

Chlorins e6 loaded silica nanoparticles coated with gastric cancer cell membrane for tumor specific photodynamic therapy of gastric cancer

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Background: Photodynamic therapy (PDT) is widely recognized as a promising way to cure cancer. However, the limited tumor homing property of currently available drug delivery systems (DDSs) is the bottleneck for the delivery of photodynamic agents.

Purpose: In our study, we decorated silica nanoparticles (SLN) with cell membrane (CM) derived from SGC7901 cells to construct carrier (CM/SLN) which was able to specifically target the homogenous SGC7901 cells.

Materials and methods: Furthermore, the decent drug loading capability of CM/SLN was adopted to load photodynamic agent chlorins e6 (Ce6) to finally construct a DDS suitable for tumor-targeted PDT of gastric cancer.

Results: The experimental results suggested that CM/SLN/Ce6 was nano-sized particles with good dispersion and stability in physiological conditions. Moreover, due to the modification of CM, CM/SLN/Ce6 could specifically target the homogenous SGC7901 cells both in vitro and in vivo. Most importantly, further in vivo results demonstrated that the CM/SLN/Ce6 showed a better anticancer outcome compared to SLN/Ce6.

Conclusion: CM/SLN/Ce6 might be a promising platform for effective tumor targeted PDT of gastric cancer.

Keywords: cell membrane, silica nanoparticles, chlorins e6, photodynamic therapy, gastric cancer

Introduction

In the past decades, the nanoparticle-based drug delivery systems (DDS) have widely applied in cancer therapy. Compared to free drugs, they showed elevated bioavailability and decreased side effects which holds great potential for clinical applications.¹⁻⁴ Herein, previous reports have constructed different kinds of nanoparticles based DDSs using materials from organic to inorganic ones,⁵⁻⁸ among which silica nanoparticles (SLN) with versatile merits is emerging as a potential candidate.^{9,10} Therefore, many DDSs have been developed using SLN as the skeleton that achieved beneficial outcomes both in vitro and in vivo.¹¹⁻¹³

However, the targeted delivery is still unsatisfied in many currently available DDSs which requires further improvement.¹⁴ To address this dilemma, the modification of DDSs with targeting ligands is one of the most adopted ways.^{6,15} In the past decades, targeting ligands varying from small molecules to monoclonal antibodies, have been successfully integrated into DDS to improve its tumor-targeting capabilities.¹⁶⁻¹⁸ However, the ectogenic nature of

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these ligands might induce some unexpected side effects, such as immunoreaction and cytotoxicity.

Photodynamic therapy (PDT) is a rapidly developing technique that employ photodynamic agents to transfer NIR light into highly ROSTo exert cytotoxic on cells.¹⁹ As a noninvasive and harmless approach, PDT is well recognized as a preferable measure for cancer therapy.²⁰ Chlorins e6 (Ce6) is known as a commonly adopted PDT agent, both in experimental and clinical applications, which has irreplaceable advantages over other agents.^{21,22} However, Ce6 was also found to have some limitations, including irreversible degradation, poor solubility as well as rapid photobleaching, which call for the aid of multifunctional DDSs.²³

Cellular membranes (CM) were reported to be involved in many vital biological processes, which mainly ascribed to the proteins in the phospholipid bilayer.²⁴ In addition, the CM also shows high biocompatibility and inherits the merits of the source cells.²⁵ As a result, CM has been used to construct tumor-targeting nanoparticles for the treatment of various carcinomas,²⁶ in which CM have been shown to endow the modified nanoparticles with great tumor homing ability while at the same time reduce the undesired side effects of drugs in a facile and flexible way.^{27,28}

With the aim of combining CM and Ce6 in one DDS with tumor targetability for advanced PDT, amine SLN with positive surface charge was firstly synthesized and loaded with Ce6 (SLN/Ce6). Subsequently, the SLN/Ce6 was coated with negatively charged CM (derived from SGC7901 cells) to fabricate a tumor targeted platform (CM/SLN/Ce6). Compared with other currently available PDT DDSs, it was expected that CM modification could direct the CM/SLN/Ce6 to the homogenous SGC7901 cell line to increase its tumor targeted accumulation. Afterwards, the released Ce6 molecule within cancer cells could achieve sufficient PDT of gastric cancer upon the irritation of external NIR light.

Materials and methods

Materials

Indocyanine green (ICG), Ce6, Triton X-100, N-(2-aminoethyl)-3-aminopropyltrimethoxysilane (AEAPS), tetraethyl orthosilicate (TEOS), 2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA), 1,3-diphenylisobenzofuran (DPBF) and methylthiazolotetrazolium (MTT) were obtained from Sigma-Aldrich (St Louis, MO, USA). Chemicals and reagents otherwise stated were purchased from Aladdin Co., Ltd (Shanghai, China) and analytically pure.

Cell culture and animal model

The SGC7901 cell line (human gastric carcinoma) was purchased from American Type Culture Collection (ATCC, Manassas, VA, USA) and cultured using standard protocol according to previous report.²⁹

Female Balb/c nude mice (6~8 week) were obtained from Model Animal Research Center of Nanjing University (Nanjing, China) and the SGC7901 tumor xenograft model was established as previous reported.²⁹ All animal procedures strictly followed the NIH guidelines for the care and protection of laboratory animals and were confirmed by the Ethics Committee of the First Hospital of Jilin University.

