

Ionically Cross-Linked Chitosan Nanoparticles for Sustained Delivery of Docetaxel: Fabrication, Post-Formulation and Acute Oral Toxicity Evaluation

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Introduction: Polymeric nanoparticles are potential carriers for the efficient delivery of hydrophilic and hydrophobic drugs due to their multifaceted applications. Docetaxel is relatively less hydrophobic and twice as potent as paclitaxel. Like other taxane chemotherapeutic agents, docetaxel is not well tolerated and shows toxicity in the patients. Nanoencapsulation of potent chemotherapeutic agents has been shown to improve tolerability and therapeutic outcome. Therefore, the present study was designed to fabricate chitosan and sodium tripolyphosphate (STPP) based on ionically cross-linked nanoparticles for sustained release of docetaxel.

Methods: Nanoparticles were prepared by the ionic-gelation method by dropwise addition of the STPP solution into the chitosan solution in different ratios. CNPs were characterized for post-formulation parameters like size, zeta potential, scanning electron microscope (SEM), FTIR, DSC/TGA, pXRD, and in-vitro drug release, as well as for acute oral toxicity studies in Wistar rats.

Results and discussion: The optimized docetaxel loaded polymeric nanoparticles were in the size range (172.6nm–479.65 nm), and zeta potential (30.45–35.95 mV) required to achieve enhanced permeation and retention effect. In addition, scanning electron microscopy revealed rough and porous surface, whereas, FTIR revealed the compatible polymeric nanoparticles. Likewise, the thermal stability was ensured through DSC and TG analysis, and powder X-ray diffraction analysis exhibited solid-state stability of the docetaxel loaded nanoparticles. The in-vitro drug release evaluation in phosphate buffer saline (pH 7.4) showed sustained release pattern, i.e. 51.57–69.93% within 24 hrs. The data were fitted to different release kinetic models which showed Fickian diffusion as a predominant release mechanism ($R^2 = 0.9734$ – 0.9786 , $n = 0.264$ – 0.340). Acceptable tolerability was exhibited by acute oral toxicity in rabbits and no abnormality was noted in growth, behavior, blood biochemistry or histology and function of vital organs.

Conclusion: Ionically cross-linked chitosan nanoparticles are non-toxic and biocompatible drug delivery systems for sustained release of chemotherapeutic agents, such as docetaxel.

Keywords: tripolyphosphate, ionic gelation, sustained release, targeting, biocompatible, biodegradable

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Introduction

Cancer is a disease characterized by uncontrolled growth of cells that are capable of invading, and metastasizing the surrounding tissue. It is the second leading cause of

mortality and accounts for 1 in every 6 deaths in the world. The majority of the cancer-related deaths occur in low- and middle-income countries, it is posing a higher impact on the total economy of the world.^{1,2} Chemotherapy is the main component of cancer treatment either alone or in combination with other treatment options.³ Although anti-cancer agents are available for chemotherapy of various types of cancer, yet inaccessibility to the target site leads to insufficient amount of drug in cancerous cells. Passive targeting facilitates drug delivery at the site of action without requiring any surface decoration or ligand attachment (active targeting). Cancer has leaky vasculature which facilitates the entry of nanoparticles and an increased amount of vascular permeability factors (bradykinin, peroxynitrite & nitric oxide). Additionally, the immature lymphatic system is incapable of drainage from interstitial spaces. Collectively, these factors lead to prolonging the stay of drug carriers at cancer site which is also known as enhanced permeability and retention (EPR) effect. Previous literature reports that drug carriers with an average diameter of approximately 400 nm are good candidates of passive targeting by the EPR effect.^{4,5}

Polymeric nanoparticles (PNPs) of a particular size range have relatively higher capability to accumulate at the tumor site without affecting normal tissues.⁶ For biomedical applications, biodegradable polymers based PNPs are used to modify solubility, permeability and bioavailability of therapeutic agents.⁷ Chitosan is a nontoxic biopolymer of natural origin derived by the deacetylation of chitin and is widely employed for various drug delivery purposes.⁸ It is obtained from crustacea shells, insect cuticles, and cell walls of some fungi and is straight-chain amino polysaccharide (poly- 1,4-D-glucosamine) with many features which are favorable for its use in drug delivery.⁹ It is being highly explored for its application in oral delivery.¹⁰ Not only chitosan, but its derivatives like glycol chitosan, thiolated chitosan, arginine-chitosan, carboxymethyl chitosan, aminated chitosan are also being explored for the drug delivery application.^{11,12} Chitosan undergoes protonation in an acidic environment and provides favorable release at low pH. As the tumor micro-environment has acidic pH due to poor drainage of wastes, chitosan may be employed for tunable drug delivery to cancer.¹³

Docetaxel (DTXL) has a taxane ring that promotes the assembly of microtubules and stabilizes them, thus, preventing them from depolarization.¹⁴ It exhibits higher cellular uptake and intracellular retention than paclitaxel

due to its broader cell cycle activity and more binding affinity towards β -tubulin. It used for chemotherapy of ovarian, prostate, breast, gastric and non-small cell lung cancer.¹⁵ It is twice as potent and relatively less hydrophobic than the paclitaxel.¹⁶ Docetaxel is used as injection via intravenous route but limited aqueous solubility and systemic toxicity are main problems associated with clinical use of DTXL. These features of DTXL make it a good candidate for delivery through PNPs to eradicate any of the said problems.^{17,18} Therefore, the present study was aimed to formulate the chitosan-based PNPs to enhance solubility and sustain delivery of docetaxel for the treatment of various types of cancer.

Materials and Methods

Materials

Docetaxel (DTXL) was purchased from Mesochem Technology Co., Ltd, Beijing, China. Low molecular weight chitosan and sodium tripolyphosphate (STPP) were procured from Sigma-Aldrich, India. Ethanol was obtained from Merck, Germany, and acetic acid from VWR chemicals, USA. Distilled water was prepared by using the distillation apparatus in the Pharmaceuticals Research Laboratory, The Islamia University of Bahawalpur, Bahawalpur, Pakistan. All the chemicals used were of analytical grades and used as received without further processing.

