

Long Non-Coding RNA GAS5 Targeting microRNA-21 to Suppress the Invasion and Epithelial–Mesenchymal Transition of Uveal Melanoma

This article was published in the following Dove Press journal:
Cancer Management and Research

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Objective: Human uveal melanoma (UM) is a common ocular malignant tumor with a high risk of metastasis. Emerging evidence indicates that long non-coding RNAs (lncRNAs) are correlated with the development of UM. Here, we aimed to determine the biological significance of lncRNA growth arrest-specific transcript 5 (GAS5) in UM.

Methods: The expression levels of GAS5 and microRNA-21 (miR-21) in UM tissues and cells were detected by qRT-PCR analysis. CCK-8 assay was performed to investigate the viability of UM cells after cell transfections, and the migration and invasion of UM cells were determined by transwell assay. The protein expression levels were detected by Western blot assay. The relationship between miR-21 and GAS5 in UM cells was confirmed by bioinformatics prediction and luciferase report assay.

Results: Our experiments demonstrated that GAS5 was markedly downregulated in UM cells and clinical specimens. Overexpression of GAS5 inhibited, whereas knockdown of GAS5 promoted the viability, migration, and invasion of UM cells. The epithelial-to-mesenchymal transition (EMT) process of UM cells was also suppressed by upregulating of GAS5 and enhanced by downregulating of GAS5. Additionally, as a competitive endogenous RNA (ceRNA), GAS5 directly binded to the oncogenic miR-21 in UM cells, and overexpression of miR-21 attenuated the EMT-suppressing effect of GAS5.

Conclusion: Taken together, our findings suggest that GAS5/miR-21 axis is implicated in the pathogenesis of UM and might serve as a potential therapeutic target.

Keywords: GAS5, uveal melanoma, invasion, epithelial–mesenchymal transition, microRNA-21

Introduction

Uveal melanoma (UM), arising from the iris, ciliary body, or choroid of the eye, is the most common primary intraocular tumor in adults.¹ UM has a high ability to metastasize, most commonly to the liver initially.² Despite advances in the diagnosis and therapy of UM, the prognosis of those diagnosed with metastatic UM remains disappointing, with the median overall survival of only 2–15 months.³ Accordingly, investigation of the underlying mechanisms involved in UM development and metastasis is urgently required.

Long non-coding RNAs (lncRNAs), a class of noncoding transcripts longer than 200 nucleotides with limited or no protein-coding capacity, are emerging as new

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players in various human cancers through acting as molecular sponges or competing endogenous RNAs (ceRNAs) to weaken the suppressive effect of microRNAs (miRNAs) on protein-coding genes.⁴ Growth arrest-specific transcript 5 (GAS5), a 651-bp lncRNA located on chromosome 1q25.1 (https://www.ncbi.nlm.nih.gov/nucore/NR_002578.2), has previously been reported to be under-expressed and function as a tumor suppressor in various human cancers, including gastric cancer,⁵ ovarian cancer,⁶ non-small cell lung cancer⁷ and cervical cancer.⁸ In detail, GAS5 inhibits tumorigenesis and enhances radiosensitivity in non-small cell lung cancer cells via suppressing miR-135b expression.⁷ And, GAS5 functions as a ceRNA of miR-222-3p to increase *PTEN* level, leading to the inhibition of papillary thyroid carcinoma progression.⁹ However, the expression pattern, biological function and the molecular mechanisms of GAS5 in UM progression have not been fully elucidated.

In the present study, we report for the first time, the potential role and underlying mechanisms of GAS5 in UM. Our results demonstrate that GAS5 is downregulated in UM, and overexpression of GAS5 inhibits the viability, migration, and invasion of UM cells. Thus, we suggest that GAS5 might be a promising therapeutic target for UM.

Materials and Methods

Clinical Samples

Eight enucleated eyes were collected from the patients with large UM lesions and stored in a liquid nitrogen tank (−196°C). Normal uveal samples were obtained from the Beijing Tongren Eye Bank (Beijing, China). The use of human tissue samples was approved by the Ethic Committee of the First Affiliated Hospital of Zhengzhou University. The informed written consents were obtained from patients or their relatives as the eyesight for some patients was too low.

