

# Gluconokinase IDNK Promotes Cell Proliferation and Inhibits Apoptosis in Hepatocellular Carcinoma

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**Purpose:** Hepatocellular carcinoma (HCC) is one of the deadliest cancers globally with a poor prognosis. Breakthroughs in the treatment of HCC are urgently needed. This study explored the role of IDNK in the development and progression of HCC.

**Methods:** *IDNK* expression was suppressed using short hairpin (shRNA) in BEL-7404 and Huh-7 cells. The expression of *IDNK* in HCC cells after *IDNK* knockdown was evaluated by real-time quantitative RT-PCR analysis and Western blot. After *IDNK* silencing, the proliferation and apoptosis of HCC cells were evaluated by Celigo cell counting, flow cytometry analysis, MTT assay, and caspase3/7 assay. Gene expressions in BEL-7404 cells transfected with *IDNK* shRNA lentivirus plasmid and blank control plasmid were evaluated by microarray analysis. The differentially expressed genes induced by deregulation of *IDNK* were identified, followed by pathway analysis.

**Results:** The expression of *IDNK* at the mRNA and protein levels was considerably reduced in shRNA *IDNK* transfected cells. Knockdown of *IDNK* significantly inhibited HCC cell proliferation and increased cell apoptosis. A total of 1196 genes (585 upregulated and 611 downregulated) were differentially expressed in *IDNK* knockdown BEL-7404 cells. The pathway of tRNA charging with Z-score = -3 was significantly inhibited in BEL-7404 cells with *IDNK* knockdown.

**Conclusion:** *IDNK* plays a key role in the proliferation and apoptosis of HCC cells. *IDNK* may be a candidate therapeutic target for HCC.

**Keywords:** hepatocellular carcinoma cells, shRNA *IDNK*, cell proliferation, cell apoptosis, microarray, differentially expressed gene

## Introduction

Hepatocellular carcinoma (HCC) is the main type of primary liver cancer and is the leading cause of cancer-related death in patients with cirrhosis.<sup>1</sup> HCC is one of the most common cancers in Southeast Asia and it is more commonly diagnosed in men than women. Chronic liver disease is the main cause for HCC. Other risk factors include viral hepatitis, aflatoxin, alcoholism, and other diseases.<sup>2-4</sup> An estimated 662,000 deaths are attributed to HCC annually. Of these global deaths, half are in China.<sup>5</sup> HCC is the one of the deadliest tumors in China and 90% of cases involve hepatitis virus B infection. The prognosis for HCC patients is poor. The median survival time of 3 months several decades ago has been improved to 1 year with advancements in therapy.<sup>6</sup> HCC is an increasing global burden and breakthrough treatment management strategies are urgently needed.

Gluconate is a glucose derivative that is abundant in fruits, honey, vegetables, and wine. Clinically, gluconate can be used as a neutral carrier of ions for treatment

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of ion deficiencies and can be combined with other drugs as an antiseptic in surgery.<sup>7,8</sup> Citrate is reportedly a central substrate that is beneficial for metabolism of cancer cells. Citrate is taken up by cancer cells through plasma membrane citrate transporter (pmCiC).<sup>9</sup> Recent evidence indicated an inhibitory effect of gluconate on tumor growth by competitive blocking of pmCiC.<sup>10</sup> Gluconokinase has a key role in gluconate metabolism in humans.<sup>11</sup> The *IDNK* gene encodes human gluconokinase, which is involved in the phosphorylation of gluconate.

The present study was grounded in our idea that inhibition of *IDNK* expression could prevent gluconate metabolism and promote the role of gluconate in suppressing cancer growth. However, the role of *IDNK* in cancer growth has not been clarified. Presently, we first explored the potential role of *IDNK* in HCC cell lines. *IDNK* knockdown in BEL-7404 and Huh-7 cells was performed using lentivirus vectors encoding short hairpin RNA (shRNA). The proliferation and apoptosis of HCC cells were measured after *IDNK* knockdown. Gene expression in BEL-7404 cells with *IDNK* knockdown was measured using a microarray. The differentially expressed genes (DEGs) induced by *IDNK* inhibition were identified, followed by the prediction of the related pathways. The findings should inform new liver cancer treatments.

## Materials and Methods

### RNA Interference (RNAi) Recombinant Plasmid Construction and Lentivirus Packaging

The small interfering RNA (siRNA) of *IDNK* was designed and synthesized by Shanghai Jirui Biological Engineering Co. Ltd. (Shanghai, China). The target sequences were sense 5' AAACAGAACTAAGCATAAA 3' and anti-sense 5' TTTATGCTTAGTTCTGTTT 3'. *IDNK* shRNA was inserted into the GV115 vector using *AgeI* and *EcoRI* restriction enzymes and T4 DNA Ligase (Fermentas, Waltham, MA, USA). The recombinant plasmids were transferred into competent *Escherichia coli* cells. Positively transfected cells were selected by growth in Luria-Bertani medium supplemented with ampicillin. The effective *IDNK* shRNA plasmid was determined by PCR analysis and isolated using the EndoFree<sup>®</sup> Maxi Plasmid Kit (Qiagen, Carlsbad, CA, USA) according to the manufacturer's instructions.