Preparation of Polymeric Nanoparticles

Chitosan polymeric nanoparticles (CNPs) were fabricated by the ionic-gelation method with minor modifications.¹⁹ Chitosan was dissolved overnight in 1.5% v/v acetic acid solution in distilled water. Varied concentrations were made of STPP in water and DTXL in ethanol in separate vessels (Table 1). DTXL and STPP solutions were mixed followed by drop-wise addition of the aforementioned mixture into chitosan solution with continuous stirring at room temperature. The mixture was subjected to stirring (900 rpm, 45 mins) followed by centrifugation (11,000 rpm, 30 mins) and nanoparticles were obtained as a fine pellet. The obtained nanoparticles were lyophilized for further characterization.

Percentage Yield (%)

Percentage yield (%) of the nanoparticles produced by a specific method is an important parameter with the pharmaceutical point of view as it may be helpful to predict the probability of industrial scale-up and commercial feasibility of a process. A process with the highest

yield is considered preferable. The percentage yield was calculated to determine the efficiency of methods to generate the product. The total amount of powder obtained after lyophilization (CHRIST alpha 1–4 LD, UK) was weighed using analytical weighing balance (Shimadzu, Japan, AUW 220) and percentage yield was calculated using the following formula.²⁰

$$\text{percentage yield} = \frac{\text{Quantity of CNPs obtained}}{\text{total mass of all ingredients (Chitosan + STPP + Drug)}} \times 100 \quad (1)$$

Particle Size, Polydispersity Index (PDI), and Zeta Potential Analysis

The particle size of PNPs is the performance determinant of their efficiency as passive targeting to cancer site while PDI and zeta potential are indicators of homogeneity and colloidal stability, respectively.²¹ For the evaluation of particle size, PDI, and zeta potential Zeta Sizer Nano (ZS-90, Malvern, UK) was used.

Entrapment Efficiency and Drug Loading Capacity

Entrapment efficiency (EE) and loading capacity (LC) were calculated by the indirect method. In this method, PNPs formulation is subjected to centrifugation and the un-entrapped fraction of drug was quantified in the supernatant by using a calibration curve. The whole analysis was done in triplicate to minimize handling errors. EE is the percentage of the drug, from total drug initially added the drug, that is successfully encapsulated within the system, whereas, LC is the percentage of the drugs that is successfully loaded on the specific mass of CNPs.^{7,22}

$$\text{L.C} = \frac{\text{total amount of drug added} - \text{amount of untrapped drug}}{\text{total mass of CNPs}} \times 100 \quad (2)$$

$$\text{E.E} = \frac{\text{total amount of drug added} - \text{amount of untrapped drug}}{\text{total amount of drug added}} \times 100 \quad (3)$$

Scanning Electron Microscopic (SEM) Studies

Scanning Electron Microscopic (SEM) studies were used to evaluate the morphology and surface characteristics of DTXL

loaded CNPs.²³ The lyophilized powder was placed on a piece of the electro-conductive chip of silicon over the top of the metal stub (aluminum). The samples were coated on the stubs and examined under Field Emission Scanning Electron Microscope (FESEM, JSM-5910, JEOL, Japan) at different magnifications using an electron acceleration voltage 10kV.

Thermal Stability (DSC and TGA)

The thermal stability of the DTXL, chitosan, STPP, physical mixture and selected formulation (CNP3) was estimated through differential scanning calorimetry (DSC) and thermogravimetric analysis (TGA). The DSC analysis was done to evaluate phase transition in the sample as a function of time and temperature in a controlled atmosphere through the graph between temperature (°C) and flow of heat (w/g). The thermogravimetric analysis was done to evaluate the percent weight loss by increasing temperature through the graph between temperature (°C) and percent weight loss.²¹

Powdered X-Ray Diffraction Analysis (PXRD)

PXRD analysis of DTXL, chitosan, STPP, physical mixture comprising equal amounts of components and the selected formulation (CNP5) was performed to elucidate the physical state of the individual components, physical mixture and the formulation. The X-ray diffractograms based on the Bragg's law were obtained by using a PXRD diffractometer (Bruker Axs, D8 Advance, Germany) with CuK α radiation, 45kV monochromatic voltage and 40 mA electric current. The range for 2 θ diffraction angle was from 10° to 60°.²⁴

Fourier Transform Infrared Spectroscopy (FTIR) Analysis

The FTIR analysis was performed to evaluate interactions or compatibility of the formulation components which in turn highlights the stability of the developed system.²⁵ FTIR spectroscopic analysis of DTXL, chitosan, STPP, physical mixture (DTXL, chitosan and STPP in 1:1:1) and a selected formulation (CNP3) was performed by Attenuated Total Reflectance (ATR) assembly with diamond crystal (Bruker, Tensor 27 series, USA). The system was set to 32 scans for each sample for a better scan and reliable results.

In-vitro Drug Release Studies and Kinetic Modeling

All formulations were evaluated for in-vitro release behavior by dissolution studies and release mechanism by application of

the kinetic modeling approach. For this purpose, developed formulations were taken equivalent to 2 mg DTXL and were dispersed in 2 mL of the Phosphate Buffer Saline (PBS) of pH 7.4. Dialysis membrane (Spectrum®) with 8–10 KDa cut off value was used in this study. The membrane was soaked for 3 hrs in distilled water before the initiation of experimentation to clear all the pores. It was cut to a suitable size and formulation equivalent to 2 mg DTXL suspended in PBS 7.4 is added in it. Both ends were tied and it was placed in 600 mL dissolution medium (PBS 7.4 containing 0.5% tween 80 to maintain sink conditions) at $37 \pm 0.5^\circ\text{C}$. USP type II apparatus (paddle apparatus) was used for the study and paddles were rotated at 50 rpm. Samples of 3 mL were withdrawn at predefined intervals and dissolution vessels were replenished with an equal volume of fresh dissolution medium to maintain sink conditions. Taken samples were filtered through $0.45 \mu\text{m}$ Nylon syringe filters (Membrane Solution, USA) and analyzed using UV-Spectrophotometer (IRMECO-U2020) at λ_{max} 230 nm. Readings were taken in triplicate to minimize error.²⁶ Calculated cumulative percentage release was evaluated for each sample using different kinetic models, such as zero order, first order, Higuchi and Korsmeyer-Peppas model, with DDSolver.xla extension of Microsoft Excel.