Cell Culture and Transfection

Human UM cell lines MUM-2B and C918 were purchased from Chinese Academy of Sciences Cell Bank (Shanghai, China) and cultured in RPMI 1640 medium (KeyGene, Nanjing, China) with 10% fetal bovine serum (FBS; HyClone, Logan, UT, USA) and 50 µg/mL penicillin/streptomycin in humidified air at 37°C with 5% CO₂. Human melanocytes (D78) obtained from Chinese Academy of Sciences Cell Bank were culture in Dulbecco's modified Eagle's medium (DMEM; KeyGene).

The GAS5 sequence was synthesized according to the full-length GAS5 sequence and then sub-cloned into a pcDNA3.1 vector (Invitrogen, Shanghai, China). GAS5 siRNAs negative control siRNA (si-NC) were specifically synthesized by Shanghai GenePharma Co, Ltd. (Shanghai, China). The sequences of the GAS5 siRNAs were as follows: si-GAS5 (#1), AGTGTGGCTCTGGATAGCAC CTTAT; si-GAS5 (#2), AGGAAGGATGAGAATAG CTACTGAA; si-GAS5 (#3), CAGTGTGGCTCTGGA TAGCACCTTA. miR-21 mimics and mimics negative control were purchased from Genecopoeia (Guangzhou, China). Cells at 70~80% confluence were selected for transfection using Lipofectamine 2000 reagent (Invitrogen). Forty-eight hours after transfection, the cells were collected for further analysis.

RNA Isolation and qRT-PCR Analysis

Total RNA was extracted from the tissues and cells using TRIZOL reagent (Invitrogen). For mRNA and lncRNA, RNA was reverse transcribed to cDNA using the PrimeScript™ RT reagent kit (Takara, Dalian, China). For miRNA, reverse transcription was performed using the TaqMan MicroRNA RT kit (Applied Biosystems, Foster City, CA, USA). qPCR was then performed using SYBR® Premix Ex Taq™ II (Takara) on the ABI PRISM 7000 Fluorescent Quantitative PCR System (Applied Biosystems). U6 or GAPDH was used as an internal control. The relative expression of genes is presented as fold change and calculated using the $2^{-\Delta\Delta C_t}$ method.¹⁰ The primer sequences are listed in Table 1.

Western Blot Assay

Proteins were extracted from the cells using RIPA protein extraction reagent (Sigma, St. Louis, MO, USA) containing the protease inhibitor cocktail (Roche, Mannheim, Germany). Equal quantity of proteins was then separated by SDS-PAGE gels, and transferred to PVDF membranes (Millipore, Billerica, MA, USA). Then, the membranes were blocked with 5% non-fat milk and incubated with specific primary antibodies against E-cadherin (1:1000, Cell Signaling, Beverly, MA, USA), Vimentin (1:1000, Cell Signaling), N-cadherin (1:1000, Cell Signaling), and β -actin (1:500, Santa Cruz, Dallas, TX, USA) at 4 °C overnight. Thereafter, the membranes were incubated with HRP-conjugated secondary antibodies (1:10,000, Santa Cruz) at room temperature for 1 hour. The bands were visualized using the enhanced chemiluminescence kit (Amersham, Little Chalfont, UK). Protein expression was

Table I Sequences of PCR Primers

Gene Name	Primer Sequences
GAS5 Forward primer	CTTCTGGGCTCAAGTGATCCT
GAS5 Reverse primer	TTGTGCCATGAGACTCCATCAG
GAPDH Forward primer	GTCAACGGATTTGGTCTGTATT
GAPDH Reverse primer	AGTCTTCTGGGTGGCAGTGAT
miR-21-RT	GTCGTATCCAGTGCAGGGTCCGAGGTATTTCGCACTGGATACGACTCAACA
U6-RT	CGCTTCACGAATTTGCGTGTTCAT
miR-21 Forward primer	TAGCTTATCAGACTGA
miR-21 Reverse primer	CTGGAGCAGCACAGCCAATA
U6 Forward primer	GCTTCGGCAGCACATATACTAAAAT
U6 Reverse primer	CGCTTCACGAATTTGCGTGTTCAT

quantified by densitometry (Quantity One software; Bio-Rad, CA, USA), with β -actin used as a control.

CCK-8 Assay

Cell viability was determined over the next 24–96 hours following cell transfections using the Cell Counting Kit-8 (CCK-8; Beyotime, Haimen, China). Cells were plated at 3000 cells per well in 96-well plates. At the indicated time after transfection, 10 μ L CCK-8 solution was added into each well, and wells were incubated for another 2 hours. The absorbance was measured at 450 nm using a micro-titer plate reader.

