

The Biological Activity Research of the Nano-Drugs Based on 5-Fluorouracil-Modified Quantum Dots

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Purpose: Over the past decades, quantum dots (QDs) have shown the broad application in diverse fields, especially in intracellular probing and drug delivery, due to their high fluorescence intensity, long fluorescence lifetime, strong light-resistant bleaching ability, and strong light stability. Therefore, we explore a kind of therapeutic potential against cancer with fluorescent imaging.

Methods: In the current study, a new type of QDs (QDs@L-Cys-TAEA-5-FUA) capped with L-cysteine (L-Cys) and tris(2-aminoethyl)amine (TAEA) ligands, and conjugated with 5-fluorouracil-1-acetic acid (5-FUA) has been synthesized. Ligands were characterized by electrospray ionization mass spectrometry and H-nuclear magnetic resonance (^1H NMR) spectroscopy. The modified QDs were characterized by transmission electron microscopy, ultraviolet and visible spectrophotometry (UV-Vis), and fluorescence microscopy. And the biological activity of modified QDs was explored by using MTT assay with HeLa, SMMC-7721 HepG2, and QSG-7701 cells. The fluorescence imaging of modified QDs was obtained by fluorescence microscope.

Results: The modified QDs are of controllable sizes in the range of 4–5 nm and they possess strong optical emission properties. UV-Vis and fluorescence spectra demonstrated that the L-Cys-TAEA-5-FUA was successfully incorporated into QD nanoparticles. The MTT results demonstrated that L-Cys-TAEA-5-FUA modified QDs could efficiently inhibit the proliferation of cancer cells as compared to the normal cells, illustrating their antitumor efficacy. The mechanistic studies revealed that the effective internalization of modified QDs inside cancer cells could inhibit their proliferation, through excessive production of intracellular reactive oxygen species, leading to apoptosis process.

Conclusion: The present study suggests that modified QDs can enter cells efficiently and could be employed as therapeutic agents for the treatment of various types of cancers with fluorescent imaging.

Keywords: quantum dots, 5-fluorouracil, nano-drugs, apoptosis, antitumor activity

Introduction

Quantum dots (QDs)-based nanomaterials have special photochemistry properties due to their unique electrochemical and photophysical properties, facilitating the wide applications of QDs, especially as nano-fluorescent probes and antitumor drug carriers.^{1–5} Although issues regarding the cytotoxicity of QDs have been raised, the surface of QDs can be modified to reduce their toxicity towards cells, while improving their water solubility and tumor cell targeting ability.^{1,6–9} The researchers have employed various biological molecules including peptides, antibodies, proteins, and

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DNA to coat the surface of QDs, and these modified QDs have intensely been studied as a new class of nanoparticle probe and drug carrier in diverse biomedical research areas, ranging from cellular fluorescence imaging and diagnostics in biomedicine to environmental monitoring for public health and security.^{10–15} However, the major hurdle for in vivo applications of QDs based nanomaterials is their organelle-specific targeted delivery.

Amino acids, containing amino and carboxyl groups, are the basic components of protein.^{16,17} Since amino acids are mostly water-soluble and inexpensive, they have frequently been used as capping agents for QDs.^{18–21} The amino acids are often linked to the surface of QDs, followed by attachment of other drugs with antitumor activity and the biomolecule with tumor cells targeting ability to form targeted drug delivery systems.^{22–24} 5-Fluorouracil (5-FU) is an anti-metabolite and a constituent of RNA.^{25–28} Following intracellular conversion to fluorouracil deoxynucleotide, 5-FU can inhibit the DNA synthesis via blocking the conversion of deoxyribonucleate to thymidylic acid by intracellular thymidylic acid synthase.^{29,30} Moreover, 5-FU can be incorporated into RNA and it can interfere with the protein synthesis after conversion to 5-Fluorouridine in vivo.^{31,32} 5-FU also inhibits the activity of exosome complexes (an exonuclease). Furthermore, it is a potent anti-cancer drug that can be used to treat a number of cancers including gastric, colon, lung, and breast cancers.^{33–36} However, the water solubility of 5-FU is very poor and intravenous administration of this drug is associated with the toxicity in bone marrow and gastrointestinal tract.^{37–39} To address the limitations related to the administration of 5-FU, the use of QDs was investigated to aide its pharmacological actions.^{40–44} To date, a few studies on L-cysteine and 5-FU-modified QDs have been reported.

In the present study, we synthesized a new type of QDs with controllable sizes (4–5 nm) and strong optical emission properties. In this nanoplatfrom, QDs were coated with the ligands L-Cys- and TAEA-, followed by conjugation of 5-FUA. The biological activity of the modified QDs was checked in different types of cells (HeLa, QSG-7701, SMMC-7721, and HepG2 cells) through cell viability assay using MTT reagent. The results demonstrated that L-Cys-TAEA-5-FUA-modified QDs effectively inhibited the proliferation of cancerous cells when compared to the normal cells. The cell morphology analysis, flow cytometry (using annexin

V-FITC), and fluorescence microscopy (AO/EB staining, mitochondrial membrane potential (MMP), and reactive oxygen species (ROS) determination) further reveal that the modified QDs could induce apoptosis to cancer cells through the formation of excessive intracellular ROS. Overall, the L-Cys-TAEA-5-FUA-modified QDs were found to enter intracellular and can be as potent antitumor agents for the treatment of cancers via apoptosis.

