

Frequency of heterozygous *TET2* deletions in myeloproliferative neoplasms

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Abstract: The Philadelphia chromosome (Ph)-negative myeloproliferative neoplasms (MPNs), including polycythemia vera, essential thrombocythemia, and primary myelofibrosis, are a group of clonal hematopoietic stem cell disorders with overlapping clinical and cytogenetic features and a variable tendency to evolve into acute leukemia. These diseases not only share overlapping chromosomal abnormalities but also a number of acquired somatic mutations. Recently, mutations in a putative tumor suppressor gene, ten-eleven translocation 2 (*TET2*) on chromosome 4q24 have been identified in 12% of patients with MPN. Additionally 4q24 chromosomal rearrangements in MPN, including *TET2* deletions, have also been observed using conventional cytogenetics. The goal of this study was to investigate the frequency of genomic *TET2* rearrangements in MPN using fluorescence *in situ* hybridization as a more sensitive method for screening and identifying genomic deletions. Among 146 MPN patients, we identified two patients (1.4%) who showed a common 4q24 deletion, including *TET2*. Our observations also indicated that the frequency of *TET2* deletion is increased in patients with an abnormal karyotype (5%).

Keywords: *TET2*, myeloproliferative neoplasms, fluorescence *in situ* hybridization, cytogenetics

Introduction

The Philadelphia chromosome negative (Ph-) myeloproliferative neoplasms (MPNs) are a group of clonal hematopoietic stem cell disorders with overlapping clinical and cytogenetic features and a variable tendency to evolve into acute leukemia. The Ph- MPNs include polycythemia vera, essential thrombocythemia, and primary myelofibrosis. The current unifying concept of cytogenetic instability of Ph- MPN is a loss or gain of chromosomal regions. The most frequent recurrent chromosomal abnormalities occurring in the Ph- MPNs include trisomy 9/+9p, deletion (20) (q11q13), deletion of 13q, and trisomy of chromosomes 8 and 9. The prognostic significance of the abnormalities in MPNs remains uncertain. However, recent attempts have been made to classify cytogenetic findings in primary myelofibrosis patients as being associated with favorable [isolated deletion (20q) and 13q, trisomy 9 as well as normal karyotype] outcomes and unfavorable outcomes (all other chromosome abnormalities). The number of patients remains too small and the time of follow-up too limited for a definitive validation of the outcomes associated with chromosomal risk profiles.¹ Nevertheless, it appears that the presence of cytogenetic abnormalities in primary myelofibrosis is associated with shorter survival, but their independent contribution to patient prognosis remains unknown.²

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Besides cytogenetic abnormalities, these diseases also share acquired somatic mutation of *JAK2* in exon 14, resulting in a valine to phenylalanine substitution in codon 617, *JAK2V617F*, present in over 90% of patients with polycythemia vera, 60% of patients with essential thrombocythemia, and 50% of patients with primary myelofibrosis.^{3–6} Although most MPNs carry mutation on one of two alleles, 2.8% of MPNs acquire *JAK2V617F* on two *JAK2* alleles,⁷ and 3% of *JAK2V617F*-negative patients have mutations in exon 12 of *JAK2*.⁸ Coexistence of these two mutations have also been described.⁹ *MPL* mutations leading to *MPLW515L* occur in 5%–10% of patients with primary myelofibrosis and in 2%–5% of patients with essential thrombocythemia who are negative for *JAK2V617F*.^{10,11} Other mutations described in MPN patients include mutations of the *CBL* and *ASXL1* genes.^{12,13}

Recently, somatic mutations and deletions of the ten-eleven translocation 2, or *TET2*, a putative tumor suppressor gene located on chromosome 4, band q24, have been described in 12% of patients with MPN.¹⁴ *TET2* mutations may predate the *JAK2V617F* mutation, may occur after the acquisition of *JAK2V617F*, or may occur simultaneously in two different clones.^{14,15} Following the original description, *TET2* nonsense and splice mutations, deletions, and out-of-frame insertions were identified in other myeloid malignancies, including myelodysplastic syndrome (19%–26%), *de novo* acute myeloid leukemia (12%–20%), therapy-related acute myeloid leukemia (24%–43%), chronic myelomonocytic leukemia (20%–51%), and systemic mastocytosis (29%).^{14,16–24} Although the current prognostic value of *TET2* mutations in MPNs remains unknown, *TET2* mutations were reported in 6/14 patients (43%) with MPN who had transformed to acute myeloid leukemia.²⁵