Transwell Assay

Cell migration and invasion were determined by using transwell chambers (24-well insert; 8- μ m pore size; BD Biosciences, Franklin Lakes, NJ, USA). In brief, after transfection, 1×10^5 cells in 200 μ L serum-free medium were seeded in the uncoated or Matrigel (30 μ L; BD Biosciences)-coated upper chambers, and the lower chambers were filled with medium containing 10% FBS. After 48 hours of incubation, the cells on the bottom surface of the membrane were fixed with methanol, stained with 0.1% crystal violet (Solarbio, Beijing, China) for 10 min at room temperature, and then counted using a light microscope (Olympus, Tokyo, Japan).

Luciferase Report Assay

The GAS5 fragment containing the putative miR-21 binding site was amplified by PCR and cloned into a pGL3 promoter vector (Promega, Madison, WI, USA). The QuickChange[®] Site-Directed Mutagenesis Kit (Stratagene, La Jolla, CA, USA) was used to mutate the binding site of GAS5 and miR-21. C918 cells were seeded into 24-well plates and co-transfected with luciferase reporter vectors

(WT-GAS5 or MUT-GAS5) and miR-21 mimics or mimics negative control using Lipofectamine 2000 reagent (Invitrogen). Forty-eight hours post transfection, the cells were harvested, and the relative luciferase activities were measured using the Dual Luciferase Reporter Assay System (Promega). Renilla luciferase activity was normalized to firefly luciferase activity.

Statistical Analysis

Data are expressed as the mean \pm standard deviation (SD) from at least three separate experiments. Data analysis, graph generation, and statistical analysis were performed using Graphpad Prism (version 6.01) software (GraphPad Software, Inc., La Jolla, CA, USA). Differences between groups were assessed using either Student's *t*-test or one-way analysis of variance (ANOVA). The correlation between GAS5 and miR-21 expression was evaluated by Spearman correlation analysis. *P*-values of less than 0.05 were considered to be statistically significant.

Results

GAS5 is Downregulated in UM and Predicts a Poor Prognosis

To determine whether GAS5 plays a role in the development of UM, qRT-PCR analysis was performed to measure the expression of GAS5 in UM cells. As shown in [Figure 1A](#), the expression of GAS5 was significantly decreased in UM cell lines compared to that in D78 cells. Furthermore, GAS5 was also downregulated in UM tissues compared with that in normal uveal samples ([Figure 1B](#)). In addition, UM patients with GAS5 low expression presented with lower overall survival rates than patients with GAS5 high expression ([Figure 1C](#)).

