Effects of NPs on the adherens junction of HUVECs were observed from confocal microscope. Scale bar is 20 μm . (B) Western-blot result of VE-cadherin protein expression by endothelial cells incubated with NPs. The concentration of NPs was listed below: Dex-Fe₂O₃: 120 $\mu\text{g Fe mL}^{-1}$, DMSA-Fe₂O₃: 60 $\mu\text{g Fe mL}^{-1}$, SiO₂ (100 nm): 25 $\mu\text{g mL}^{-1}$, TiO₂ (25 nm): 25 $\mu\text{g mL}^{-1}$, Pt (70 nm): 1 $\mu\text{g mL}^{-1}$, Au (50 nm): 200 $\mu\text{g mL}^{-1}$, and MWCNTs: 7.5 $\mu\text{g mL}^{-1}$. (C) Gap widths between cells treated by NPs which calculated from 10 groups. **: $P < 0.01$ compared with the control group.

Abbreviation: HUVECs, human umbilical vein endothelial cells.

mediated ROS production effectively, and the gaps between the cells were reduced as well (Figure 6), which strongly suggested that the NP-induced excess ROS in the HUVECs led to the reduction of cell-cell adherens junction and, lastly, induced gap formation and resulted in the permeability of the endothelial layer. At the same time, these results suggested that the NPs induced gap formation could be rescued by the treatment of antioxidant.

Discussion

Due to their specific and unique nanoscale physicochemical properties, NPs have emerged as powerful tools for a wide scope of applications such as in nanomedicine. For example, plasmonic NPs, including Au and Pt, were employed in sensing, drug delivery, photoacoustic imaging, photothermal therapy, and photodynamic therapy.³⁴

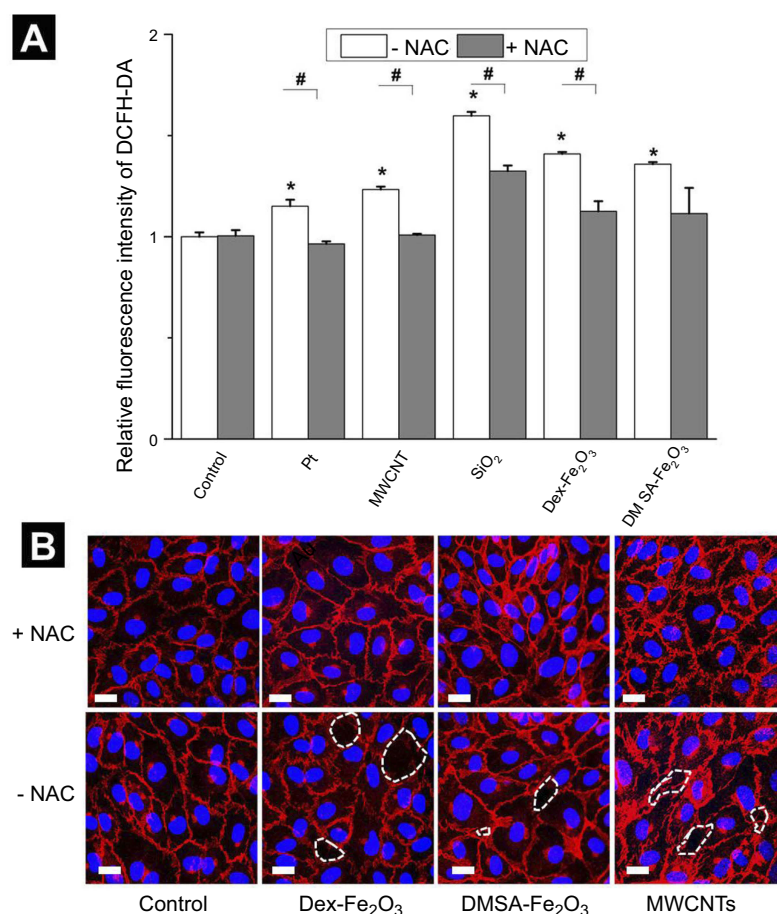


Figure 6 (A) The intracellular ROS level caused by NPs in the absent and present of NAC. The * represents a significant difference comparing with the control group (*: $P < 0.05$), and the # indicates a significant difference between the NAC group and the without NAC pre-treatment group (#: $P < 0.05$). **(B)** Confocal images in the absent and present of 3 mM NAC. The scale bar is 20 μm for all. The concentration of NPs was following: Pt (70 nm): 1 $\mu\text{g mL}^{-1}$, MWCNTs: 7.5 $\mu\text{g mL}^{-1}$, SiO₂ (100 nm): 25 $\mu\text{g mL}^{-1}$, Dex-Fe₂O₃: 30 $\mu\text{g Fe mL}^{-1}$, and DMSA-Fe₂O₃: 30 $\mu\text{g Fe mL}^{-1}$.