Preparation of CM/SLN/Ce6

The amine SLN was prepared using a water-in-oil microemulsion according to a previous report.³⁰ In brief, the water-in-oil microemulsion was firstly prepared and stirred properly to obtain transparent solution. Afterwards, TEOS, AEAPS and NH₄OH were successively added into the microemulsion to initiate the reaction. After being stirred at room temperature for 24 hours, the solution was charged with additional ethanol, followed by centrifugation (3,000 rpm, 10 minutes, CR21, Hitachi Ltd, Tokyo, Japan) to collect amine SLN as precipitate. In order to obtain Ce6 loaded SLN (SLN/Ce6), the collected SLN was suspended in the proper amount (5 mL) of DMSO in which Ce6 was dissolved to achieve drug concentration of 5 mg/mL. The solution was agitated under room temperature for 24 hours and then centrifuged to collect SLN/Ce6 as mentioned above. The product was washed with ethanol several times and stored at 4°C until used.

The isolation of CM from SGC7901 cells was performed according to a previous report.³¹ In brief, the SGC7901 cells were seeded into culture dish (200 mm) and allowed to grow to 90% confluence. Afterwards, cells were washed with PBS, detached with trypsin and concentrated in 3 mL of dispersion buffer. The cells were centrifuged at 1,000 rpm for 5 minutes, and the collected pellet was homogenized in 1 mL of extracting buffer. Afterwards, the mixture was further centrifuged at 10,000 rpm for 10 minutes. The supernatant was again ultracentrifuged (100,000 rpm for 60 minutes). The CM pellet was resuspended and stored. The protein in CM was quantified by BCA kit (Beyotime, Shanghai, China).

The coating of CM onto SLN/Ce6 was performed according to previously reported method.³² Briefly, 250 µL of SLN/

Ce6 (nanoparticle concentration: 1 mg/mL) was mixed with different volume SGC7901 CM solution under vortex. Afterwards, the mixture was subjected to probetype sonication (100 W, 5 minutes). The mixture was further centrifuged (10,000 rpm, 10 minutes) to collect CM/SLN/Ce6.

The drug loading content (DLC) of CM/SLN/Ce6 was determined by immersing the prepared nanoparticles in methanol for 48 hours. After being centrifuged at 10,000 rpm for 30 minutes, the content of Ce6 in the supernatant was determined using HPLC with the method described as follows: Thermo-Fisher UltiMate™ 3000 (Thermo Fisher Scientific, Waltham, MA, USA) equipped with Phenomenex-C₁₈ column (250 mm×4.6 mm, 5 µm), mobile phase consisted of mixed methanol- ammonium acetate aqueous buffer (0.05 M, pH 5.5 and was adjusted to 3.0 by acetic acid); the wavelength was set at 405 nm, flow rate was 1 mL/minute and temperature was 30°C

Characterization

The size distribution of CM/SLN/Ce6 was measured by size analyzer (ZS90, Malvern). The morphology was observed using a transmission electron microscope (TEM, JEM-2100, JEOL, Japan).

Total protein from the CM was extracted using RIPA lysis buffer (Thermo Fisher) following the manufacturer's instructions, followed by concentration quantification using the BCA kit. Afterwards the Western blot analysis was performed strictly according to a previous report.³³ Briefly, samples were resolved gel electrophoresis, transferred onto PVDF membrane, stained with corresponding first antibodies and followed by incubation with IRDyeR680CW-labeled second antibody. Finally, the blots were observed using a densitometer (E-Gel Imager, Thermo-Fisher). Each sample was repeated in triplicate and beta-actin was employed as control.

Stability assay

The CM/SLN/Ce6 was diluted using PBS and mouse plasma, respectively. The changes of particle size was recorded for 48 hours. In addition, the fluorescence intensity of CM/SLN/Ce6 against Ce6 in solution was also monitored using fluorescence spectrophotometer (F-7000, Hitachi) for 6 days.³⁴

In vitro release and in vitro ROS generation

The Ce6 release from CM/SLN/Ce6 was studied using previous reported protocol.²⁹ In brief, CM/SLN/Ce6 was

loaded into a dialysis bag, immersed in 25 mL of PBS (pH of 7.4 and 5.5), and fixed in a thermostatic shaker (SY-2230; Crystal Technology, Texas, USA). At different time points, aliquot buffer was withdrawn to determine the drug concentration using HPLC as mentioned above.

The DPBF was adopted to test the generation of singlet oxygen as reported in a previous article.³⁵ In brief, 20 µL of DPBF (10 mM in DMSO) solution was added to CM/SLN/Ce6 under gentle agitation. The mixture was irradiated with a 680 nm laser beam (1 W/cm²). The time or concentration dependent variations of absorption peak at 418 nm of DPBF were measured by UV spectrophotometer.

Cytotoxicity assay

Drug free nanoparticles (5–100 µg/mL) or CM/SLN/Ce6 (Ce6 concentration, 0.25–5 µg/mL) were cultured with SGC7901 cells for 48 hours and the subjected to a standard MTT assay. The irradiation wavelength was 680 nm at 1 W/cm² for 5 minutes in all Ce6 containing groups.

Cellular uptake and ROS detection

In order to observe the internalization profile of CM/SLN/Ce6 in SGC7901 cells. Cells were firstly seeded in 6-well plates with 70% confluence and then cultured with free Ce6, SLN/Ce6 and CM/SLN/Ce6 at the Ce6 concentration of 2 µg/mL. For competitive binding assay, cells were pretreated with excessive CM for 2 hours prior to the addition of formulations. At prearranged time intervals, cells were rinsed with fresh HBSS and then subjected to confocal laser scanning microscope (CLSM, BX61W1-FV1000, Olympus Corporation, Tokyo, Japan) observation. In addition, cells were harvested and analyzed by flow cytometer (FCM, Attune NxT, Thermo Fisher).

DCFH-DA is a cell permeable derivative of DCF which remains nonfluorescent during endocytosis. However, the oxidization product of DCFH-DA can exert greatly enhanced fluorescent intensity upon exposure to ROS. To detect the intracellular ROS levels, SGC7901 cells were firstly incubated with different formulations for 4 hours (Ce6 concentration: 0.1 mg/mL). Afterwards, DCFH-DA (25 mM) was added to incubated with cells at 37°C in the dark for 30 minutes. Thereafter, cells were exposed to 680 nm irradiation (1 W/cm², 5 minutes), followed by CLSM observation as mentioned above.