Deletions of the 4q24 region, including the *TET2* gene, have been identified by conventional cytogenetics. In 2005, Viguie et al reported a common deletion of 4q24 in four patients with acute myeloid leukemia/myelodysplastic syndrome.²⁶ Additional cytogenetic and high-resolution single-nucleotide polymorphism karyotyping revealed that 20/886 (2%) patients with myeloid disorders exhibited either partial or complete deletion of *TET2*.^{14,18–20} In at least four patients with acute myeloid leukemia/myelodysplastic syndrome, the remaining *TET2* copy harbored a somatic mutation.¹⁹ More recently, Hussein compared cytogenetic findings with *TET2* status and concluded that unmutated *TET2* MPN patients were not cytogenetically different from those MPN patients who harbor *TET2* mutations, but there was no information provided regarding deletions of 4q24/*TET2*.²⁷

Materials and methods

Patients

A total of 146 patient samples (48 were obtained from the Myeloproliferative Disorders-Research Consortium Tissue Bank) were entered in this study, including myelofibrosis (n = 52), polycythemia vera (n = 47), essential thrombocythemia (n = 21), myeloproliferative neoplasm (unclassified, MPNu, n = 20), and MPNu/myelodysplastic syndrome (n = 6). After patient informed consent was obtained, peripheral blood and bone marrow specimens were collected according to the Institutional Review Board guidelines of the Mount Sinai School of Medicine.

Bacterial artificial chromosome clones

Three bacterial artificial chromosome (BAC) clones, RP11-912N16 (137 kb), RP11-16G16 (175 kb), and RP11-45L9 (162 kb) obtained from BACPAC Resources, Oakland, CA, were grown on chloramphenicol agar plates. The BAC clone RP11-16G16 contains the commonly deleted region on 4q24, including *TET2*. To determine the extent of the deletion or possible rearrangements of the 4q24 region, two additional BAC clones, RP11-912N16 and RP11-45L9, containing DNA segments centromeric and telomeric to this commonly deleted region, respectively, were also used. Selected positive colonies were harvested after overnight growth using the Qiagen Large-Construct kit (Qiagen, Valenica, CA). DNA labeling was achieved using the CGH Nick Translation Kit (Abbott Molecular, Des Plaines, IL) and was fluorescently labeled with spectrum green (RP11-912N16), spectrum aqua (RP11-16G16), and spectrum red (RP11-45L9) according to the Abbott Molecular CGH protocol (Abbott Molecular). Validation fluorescence *in situ* hybridization (FISH) studies for all three BAC clones were performed on normal controls by evaluating 2000 interphase nuclei. The cutoff percentage for these probes was 3.5% and was calculated by using the mean \pm three standard deviations. All three BAC probes mapped to the 4q24 region in 40 examined metaphase cells, demonstrating the 100% specificity of these probes. In interphase nuclei of normal cells, the expected normal probe signals appear as two tricolor (green, red, aqua) fusions.

Cytogenetic and FISH analysis

FISH analysis was performed as previously described with pepsin modification treatment (100 μ L 10% pepsin and

2 mL 1% HCl) for five minutes.²⁸ Codenaturation of cells and FISH probes were carried out on a Thermobrite (Abbott Molecular) for three minutes at 73°C and hybridized at 37°C overnight. A total of 200 nuclei were scored in each of the 146 specimens. Metaphase cells were obtained using standard cytogenetic methods. Chromosomal abnormalities were described according to the International System of Human Cytogenetic Nomenclature (2009).²⁹

Results

We investigated 146 patients with Ph- MPN to determine the frequency of *TET2* genomic rearrangements. The summary of FISH studies, using the three *TET2* BAC FISH probes as well as the cytogenetic results, are provided in Table 1. Chromosomal analyses were available for 83/146 (57%) patients; 44 (53%) patients had a normal karyotype and 39 (47%) had an abnormal karyotype. The frequency of chromosomal abnormalities among the 39 patients with an abnormal karyotype were: deletion of the long arms of chromosome 20 in 11/39 (29%), followed by +9/+i(9p) in 9/39 (24%), trisomy of chromosome 8 in 7/39 (18%), and trisomy of the long arms of chromosome 1 in 6/39 (16%). Abnormalities of 4q24 (deletions, translocations, or dupli-

cations) were not detected in any of the 39 patients with an abnormal karyotype by conventional cytogenetics. Interphase FISH analysis using these probes revealed deletion of *TET2* in 2/83 patients (2% of total, and 5% among cytogenetically abnormal).