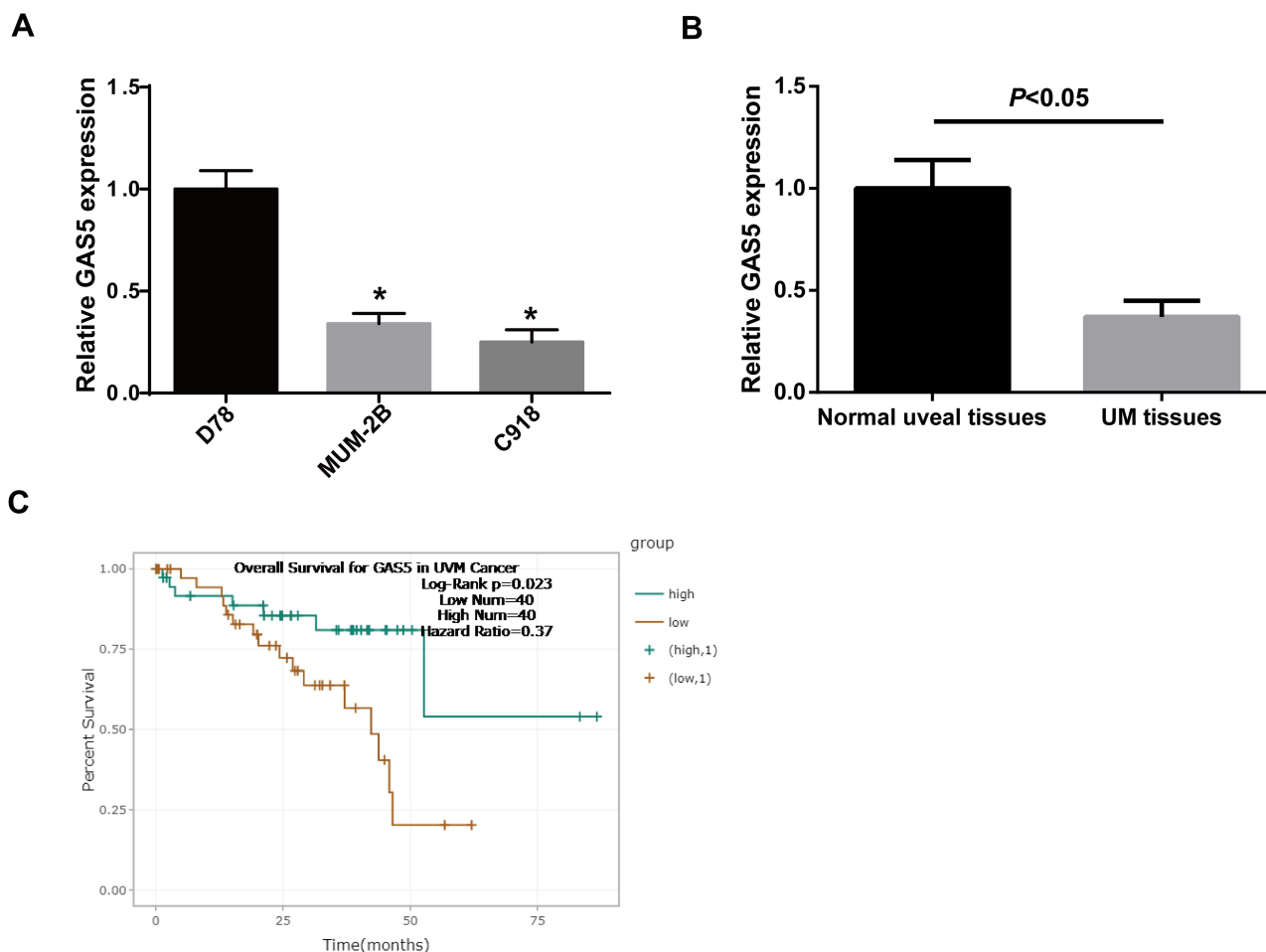


Figure 1 GAS5 is downregulated in UM. **(A)** qRT-PCR analysis was conducted to quantify the expression levels of GAS5 in UM cell lines and D78 cells. **(B)** Expression levels of GAS5 in UM tissues and normal uveal tissues were determined by qRT-PCR analysis. **(C)** Analysis of the relationship between GAS5 expression levels and the overall survival rates of UM patients using starBase. Data are shown as mean \pm SD. * $P<0.05$.

GAS5 Inhibits UM Cell Viability and Invasion

To further explore the biological role of GAS5 in UM progression, GAS5 was knocked-down in MUM-2B cells by using GAS5 siRNAs (Figure 2A), and overexpressed by transfecting pcDNA3.1-GAS5 into C918 cells (Figure 2B) as C918 cells showed the lower expression level of GAS5 than MUM-2B. si-GAS5 (#2) was used for all the RNAi experiments in this study. UM cell viability was then detected by CCK-8 assay at 24, 48, 72, and 96 hour time points and found that increased GAS5 expression significantly inhibited the viability of C918 cells, whereas knockdown of GAS5 in MUM-2B cells resulted in a significant increase in cell viability (Figure 2C). GAS5 effects on migratory and invasive abilities of UM cells were then assayed. After knockdown of GAS5, the migratory and invasive abilities were remarkably enhanced in MUM-2B cells, but was suppressed by ectopic expression of GAS5 in C918 cells (Figure 2D).

GAS5 Suppresses EMT Process in UM Cells

EMT is vital for cell invasion, therefore we determined the effects of GAS5 on EMT in UM cells. The results show that the expression levels of Vimentin and N-cadherin were significantly increased whereas E-cadherin expression was reduced when GAS5 was knocked down in MUM-2B cells (Figure 3A). Ectopic expression of GAS5 in C918 cells inhibited the expression levels of N-cadherin and Vimentin, but promoting the E-cadherin expression (Figure 3B).