Abbreviation: NAC, N-acetylcysteine.

MWCNT, with the characters of lightweight, extreme strength and tensile forces, electronic conductivity, and strong capillary forces, have a broad application in supercapacitors, automotive industry, aerospace industry, pharmaceuticals, and medical devices.^{11,35} With large surface area, high structural stability, easy surface functionalization, low cost of production, and excellent biocompatibility, metal oxide NPs (such as SiO₂, TiO₂, and Fe₂O₃) were researched and used in drug, protein or gene delivery, cosmetics doping, and imaging enhancements.^{18,34,36} Therefore, it is necessary to systematically and comparatively evaluate the effects of above typical NPs in biological systems.

As mentioned in the introduction, major previous studies were performed upon vascular endothelial cells at cytotoxic concentrations.^{9,12,15,37} In our opinions, effects of NPs at acutely non-cytotoxic concentration on

endothelial cells are particularly worthy to note, because there may be some unseen biological effects. Therefore, non-acute cytotoxic concentrations were set for each kind of NPs employed in this study to assure the relative viability of cells was higher than 80% (comparing with the control group), which is considered as non-toxic or slight cytotoxic and clinically acceptable according to the guidance of ISO for biological evaluation of medical devices.³⁸

Our results suggested that even at a concentration of not inducing cell death significantly, the NPs were able to affect the expression of VE-cadherin between vascular endothelial cells by inducing oxidative stress, though this effect is reported in previous literatures with cytotoxic exposure. The downregulation of VE-cadherin induced by the NPs led the gap formation between the HUVECs, might lead a dysfunction of ECs.¹⁹ Nevertheless, it could

be noticed that the decrease of VE-cadherin could be rescued by employing the antioxidant, which is in consistency with the results reported previously.^{39,40} Furthermore, the decrease of VE-cadherin and the gap formation between the vascular endothelial cells exerted by the NPs were associated with the morphology, size, and coatings of NP, which was ubiquitous, but there was a matter of degree.

HUVECs that cover the lumen of blood vessels have been considered as a general model for access the toxicity of NPs.⁴¹ There are two prevailing paradigms on how NPs traverse across endothelial barriers.⁴² One is a transcellular process with energetically costly and slow, the other is a paracellular transport process with crossing cell barriers by diffusion between cells through the cell–cell junctions. In drug-based therapies, the alteration of permeability barrier integrity plays a major role when drugs either small biomolecules or NPs employ the paracellular transport process, the leakiness may provide an easier journey for the NPs. Thus, this tuneable influence with size and/or coating related to adherens junction of endothelial cells might contribute to the regulation on enhanced permeability and retention effect in drug delivery system.

In this work, we proposed the generality and specificity of various NPs to the endothelial cells, which provided a new insight into the safety evaluation of NPs in applications, especially would be benefit to the establishment of standard assays in the future to predict or evaluate the NP-mediated potential risks to vascular endothelial cells as well as monolayers by measuring oxidative stress (ROS or/and CAT detection) simply, and those effects may not be detected by using conventional cytotoxicity assays. Nevertheless, gene-related studies need to be further conducted in the future. So far there are only a few of studies from gene level for NPs upon endothelial cells have been reported.^{43–46} For example, Ag NPs induce excess ROS production of HUVECs by activation of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase activity through upregulation of NADPH oxidase 4 (NOX4) gene,⁴³ increase the permeability of rat brain microvascular endothelial cells by inhibiting the antioxidant defence of the astrocytes that inhibits the Trx system, and decreasing Nr4a1 and Dusp1 gene.⁴⁵ Hydroxyapatite NPs down-regulated VE-cadherin and CD31 gene of ECs.⁴⁴

Conclusion

In summary, we employed a small library of NPs to show that the exposure of NPs to HUVECs at the concentration of low cytotoxicity induced the upregulation of intracellular ROS

and increased the antioxidant (CAT) activity at the same time, which led to the downregulation of VE-cadherin. These effects resulted in the formation of gaps between the cells and induced endothelial leakiness. Thus, even if the NPs applied at non-cytotoxic concentration, NP-mediated ROS should be prevented to reduce the negative effects on the integrity of endothelial barriers and risk of vascular diseases.