Acute Oral Toxicity Study Evaluation

The oral acute toxicity study was conducted according to the OECD (Organization for Economic Co-operation and Development) animal welfare guidelines and Declaration of Helsinki for ethical principals in medical research in humans and, where appropriate, on animals experimentation.²⁷ The approval of the study was obtained from the Pharmacy Research Ethics Committee (PREC), Faculty of Pharmacy and Alternative Medicine, The Islamia University of Bahawalpur (letter no. 90/S-2018/PREC dated 15-01-2018). Eight (08) male healthy Wistar rats were selected for the study and kept in clean cages for 14 days. All the rats were given free access to water and diet throughout the study. The weight of the rats was determined before dosing. Rats

were divided into two groups randomly, each group containing four rats. Group I was administered with Chitosan nanoparticles at 2 g/kg of the body weight. Group II was used as the control for the study, as this group was administered with water and food only. The parameters checked were mortality rate, physical appearance, and body weight of the animals. After the time suggested by the OECD guidelines, i.e. 14 days animal's blood was taken for the biochemical analysis and the rats were sacrificed for the histopathological examination of the vital organs.

Statistical Analysis

One-tailed *t*-test was applied for the statistical analysis ($n=3$). The statistical analysis was applied on percentage yield, entrapment efficiency, loading capacity, particle size, PDI, and zeta potential studies. The *p* values were calculated to evaluate the effect of varying concentrations of STPP on the CNPs (CNP1-CNP5).

Results and Discussion

In the present study, DTXL loaded chitosan nanoparticles (CNPs) were prepared by ionic-gelation method by varying ratio of STPP (Table 1), and then characterized for various parameters.

Preparation of Polymeric Nanoparticles

Ionic-gelation method relies on the instituting a sol–gel transition of chitosan in a suitable solution under the threshold of macroscopic gelation. The stated method is a spontaneous and CNPs are formed immediately upon addition of STPP. Previously, a 3:1 volume ratio of Chitosan and STPP as well as the weighed ratio of chitosan and STPP as 3:1 to 6:1, were regarded as best suitable for the said purpose.²⁸ In the current study, the volume ratio of 2.5:1 was used that lies close to the previously stated value and the weight ratio was from 2.5:1 to 7.5:1. Additionally, parameters of stirring speed and time were set at 900 rpm and 35 mins.

Table 1 Composition of Chitosan Loaded Nanoparticles Containing Docetaxel

Formulation Code	Chitosan Concentration ^a	DTXL Concentration ^b	STPP Concentration ^c
CNP1	75 mg/25 mL, i.e. 3 mg/mL	10 mg/3 mL	10 mg/10 mL
CNP2	75 mg/25 mL, i.e. 3 mg/mL	10 mg/3 mL	15 mg/10 mL
CNP3	75 mg/25 mL, i.e. 3 mg/mL	10 mg/3 mL	20 mg/10 mL
CNP4	75 mg/25 mL, i.e. 3 mg/mL	10 mg/3 mL	25 mg/10 mL
CNP5	75 mg/25 mL, i.e. 3 mg/mL	10 mg/3 mL	30 mg/10 mL

Notes: ^aIn 25 mL of 1.5% acetic acid solution in water to produce the concentration of 3mg/mL. ^bIn 3 mL of ethanol. ^cIn 10 mL of distilled water.

Particle Size, PDI, and Zeta Potential Analysis

A size of 200 to 300 nm is regarded as suitable for crossing biological barriers, avoid RES and to escape glomerular filtration.²⁹ Size up to 400 nm is also considered acceptable (evaluated in case of liposomes).³⁰ A study reported suitable size for EPR as 10–500 nm.³¹ In this study, particle sizes of developed CNPs were in nanosize range (172.6 nm to 479.65 nm) and fulfill the requirement of EPR to enhanced antitumor efficacy. The size of nanoparticles was found to be directly related to the varied ratio of chitosan and STPP as increasing the concentration STPP leads to increased particle size ($p < 0.05$). The obtained results were in accordance with the results reported by Sreekumar et al, 2018, who reported that proportions of chitosan and STPP collectively affect the particle size and the size of nanoparticles increases by increasing any of the two components.³² The PDI of the developed nanoparticles was less than 0.5 in all cases. However, relatively more monodisperse systems ($PDI = 0.219 \pm 0.004$) were achieved in CNP3 prepared with 20 mg STPP.³³ PDI increased at both above and below this concentration. Zeta potential of the developed chitosan nanoparticles was found to be 30.45 mV - 35.95 mV. The zeta potential value of greater than 30 mV depicts excellent colloidal stability wherein positive potential is due to the presence of positively charged chitosan in CNPs.^{31,34,35} It can be seen in the results that the zeta potential value decrease by increasing the STPP concentration which is similar to previous reports.³⁶ This may be due to higher degree of gelation of the nitrogen of chitosan to the phosphate of STPP, as reported previously in case of nitrogen of chitosan and phosphate of DNA molecules.³⁷