GAS5 Functions as a ceRNA for miR-21 in UM

Using an online database (<http://cbcsrv.watson.ibm.com/rna22.html>), we found that GAS5 has one predicted miR-21 target binding site (Figure 4A). Additionally, as shown in

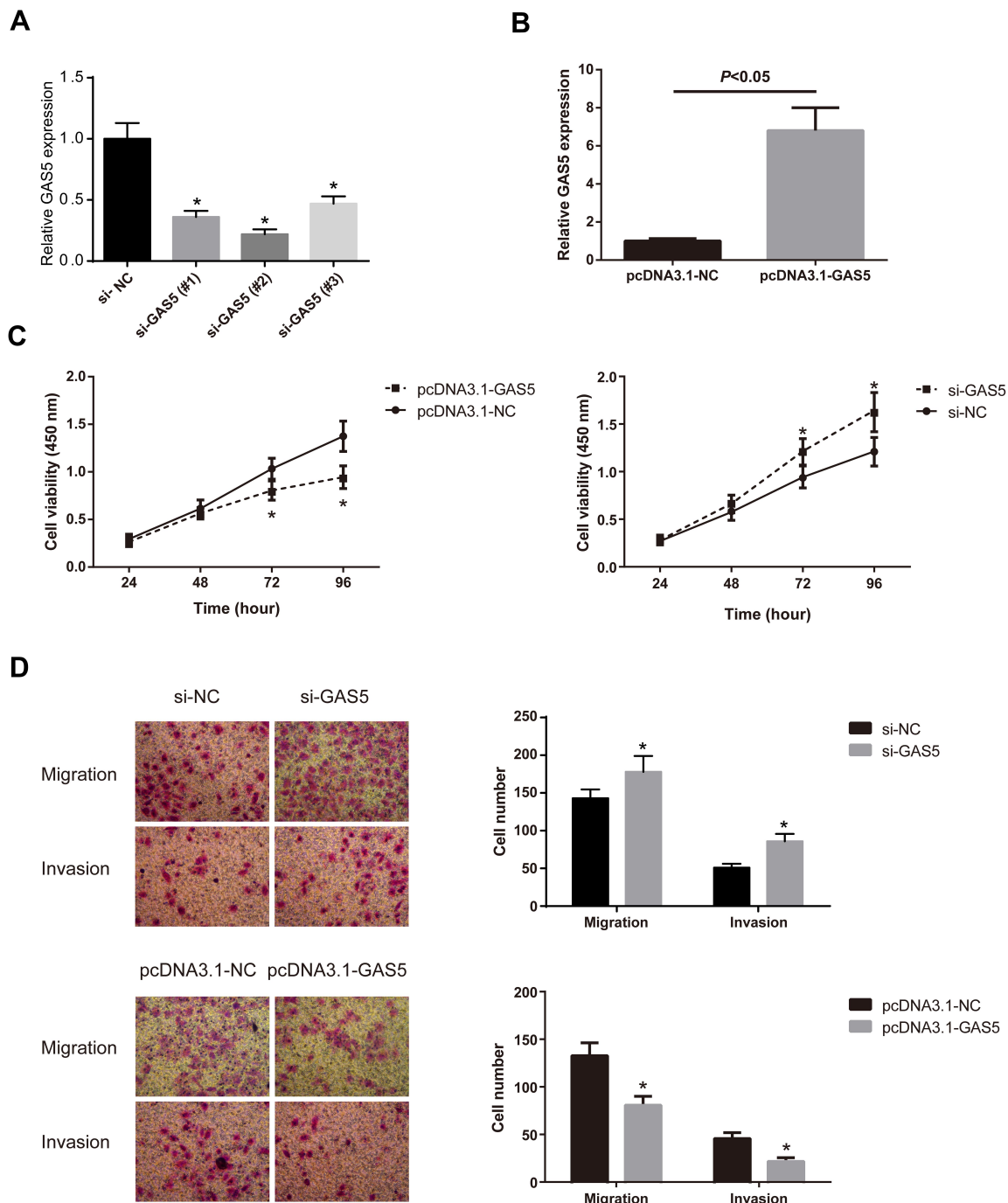
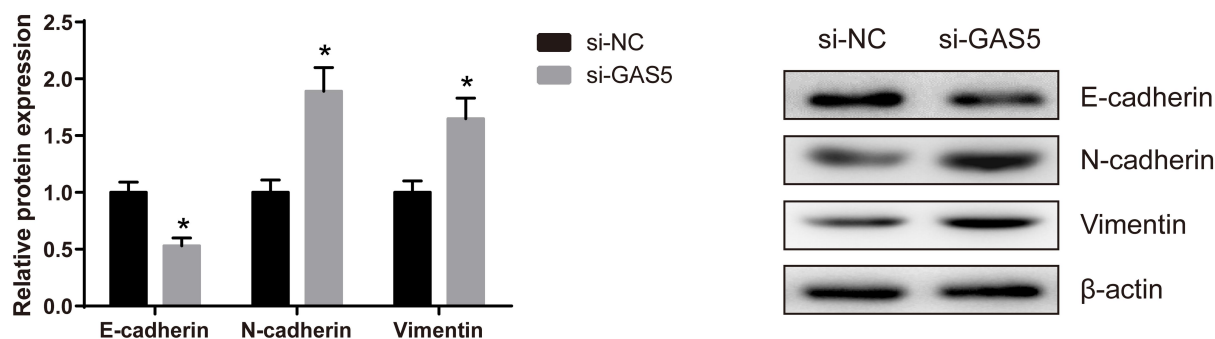