### Lentiviral Infection

The BEL-7404 and Huh-7 HCC cell lines obtained from Shanghai Cancer Institute (Shanghai, China) were maintained

in RPMI 1640 medium (Gibco, Grand Island, NY, USA) supplemented with 10% fetal bovine serum at 37°C in an atmosphere of 5% CO<sub>2</sub>. Cells were seeded in wells of six-well plates (2×10<sup>5</sup> cells/well) and infected with lentivirus containing either *IDNK* shRNA plasmid or blank control plasmid in the shCtrl and shIDNK group, respectively. After lentiviral infection for 72 h, positive infection efficiency was observed by fluorescence microscopy using a model IX71 microscope (Olympus, Tokyo, Japan).

### RNA Isolation and Real-Time Quantitative PCR Analysis

Total RNA was extracted from HCC cells using Trizol reagent (Invitrogen, Carlsbad, CA, USA). cDNA was synthesized by reverse transcription with 2 µg RNA using an M-MLC kit according to the manufacturer's instructions (Promega, Madison, WI, USA). The amplification reaction was performed in a 12-µL reaction system containing 0.3 µL primer mix (5 µM), 0.6 µL cDNA, 6.0 µL SYBR premix ex taq, and 5.1 µL RNase-Free water. The primer sequences were: *IDNK* forward primer 5' GAAAAGGCATACCGCTCAAT-3' and reverse primer 5'-GAGATGACCTCAAACGACCC-3'; *glyceraldehyde-3-phosphate dehydrogenase (GAPDH)* forward primer 5'-TGACTTCAACAGCGACACCCA-3' and reverse primer 5'-CACCTGTTGCTGTAGCCAAA-3'. Real-time PCR was performed using the LightCycler 480 instrument (Roche, Mannheim, Germany). The reaction conditions were 95°C for 30 s and 40 cycles of 95°C for 5 s and 60°C for 30 s. The *IDNK* gene expression relative to *GAPDH* was analyzed by the 2<sup>-ΔΔCT</sup> method.<sup>12</sup>

### Western Blot Analysis

After infection, cells in shCtrl and shIDNK groups were lysed using 2× Lysis Buffer and the supernatant was collected by centrifugation at 12,000 ×g, 4°C for 15 min. The protein concentration was measured by the BCA Protein Assay Kit (P0010S; Beyotime, Shanghai, China). A total of 20 µg of protein was used for 10% SDS-PAGE. The resolved proteins were transferred to a polyvinylidene fluoride (PVDF) membrane (IPVH00010; Millipore, Billerica, MA, USA). Each membrane was blocked with 5% skim milk and incubated with the primary antibodies. The antibodies and dilutions used were: mouse anti-IDNK (1:2000, F1804; Sigma-Aldrich, St. Louis, MO, USA), and mouse anti-GAPDH (1:2000, sc-32233; Santa Cruz Biotechnology, Dallas, TX, USA) at 4°C overnight. This was followed by incubation with secondary antibody (goat anti-mouse IgG,

1:2000, sc-2005; Santa Cruz Biotechnology) for 2 h at room temperature. The immunoreactive bands were visualized by electrochemiluminescent (ECL) staining (M3121/1859022; Thermo Fisher Scientific, Waltham, MA, USA).

## Cell Growth Assay

After infection for 2 h, cells were collected and seeded in wells of 96-well plate (1000 cells/well) at 37°C in an atmosphere of 5% CO<sub>2</sub>. The number of surviving cells was counted daily beginning after 24 h incubation for 5 consecutive days. The cell growth curves were constructed by measuring green fluorescence emitted from cells using a Celigo Imaging Cytometer (Nexcelom, Lawrence, MA, USA).

## Flow Cytometry Analysis

Flow cytometry allows the evaluation of the cell cycle and apoptosis in different models. The infected cells were seeded in wells of 6-well plate (2 mL/well) and maintained in an incubator at 37°C in an atmosphere of 5% CO<sub>2</sub>. When the cells grew to 85% confluence, they were collected by centrifugation at 1300 rpm for 5 min at 4°C and washed three times using cooled D-Hanks solution (pH 7.2~7.4). The cells were stained with propidium iodide (PI) or Annexin V to determine cell cycle distribution and cellular apoptosis by flow cytometry using a Guava easyCyte HT flow cytometer (Millipore).

## MTT (3-[4, 5-Dimethylthiazol-2-Yl]-2, 5-Diphenyl Tetrazolium Bromide) Cell Viability Assay

The viability of cells infected with IDNK shRNA lentivirus plasmid and blank control lentivirus plasmid was evaluated using the MTT assay. Briefly, the infected cells (1500 cells/well) were seeded in wells of a 96-well plate and maintained for 5 days. Each day, 20 µL 5 mg/mL MTT was added to each well and incubated for 4 h. The cell supernatants were removed and 100 µL dimethyl sulfoxide (DMSO) was added to each well. After 5 min, the optical density (OD) of cells was detected at 540 nm using a M2009PR microplate reader (Tecan, Männedorf, Switzerland).