In vivo tumor targeting of CM/SLN

ICG was loaded into SLN and CM/SLN in place of Ce6. Mice bearing SGC7901 tumor were intravenously administered with SLN/ICG and CM/SLN/ICG (20 $\mu\text{g}/\text{mouse}$ of ICG) and the in vivo distribution of ICG signal at 6 and 12 hours was recorded by imaging system (ZEWTON 7.0, Vilber, Marne-la-Vallée, France). Moreover, the mice were sacrificed at 48 hours post injection and the fluorescence intensity of the both formulations in tumor tissues and major organs was assessed using the same system.

In vivo antitumor assay

Mice were randomly divided into 4 groups ($N=6$) and administered with saline (as control), free Ce6, SLN/Ce6 and CM/SLN/Ce6 (5 mg/kg Ce6). The irradiation wavelength was 680 nm at 1 W/cm^2 for 5 minutes in all Ce6 containing groups. The administration and other protocols follow the previous report.³⁶

Statistical analysis

Statistical analysis was performed according to a previous report.³⁷ In brief, experiments were conducted in a parallel manner and data were presented as mean \pm SD. Statistical significance was tested by two-tailed Student's *t*-test or

one-way ANOVA $P<0.05$ or $P<0.01$ was considered as statistically significant.

Results and discussion

Characterization of CM/SLN/Ce6

In order to combine decent drug-loading capacity and biocompatibility in one DDS, SLN as a widely adopted biomaterial was prepared by a previous reported water-in-oil microemulsion method and employed as the skeleton material. Afterwards, the optimal ratio between CM protein and SLN/Ce6 was explored by mixing SLN/Ce6 with different ratios of CM (SLN/Ce6 to CM protein, w/w). The particle size and zeta potential of different ratios were recorded. As indicated in Figure 1A, the SLN/Ce6 had a hydrodynamic size of 90.4 ± 3.2 nm and a zeta potential of $+32.4 \pm 1.7$ mV. Upon CM coating, the particle size of CM/SLN/Ce6 gradually increased with the increase of CM while the zeta potential decreased. This phenomenon suggested that SLN/Ce6 was successfully coated with CM. Interestingly, when the mass ratio excess 6, both the particle size and zeta potential of CM/SLN/Ce6 reached a balance with additional CM resulted insignificant changes. In detail, the hydrodynamic diameter of CM/SLN/Ce6 reached 115.6 ± 1.6 nm while the surface charge of CM/SLN/Ce6 reversed to -30.4 ± 2.3 mV, which was similar to that of the CM

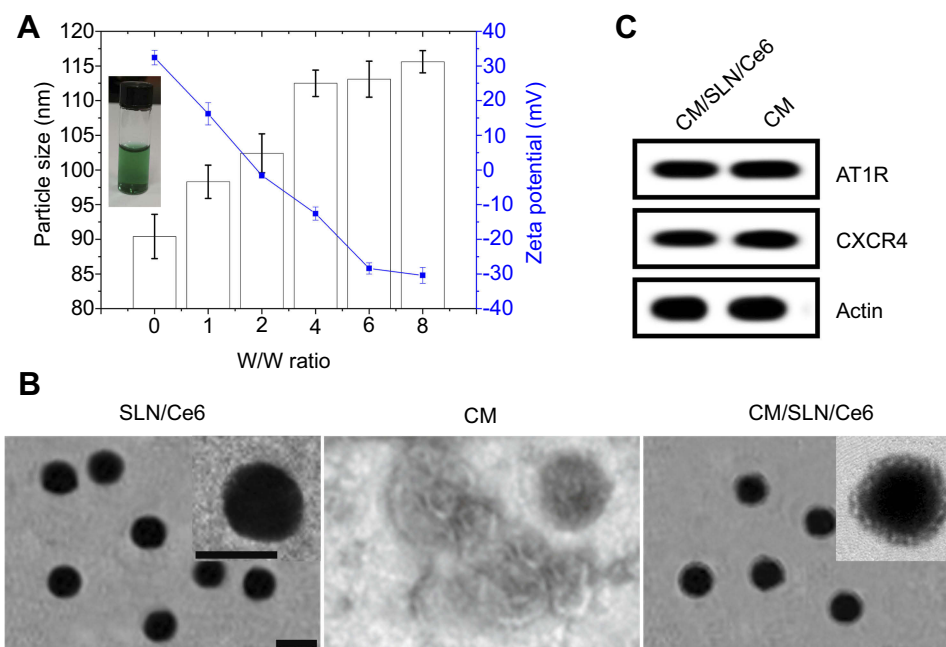


Figure 1 (A) Variation of particle size and zeta potential as a function of CM ratio (SLN/Ce6 to CM protein, w/w). Inset image was the optical image of CM/SLN/Ce6 at the optimal w/w ratio, which provides direct evidence of successful Ce6 loading. (B) Western blot analysis of two proteins (AT1R and CXCR4) in CM and CM/SLN/Ce6. (C) TEM images of SLN/Ce6, CM and CM/SLN/Ce6. Scale bar: 100 nm.