Patient 34 with primary myelofibrosis was *JAK2V617F*+ and had a complex karyotype, but 4q24 rearrangement was not identified (see Table 2). Interphase FISH with *TET2* revealed a deletion of all three BAC probes in 5% of interphase nuclei. The follow-up peripheral blood specimen eight months later revealed 30% of interphase cells with a *TET2* deletion, as well as a clone with a cytogenetically identified deletion of the 4q24 region (see Figure 1). Patient 8, with primary myelofibrosis, was *JAK2V617F*+, had a normal karyotype (see Table 2), and was disomic for the following loci: 7p11.1–q11.1, 8p11.1–q11.1, 9p11.1–q11.1, 1q12, 1q21, 1p36, 1q25, 5p15.2, 5q31, 7q31, 9p21 13q14, and 20q12 using interphase FISH. These loci are most frequently rearranged in MPN. In contrast, 90% of interphase cells revealed a deletion of only the RP11-16G16 BAC probe, and not RP11-912N16 and RP11-45L9, indicating a small interstitial *TET2* deletion of at least 175 kb localized between the centromeric and telomeric BAC FISH probes (see Figure 1).

Based on these initial studies, we screened an additional 63 patients for *TET2* genomic rearrangements, as shown in Table 1, and none were identified.

Table 1 also shows *JAK2* status in 42 patients and *MPLW515L* mutational status in 16 patients. Of the three patients who had *TET2* deletion, one was *MPLW515L* mutation-negative, and the *MPL* mutational status of the remaining two patients was not available.

Among 22,500 cytogenetically examined patients at our institution we identified only three patients with 4q24 chromosomal rearrangements, two with myelodysplastic syndrome and one with primary myelofibrosis (#34), indicating their rare occurrence. Patient 34 was identified among 512 patients with MPN (polycythemia vera = 361, myelofibrosis = 151). We therefore used interphase FISH as a more sensitive method to detect 4q24 chromosomal rearrangements and identified an additional patient of the 146 tested patients with *TET2* deletion, confirming the published frequency of 1%–2% in MPNs.^{14,18} However, among patients with available conventional cytogenetic results, the frequency was 2% overall (2/83), or 5% (2/39) among the cytogenetically abnormal patients. Two patients had primary myelofibrosis, while none of the 47 patients with polycythemia vera nor 21 patients with essential thrombocythemia had *TET2* deletions.

Table 1 Interphase FISH detection of *TET2* deletion

	PMF	PV	ET	MPNu	MPNu/ MDS	Total
Cytogenetics						
Normal	15	14	6	8	1	44
Abnormal	25	8	2	2	2	39
ND	12	25	13	10	3	63
Total	52	47	21	20	6	146
<i>JAK2</i> status						
<i>JAK2V617F</i>	4	16	9	0	0	29
<i>JAK2</i> wt	6	1	5	0	0	12
ND	42	30	7	20	6	105
Total	52	47	21	20	6	146
<i>MPL</i> status						
<i>MPLW515L</i>	0	1	1	0	0	2
<i>MPL</i> wt	2	4	7	0	0	13
ND	50	42	13	20	6	131
Total	52	47	21	20	6	146
<i>TET2</i> status						
<i>TET2</i> +	2	0	0	0	0	2
<i>TET2</i> –	50	47	21	20	6	144
Total	52	47	21	20	6	146

Notes: (+) deletion; (–) I-FISH did not detect deletion of three bacterial artificial chromosomes.

Abbreviations: FISH, fluorescence *in situ* hybridization; MPNu, myeloproliferative neoplasm (unclassified); ND, not done; wt, wild type; MDS, myelodysplastic syndrome; ET, essential thrombocythemia; PV, polycythemia vera; PMF, primary myelofibrosis.

Table 2 Summary of results from two patients with *TET2* deletions

Pt #	Dx	Mutational status		<i>TET2</i> BACs			% of nuclei	Karyotype
		<i>JAK2V617F</i>	<i>MPLW515L</i>	<i>RP11-912N16</i>	<i>RP11-16G16</i>	<i>RP11-45L9</i>		
8	PMF	+	ND	+	–	+	90	46,XY
34*	PMF	+	ND	–	–	–	5	45,XY,-7,del(10)(q21),inv(12)(?p12.1;q13.3),der(12)t(12;?),add(17)(p11.24)

Notes: On a subsequent follow-up specimen a clone with the karyotype 45,XY,der(4)del(4)(p14)del(q24),-7,del(8)(p21)del(10)(q22),add(17)(p11.2),del(20)(q11q13) had developed.

Abbreviations: ND, not done; PMF, primary myelofibrosis; BACs, bacterial artificial chromosomes.