## Caspase 3/7 Assay

The active caspase 3/7 in infected cells was evaluated using the Caspase-Glo<sup>®</sup> 3/7 assay kit (Promega). Huh-7 cells (1×10<sup>4</sup> cells/well) infected with recombinant plasmid for 3 days in shCtrl and shIDNK group were seeded in wells of 96-well plate at 37°C in an atmosphere of 5%

CO<sub>2</sub>. After adding 100 µL Caspase-Glo reagent to each well, the plate was shaken at 300 rpm for 30 min and then incubated for 2 h at 20°C. Caspase 3/7 activity was measured using the aforementioned plate reader (Tecan).

## Microarray Data Analysis

Gene expression in BEL-7404 cells infected with IDNK shRNA lentivirus plasmid (KD) and blank control lentivirus plasmid (NC) was measured using the PrimeView GeneChip (Affymetrix, Santa Clara, CA, USA) according to the manufacturer's protocol. The arrays were scanned using a GeneChip Scanner with the use of GeneChip Operating Software. DEGs were sorted by empirical Bayes procedure with |Fold Change|≥1.3 and False discovery rate (FDR)<0.05. Ingenuity Pathway Analysis (IPA) was performed to globally analyze the DEGs in the pathways.

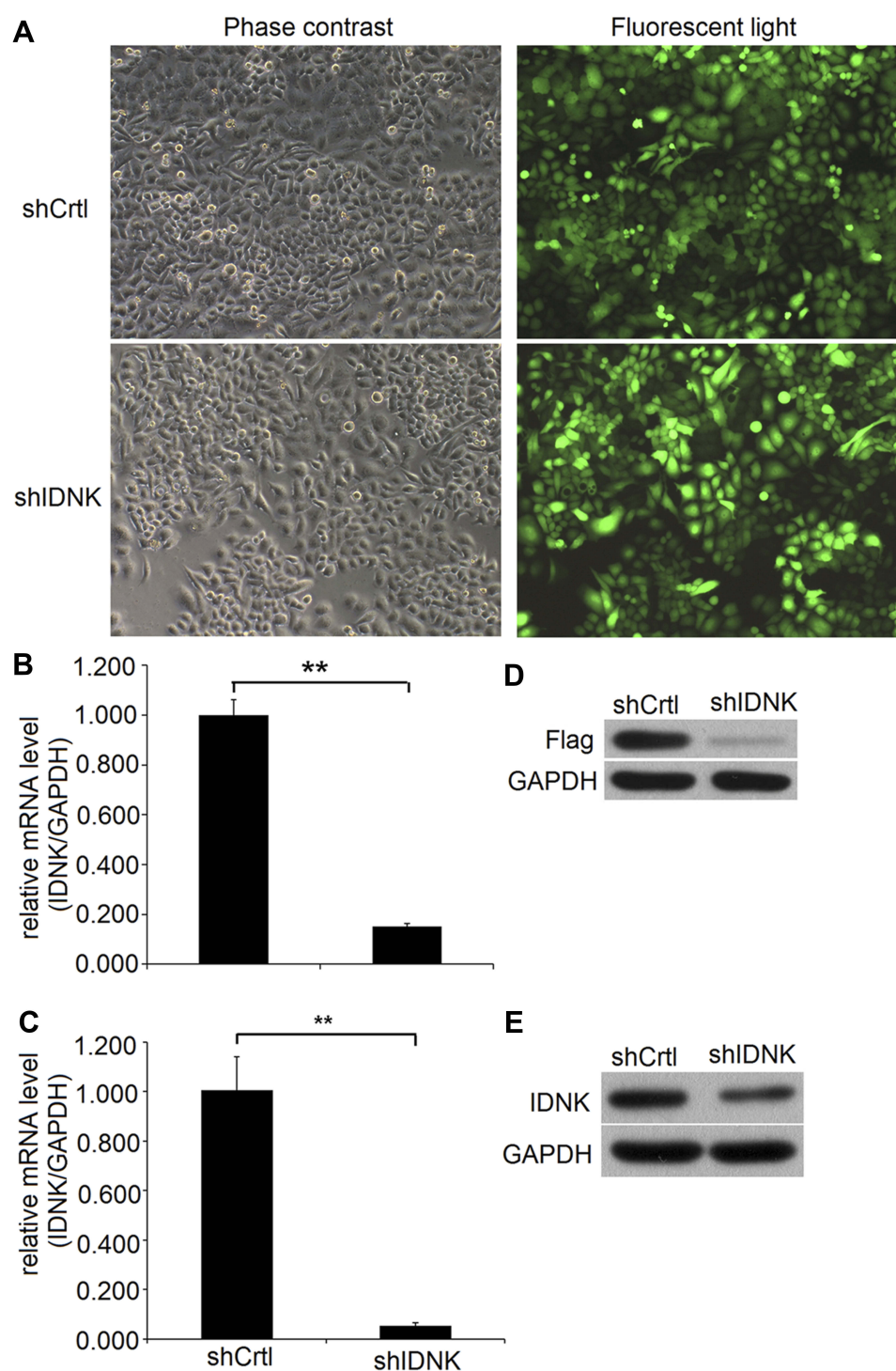
## Statistical Analysis

All the data are expressed as mean ± standard deviation (STDEV). Data were analyzed using SPSS software (SPSS, Inc., Chicago, IL, USA). The comparison between groups was performed by *T*-test. Differences with a *P*-value <0.05 were considered significant.