Materials and Methods

Materials

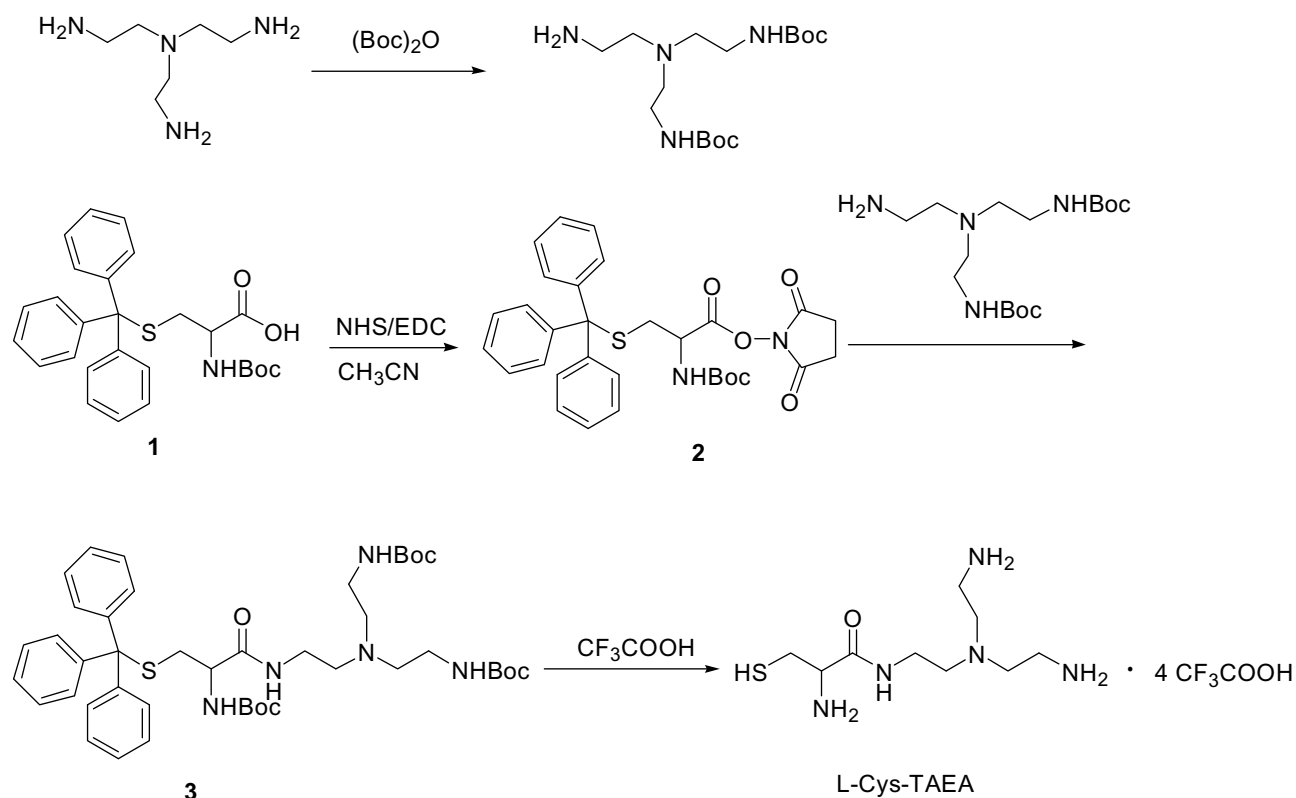
All the reagents were purchased from commercial suppliers and used without further purification. Solvents used were purified by standard methods prior to use. Cervical cancer cells (HeLa), human normal hepatocytes (QSG-7701), and human hepatoma cells (SMMC-7721 and HepG2) were procured from Shanghai Cell Bank of Chinese Academy of Sciences.

¹H NMR spectra were recorded on a Bruker AV-400 spectrometer. UV-Vis spectra were monitored with a Varian Cary 300 BIO UV-Vis spectrophotometer. Fluorescence spectra were measured by using a Varian Cary Eclipse fluorescence Spectrophotometer (American, Agilent, Co.). Fluorescence images were obtained employing a fluorescence microscope (Leica, Germany). The cell fluorescence intensity was recorded by using a FACSCalibur flow cytometer (Becton Dickinson & Co., Franklin Lakes, NJ).

Synthesis of L-Cys-TAEA (Scheme 1)

Tris(2-aminoethyl)amine (TAEA) (2.0 g, 13.7 mmol) dissolved in 70 mL of dichloromethane was taken in a round-bottomed flask equipped with a magnetic stirring bar. Then, (Boc)₂O (1.0 g, 4.6 mmol) dissolved in dichloromethane was slowly added dropwise to the flask using a constant pressure funnel for 1 h at 0°C, followed by stirring at room temperature overnight. The resulting reaction mixture was evaporated to dryness under reduced pressure, and the residue was extracted using ethyl acetate and distilled water. Removal of the solvent in the organic phase resulted in a solid material. After drying under vacuum, the solid material was purified by silica gel column chromatography using a CH₂Cl₂:CH₃OH (v:v = 50:1) mixture as the eluent to obtain the colorless grease protected TAEA.

Compound 1 (2.0 g, 3.5 mmol) in 100 mL of acetonitrile and NHS (0.59 g, 3.5 mmol) were taken in a 250 mL round-bottomed flask, placed on a magnetic stirrer. EDC (0.89 g, 3.5 mmol) was then added into the flask under ice-cooling



Scheme 1 Schematic illustration for the synthesis of L-Cys-TAEA.

condition, followed by stirring for 2 h at 0°C . The reaction mixture was then further stirred under N_2 at room temperature overnight. The resulting precipitate was filtered and extracted with dichloromethane and distilled water. Removal of solvent from the organic phase resulted in a white foamy solid **2**.

Protected TAEA (0.5 g, 1.5 mmol), compound **2** (0.84 g, 1.5 mmol), and NaHCO_3 (0.15 g, 1.8 mmol) were added to a 100 mL round-bottomed flask and the mixture was stirred for 12 h at 60°C . The reaction mixture was evaporated to dryness under reduced pressure, and the residue was extracted using dichloromethane and distilled water. Removal of the solvent from the organic phase resulted in a solid material. After vacuum drying, the solid material was purified by silica gel column chromatography with a CH_2Cl_2 : CH_3OH (v:v = 10:1) mixture as the eluent to obtain the yellow oil compound **3**.

Compound **3** was dissolved in 10 mL CH_2Cl_2 in a 50 mL round-bottom flask and placed under N_2 vacuum at 0°C . Next, trifluoroacetic acid (9.5 mL) and triethylsilane (0.5 mL) were added to the flask, followed by stirring at 25°C for 6 h. The reaction mixture was evaporated to dryness under reduced pressure and residue was extracted

with methylene chloride and distilled water. Evaporation of solvent in the aqueous phase resulted in L-Cys-TAEA.

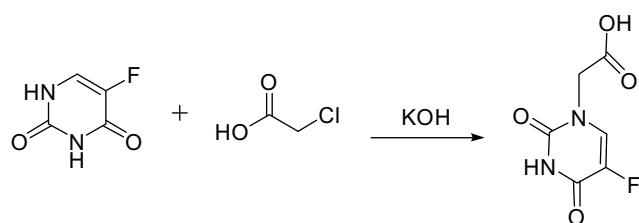
1. Cys-TAEA, yield: 60%; ^1H NMR (300 MHz, deuterium oxide) δ 4.10 (t, J = 6.0 Hz, 1H), 3.40 (t, J = 7.5 Hz, 2H), 3.14 (t, J = 6.0 Hz, 4H), 3.00 (t, J = 7.5 Hz, 6H), 2.85 (t, J = 7.5 Hz, 2H). ^{13}C NMR (75 MHz, D_2O) δ 170.06, 163.55, 163.07, 162.60, 162.13, 122.00, 118.14, 114.27, 110.41, 54.31, 49.75, 36.22, 29.68, 23.77. ESI-MS, m/z : 250.15 $[\text{M}+\text{H}]^+$.

Synthesis of QDs

CdSe , CdSe/CdS , and CdSe/ZnS QDs capped with tri- n -octylphosphine oxide (TOPO) were synthesized employing organometallic procedures as described in our previous report.⁴⁵ The QDs synthesized using this procedure exhibit high crystallinity, narrow emission spectra, and narrow size distributions.