Entrapment Efficiency and Drug Loading Capacity

The entrapment efficiency of the chitosan/STPP nanoparticles was in the range of 56.39% to 72.73% and loading

efficiency ranged from 7.88% to 9.35% (Table 2). Initially, entrapment efficiency was directly proportional to the concentration of STPP as it increased significantly ($p < 0.05$) by increasing the STPP concentration up-to a certain level, i.e. CNP3 (20 mg/10 mL), which corresponds optimized ionic gelation of the components. Above this value, entrapment efficiency decreased which can be attributed to the shells becoming thicker with the STPP concentration and relative competition between the DTXL and STPP within the nanoparticles.³⁸ A similar response was also observed for the encapsulation of protein in CNPs.³⁹ By increasing drug concentrations, the entrapment efficiency and loading efficiency were decreased.¹³

Scanning Electron Microscopic (SEM) Studies

The scanning electron microscopy showed round shape with the porous and rough surface of two PNP formulations at different magnifications (Figure 1). This is in agreement with previous studies that chitosan PNPs are round in shape and their surface characteristics depend upon the ratio of components.⁴⁰ The morphology of PNPs is also important from pharmacokinetics perspective and round shape PNPs are desirable.^{41,42}

Thermal Analysis (DSC and TGA)

The initial mass loss was due to the moisture loss at 58.01°C and 92.37°C in the physical mixture, and formulation (Figure 2). The differential scanning calorimetric analysis demonstrated that transition of phase from hard solid powder form into plastic flakes takes place at 174.01°C to 299.33°C, whereas, the abrupt declination of weight due to the phase transition temperature of drug was detected at 224.59°C by the thermogravimetric analyzer. The destruction of the DTXL was observed at 419.01°C and 431.80°C by differential scanning calorimetry and thermogravimetric analysis (Figure 2). Phase transition temperature of the physical mixture was observed in the range of

Table 2 Evaluation of Different Formulation Parameters of Docetaxel Loaded Nanoparticles

Formulation Code	Percentage Yield (%)±SD	Entrapment Efficiency (%)±SD	Loading Capacity (%)±SD	Particle Size ±SD	PDI ±SD	Zeta Potential ±SD
CNP1	66.79±0.008	60.124 ± 0.93	9.002 ± 0.14	172.6 ± 4.08	0.319 ± 0.013	35.95 ± 0.69
CNP2	73.79±0.012	66.758 ± 1.31	9.025 ± 0.18	195.3 ± 1.92	0.448 ± 0.012	33.65 ± 0.37
CNP3	77.81±0.022	72.728 ± 1.12	9.347 ± 0.14	270.9 ± 5.31	0.219 ± 0.004	33.0 ± 0.82
CNP4	69.20±0.016	64.022 ± 0.82	9.252 ± 0.12	337.4 ± 2.45	0.398 ± 0.016	33.5 ± 0.57
CNP5	71.57±0.005	56.393 ± 0.61	7.880 ± 0.09	479.65 ± 11.47	0.487 ± 0.011	30.45 ± 0.37

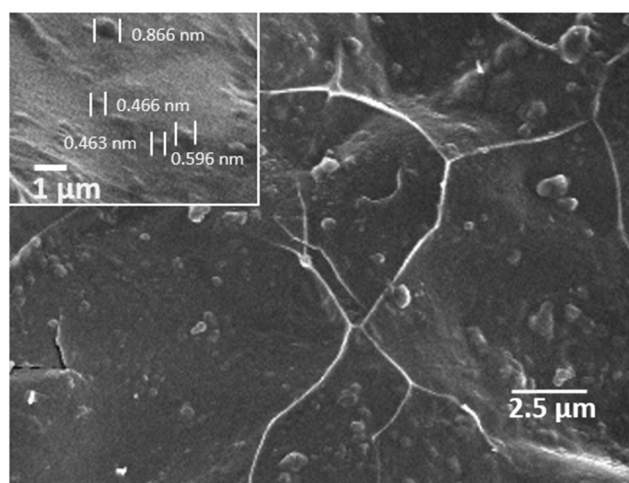


Figure 1 Scanning electron micrographs of docetaxel loaded chitosan nanoparticles.

260.75–320.27°C. Likewise, the drug peak of phase transition temperature was also found in the physical mixture at 261.75°C of TGA. The destructive peak in the DSC graph of the physical mixture was almost identical to the drug. The formulation showed phase transition temperature in the

range of 234.90–323.34°C in DSC analysis and did not show prominent drug peak. Likewise, the TGA graph of the PNPs showed the DTXL peak with slight shifting. The absence of drug peak in DSC and slight shifting in TGA reveals the efficient entrapment of the drug within the nanoparticulate system and thermal stability of the developed nanoparticles as shown in Figure 2.⁴³

Powdered X-Ray Diffraction Analysis (PXRD)

The distinct and sharp peaks of DTXL, chitosan, and STPP were observed which represented their crystalline nature with insignificant amorphous contents (Figure 3) in agreement with previous literature on chitosan-based PNPs. Likewise, the distinct and sharp peaks of DTXL and STPP were also observed in the physical mixture in the range of 11–17° and 33–37°, respectively. The sharp peaks of DTXL were also found in the formulation CNP3 in the range of 11–17° that depicted the crystalline nature of the formulation as shown in Figure 3.⁴⁴