Abbreviations: Ce6, chlorins e6; CM, cellular membranes; TEM, transmission electron microscope; SLN, silica nanoparticles.

vesicles (-29.7 ± 1.5 mV). These results indicated that CM was successfully coated onto SLN/Ce6 and the coating was saturated at the mass ratio of 6. The negative surface charge of CM/SLN/Ce6 was expected to protect CM/SLN/Ce6 from nonspecific absorption of plasmatic proteins, which in turn enhance the stability of CM/SLN/Ce6 during circulation.³⁸

The formation of CM/SLN/Ce6 from SLN/Ce6 was also verified using TEM. As shown in Figure 1B, TEM revealed that SLN/Ce6 exhibited uniform and spherical nanostructure with homogeneous diameter dispersion at around 90 nm. CM vesicles showed irregular shape of particles with diameter ranging from nanometer to micrometer. The finally obtained hybrid CM/SLN/Ce6 was characterized by a spherical core-shell structure with a clearly observed lipid bilayer on the surface.

It has been demonstrated that proteins on CM are essentially responsible for the tumor-targeting capability of the modified nanoparticles.³⁹ As a result, we selected two different membrane proteins (AT1R and CXCR4) and compared their expression pattern in both CM and CM/SLN/Ce6 using Western blot assay. As displayed in Figure 1C, the expression levels of both proteins in both groups were comparative, which suggested that CM/SLN/Ce6 inherited the integrated protein profiles of CM without loss or degradation during the coating process. It was also considered as a solid evidence to verify the successful preparation of CM/SLN/Ce6.

The DLC of Ce6 in CM/SLN/Ce6 determined by HPLC was 15.34%.

Stability assay

The stability of nanoparticles in physiological environment is generally considered as one of the basic requirements if a DDS is intended to successfully deliver the cargo.⁴⁰ According to previous report, the *in vivo* fate of the DDS is strongly related to its size.⁴¹ As a result, the capability to maintain its size without significant variation is vital for the DDS to bypass the complex extracellular barriers before arriving at the targeted sites. Herein, the size change of CM/SLN/Ce6 in PBS (pH 7.4) and mouse plasma was monitored for 48 hours to assess its colloidal stability. As shown in Figure 2A, the size of CM/SLN/Ce6 in both PBS and plasma at all tested time points showed insignificant changes, which suggested that CM/SLN/Ce6 might be able to withstand the irritation during circulation. Alternatively, Ce6 is prone to degrade upon light irradiation in aqueous solution, which was reflected as photo bleaching. The DDS was expected to retard this process and increase the resistance of Ce6 to light irradiation. As a proof of concept, the comparative fluorescence stability of CM/SLN/Ce6 and free Ce6 under sunlight irradiation for 6 days was studied. As displayed in Figure 2B, free Ce6 showed immediate photo bleaching as the fluorescence intensity dropped dramatically to 61.5% only 1 day post irradiation, which continuously declined to 15.6% at the end of the test. In contrast, the fluorescence intensity in CM/SLN/Ce6 group showed much slower decrease as only 20.1% of the Ce6 suffered from photo bleaching at the end of the test. These results suggested that CM/SLN/Ce6 was capable of protecting the encapsulated Ce6 from light induced degradation, which is

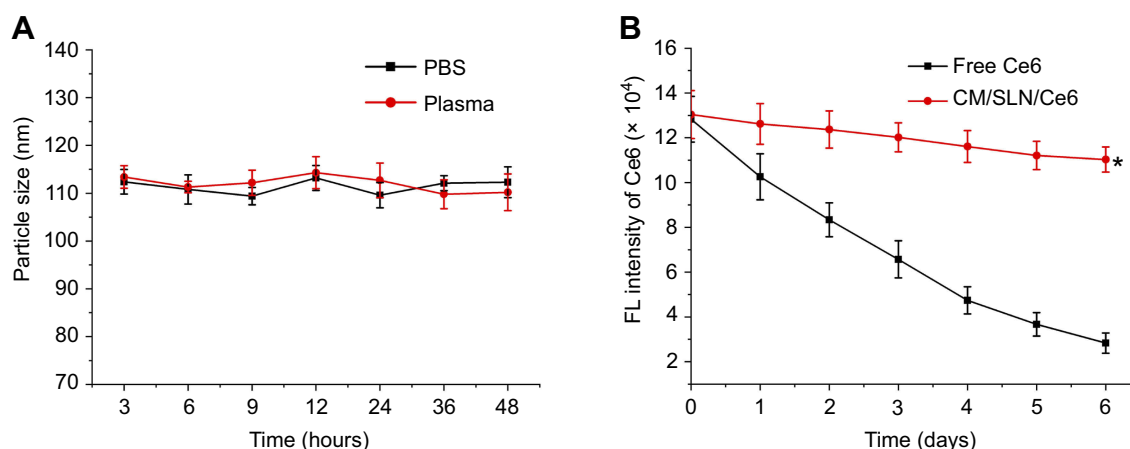


Figure 2 (A) Colloidal stability of CM/SLN/Ce6 in PBS (pH 7.4) and mouse plasma at 37°C for up to 48 hours. **(B)** Comparative fluorescent stability of CM/SLN/Ce6 and free Ce6.

Note: Data were shown as mean \pm SD (N=3). * $p < 0.05$.

Abbreviations: FL, fluorescent; Ce6, chlorins e6; CM, cellular membranes; SLN, silica nanoparticles.

favorable for CM/SLN to be developed as a stable system to satisfy further applications.⁴²

In vitro drug release and in vitro ROS generation

The drug release pattern of CM/SLN/Ce6 was evaluated in two different pH conditions (pH 7.4 and pH 4.5) to mimic the extracellular physiological environment and intracellular endolysosomal compartment. As illustrated in Figure 3A, after 120 hour of incubation, the accumulated drug release of CM/SLN/Ce6 at pH 7.4 was relatively slower (32.96%) compared to that of pH 5.5 (74.61%). It was inferred that the acidic environment could facilitate the drug release from CM/SLN/Ce6, which was beneficial for CM/SLN/Ce6 to serve as a responsive platform for advanced drug delivery.