The size of the deletions varied. In one patient (#8) with a normal karyotype, a deletion of approximately 175 kb in 90% of interphase nuclei containing the commonly deleted region of 4q24 (the other two BACs outside the region remained intact) was identified. This observation suggests a submicroscopic (cryptic) deletion beyond the resolution of conventional cytogenetic analysis and confirms the importance of using a higher resolution methodology for identifying *TET2* deletion. Moreover, metaphase FISH analysis confirmed a submicroscopic 4q24 deletion that was not detectable by conventional cytogenetics. The other patient (#34) had a deletion of all three BACs, resulting in a loss of at least 1.6 Mb. Interestingly, patient 34 had initially 5% of interphase cells with *TET2* deletion that was not detectable by conventional cytogenetics. When 30% of his peripheral blood cells showed loss of *TET2*, a new complex cytogenetic

clone emerged showing der(4)del(4)(p14)del(q24), resulting in a complete 4q24 deletion.

Heterozygous *TET2* mutations have been rarely identified in patients with chronic myelogenous leukemia but were detected in those patients who progress to blast crisis or accelerated phase.³⁰ Interpretation of these observations suggested that *TET2* mutations represent secondary lesions which contribute to the progression of the disease.³⁰ We identified one patient with chronic myelogenous leukemia, not included in this study, who had complex chromosomal aberrations consistent with blast crisis of chronic myelogenous leukemia. FISH analysis revealed a complete deletion of all three BAC probes in 17% of interphase nuclei.

In summary, *TET2* deletions are not a frequent genomic event in MPN. Their deletions are present in about 2% of MPN patients and, in those with increased karyotype

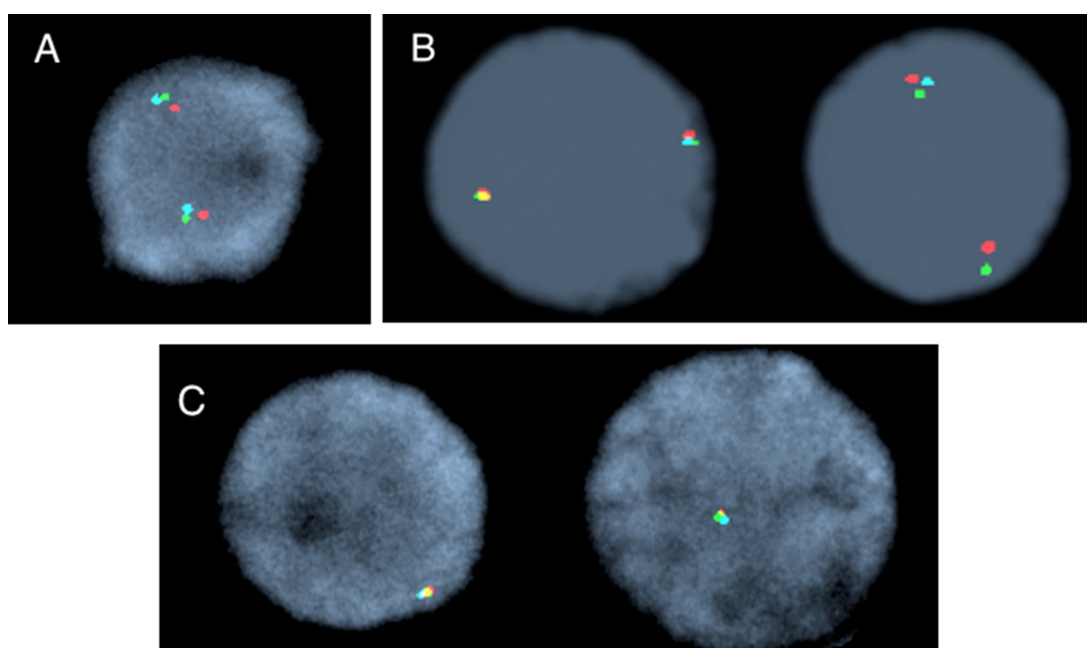


Figure 1 Interphase nuclei representing *TET2* FISH signal patterns. **A)** Normal nuclei with two triple color fusion signals. **B)** Deletion of the 175 kb RP11-16G16 BAC probe (aqua). **C)** Deletion of all three bacterial artificial chromosome probes; note only one triple color fusion signal.

instability, they may occur in about 5% of patients. These observations may add supporting evidence that *TET2* mutations, as well as deletions, may contribute to leukemic evolution in patients with MPN that transform into leukemia.

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Disclosure

The authors report no conflicts of interest in this work.

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