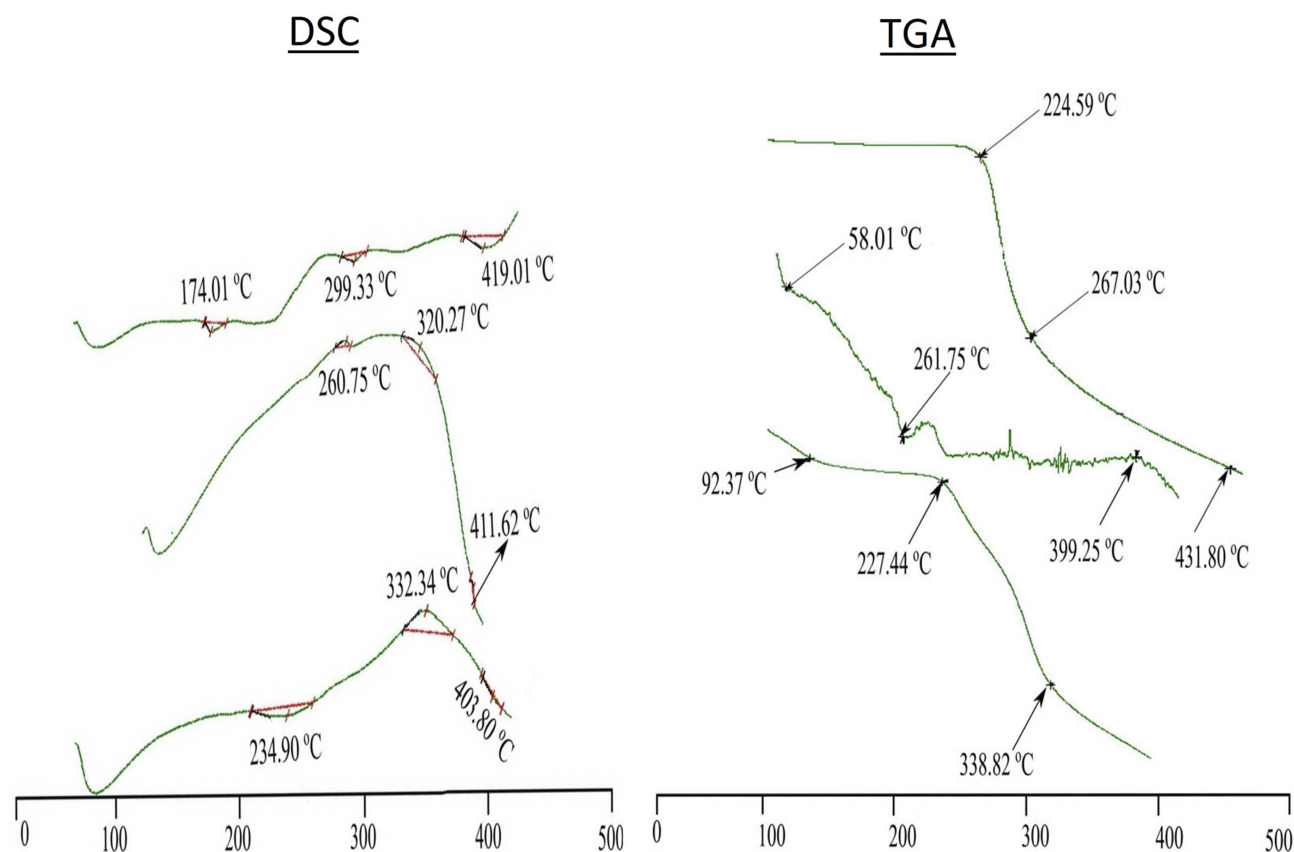


Figure 2 DSC and TGA (Drug, physical mixture, formulation CNP3).