Acknowledgments

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Author contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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Supplementary materials

Experimental Section

Synthesis of Gold Nanoparticles

1. Small size-10 nm: 150 mL of reduced solution of sodium citrate (SC, 2.2 mM, Sinopharm Chemical Reagent Co., Ltd) containing 0.1 mL tannic acid (TA, 2.5 mM, Aladdin, Shanghai) and 1 mL potassium carbonate (K_2CO_3 , 150 mM, Aladdin, Shanghai) was heated (temperature set at 70°C) in a 250 mL three-necked bottom flask under severe stirring. One milliliter tetra chlorauric acid ($HAuCl_4$, 25 mM, Sinopharm Chemical Reagent Co., Ltd) was slowly injected when the temperature was reached. The color of the solution quickly turns into black gray (less than 10 s) and then turns into orange red subsequently. The solution was kept at 70°C for 10 mins to guarantee complete reaction of the gold nanoparticle.¹
2. Large size-30 nm: 150 mL of the reduced solution of sodium citrate (SC, 2.2 mM) was heated (temperature set on 100°C) in a heating mantle in a 250 mL three-necked bottom flask under severe stirring. When the temperature reached 96°C, 1 mL of tetra chlorauric acid ($HAuCl_4$, 25 mM) was injected slowly. The color of the solution quickly turns into black gray (less than 10 s) and then turns into orange red subsequently. The solution was kept at 100°C for 20 mins to ensure complete reaction of the gold precursor. After 20 mins, the reaction was cooled down until the temperature of the solution reached 90°C. Then, 1 mL of sodium citrate (60 mM) and 1 mL of $HAuCl_4$ solution (25 mM) were sequentially injected (time delay about 2 mins). 30 mins later, repeated once, then turn off the heat and stir for another 20 mins.²
3. Large size-50 nm: Gold nanoparticles were prepared just like 30 nm. Repeat subsequent steps four times, then turn off the heat and stir for another 20 mins.

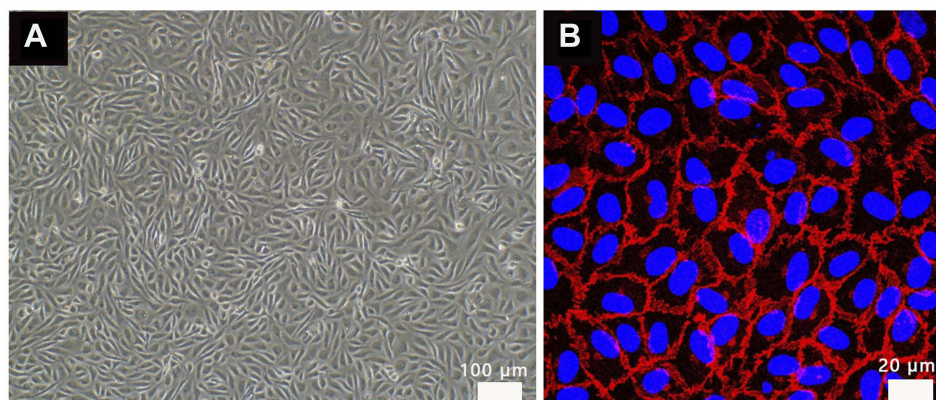


Figure S1 Images of HUVEC in endothelial cell medium from (A) optical microscope, HUVECs were paved stone-like growth and (B) confocal labeled with VE-cadherin antibody (red) and DAPI (blue). Normal umbilical vein endothelial cells connected closely.

Abbreviation: HUVECs, human umbilical vein endothelial cells.