Figure 2 GAS5 inhibits UM cell viability and invasion. (A) qRT-PCR analysis of GAS5 expression in MUM-2B cells transfected with GAS5 siRNAs. (B) qRT-PCR analysis of GAS5 expression in C918 cells transfected with pcDNA3.1-GAS5. (C) CCK-8 assay was performed to detect the viability of pcDNA3.1-GAS5-transfected C918 cells and si-GAS5-transfected MUM-2B cells. (D) Transwell assay was performed to detect the migration and invasion abilities of pcDNA3.1-GAS5-transfected C918 cells and si-GAS5-transfected MUM-2B cells. Data are shown as mean \pm SD. * $P < 0.05$.

Figure 4B, GAS5 could negatively regulate the expression of miR-21 in UM cells. The luciferase reporter assay manifested that miR-21 significantly suppressed luciferase activity that carried WT but not MUT 3'-UTR of GAS5

(Figure 4C). Further analysis revealed that the expression of miR-21 was increased in UM tissues (Figure 4D), and the increased miR-21 expression was closely correlated with the decreased GAS5 expression in UM tissues (Figure 4E).

A



B

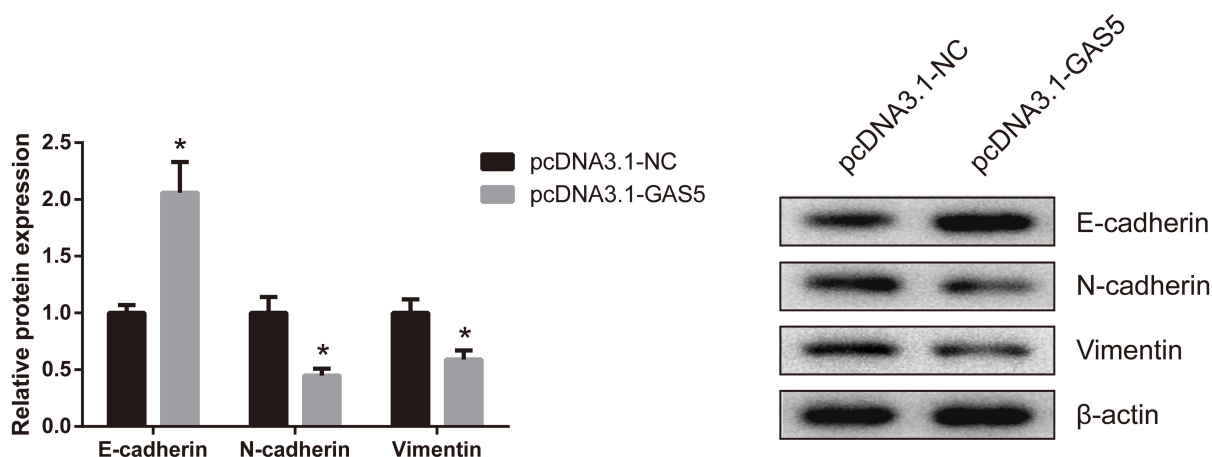
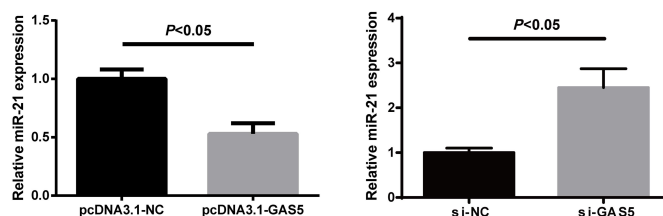


Figure 3 GAS5 suppresses EMT in UM cells. **(A)** Expression levels of E-cadherin, N-cadherin and Vimentin were measured by Western blot assay in si-GAS5-transfected MUM-2B cells. **(B)** Expression levels of E-cadherin, N-cadherin and Vimentin were measured by Western blot assay in pcDNA3.1-GAS5-transfected C918 cells. Data are shown as mean \pm SD. * $P < 0.05$.