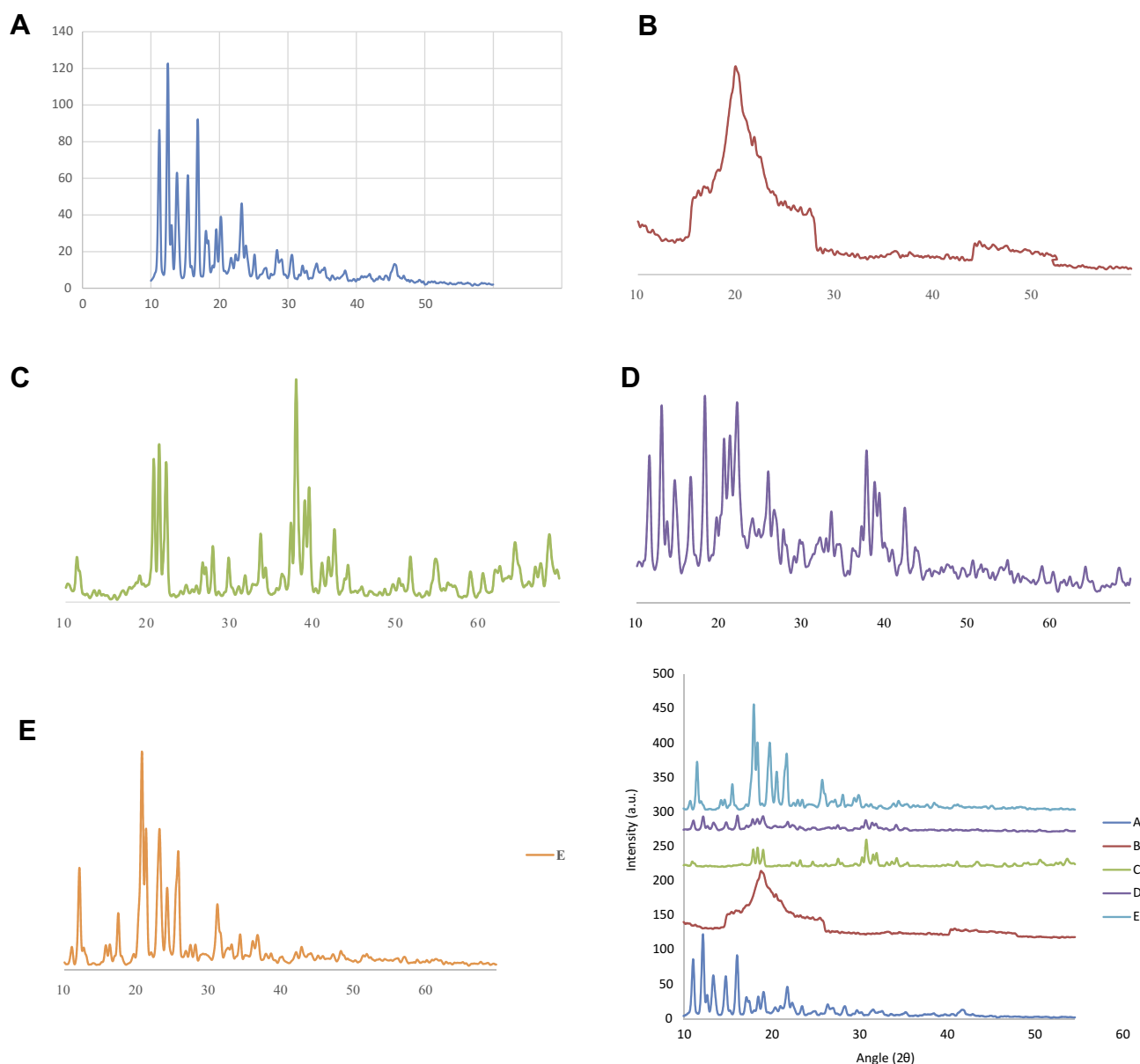


Figure 3 XRD of DTXL (A), chitosan (B), STPP (C), physical mixture (D) and formulation CNP3 (E).

Fourier Transform Infrared Spectroscopy (FTIR) Analysis

DTXL showed characteristic peaks at 3477 cm^{-1} mimicking with N-H stretching of alkanes,²⁵ 2990 cm^{-1} due to C-H stretching,⁴⁵ 1713 cm^{-1} due to C=O stretching,²⁵ 1490 cm^{-1} associated with C=C stretching,⁴⁵ and 1246 cm^{-1} indicated the C-N stretching vibration of DTX (Figure 4).⁴⁶ The characteristic peaks of chitosan were observed at 3296 cm^{-1} depicted stretching vibration of -OH groups, 1644 cm^{-1} described C=O of acetamide group,^{47,48} 1526 cm^{-1} represented -NH bending vibration of NH_2 group⁴⁸ and 1063 cm^{-1} showed -CN and -CO stretching

vibration.⁴⁷ The characteristic peaks of STPP at the wavenumber of 1129 cm^{-1} provided the recognition of the occurrence of phosphate group.^{49,50} Physical mixture also showed characteristic peaks of drug and STPP with slight shifting at 2900 cm^{-1} and 1711 cm^{-1} , respectively. The PNPs showed the peak at 2767 cm^{-1} that might be due to the overlapping of the wavenumber peaks between drug and chitosan. Similarly, PNPs also showed the characteristic drug peak of STPP at 1708 cm^{-1} .

The presence of peaks of DTXL in the physical mixture and optimized formulation (CNP3) ensures the absence of chemical interaction of the drug with formulation components and stability of the developed PNPs.⁵¹ Together with

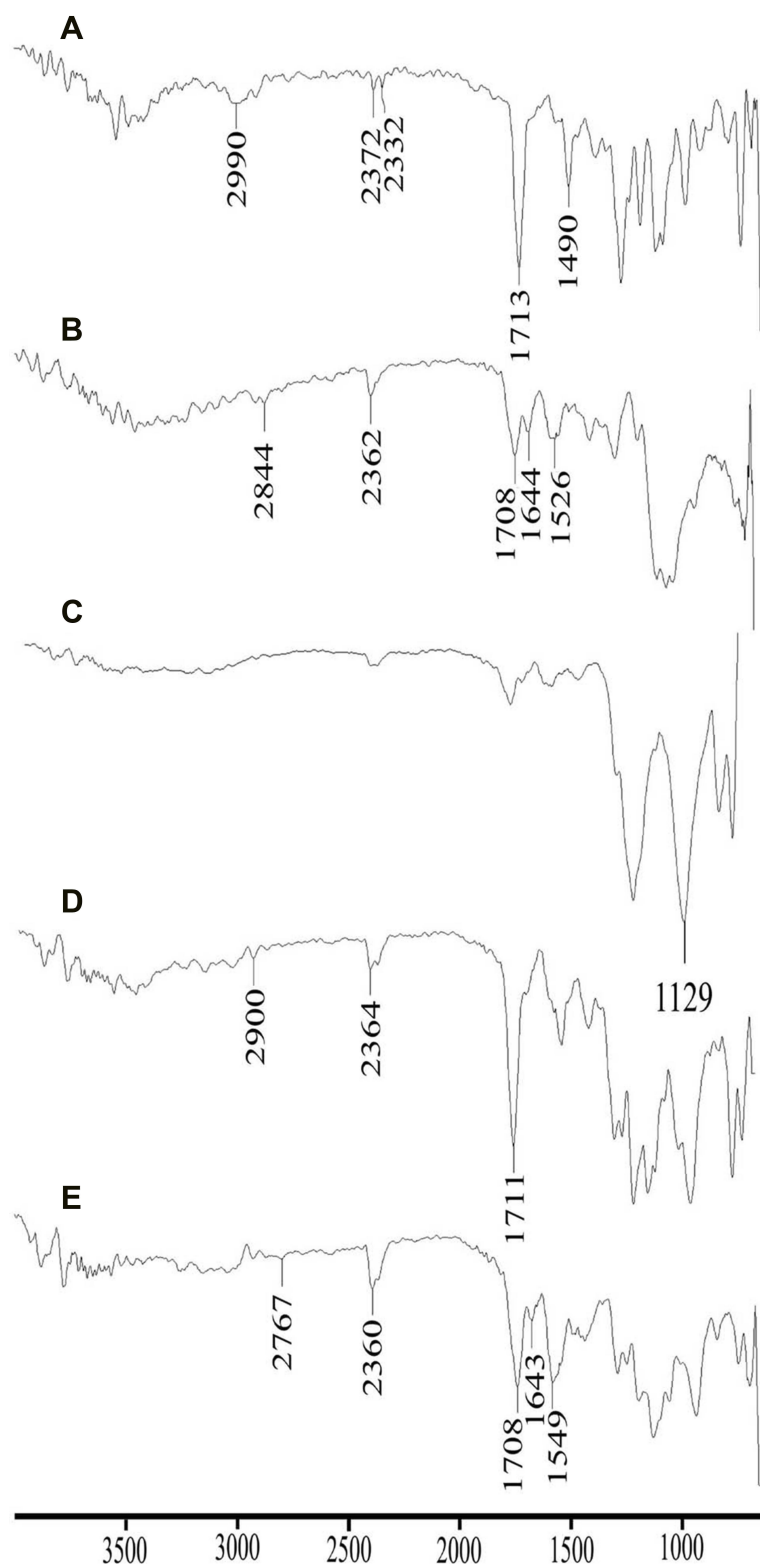


Figure 4 Comparative FTIR spectra of DTXL (A), chitosan (B), STPP (C), physical mixture (D) and formulation CNP3 (E).