Synthesis of Water-Soluble L-Cys-TAEA-Modified QDs

One-milliliter QDs (CdSe or CdSe/ZnS or CdSe/CdS) in 1 mL of chloroform, L-Cys-TAEA (300 mg) in 3 mL of



Scheme 2 Schematic illustration for the synthesis of 5-FUA.

triple distilled water, and 10 drops of triethanolamine were placed in a bottle and sonicated for 4 h. The reaction was considered complete when red QDs from the lower organic phase diffused into the upper aqueous phase. Ethyl acetate was then added to precipitate nanocrystal complexes and the nanocrystals were purified from side products and unreacted precursors. The solid was filtered from the solution and dried under vacuum to generate three water-soluble QDs, namely, CdSe@L-Cys-TAEA, CdSe/ZnS@L-Cys-TAEA, and CdSe/CdS@L-Cys-TAEA.

Synthesis of 5-FUA (Scheme 2)

5-FU (2.6 g, 20 mmol) and KOH (2.24 g, 40 mmol) in 16 mL of water and chloroacetic acid (3.4 mL, 36 mmol) in 8 mL of water were placed in a vessel. The solution mixture was stirred at room temperature for 30 min. Then, the pH of the reaction was adjusted to 10 with a 10% KOH solution and the reaction was refluxed for 2 h. After cooling the reaction liquid, the pH of the reaction mixture was adjusted to 2 with concentrated hydrochloric acid. Then, the reaction mixture was evaporated under reduced pressure, and a small amount of water was added to dissolve the solid. Then, the solution was placed in ice bath to yield the precipitation, which was filtered to obtain the solid 5-Fluorouracil-1-acetic acid (5-FUA).

5-FUA, yield: 82%; ^1H NMR (300 MHz, DMSO- d_6) δ 11.94 (d, $J=3.0$ Hz, 1 H), 8.08 (d, $J=4.5$ Hz, 1 H), 4.36 (s, 2 H).

Synthesis of QDs@L-Cys-TAEA-5-FUA

5-FUA (109 mg, 0.58 mmol) in 2 mL of DMSO, EDC (111 mg, 0.58 mmol), NHS (66.7 mg, 0.58 mmol), and the above prepared QDs@L-Cys-TAEA were taken in a 50-mL round-bottomed flask, placed on a magnetic stirrer. The solution was stirred constantly at room temperature for 4 h. After completion of the reaction, the precipitate was centrifuged and washed by DMSO and $\text{C}_2\text{H}_5\text{OH}$ to obtain three target products CdSe@L-Cys-TAEA-5-FUA, CdSe/ZnS@L-Cys-TAEA-5-FU, and CdSe/CdS@L-Cys-TAEA-5-FUA.

Cell Culture

HeLa, QSG-7701, SMMC-7721, and HepG2 cells were cultured in RMPI-1640 medium and maintained at 37°C incubator with 5% CO_2 . Cells were allowed to grow in a monolayer in the tissue culture flask.

Cell Viability Assays

The antitumor ability of the modified QDs was investigated in HeLa, QSG-7701, SMMC-7721, and HepG2 cells using a modified cell viability assay using 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyltetrazolium bromide (MTT) method. Briefly, cells were seeded into 96-well plates at 5×10^4 cells per well. After 12 h, the cells were incubated with the samples (modified QDs: 50, 100, 200, 500, and 1000 $\mu\text{g/mL}$, original QDs: 5, 10, 20, 50, 100 $\mu\text{g/mL}$) for 48 h. Then, 20 μL MTT (2.5 mg/mL) was added to each well. After 4 h incubation, the medium was removed and 100 mL DMSO was added to each well of the plates to dissolve the formazan products. Absorbance of the solution in each well was read at 570 nm using a Tecan Infinite F200 M200 multimode reader. The inhibition rate was calculated from plotted results considering the untreated cells as 100%.

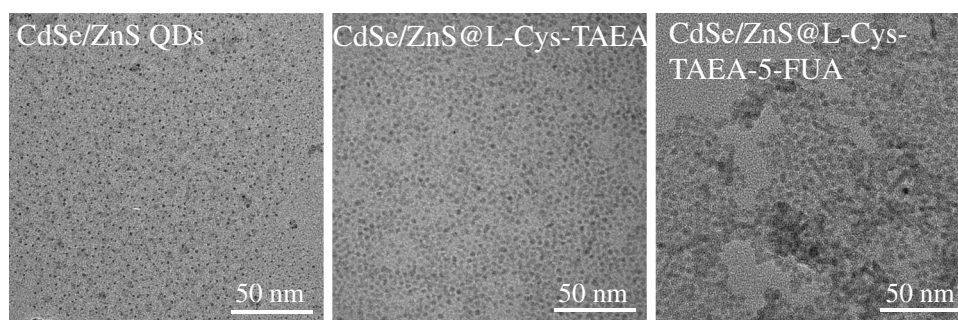


Figure 1 Representative TEM images of CdSe/ZnS QDs, CdSe/ZnS@L-Cys-TAEA, and CdSe/ZnS@L-Cys-TAEA-5-FUA. Scale bars = 50 nm.

Hoechst Staining Study for the Detection of Apoptosis

Cells were seeded in 6-well plates and allowed to adhere for overnight. Then, the cells were incubated with CdSe/ZnS@L-Cys-TAEA-5-FUA (200, 100, 50, 20, and 10 $\mu\text{g}/\text{mL}$) for respective time points and stained with Hoechst 33342 (10 mg/mL) at 37°C for 30 min. The fluorescence images of cells were captured using a fluorescence microscope. The rate of the abnormal (condensed or fragmented) nucleus was obtained by counting eight randomly chosen fields per dish for each experimental group from three independent experiments and expressed as percent apoptotic nuclei. Cell nuclei stained with Hoechst 33342 were observed at a wavelength of 405 nm and emission wavelength between 430 and 480 nm.

Apoptosis Assay Using Annexin V-FITC Staining by Flow Cytometry

Apoptosis assay was carried out using an annexin V-FITC apoptosis detection kit by a FACSCalibur flow cytometer. Briefly, cells were seeded in 6-well plates for 24 h. The cells were incubated with the treatments (30, 100 $\mu\text{g}/\text{mL}$) for 6 h. The cells were then carefully washed with PBS and resuspended in PBS buffer. After that, the cells were incubated with annexin V-FITC for 30 min at 37°C, followed by flow cytometry analysis. Data were analyzed using the BD Cell Quest software. Percentage of live and apoptotic cells was determined using negative controls, prepared by staining control experiments with propidium iodide (PI) or annexin V.