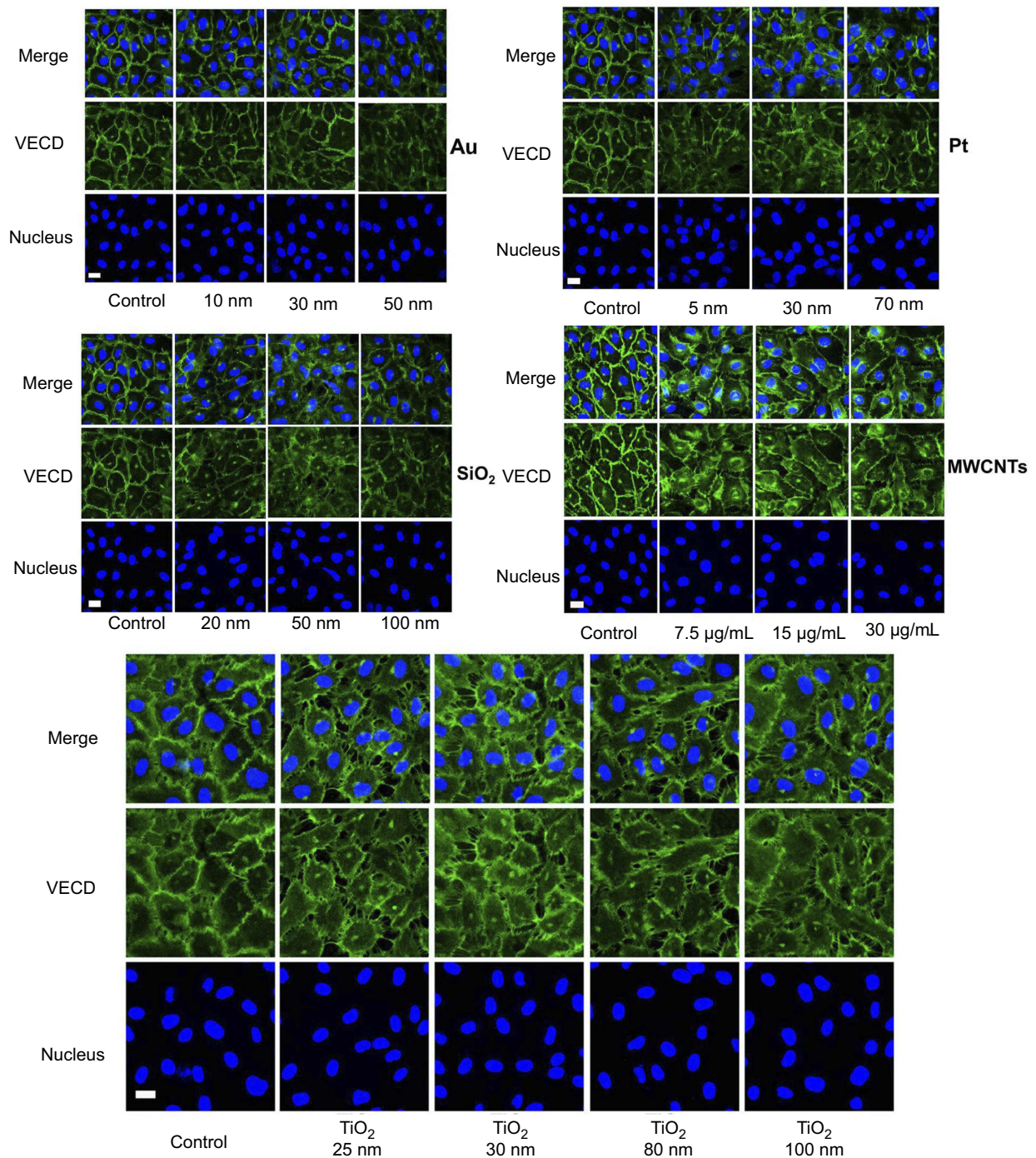


Figure S2 Effects on the adherens junction of HUVEC with various nanoparticles of different size, coating, and concentration. Images are taken from confocal labeled with VE-cadherin antibody (green) and cell nucleus (DAPI, blue). The concentration of various nanoparticles (Au: 200 µg mL⁻¹, Pt: 1 µg mL⁻¹, SiO₂: 25 µg mL⁻¹, and TiO₂: 25 µg mL⁻¹). **Abbreviations:** VECD, VE-cadherin; DAPI, 4',6-diamidino-2-phenylindole.