the results of zeta potential, PXRD and FTIR, it was assumed that the PNP formulations prepared in this study

are stable and will also form a stable colloidal dispersion which is a prerequisite for biomedical applications.

In-vitro Drug Release Studies and Kinetic Modeling

Cumulative drug release over 24 hrs study period was found to be in the range of 51.57% (CNP5) to 69.93% (CNP3) which depicts a sustained release behavior (Figure 5). Initially, the drug release is faster³¹ due to the rapid dissolution of surface entrapped/adhered DTXL followed by a second sustained release phase.⁵² The final slower release phase may correspond to the release of DTXL from the core as a result of chitosan hydration and swelling. Higher values of the correlation coefficient are observed for the Korsmeyer-Peppas model so it was concluded that the release of DTXL from CNPs predominantly followed the Korsmeyer-Peppas model. According to this model, the value of release exponent “n” was below 0.45 that depicts drug release from CNPs by diffusion mechanism (Table 3). Comparable results of the drug release mechanism were previously reported.⁵³

Acute Oral Toxicity Study Evaluation

Improved tolerability of potent drugs is the main advantage of different nanoscale drug delivery systems available in market.⁵⁴ Also, the drug delivery systems must lack the toxicity of their own.²⁹ Therefore, the acute oral toxicity studies were performed as shown in Table 4.

Biochemical Analysis of Blood

The components used for the synthesis of polymeric nanoparticles including polymer and surfactant formed by solvent evaporation technique undergo various challenges when introduced into the body. Besides releasing the drug in-vivo, the formulation components might leach

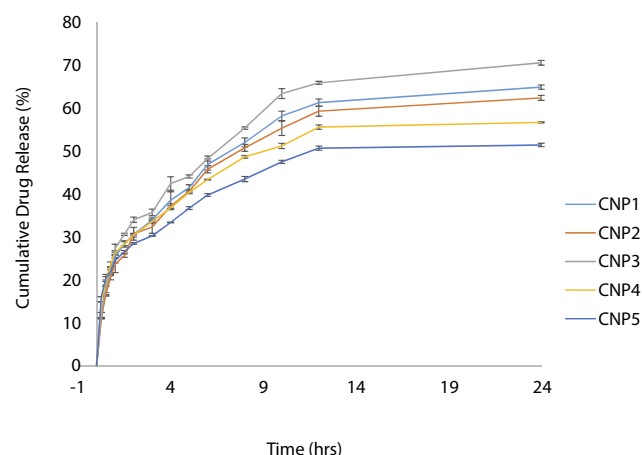


Figure 5 Cumulative drug release of polymeric nanoparticles.

Table 3 Kinetic Modeling of Drug Release Profile of Chitosan Nanoparticles

Formulation Code	Zero Order	First Order	Higuchi Model	Korsmeyer-Peppas Model	
	R ²	R ²	R ²	R ²	n
CNP1	0.1796	0.6131	0.8573	0.9734	0.338
CNP2	0.1639	0.5962	0.8616	0.9734	0.340
CNP3	0.1598	0.6739	0.8624	0.9744	0.340
CNP4	0.7360	0.2047	0.7018	0.9769	0.279
CNP5	0.9048	0.0139	0.6478	0.9786	0.264

Table 4 Clinical Observations of an Acute Oral Toxicity Study for Prepared Chitosan Loaded Nanoparticles

Observation	Group I (Treated with CNPs) Mean ± SD	Group II (Control) Mean ± SD
Signs of illness	Nil	Nil
Body Weight (in grams)		
• Day 1	172.42 ± 0.42	167.25 ± 0.35
• Day 7	193.08 ± 0.31	197.17 ± 0.31
• Day 14	206.83 ± 0.47	207.83 ± 0.66
Any visible skin irritation/ toxicity	Nil	Nil
Ocular toxicity	Nil	Nil
Mortality	Nil	Nil

out of the formulation. Therefore, toxicity study was carried out to check the various parameters including hematological, biochemical and histopathological aspects. The study was conducted using the OECD guideline 420.

For the hematological analysis, blood samples were collected from the rats via retro-orbital plexus, immediately transferred to EDTA (ethylene diamine tetraacetate) tubes to prevent coagulation of collected blood. The parameters checked in the hematological examination are given in Table 5. Hematological values of CNPs treated rats were comparable with control group which depicts acceptable biocompatibility and bio acceptance of developed formulations (Table 5).

Liver and Kidney Function Tests

The liver and kidney function tests of both groups have been presented in Table 6. ALT, alkaline phosphate, bilirubin, AST, urea, and creatinine showed comparable results for test and control group depicting the safety and biocompatibility of developed CNPs.