## Results

### Knockdown of IDNK in HCC Cell Lines

After cells were infected with the recombinant lentivirus shRNA IDNK plasmid for 72 h the infective efficiency was determined to be more than 80% by observing green fluorescent protein using a fluorescence microscope (Figure 1A). The effect of shRNA IDNK on BEL-7404 and Huh-7 cells was determined by real-time quantitative PCR analysis and Western blot. After infection, the relative expression of IDNK gene in BEL-7404 cells in the shIDNK group (0.15±0.01) was considerably decreased compared to the shCtrl group (1.00 ± 0.06) (*p*<0.001) (Figure 1B). The relative IDNK expression in Huh-7 cells was 1.00 ± 0.14 in shCtrl group and was significantly less in the shIDNK group (0.05 ± 0.01) (*p*=0.006) (Figure 1C). As shown in Figure 1D and E, the protein expression of IDNK was obviously reduced in the shIDNK group of the two cell lines. The expression of IDNK at the mRNA and protein levels was considerably reduced in shRNA IDNK infected cultures, indicating the effective knockdown of IDNK target sequencing.



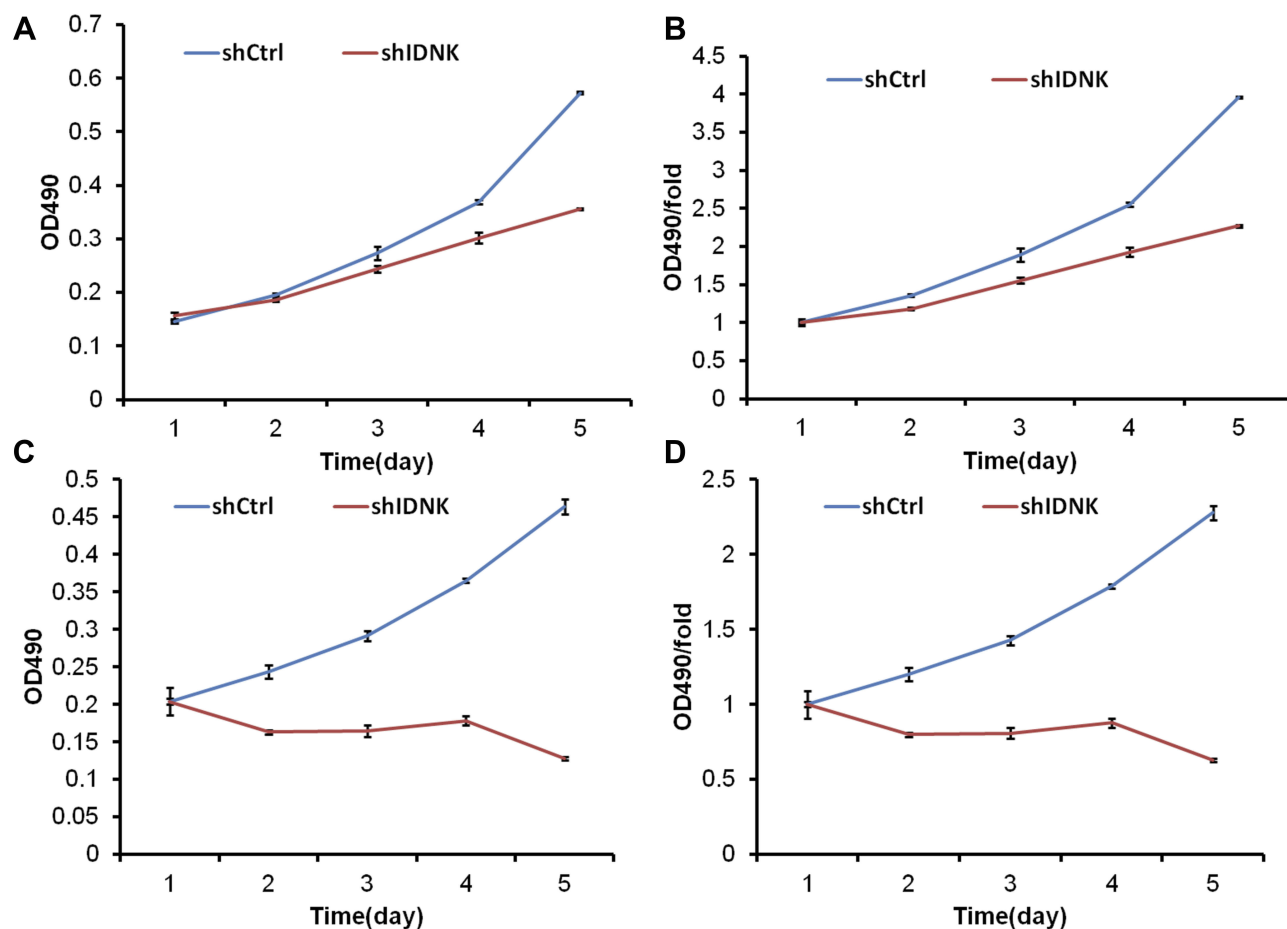
**Figure 1** The efficiency of infection of HCC cells with shIDNK and shCtrl lentivirus plasmid. **(A)** The expression of *IDNK* in BEL-7404 cells by real-time quantitative RT-PCR analysis. **(B)** The expression of *IDNK* in Huh-7 cells by real-time quantitative RT-PCR analysis. **(C)** The expression of *IDNK* protein in BEL-7404 cells by Western blot analysis. **(D)** The expression of *IDNK* protein in Huh-7 cells by Western blot analysis **(E)**. \*\* $P < 0.01$ .