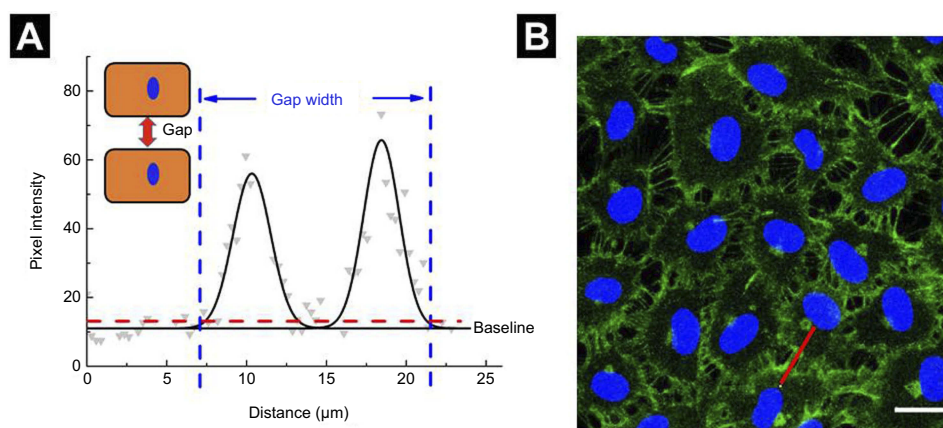


Figure S3 Measurement of gap-width. (A) Gray dots represent the pixel intensity profile of the red line drawn in (B), whereas the black line represents a two-Gaussian fit. The junction width was considered as the width of the fitted curve at 20% above baseline intensity, as indicated by the dashed red line. (B) Images from a confocal microscope. Scale bar is 20 μm .

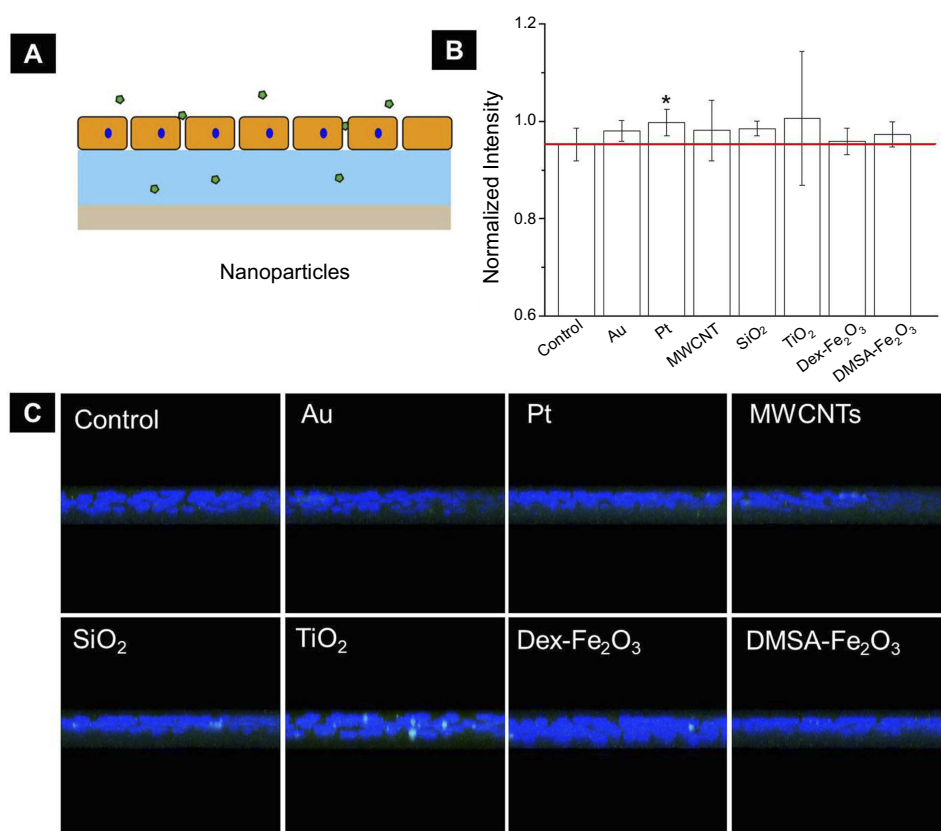


Figure S4 (A) The schematic diagram shows FITC-dextran permeating the collagen layer through a monolayer of endothelial cells. (B) Monolayer permeability was measured in samples that were untreated or treated with NPs for 1 hr. Data were shown as average mean $\pm$ SE from five groups of confocal images. (C) The representative YZ confocal cross-section images of the dextran captured in the 2 $\mu\text{g cm}^{-2}$ collagen, fluorescent images showing nuclei (blue), and FITC-dextran (green) of confluent HUVECs on collagen. The concentration of NPs was following: Au (50 nm): 200 $\mu\text{g mL}^{-1}$, Pt (70 nm): 1 $\mu\text{g mL}^{-1}$, MWCNTs: 7.5 $\mu\text{g mL}^{-1}$, SiO₂ (100 nm): 25 $\mu\text{g mL}^{-1}$, TiO₂ (80 nm): 25 $\mu\text{g mL}^{-1}$, Dex-Fe₂O₃: 30 $\mu\text{g Fe mL}^{-1}$, and DMSA-Fe₂O₃: 30 $\mu\text{g Fe mL}^{-1}$.

Abbreviation: FITC, fluorescein isothiocyanate.

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