Table 5 Biochemical Blood Analysis

Hematology	Group I (CNPs) Mean $\pm$ SD	Group II (Control) Mean $\pm$ SD
Hemoglobin (g/dL)	13.3 $\pm$ 0.01	13.20 $\pm$ 0.14
Red blood cells $\times 10^6$ (μ L)	6.99 $\pm$ 0.07	6.94 $\pm$ 0.05
Platelets $\times 10^3$ (μ L)	924.75 $\pm$ 3.96	928.50 $\pm$ 4.72
White blood cells $\times 10^3$ (μ L)	7.58 $\pm$ 0.38	7.93 $\pm$ 0.33
Lymphocytes (%)	89.08 $\pm$ 1.06	88.38 $\pm$ 0.41
Neutrophils (%)	8.23 $\pm$ 0.59	8.75 $\pm$ 0.38
Mean corpuscular volume (fL)	67.38 $\pm$ 0.71	67.83 $\pm$ 0.48
Mean corpuscular hemoglobin concentration (g/dL)	28.34 $\pm$ 0.03	28.41 $\pm$ 0.06
Mean corpuscular hemoglobin (pg)	18.95 $\pm$ 0.11	19.20 $\pm$ 0.22

Note: All values are given as mean plus or minus the standard deviation (n=3).

Table 6 Liver and Kidney Function Tests

Liver and Kidney Tests	Group I (CNPs) Mean $\pm$ SD	Group II (Control) Mean $\pm$ SD
ALT (U/L)	73.88 $\pm$ 0.74	73.13 $\pm$ 1.88
Alkaline phosphatase (g/dL)	115.75 $\pm$ 3.77	113.25 $\pm$ 4.66
Bilirubin (mg/dL)	0.31 $\pm$ 0.01	0.37 $\pm$ 0.02
AST (U/L)	122 $\pm$ 2.24	123 $\pm$ 2.45
Urea (mg/dL)	46.75 $\pm$ 2.28	45.75 $\pm$ 1.92
Creatinine (mg/dL)	0.38 $\pm$ 0.06	0.41 $\pm$ 0.04

Note: All values are given as mean plus or minus the standard deviation (n=3).

Histopathological Examination

Histological examination is used to visualize any unusual change in biological tissues due to toxicity.⁵⁵ In this study, no animal died during the study as shown in Figure 6. Furthermore, no pathological change, lesion, disruption or deformation were observed in the vital organs of the control as well as group I administered with CNPs. The lack of abnormal changes also shows the safety and compatibility in agreement with previous results.⁵⁶ Therefore, the result of our acute oral toxicity study indicated that the CNPs developed were not toxic to Wistar rats and are acceptable for the delivery of various drugs.⁵⁷ Results are in agreement with previous findings while evaluation of CNPs safety with the aim to deliver quercetin.⁵⁸

Conclusion

The docetaxel loaded polymeric nanoparticles were successfully developed by the ionic-gelation method and characterized for post-formulation parameters including physicochemical analysis, in-vitro drug release, and acute oral toxicity study. All the CNP formulations showed desirable size with good mono-dispersity, EE and LC depicts the effect

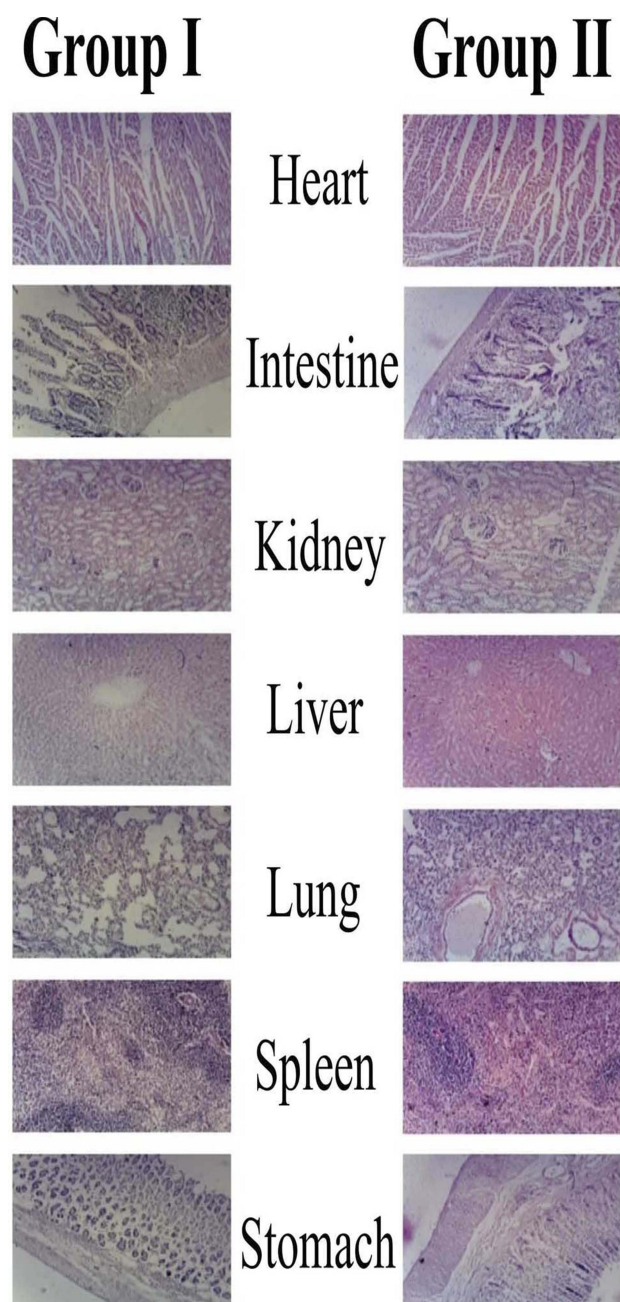


Figure 6 Histopathological microscopic examination of different tissues of wistar rats of group-I (test), and group-II (control).

of STPP concentration, in-vitro drug release studies showed docetaxel release up-to 24 hrs. DSC, TGA, PXRD, and FTIR studies confirmed thermal stability, physical state, and compatibility of DTXL with formulation components. Acute oral toxicity studies showed that chitosan-based PNPs were non-toxic and biocompatible for the delivery of docetaxel. Therefore, the chitosan NPs provide a potential drug delivery platform for the delivery of DTXL for the treatment of various types of cancers.

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Disclosure

The authors declare no conflicts of interest in this work.

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