## Knockdown of IDNK Inhibits HCC Cell Proliferation

The viability of BEL-7404 IDNK knockdown cells was determined using the MTT assay. The growth rate of both

BEL-7404 and Huh-7 cells in the shRNA IDNK group was slower than that in the shCtrl group at days 3, 4, and 5 after plasmid infection (Figure 2). The OD values of BEL-7404 cells in shIDNK group were  $0.302 \pm 0.010$  at day 4 and  $0.356$





**Figure 2** HCC cell proliferation is significantly inhibited by *IDNK* knockdown in BEL-7404 cells (**A, B**) and in Huh-7 cells (**C, D**). The optical density (OD) value of surviving cells was measured in the MTT assay after *IDNK* knockdown for consecutive 5 days.

$\pm 0.002$  at day 5, which were significantly lower than the value in the shCtrl group ( $0.369 \pm 0.0045$  at day 4 and  $0.573 \pm 0.0017$  at day 5, both  $p < 0.001$ ) (Figure 2A and B). The fold-changes of the OD values in BEL-7404 cells were also significantly reduced at day 4 ( $1.924 \pm 0.0611$ ) and day 5 ( $2.267 \pm 0.0125$ ) following infection (both  $p < 0.001$ ) (Figure 2C and D).

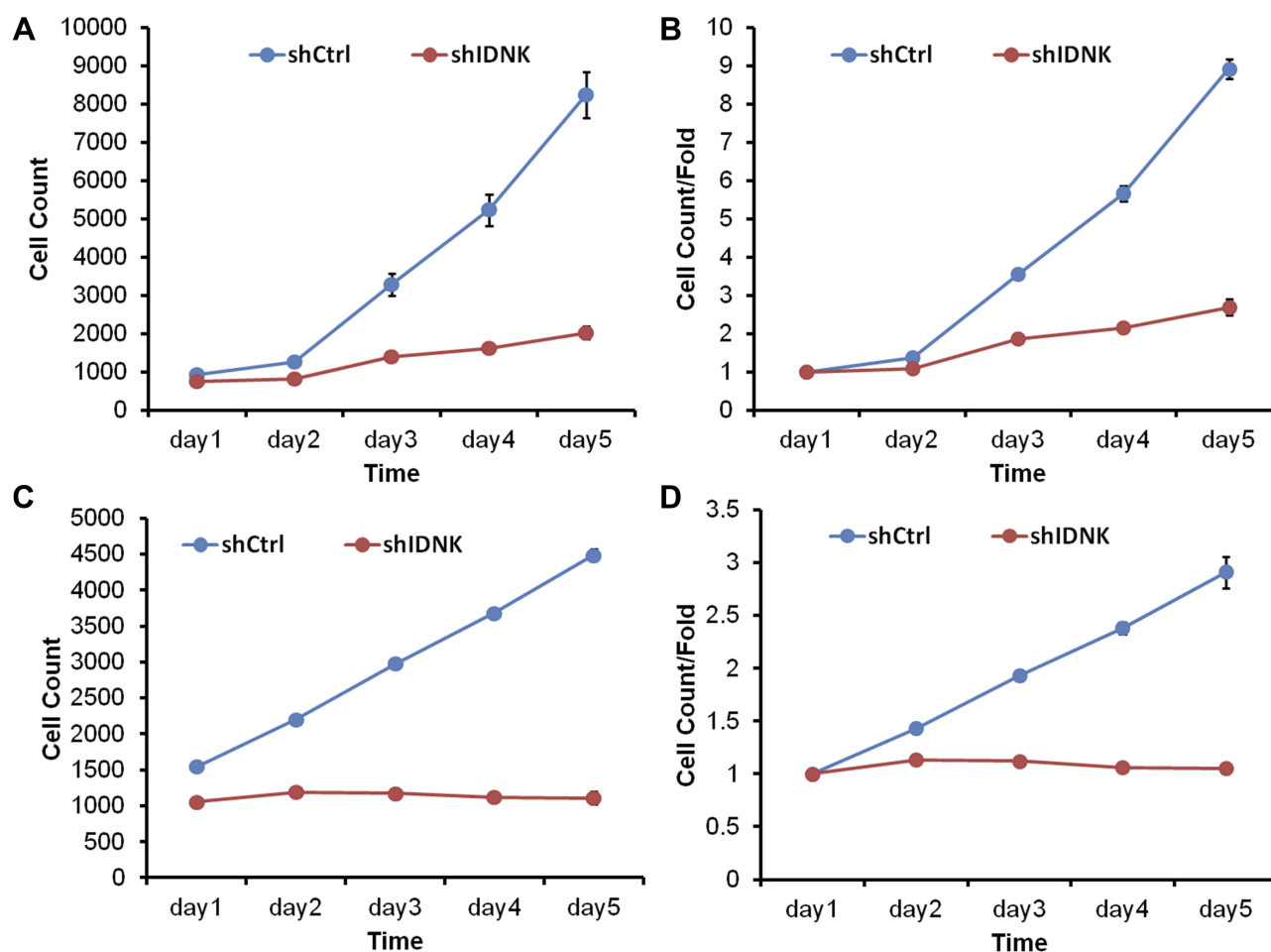
Similar findings were observed by Celigo cell counting (Figure 3). The growth curve of cells revealed that compared with the shCtrl group, the cell count and growth fold-change of BEL-7404 and Huh-7 cells following *IDNK* knockdown were significantly decreased at day 4 and 5 (all  $p < 0.001$ ). These results indicated that *IDNK* may play a key role in promoting the proliferation of HCC cells.