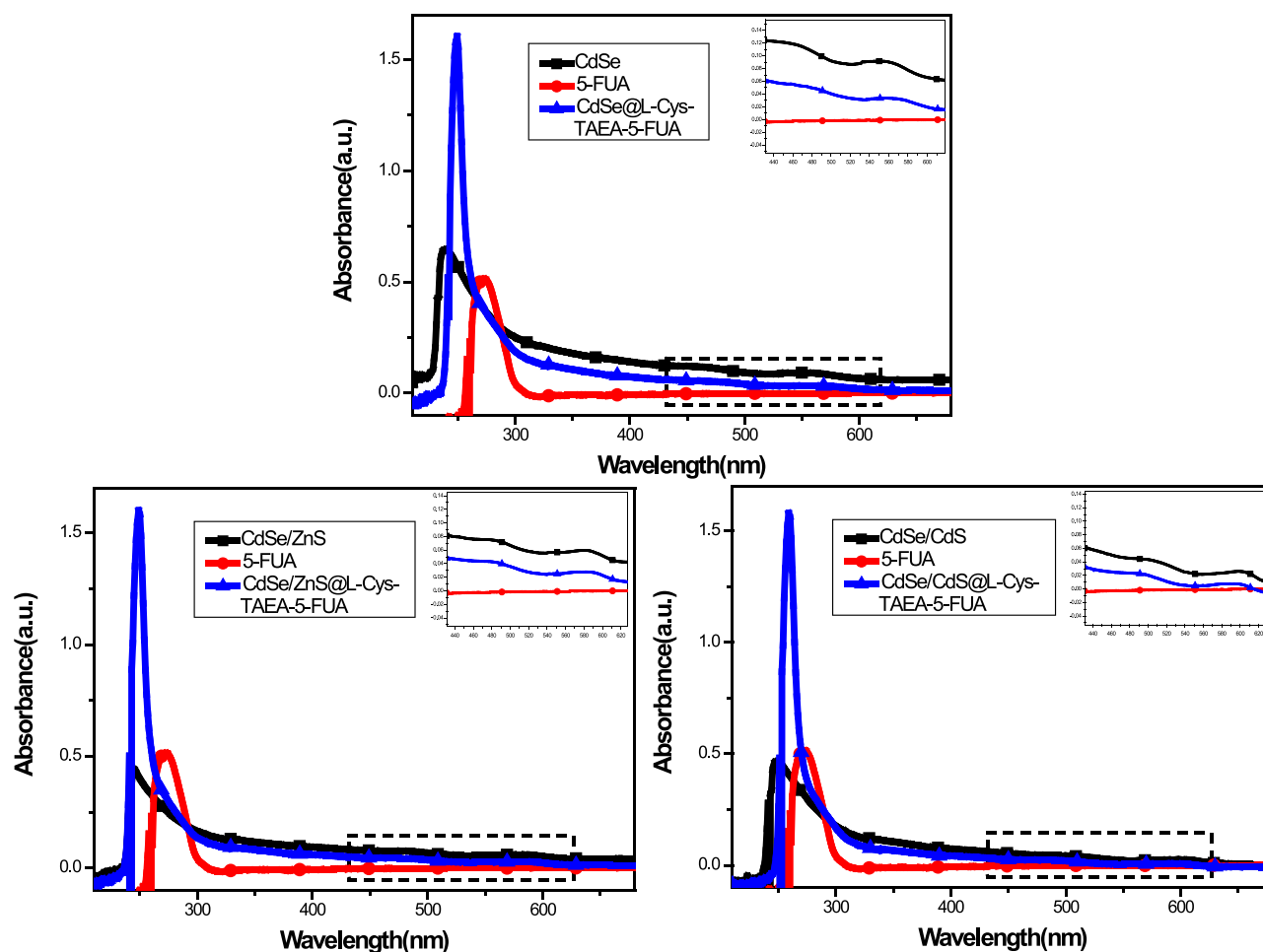


Figure 2 UV-Vis spectra of the original QDs, 5-FUA, and QDs@L-Cys-TAEA-5-FUA.

AO/EB Staining Study for Determination of Apoptosis

Cellular apoptosis was evaluated using the AO/EB apoptosis detection kit employing fluorescence microscopy. HeLa cells were first seeded in 6-well plates for 24 h. The cells were then incubated with the samples (30, 100 $\mu\text{g/mL}$) for 6 h. The cells were then thoroughly washed with PBS, followed by incubation with AO/EB for 30 min at 37°C. The fluorescence images of the cells were captured using a fluorescence microscope.

Analysis of Mitochondrial Membrane Potential (MMP)

MMP of cancer cells in presence of the QDs was assessed using flow cytometry. In brief, HeLa cells were first incubated with the samples for 24 h. The cells were then collected via trypsinization and incubated in media containing 10 $\mu\text{g/mL}$ JC-1 dye (5,5',6,6'-tetrachloro-1,1',3,3'-

tetraethyl-imidacarbocyanine iodide; Molecular Probes) for 15 min at 37°C. After that, the cells were carefully washed with PBS and the MMP of cells was measured using a FACSCalibur flow cytometer. Photomultiplier settings were adjusted to detect JC-1 monomer fluorescence signals using an FL1 detector (green fluorescence, centered around 530 nm), and JC-1 aggregate fluorescence signals employing FL2 detector (red fluorescence, centered around 590 nm). Ten thousand events were acquired for each sample, and the red and green mean fluorescence intensities were analyzed using BD Cell Quest software.

To evaluate the MMP through fluorescence microscope, HeLa cells were seeded in 6-well plates and incubated 5 $\mu\text{g/mL}$ Rh123 in complete media for 30 min at 37°C. The cells were then washed twice with PBS and the MMP was observed using a fluorescence microscope.