The preservation of Ce6 to generate ROS under laser irradiation is a prerequisite for CM/SLN/Ce6 to exert PDT. To verify this, DPBF was employed to measure the the ROS generation capacity of CM/SLN/Ce6. It is generally recognized that the UV absorbance of DPBF at 418 nm would decrease when DPBF reacts with neighboring ROS. As a result, the variation of DPBF absorption at 418 nm was identified as an easy tool to evaluate the ROS generation of Ce6. As shown in Figure 3B, under laser irradiation (1 W/cm^2), UV absorption at 415 nm showed a continuous decrease at constant CM/SLN/Ce6 concentration (Ce6: 0.1 mg/mL) with 72% of absorption quench at 6 minutes post irradiation. The concentration dependent variations of DPBF absorption at 415 nm was also investigated. Various concentrations of CM/SLN/Ce6 (Ce6 concentration: $0.1\text{--}0.5 \text{ mg/mL}$) were irradiated 1 W/cm^2 for 60 seconds

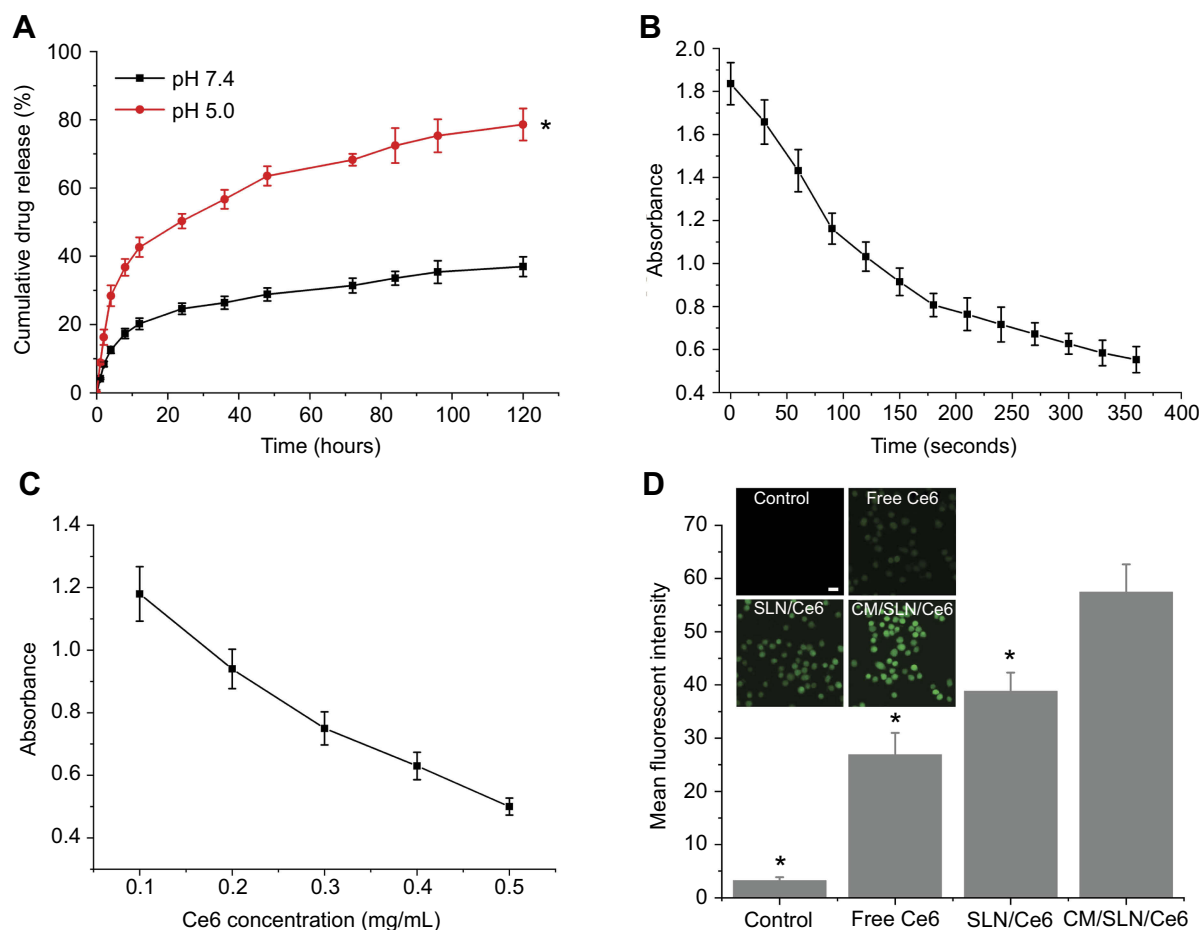


Figure 3 (A) Drug release profiles of Ce6 from the CM/SLN/Ce6 in release media under extracellular and intracellular condition of pHs (7.4 and 5.5). (B) Variation on absorbance of DPBF at 418 nm in the presence of CM/SLN/Ce6 (Ce6 concentration 0.1 mg/mL) after irradiation (1 W/cm^2) for different time intervals. (C) Variation on absorbance of DPBF in the presence of different concentrations of CM/SLN/Ce6 (Ce6 concentration: $0.1\text{--}0.5 \text{ mg/mL}$) under irradiation for 60 seconds. (D) The in vitro ROS generation analysis of CM/SLN/Ce6. Main figure illustrated flow cytometric analysis of MFI in cells treated by different formulations after 5 minutes of laser irradiation (1 W/cm^2). Inset images demonstrated the representing CLSM images of cells treated with different formulations.

Notes: Significant differences were vs CM/SLN/Ce6. Scale bar: $20 \mu\text{m}$. Data were shown as mean \pm SD (N=3). * $p < 0.05$.