In addition, PI flow cytometry assay showed that the percentage of BEL-7404 cells in the shIDNK group was considerably decreased in the S phase ( $18.43 \pm 0.693\%$  vs.  $30.23 \pm 0.771\%$ ) and significantly increased in G1 ( $63.61 \pm 0.449\%$  vs.  $57.31 \pm 0.186\%$ ) and G2/M phase ( $17.91$

$\pm 1.094\%$  vs.  $12.45 \pm 0.6227\%$ ), compared to the shCtrl group (all  $p < 0.05$ ) (Figure 4A and B). Similarly, Huh-7 cells following shRNA *IDNK* plasmid infection were significantly decreased in S phase ( $18.43 \pm 0.694\%$  vs.  $30.23 \pm 0.771\%$ ) and remarkably increased in G1 ( $63.61 \pm 0.499\%$  vs.  $57.31 \pm 0.186\%$ ) and G2/M phase ( $17.96 \pm 1.094\%$  vs.  $12.45 \pm 0.623\%$ ) (all  $p < 0.05$ ) (Figure 4C and D). The results suggested that knockdown of *IDNK* inhibited the cell cycle in S phase.

## Knockdown of *IDNK* Promotes HCC Cell Apoptosis

The apoptosis of BEL-7404 and Huh-7 cells after *IDNK* knockdown was evaluated by Annexin V staining and caspase 3/7 assay. After 5 days of infection, the apoptosis rate of BEL-7404 cells in shIDNK group ( $34.03 \pm 0.063\%$ ) was significantly higher than that in shCtrl group ( $4.24 \pm 0.170\%$ ) ( $p < 0.05$ ) (Figure 5A). To further verify the effect of *IDNK* on cell apoptosis, the activity of caspase 3/7 was evaluated



**Figure 3** Cell growth curves of HCC cells after IDNK knockdown by Celigo cell counting in BEL-7404 cells (A, B) and in Huh-7 cells (C, D). The surviving cells with green fluorescence were counted using a Celigo Imaging Cytometer.

in Huh-7 cells after infection. As shown in Figure 5B, Huh-7 cells infected with shRNA IDNK plasmid exhibited remarkable caspase 3/7 activity ( $30,681.67 \pm 418.073$ ), compared with that in cells infected with blank control plasmid ( $11,843 \pm 418.07$ ) ( $p < 0.001$ ). The results suggested that the IDNK was important in the regulation of cell viability.