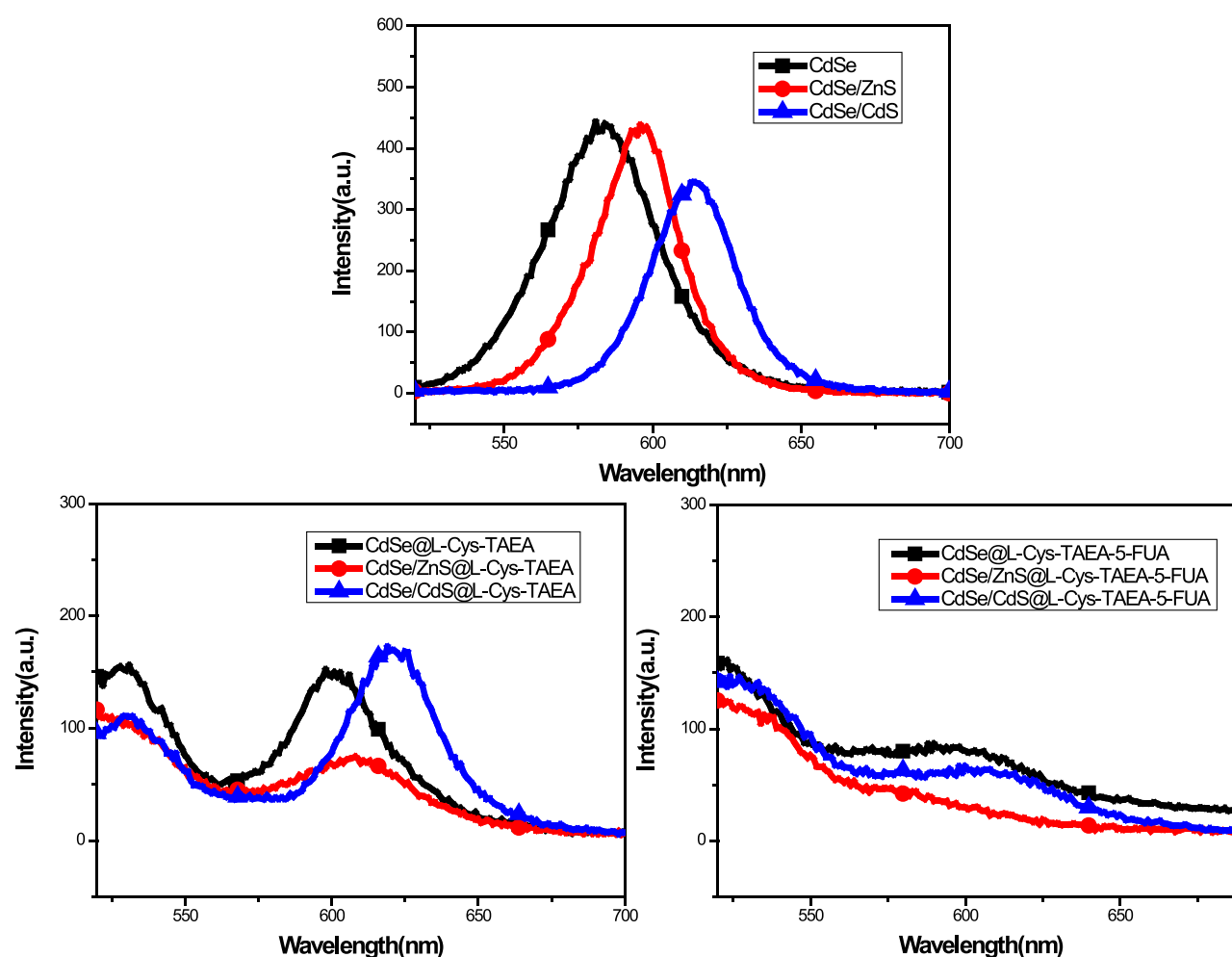


Figure 3 Fluorescence spectra of the original QDs, QDs@L-Cys-TAEA, and QDs@L-Cys-TAEA-5-FUA.

Determination of Reactive Oxygen Species (ROS)

Cellular accumulation of ROS was determined using the 2',7'-dichlorodihydrofluorescein diacetate (H₂DCF-DA) reagent. Briefly, after treatment with samples for 12 h, the cells were rinsed thrice with PBS to remove the unbound samples. The cells were then incubated with 10 μ M of H₂DCF-DA for 30 min at 37°C. Next, the wells were washed twice with PBS to remove the excess dye. The fluorescence intensity of the cells was determined using a FACSCalibur flow cytometer and fluorescence microscope.

Results and Discussion

Synthesis and Characterizations

In the present study, three different kinds of QDs (CdSe, CdSe/ZnS, and CdSe/CdS) were prepared by organic phase synthesis method. These QDs were characterized by various physicochemical techniques including TEM, UV-Vis, and fluorescence spectroscopy (Figures 1–3). The QDs exhibited high fluorescence intensity, narrow emission spectra, as well as narrow size distributions. Besides the QDs, the ligands L-cysteine-tris(2-aminoethyl)amine (L-Cys-TAEA) and 5-FUA were also synthesized and characterized by ESI-MS, ¹H NMR spectroscopy, or ¹³C NMR spectroscopy. At first, the surface of QDs was modified with L-Cys-TAEA. However, the cell growth inhibition rate of the QDs@L-Cys-TAEA is found to be lower. Therefore, 5-FUA was further conjugated with the QDs@L-Cys-TAEA to improve the cell growth inhibition rate. The size and morphology of the original QDs, QDs@L-Cys-TAEA, and QDs@L-Cys-TAEA-5-FUA were then characterized using TEM. As shown in Figure 1, the modified QDs maintained the good dispersion and uniformity with spherical shape in PBS. The modified QDs showed good water solubility and dispersibility.

The modified QDs also exhibited better optical properties in UV-Vis spectra and fluorescence spectroscopy. Figure 2 depicts the absorption spectra of original QDs, 5-FUA, and the modified QDs. It can be observed that with the addition of 5-FUA, the UV-Vis absorption spectrum of QDs@L-Cys-TAEA-5-FUA exhibits a new absorption band at around 270 nm, which could be attributed to the absorption of 5-FUA. Moreover, there was no much difference in the position or the width of absorbance bands between QDs@L-Cys-TAEA-5-FUA and original QDs, suggesting that 5-FUA was successfully incorporated into QDs.

The original QDs, QDs@L-Cys-TAEA, and QDs@L-Cys-TAEA-5-FUA were further characterized by fluorescence spectroscopy. As shown in Figure 3, the fluorescence peak of L-Cys-TAEA modified QDs exhibited a slightly red-shifted emission as compared to that of corresponding QDs, indicating that the emission arises due to the recombination of L-Cys-TAEA-5-FUA on the surface of QDs.

Cytotoxicity of QDs

The cytotoxicity of original and modified QDs was investigated in HeLa, SMMC-7721, QSG-7701, and HepG2 cells using the MTT assay. Calculated IC₅₀ values of original and modified QDs are shown in Table 1. The result demonstrated that original QDs are highly toxic towards both tumor cells and normal cells. It indicates that there is no obvious selectivity for normal cells and cancer cells. Notably, all the L-Cys-TAEA-5-FUA modified QDs exerted less toxicity on the normal liver QSG-7701 cells as compared to the three types of cancer cells. Interestingly, the CdSe/ZnS@L-Cys-TAEA-5-FUA QDs have better anti-tumor activity in comparison with 5-FUA. Therefore, it is easy to understand that CdSe/ZnS@L-Cys-TAEA-5-FUA QDs possess obvious selectivity for normal cells and cancer cells. Henceforth,