Abbreviations: Ce6, chlorins e6; CM, cellular membranes; SLN, silica nanoparticles; DPBF, 1,3-diphenylisobenzofuran; MFI, mean fluorescence intensity.

and the UV absorption at 415 nm was recorded. As shown in Figure 3C, the ROS production of CM/SLN/Ce6 was positively related to the Ce6 concentration upon laser irradiation. Both results suggested that the encapsulated Ce6 within CM/SLN/Ce6 preserved the capability to generate ROS.

The intracellular ROS generation profile of CM/SLN/Ce6 was further investigate using DCFH-DA. DCF, the oxidized product of DCFH-DA, shows green fluorescence which can be easily detected at Ex 485 nm/Em 530 nm and can be quantified using flow cytometry. As shown in Figure 3D, after laser irradiation, no observable fluorescent signal was detected in cells without Ce6 (control group), which suggested that the ROS level in these cells was weak. Observable weak fluorescent signal was found in cells treated with free Ce6. In addition, significantly enhanced ROS concentration was detected in SLN/Ce6 treated cell, indicating that the intracellular Ce6 in this group might be higher than free Ce6 group. Most importantly, fluorescent intensity of DCF in cells treated with CM/SLN/Ce6 was much more potent than other groups, suggesting effective ROS production. CLSM (inset images) also confirmed these results. Therefore, CM/SLN/Ce6 with effective delivery efficacy was suggested to be a preferable DDS for PDT of cancer.

In vitro anticancer assay

The standard MTT assay was adopted in our study to investigate the in vitro anticancer efficiency of CM/SLN/Ce6. The viability of cells exposed to drug free CM/SLN

still over 90% at the highest concentration (Figure 4A), which suggested that CM/SLN holds preferable biocompatibility with negligible cytotoxicity. The following cytotoxicity assay using drug loaded formulation was shown in Figure 4B. It was observed that under laser irradiation, CM/SLN/Ce6 showed superior anticancer efficacy than the other two groups at all tested concentrations. To further verify this conclusion, the apoptosis related proteins (cleaved caspase-3, bcl-2 and cytochrome-3) in three tested groups (Ce6 concentration: 5 $\mu\text{g/mL}$) were assessed.^{43–45} As shown in the inset images, as expected, the level of cleaved caspase-3 was higher in CM/SLN/Ce6 treated cells while bcl-2 (responsible for suppressing apoptosis) was much lower than other groups, which further confirming the preferable anticancer efficacy of CM/SLN/Ce6. In addition, the CM/SLN/Ce6 showed much higher level of cytochrome-3, which indicated that mitochondria damage was also involved in the apoptosis of cells.⁴⁶

Cellular uptake assay of cm/sln/ce6

In order to reveal the potential reasons responsible for the discriminative anticancer efficacy of different formulations, in vitro cellular uptake assay was performed to investigate if SGC7901 CM modification could increase the internalization of CM/SLN/Ce6 to SGC7901 cells.

As displayed in Figure 5A, the increase of fluorescent-signal positively related to the incubation time in both groups. However, free Ce6 showed the most inferior intracellular accumulation compared to both nanoparticles,

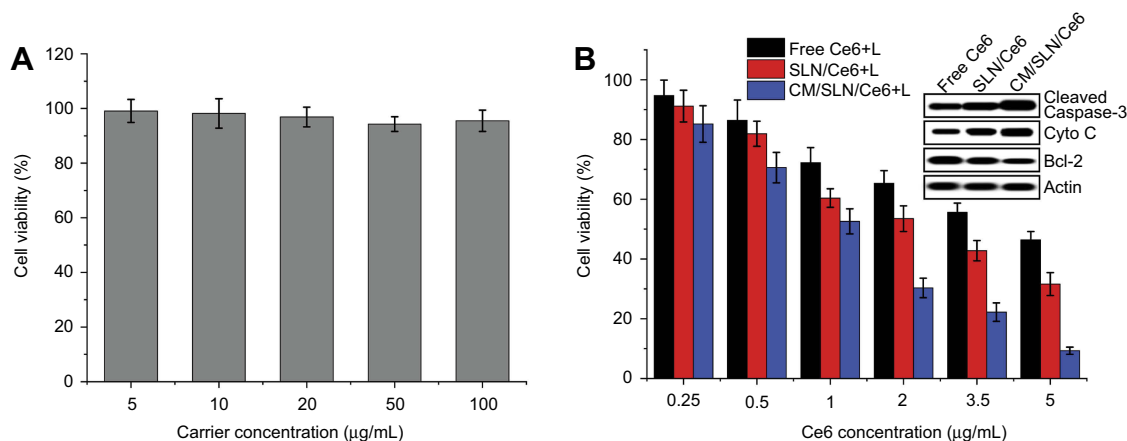


Figure 4 (A) Cytotoxicity of CM/SLN after 48 hours incubation with SGC7901 cells. **(B)** Cytotoxicity of free Ce6, SLN/Ce6 and CM/SLN/Ce6 (with laser irradiation) at different Ce6 concentrations against SGC7901 cells after 48 hours incubation. Inset images demonstrated the cleaved caspase-3, cytochrome C and bcl-2 levels in three groups (Ce6 concentration: 5 $\mu\text{g/mL}$).

Note: Data were shown as mean \pm SD (N=3).

Abbreviations: Ce6, chlorins e6; CM, cellular membranes; SLN, silica nanoparticles.