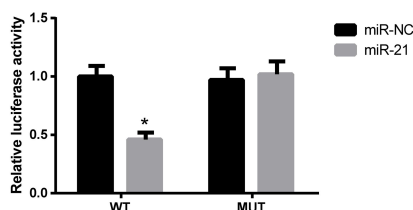
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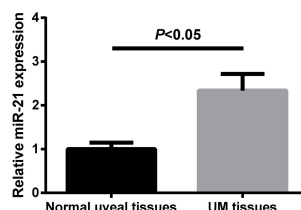
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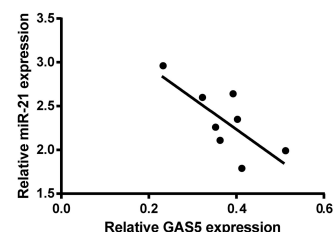


Figure 4 GAS5 functions as a ceRNA of miR-21 in UM. **(A)** The binding site of miR-21 on GAS5. **(B)** miR-21 expression in UM cells after overexpression or knockdown of GAS5 was measured by qRT-PCR analysis. **(C)** Luciferase activity in C918 cells co-transfected with miR-21 mimics and luciferase reporters containing GAS5-WT or GAS5-MUT (* $P < 0.05$). **(D)** Expression levels of miR-21 in UM tissues and normal uveal tissues were determined by qRT-PCR analysis. Data are shown as mean \pm SD. **(E)** The correlation between miR-21 and GAS5 expression in UM tissues was determined.