Table 1 IC₅₀ Values of Modified and Unmodified QDs

Compounds	IC ₅₀ (μ g/mL)			
	HeLa	SMMC-7721	HepG2	QSG-7701
CdSe@L-Cys-TAEA	389.5 \pm 3.5	425.6 \pm 4.5	434.8 \pm 3.9	463.7 \pm 2.9
CdSe/ZnS@L-Cys-TAEA	329.4 \pm 3.8	400.0 \pm 4.7	413.6 \pm 4.6	439.5 \pm 3.6
CdSe/CdS@L-Cys-TAEA	339.1 \pm 3.7	412.6 \pm 4.3	432.3 \pm 4.9	457.9 \pm 2.7
CdSe@L-Cys-TAEA-5-FUA	46.3 \pm 1.8	60.8 \pm 2.9	61.2 \pm 2.1	80.0 \pm 3.2
CdSe/ZnS@L-Cys-TAEA-5-FUA	41.8 \pm 3.2	49.9 \pm 2.3	50.7 \pm 3.1	106.5 \pm 2.3
CdSe/CdS@L-Cys-TAEA-5-FUA	44.2 \pm 4.0	56.7 \pm 1.7	55.8 \pm 3.4	97.6 \pm 2.1
5-FUA	71.1 \pm 2.6	68.3 \pm 3.1	61.4 \pm 2.7	69.4 \pm 2.9
CdSe	17.5 \pm 2.5	13.9 \pm 3.1	14.5 \pm 3.4	16.7 \pm 3.1
CdSe/ZnS	15.4 \pm 2.2	13.2 \pm 1.9	13.7 \pm 2.9	14.5 \pm 2.4
CdSe/CdS	16.2 \pm 3.1	14.4 \pm 2.7	13.8 \pm 2.8	15.4 \pm 1.7

CdSe/ZnS@L-Cys-TAEA-5-FUA QDs were used as a representative QDs for the subsequent experiments. At the same time, the cytotoxicity of L-Cys-TAEA modified QDs was found to be lower for normal cells and cancer cells. Overall, the cytotoxicity study revealed that the L-Cys-TAEA-5-FUA modified QDs can inhibit the proliferation of tumor cells effectively while showing less cytotoxicity towards normal cells.

Effect of Different Concentrations of Modified QDs on Cell Morphology

The analysis of morphology of HeLa cells in response to different concentrations of CdSe/ZnS@L-Cys-TAEA-5-FUA QDs was carried out by Hoechst 33342 staining using a fluorescence microscope. Figure 4 shows the changes of distribution and cell morphology in presence of CdSe/ZnS@L-Cys-TAEA-5-FUA QDs at different

concentrations (20 $\mu\text{g/mL}$: Figure 4A, 50 $\mu\text{g/mL}$: Figure 4B, 100 $\mu\text{g/mL}$: Figure 4C, and 200 $\mu\text{g/mL}$: Figure 4D). Upon internalization, the modified QDs remained intact in cells with a distribution in a cytoplasm. The results also exhibited that the modified QDs treatment could increase the number of apoptotic nucleus in HeLa cells in a concentration-dependent manner and these cells showed typical apoptotic changes, including stained brightness, reduction of cellular volume, fragmented nuclei, as well as condensed chromatin. However, no obvious apoptosis phenomenon was observed in QSG-7701 cells in presence of CdSe/ZnS@L-Cys-TAEA-5-FUA. These results indicate that modified QDs possess certain selectivity towards tumor cells and can inhibit their proliferation effectively by inducing early apoptosis.

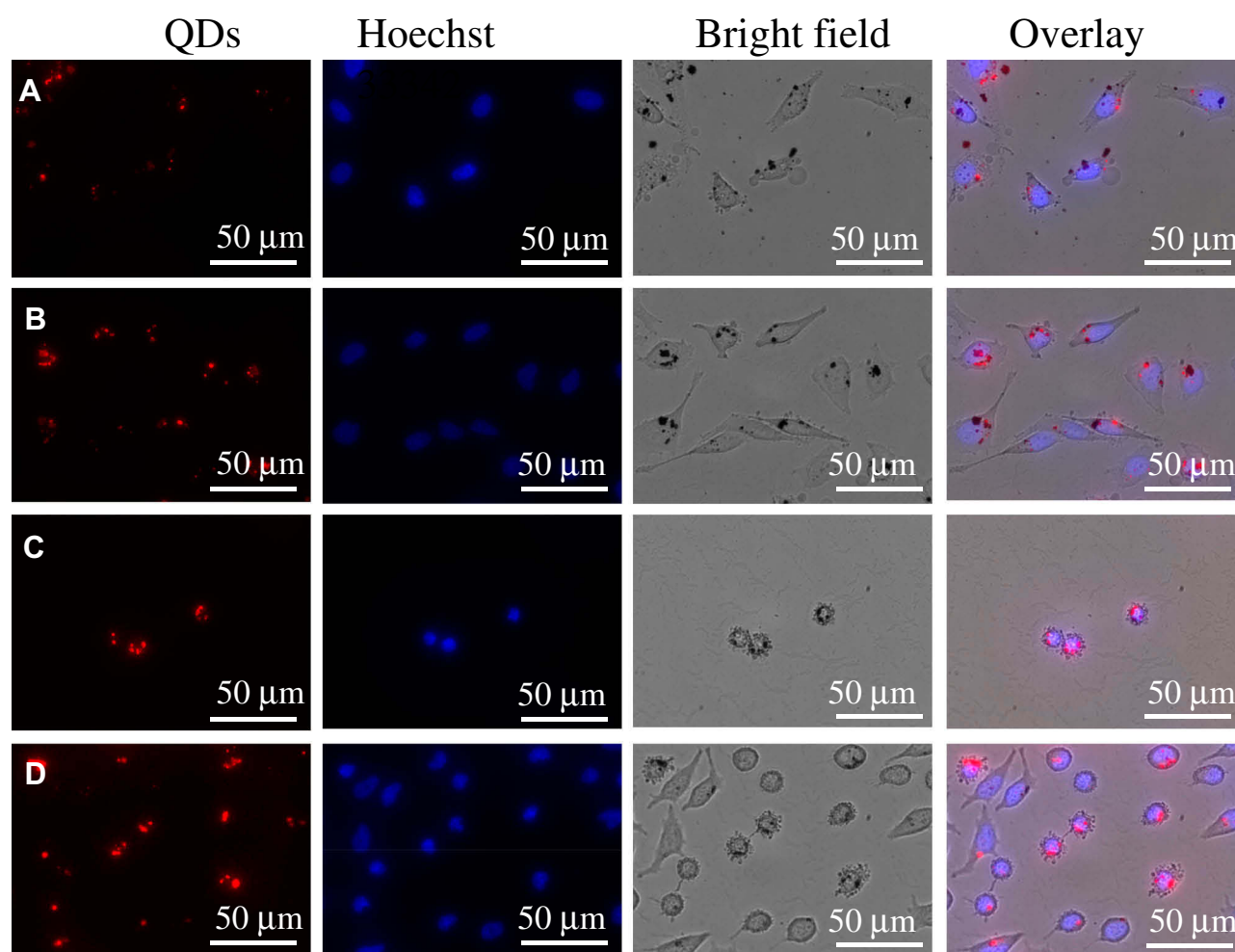


Figure 4 The distribution and cell morphology in HeLa cells in response to the treatment of CdSe/ZnS@L-Cys-TAEA-5-FUA QDs at different concentrations. (A) 20 $\mu\text{g/mL}$, (B) 50 $\mu\text{g/mL}$, (C) 100 $\mu\text{g/mL}$, and (D) 200 $\mu\text{g/mL}$. Scale bars = 50 μm .