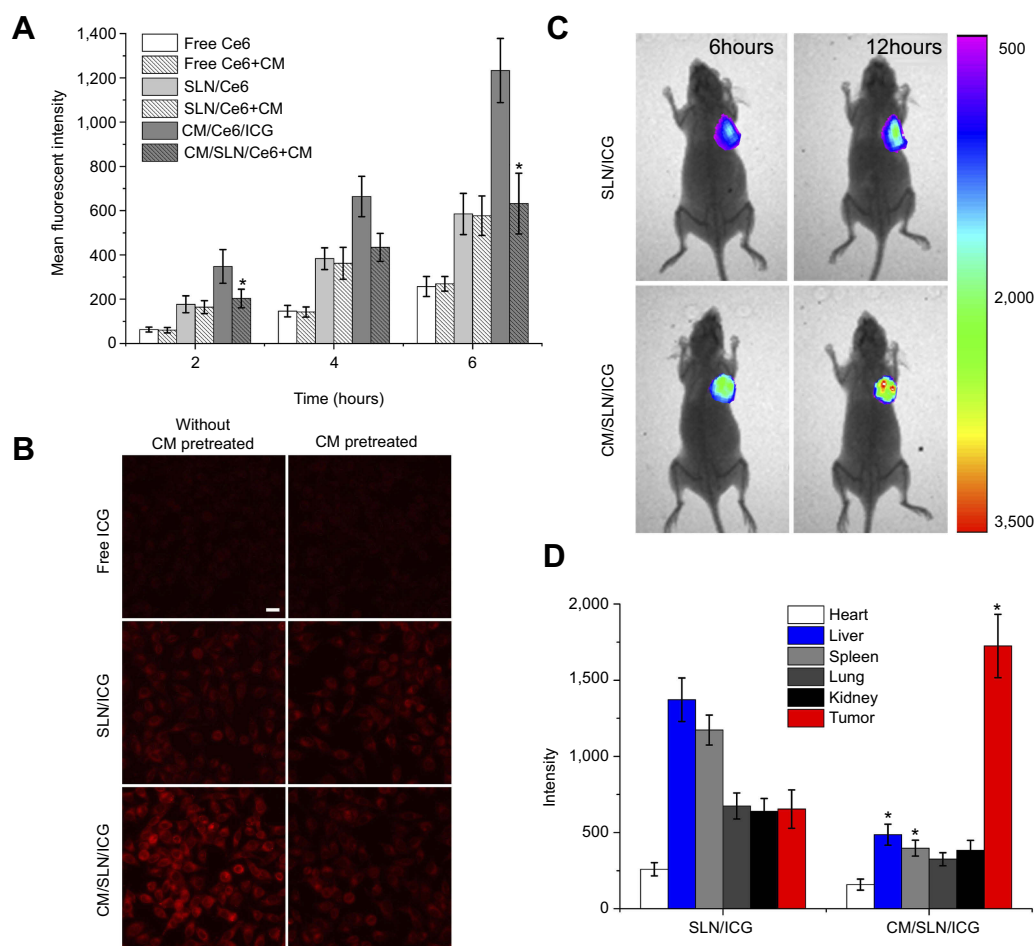


Figure 5 (A) Quantitative analysis of intracellular time-dependent uptake of different formulations in SGC7901 cells (pretreated with/without CM). Significant differences were vs groups pretreated without CM. (B) Representative CLSM images of uptake of different formulations in SGC7901 cells (pretreated with/without CM) at 4 hours postincubation. Scale bar: 20 μ m. (C) In vivo real time distribution of SLN/ICG and CM/SLN/ICG at 6 and 12 hours postincubation. (D) Mean fluorescent intensity of dissected tumors and major organs of mice treated with SLN/ICG and CM/SLN/ICG at 48 hours postinjection.

Note: Data were expressed as mean \pm SD (N=3).

Abbreviations: Ce6, chlorins e6; CM, cellular membranes; SLN, silica nanoparticles; CLSM, confocal laser scanning microscope; ICG, indocyanine green dye.

which was inconsistent with a previous report that nanoparticles can improve the uptake of drugs into cells.³⁵ In addition, it was observed that higher Ce6 signals were observed in CM/SLN/Ce6 group than SLN/Ce6 under all tested conditions. In order to reveal the role of CM in cellular uptake, cells were pretreated with excess CM prior to addition of nanoparticles. Interestingly, it was observed that the cellular Ce6 intensity in CM/SLN/Ce6 group suffered a significant decline while SLN/Ce6 group showed insignificant variations, which was also verified by CLSM imaging of cells at 4 hours postincubation (Figure 5B). These results suggested that the internalization of CM/SLN/Ce6 into SGC7901 cells was positively related to CM modification, possibly via CM mediated endocytosis.

In vivo distribution

SGC7901 CM modification was expected to assist the tumor-homing of CM/SLN/ICG to the isogenous SGC7901 cells to increasing the accumulation of nanoparticles in the tumor tissue. In order to verify the hypothesis, the real time distribution of SLN/ICG and CM/SLN/ICG was monitored by in vivo imaging system at 6 and 12 hours postadministration. As shown in Figure 5C, as expected, CM/SLN/ICG showed much effective accumulation within the tumor tissue compared to SLN/ICG at both time intervals. In addition, the fluorescent distribution of tumor and major organs obtained by ex vivo imaging also confirmed this observation. As shown in Figure 5D, due to the poor tumor targetability of SLN/ICG, the ICG signal in this group was shown to be largely located in the