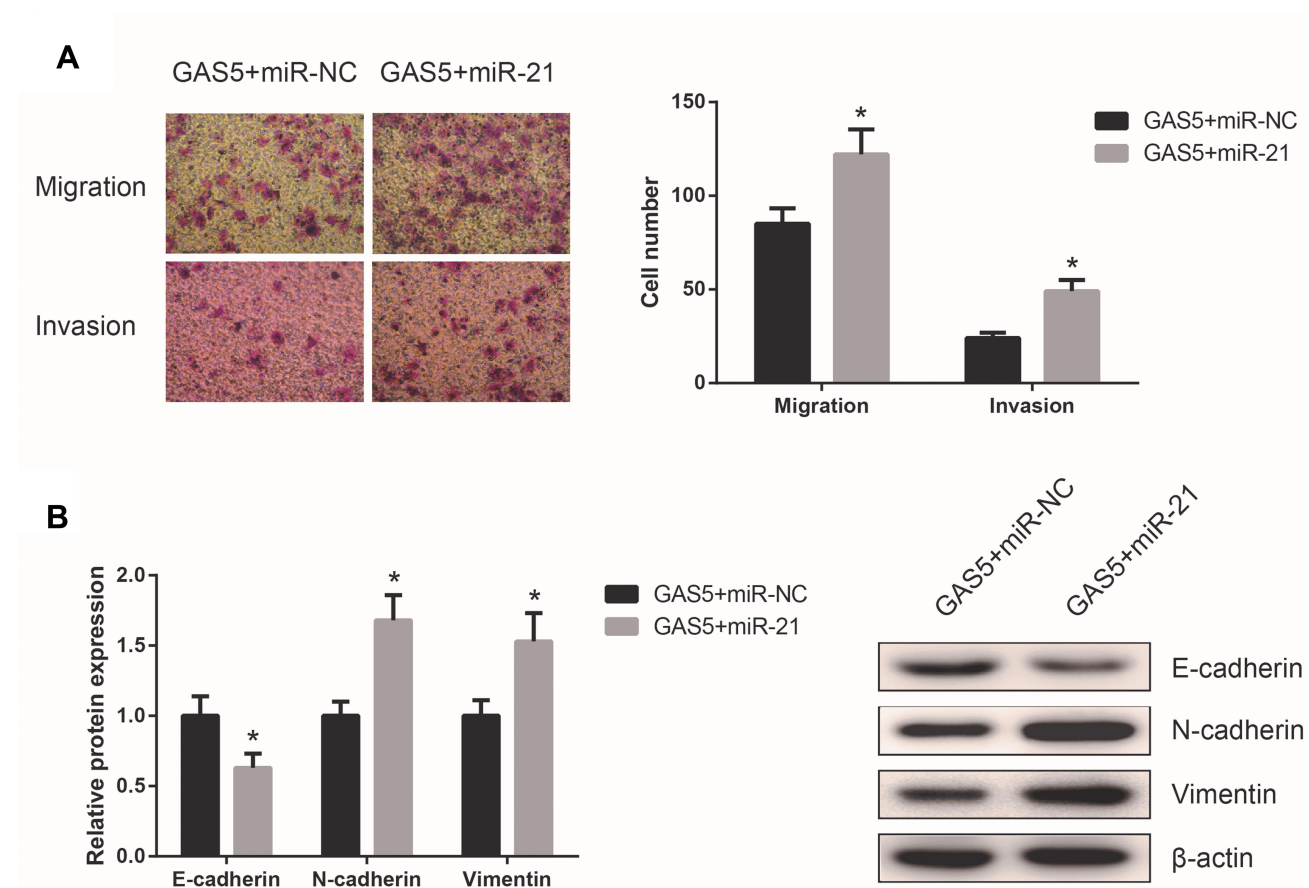


Figure 5 miR-21 attenuates the function of GAS5 in UM. **(A)** Transwell assay was performed to detect the migration and invasion of C918 cells co-transfected with pcDNA3.1-GAS5 and miR-21 mimics. **(B)** Expression levels of E-cadherin, N-cadherin, and Vimentin were measured by Western blot assay in C918 cells co-transfected with pcDNA3.1-GAS5 and miR-21 mimics. Data are shown as mean \pm SD. * $P < 0.05$.

miR-21 Attenuates the Function of GAS5 in UM

To further determine the role of miR-21 in the GAS5-mediated effects on UM cells, rescue experiments were subsequently performed. We found that upregulation of miR-21 abolished GAS5-induced inhibition of the migration and invasion of C918 cells (Figure 5A). Furthermore, miR-21 rescued repression of EMT process caused by GAS5 overexpression in C918 cells (Figure 5B).

Discussion

In addition to microRNAs, lncRNAs have emerged as critical players in UM progression.^{11–13} Altered GAS5 expression is related to many human cancers. This study might be the first direct investigation of the relationship between GAS5 expression and UM. Here, we observed that GAS5 is downregulated in UM tissue samples and cell lines, and overexpression of GAS5 markedly inhibited the viability, migration and invasion of UM cells in vitro. These findings indicate that GAS5 might function as a tumor suppressor in UM.

Cancer metastasis is a complex process involving multiple cellular steps. Featured by the loss of intracellular junctions and acquirement of mesenchymal characteristics, EMT is a key process for the dissemination of malignant cells to adjacent tissues and distant sites.^{14,15} EMT-associated factors could promote invasive properties of UM cells.¹⁶ A recent study reported that overexpression of GAS5 could inhibited the EMT of osteosarcoma cells¹⁷ In this study, we observed that the expression levels of EMT-associated proteins, including N-cadherin and vimentin, were reduced in GAS5-overexpressed UM cells, indicating that GAS5 inhibited human UM metastasis by suppressing EMT process.

Recently, a large body of experimental evidence has shown that lncRNAs can act as either ceRNAs which bind to miRNAs to influence miRNAs-induced gene silencing or natural miRNA sponges to regulate gene expression, playing critical roles in human diseases.^{18,19} The direct regulatory association between GAS5 and miR-21 was previously reported.^{20,21} As a well-known oncogenic miRNA, miR-21 was also found to be upregulated in

formalin-fixed, paraffin-embedded UM samples.²² In the present study, through bioinformatics prediction and functional assays, we lend credence to the previous study, that lncRNA GAS5 could directly bind to miR-21 in UM, and that overexpression of miR-21 could attenuate the effects of GAS5 on UM cells.

PTEN is a well-known tumor suppressor, and plays a tumor-suppressive role during UM pathogenesis.²³ Loss of *PTEN* expression is associated with shortened disease-free survival of UM patients.²³ It has been reported that miR-21 induced EMT in lung cancer cells via targeting *PTEN*.²⁴ Therefore, we conjecture that GAS5 may function as a ceRNA of miR-21 to induce *PTEN* expression, resulting in the suppressions of invasion and EMT in UM. We intend to clarify it in following assays.

In summary, this study identifies GAS5 serves as a novel potential cancer suppressor in UM. GAS5 might act as a ceRNA to inhibit the EMT process by sponging miR-21. Our findings provide a more adequate theoretical basis for understanding the molecular pathology of UM, and describe a novel GAS5/miR-21/EMT regulatory network involved in UM development.

Disclosure

The authors report no funding and no conflicts of interest for this work.

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