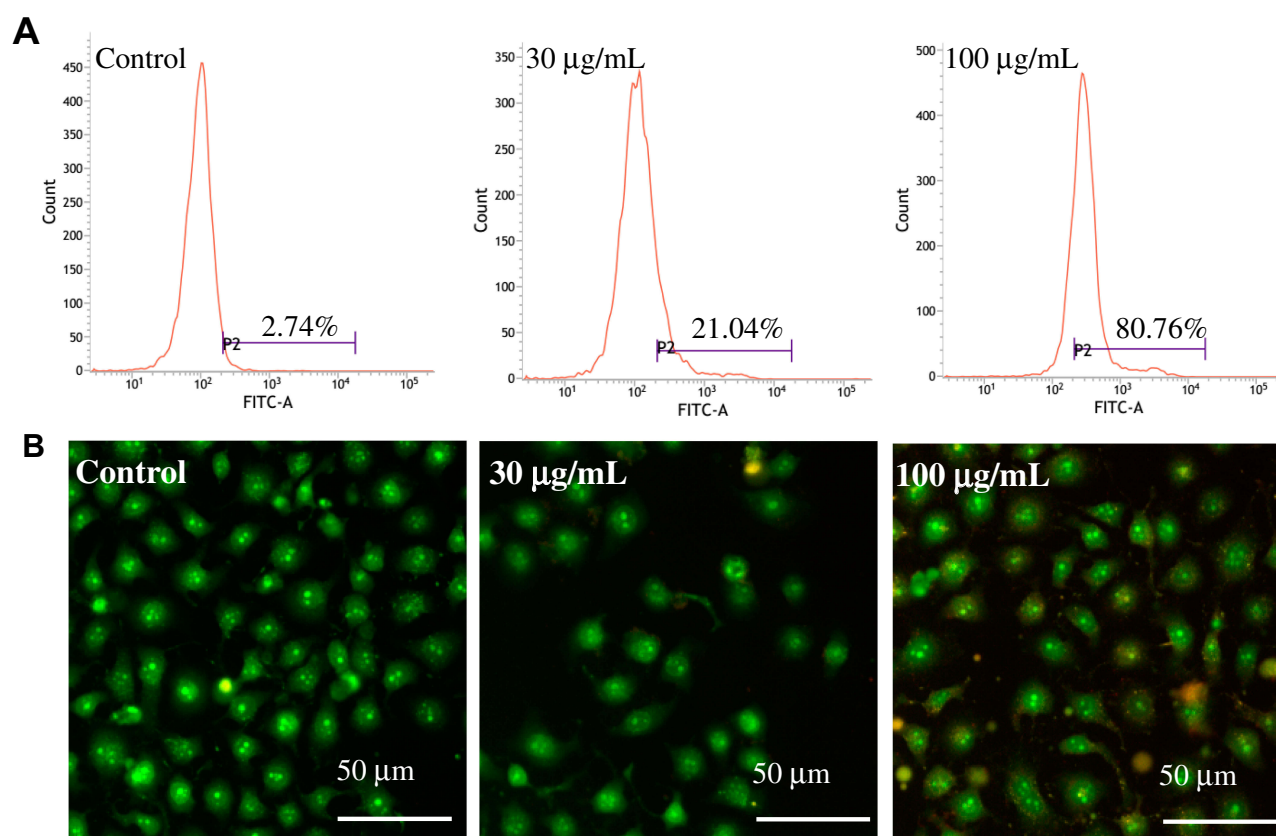


Figure 5 Apoptosis analysis in HeLa cells treated with CdSe/ZnS@L-Cys-TAEA-5-FUA (30 µg/mL and 100 µg/mL) using (A) flow cytometry data (Annexin V-FITC staining) and (B) fluorescence microscope images (AO/EB staining) of HeLa. Scale bars = 50 µm.

Induction of Apoptosis

To confirm the early apoptosis in HeLa cells in presence of CdSe/ZnS@L-Cys-TAEA-5-FUA QDs, apoptosis assay was carried out by flow cytometry using Annexin V-FITC kit. As shown in Figure 5A, the percentage of early apoptotic cells was significantly higher in response to the modified QDs treatment (30 µg/mL: 21.04% and 100 µg/mL: 80.76%) as compared to the control experiment (2.74%). In addition, the cell apoptosis was further studied through fluorescence microscopy using AO/EB staining. As can be seen from Figure 5B, the control cells were found to be structurally intact and green fluorescence was observed in cell nucleus due to the passing of AO through the intact cell membrane. Contrastingly, the cells treated with the modified QDs exhibited orange fluorescence as the EB passed through the damaged cell membrane. Furthermore, the orange fluorescence intensity in cells was gradually increased with increasing concentration of the modified QDs. Overall, these results revealed that the modified QDs can induce apoptosis in cancer cells in a concentration-dependent manner.

The Reduction of Mitochondrial Membrane Potential (MMP)

To examine CdSe/ZnS@L-Cys-TAEA-5-FUA mediated potential mitochondrial dysfunctions in HeLa cells, fluorescence microscopy was employed using JC-1 dye staining. The dye molecules are separate at low concentration and can emit green fluorescence (monomeric form), but form J-aggregates with red-shift emission. Figure 6A presents the quantitative analysis results of JC-1-stained HeLa cells employing flow cytometry. The result exhibited significant increase of red/green ratio in modified QDs-treated cells as compared to that of control cells, suggesting that modified QDs are capable of inducing apoptosis.

Additionally, the MMP of HeLa cells in presence of the modified QDs was further checked employing fluorescence microscope using the Rhodamine 123/Hoechst 33342 staining. The structurally intact control cells exhibited intense green fluorescence (Figure 6B). However, the intensity of green fluorescence was decreased in cells in the presence of modified QDs in a dose-dependent manner, indicating mitochondrial membrane damage. Altogether,