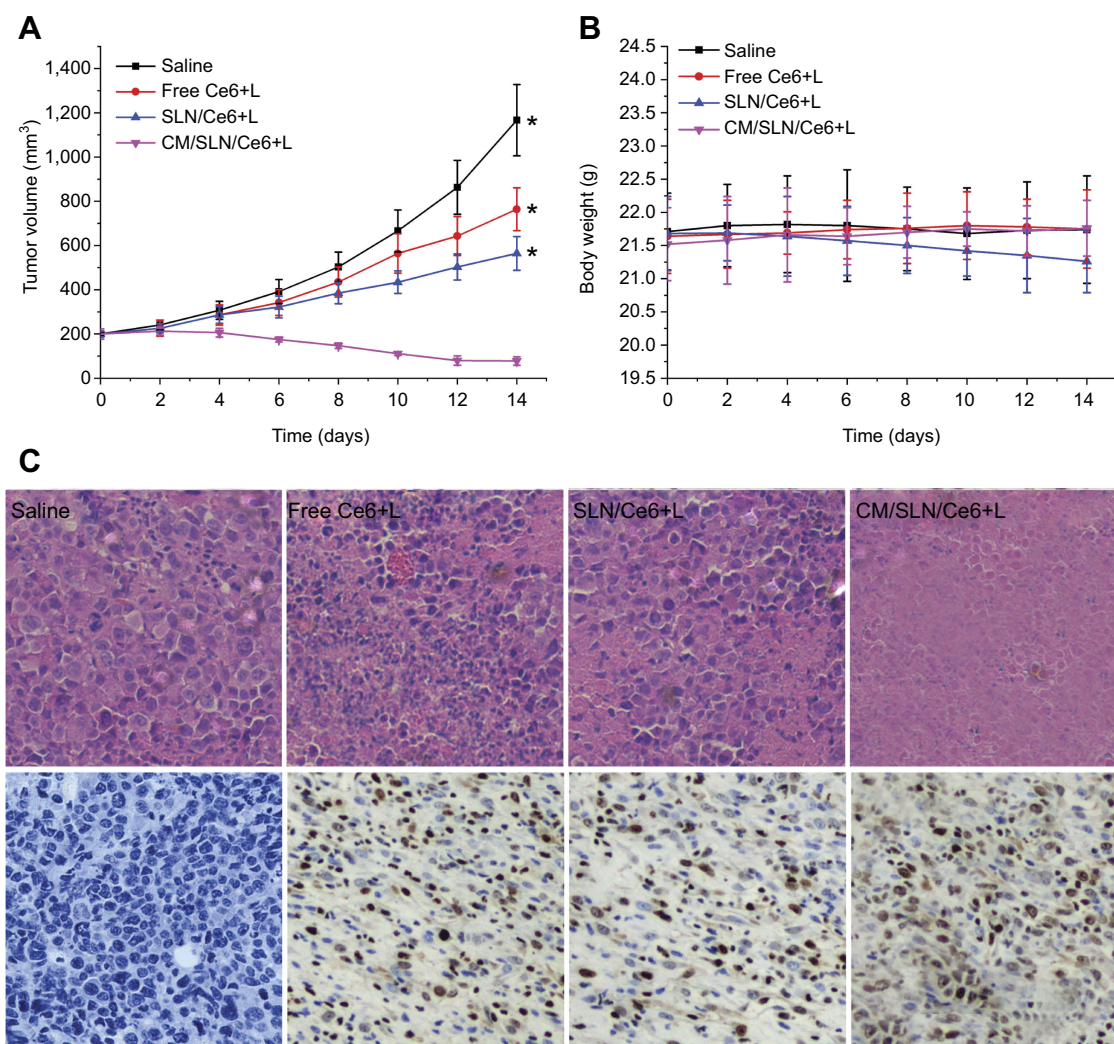


Figure 6 The tumor volume (A), body weight (B) and H&E/TUNEL staining (200×) of tumor tissue (C) analysis of SGC7901 tumor-bearing Balb/c nude mice after intravenous administration of different formulations.

Notes: Data were expressed as mean \pm SD (N=6). Significant differences were vs CM/SLN/Ce6. * p <0.05.

Abbreviations: Ce6, chlorins e6; CM, cellular membranes; SLN, silica nanoparticles.

liver and kidneys. In contrast, the distribution profile of CM/SLN/ICG is significantly changed with less accumulation in liver and kidney while enhanced homing to tumor tissue.

In vivo anticancer assay

In vivo anticancer efficacy of CM/SLN/Ce6 was assessed. The tumor growth profiles of different formulations were displayed in Figure 6A. It was observed that free Ce6 or SLN/Ce6 could exert certain suppression effects on tumor growth. By contrast, the anticancer effect of CM/SLN/Ce6 was much more potent than the others with a final tumor volume of $78 \pm 19 \text{ mm}^3$ at the end of the test. Moreover, body weight variations of mice in different groups also revealed some interesting results. As shown in Figure 6B, no significant decline in body weight

was observed in CM/SLN/Ce6 group, suggesting that the preferable tumor homing of CM/SLN/Ce6 could increase its anticancer efficacy while reduce the undesired toxic effects. In contrast, the untargeted distribution of SLN/Ce6 caused systematic toxicity to mice which was reflected by the time-dependent decrease in body weight. Moreover, H&E and TUNEL assays were conducted to assess the apoptosis profile of tumor tissues treated by different formulations (Figure 6C). Compared to the insignificant apoptosis in saline group, increased apoptosis was observed in all Ce6 containing groups with the most serious one observed in CM/SLN/Ce6 group, which was in line with results obtained in the above assays. In a word, the CM/SLN/Ce6 holds great potential to be a preferable tumor-targeting DDS for effective PDT of gastric cancer.

Conclusion

In our study, we successfully fabricated a Ce6 loaded and tumor-targeted DDS using SGC7901 CM modified SLN (CM/SLN/Ce6). In this platform, we combined the tumor-targeting nature of CM and the decent loading capacity of SLN. Experimental results revealed that CM/SLN/Ce6 was able to specifically deliver Ce6 to isogenous SGC7901 cells. Furthermore, the CM/SLN/Ce6 was capable of releasing the loaded drug in a pH dependent manner. It was noted that both in vitro and in vivo experiments revealed that CM/SLN/Ce6 hold superior tumor-targeting ability with greatly enhanced anticancer efficacy than unmodified SLN and free drug.

Disclosure

The authors report no conflicts of interest in this work.

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