## DEGs and Validation

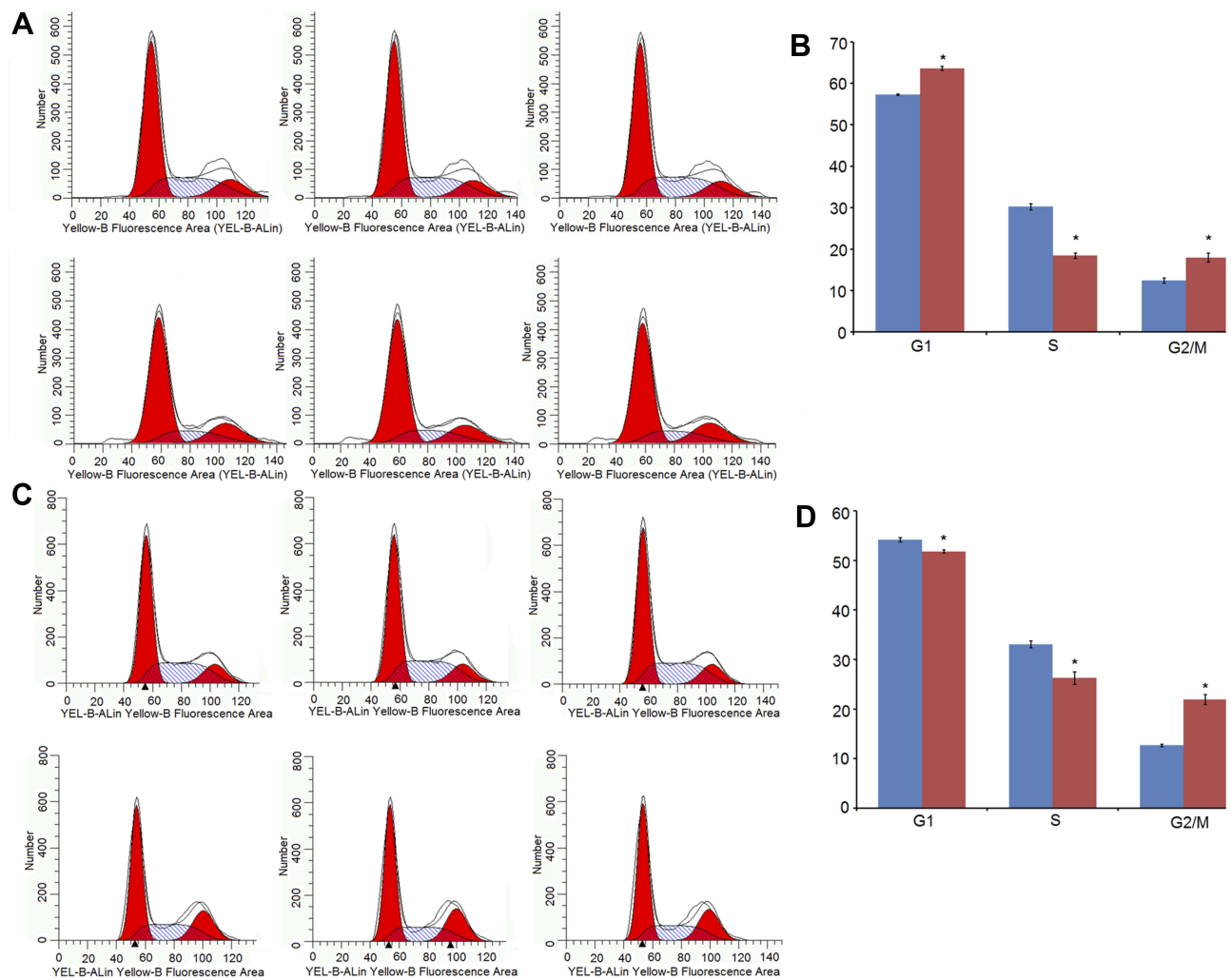
Based on microarray data, a total of 1196 genes (585 up-regulated and 611 down-regulated) were identified as DEGs in BEL-7404 cells with IDNK knockdown compared with cells infected with blank control plasmid. Hierarchical clustering showed that the samples between groups could be clearly distinguished by DEGs (Figure 6A). The differential expression of significant genes was validated by PCR analysis and Western blot. The results suggested that the differential expressions of genes at the mRNA and protein levels were relatively consistent with the microarray data (Supplemental Figure 1A and B).

## Canonical IPA Pathways

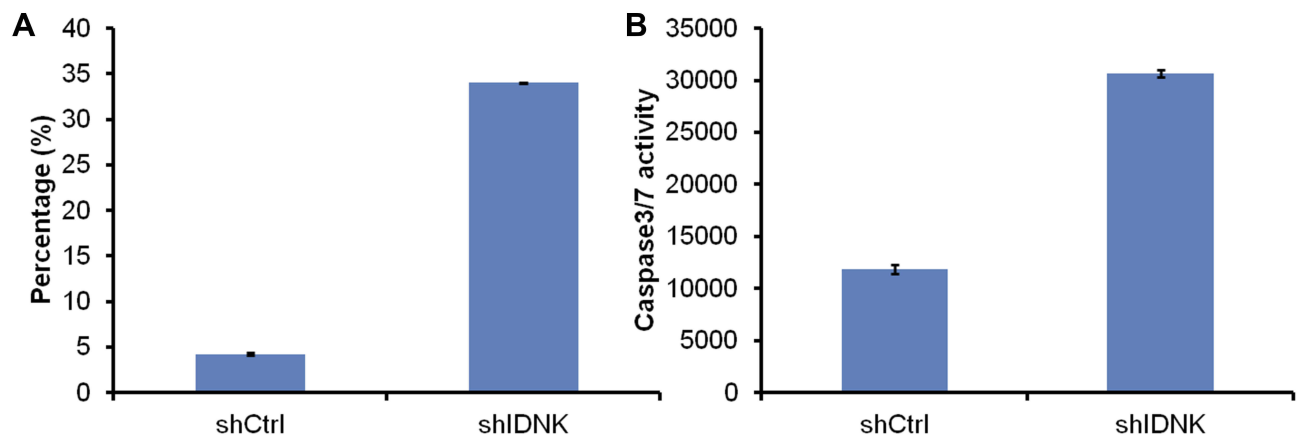
The significant pathways enriched by differentially expressed pathways were analyzed by IPA software. The top 20 IPA pathways involved with DEGs included tRNA charging (Z-score=-3), Insulin-like growth factor-1 (IGF-1) signaling (Z-score=1.155), and p53 signaling (Z-score=-1.667) (Figure 6B).

## Discussion

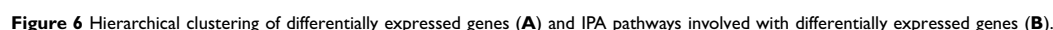
The mRNA and protein levels of IDNK were considerably reduced after IDNK knockdown. The proliferation and growth of HCC cells significantly decreased following IDNK knockdown. Additionally, knockdown of IDNK inhibited the cell cycle in S phase, and prominently promoted apoptosis of HCC cells. Microarray analysis revealed that a total of 1196 genes were differentially expressed after IDNK knockdown compared with cells infected with blank control plasmid, which were involved in the tRNA charging pathway.