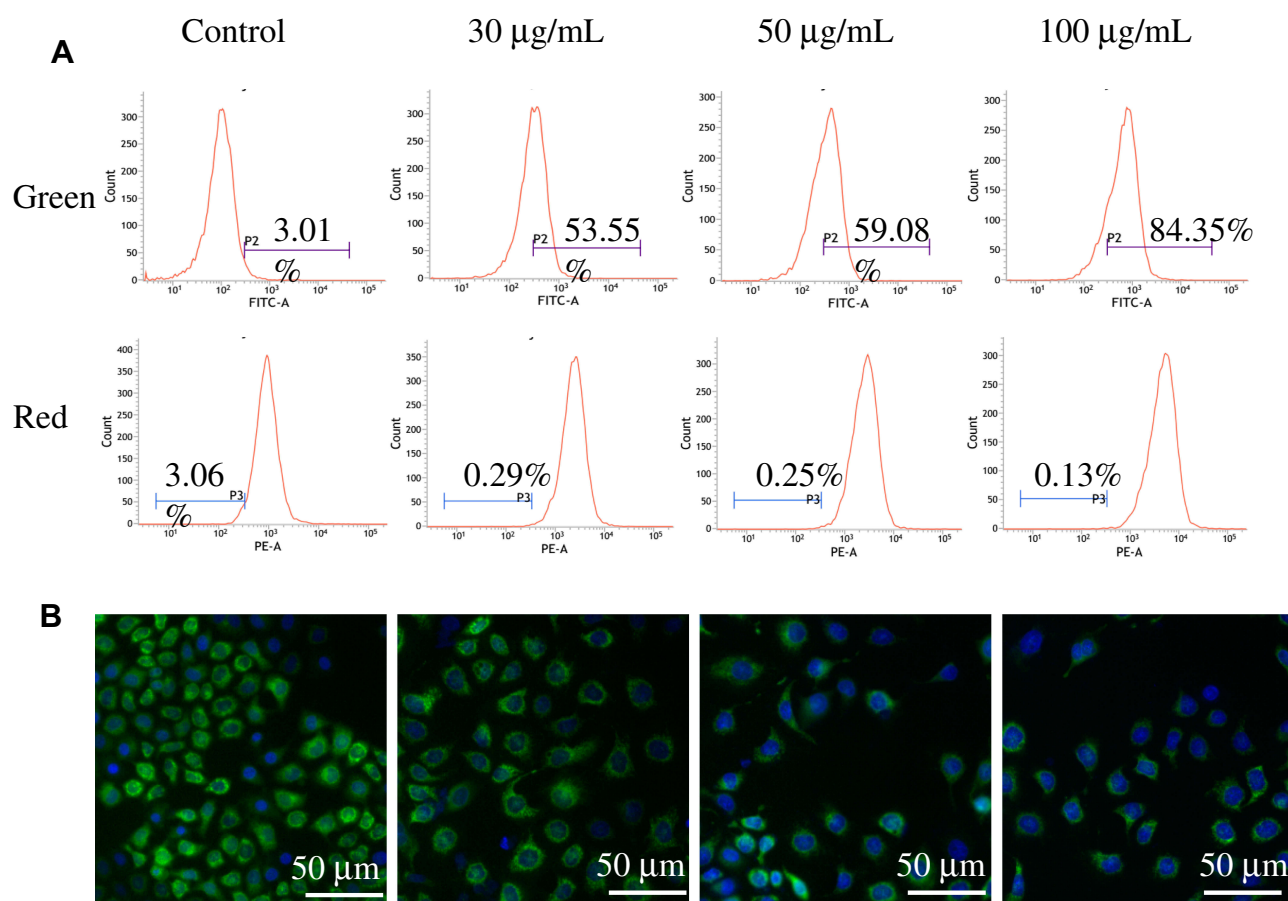


Figure 6 Effects of QDs on MMP of HeLa cells. **(A)** Flow cytometry images (JC-1 staining) and **(B)** fluorescence microscopy images (Rhodamine 123 staining).

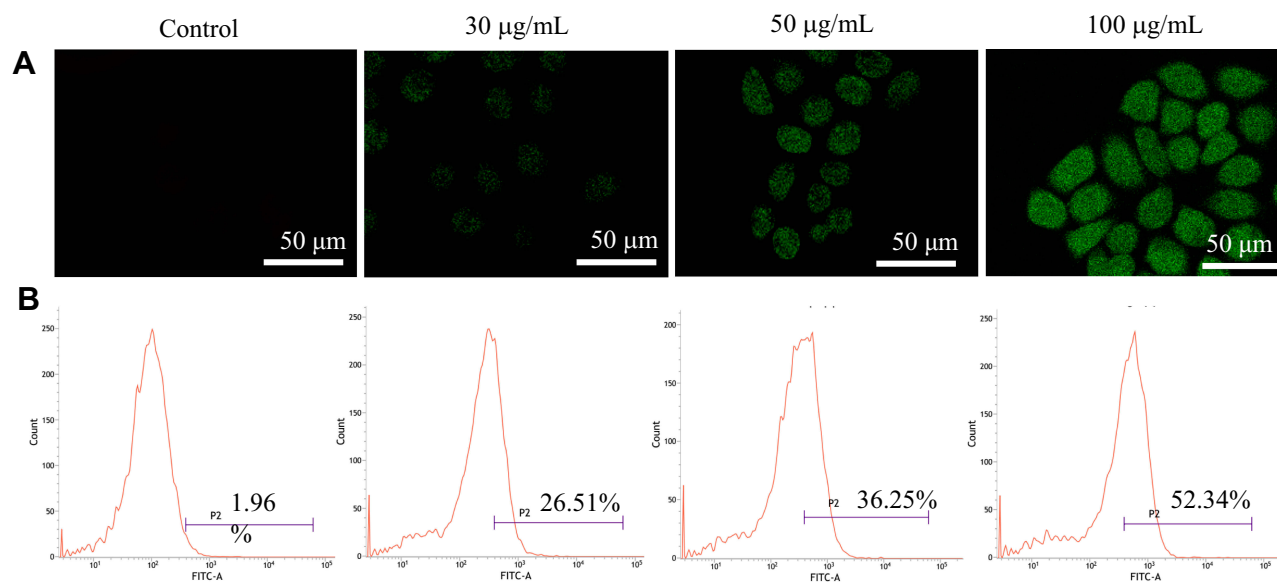


Figure 7 The determination of intracellular level of ROS in HeLa cells using H₂DCF-DA reagent through **(A)** fluorescence microscope and **(B)** flow cytometry.

the results suggest that the modified QDs could induce apoptosis in HeLa cells through depolarization of MMP.

Determination of ROS Generation in HeLa Cells Treated with Modified QDs

To investigate whether the mitochondrial membrane damage in HeLa cells in the presence of the modified QDs was mediated by the excessive intracellular ROS generation, fluorescence microscopy, and flow cytometry were employed using H₂DCF-DA reagent. As shown in Figure 7A, the intensity of green fluorescence (H₂DCF-DA staining) in modified QDs-treated cells was significantly higher as compared to the control experiment, indicating that QDs could induce the excessive formation of ROS inside cells. The flow cytometry experiment further confirmed that the intracellular ROS level was significantly increased in cells treated with the modified QDs in a dose-dependent manner as compared to the control cells (Figure 7B). These results depict that the modified QDs mediated mitochondrial dysfunction was likely due to the production of excessive intracellular ROS, leading to apoptosis process.

Conclusions

In conclusion, a new type of L-Cys-TAEA-5-FUA modified QDs was synthesized. The modified QDs exhibited improved water solubility and dispersibility. Moreover, the modified QDs exerted efficient inhibition of cancer cell proliferation as compared to the normal cells, indicating their antitumor activity. Post cellular internalization, the modified QDs could enter intracellular and they could effectively induce the formation of excessive intracellular ROS in cancer cells, leading to initiate the apoptosis process. The present study depicts that the modified QDs could be used as potent therapeutic agents to treat different types of cancers with fluorescent imaging.

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Author Contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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