**Figure 4** Cell cycle distribution of HCC cells after *IDNK* knockdown. (A) Cell cycle distribution of BEL-7404 cells after *IDNK* knockdown by flow cytometry. (B) Percentage of BEL-7404 cells after *IDNK* knockdown in the G1, S, and G2/M phase. (C) Cell cycle distribution of Huh-7 cells after *IDNK* knockdown by flow cytometry. (D) Percentage of Huh-7 cells after *IDNK* knockdown in the G1, S, and G2/M phase. \* $P < 0.05$ .



**Figure 5** *IDNK* knockdown significantly increases apoptosis of HCC cells. (A) Cell death was determined by Annexin V staining in BEL-7404 cells. (B) Apoptotic Huh-7 cells were determined by measuring caspase 3/7 activity.



Citrate is produced in mitochondria and plays a critical role in the Krebs' cycle. Excessive intracellular citrate is reported to inhibit glycolysis by suppressing phosphofructokinase-1.<sup>14</sup> Many studies have found that the level of citrate is significantly decreased in the blood, tissue, and urine of cancer patients,<sup>15–19</sup> which may be explained by the increased need for citrate in cancer cell metabolism. Glycolysis is the main

pathway used by cancer cells to generate ATP and suppression of glycolysis can inhibit cancer cell proliferation and growth.<sup>20</sup> The expression of anti-apoptotic protein in cancer cells is reduced by continuous exposure to citrate. Different from normal cells, cancer cells show pronounced citrate oxidizing capability.<sup>21</sup> A previous study has proposed that the uptake of extracellular citrate in cancer cells is associated with cell metabolism and fatty acid synthesis.<sup>9</sup> Theoretically, the inhibition of citrate cycle can slow down citrate metabolism, suppressing cancer cell proliferation and promoting cancer cell apoptosis. Another study described the behavior of gluconate as the competitive and irreversible blocker of citrate transport, with the inhibition of cancer cell growth in immunodeficient mice.<sup>13</sup> Thus, increasing the level of gluconate could inhibit the oxidation of citrate oxidizing, therefore inhibiting cancer cell proliferation and promoting apoptosis. The intriguing



discovery of *IDNK* in the control of HCC cancer cell proliferation may inspire future studies about the mechanism.

Caspase 3 is the master “executioner” during apoptotic cell death, which serves as a direct link between cell death and tumor repopulation.<sup>22</sup> Caspase-7 is considered to be redundant with caspase-3 because these related cysteine proteases share an optimal peptide recognition sequence.<sup>23</sup> It has been reported that caspase-3 and caspase-7 contribute to the majority of cleavage that takes place during apoptosis.<sup>24</sup> Thus, we detected the expression levels of caspase-3 and caspase-7 in HCC cells after *IDNK* suppression. As expected, the activities of caspase-3 and caspase-7 were significantly increased in shRNA *IDNK* group compared with control, which demonstrated that *IDNK* may inhibit the apoptosis of HCC cells by repressing the expression of caspase-3 and caspase-7.

Our microarray data suggested that tRNA charging was inhibited by the deregulated expression of *IDNK* in HCC cells. tRNAs are noncoding RNAs that play a key role in protein translation for cancer cell proliferation.<sup>25</sup> tRNAs and their derivatives are dysregulated in the progression of cancers, specifically in metastasis, invasiveness, and tumor growth.<sup>26</sup> tRNA charging is the aminoacylation of tRNAs. This event is altered in multiple myeloma, and is responsible for the increased need for protein translation.<sup>27</sup> In this study, the DEGs were significantly enriched in the tRNA charging pathway, indicating the changes of tRNA charging in HCC cells. IPA pathway analysis also showed that tRNA charging was a significant pathway, with a Z-score=-3, indicating that the tRNA charging was prominently inhibited in *IDNK* knockdown HCC cells. Thus, we suspect that the deregulation of *IDNK* expression inhibited tRNA charging, therefore inhibiting HCC cell proliferation.

There are some limitations in this study. The detailed mechanism underlying *IDNK* in cancer cells has not been clarified clearly. In addition, the role of *IDNK* in controlling cancer cell proliferation and apoptosis has not been determined in clinical samples in vivo and in vitro. Further mechanism and clinical studies are urgently needed.

## Conclusion

In conclusion, our study demonstrates that *IDNK* plays a key role in HCC cell proliferation and apoptosis. Knockdown of *IDNK* may inhibit tRNA charging, suppressing HCC cell proliferation. Our findings may indicate a novel therapeutic target for HCC. A good understanding of the role of *IDNK* may facilitate the discovery of therapies for HCC.

## Ethics Approval

Experiments performed in this study were approved by the Institutional Biosafety Committee (IBC approval reference number 022016017).

## Highlights

- *IDNK* expression was knocked down by shRNA in HCC cells.
- Proliferation of HCC cells significantly declined after *IDNK* silencing.
- Apoptosis of HCC cells significantly increased after *IDNK* silencing.
- tRNA charging was a significantly inhibited pathway in *IDNK* knockdown cells.

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## Disclosure

The authors report no conflicts of interest in this work.

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