

RETRACTED ARTICLE: Inhibition of Circulating Exosomal miRNA-20b-5p Accelerates Diabetic Wound Repair

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Purpose: Efficient approaches to reliably improving wound healing in diabetic patients remain to be developed. Exosomes are nanomaterials from which therapeutically active microRNAs (miRNAs) can be isolated. In the present report, we therefore isolated circulating exosome-derived miRNAs from patients with diabetes and assessed the impact of these molecules on wound healing.

Patients and Methods: Exosomes were isolated from the serum of control or diabetic patients (Con-Exos and Dia-Exos, respectively), after which the effects of these exosomes on cellular activity and wound healing were assessed.

Results: We determined that miR-20b-5p was overexpressed in Dia-Exos and that it functioned by impairing wound repair by suppressing vascular endothelial growth factor A (VEGFA) expression. Consistent with such a model, the administration of Dia-Exos or this miRNA both in vivo and in vitro was sufficient to slow wound repair.

Conclusion: Dia-Exos exhibit significant increases in miR-20b-5p relative to Con-Exos, and this miRNA can be transferred into HSFs wherein it can suppress VEGFA expression and thereby slow the process of wound healing.

Keywords: nanomedicine, exosome, miRNA, diabetes, wound, fibroblast

Introduction

Diabetes is an increasingly prevalent and debilitating condition that is common in older populations.¹ Diabetic foot ulcers (DFUs) are a serious complication of this metabolic disease that arises as a consequence of impaired wound healing, resulting in secondary infections and the potential need for limb amputation, thus negatively impacting patient quality of life.² Many efforts have been made to explore the mechanistic basis for DFU onset in order to improve patient healing activity.³ In one study, the lncRNA H19/miR-152-3p/PTEN axis was shown to function as a critical regulator of wound repair in individuals with diabetes, indicating that it may be a viable therapeutic target in the context of DFU development.⁴ Despite these findings, however, the mechanisms that constrain diabetic wound healing remain incompletely understood, and effective treatments for affected patients are lacking.⁵ Novel approaches to the study of diabetic wound healing are thus required to facilitate novel therapeutic strategies.

Functional fibroblasts are integral to rapid wound healing,⁶ and the proper functionality of these cells is governed by diverse regulatory factors.⁷ For sample, WNT-inducible signaling pathway protein-1 (WISP-1) can promote the migratory activity and proliferation of dermal fibroblasts, thereby bolstering wound healing

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and reducing the adverse inflammatory effects of TNF- α .⁸ Exosomes (Exos) are small membrane-enclosed vesicles that are 30–150 nm in size and that can regulate a wide range of biological processes by delivering specific proteins, nucleic acids, and microRNAs (miRNAs) to different cells and organs in vivo.^{9,10} For example, human mesenchymal stem cell (MSC)-derived Exos can deliver miR-146a-5p to other cells, thereby suppressing innate lymphoid cell-dominant allergic airway inflammation.¹¹ Such regulatory mechanisms are increasingly well understood and are linked to the ability of miRNAs, which are non-coding RNAs, to suppress target gene expression in recipient cells following Exo-mediated delivery.¹² In one recent report, miR-20b-5p levels were shown to be increased in the circulating Exos of diabetes patients, and such upregulation was associated with insulin regulation in the skeletal muscle of humans.¹³

Herein, we explored the ability of Exo-derived miR-20b-5p to alter fibroblast functionality in the context of diabetic wound repair. Vascular endothelial growth factor A (VEGFA) is known to serve as a potent regulatory cytokine capable of promoting blood vessel permeability, enhancing cellular proliferation, and suppressing apoptotic cell death.¹⁴ VEGFA signaling is also closely linked to fibroblast functionality.¹⁵ Herein, we isolated Exos from control and diabetes patients (Con-Exos and Dia-Exos, respectively) and evaluated the impact of these molecules on human skin fibroblasts (HSFs) and animal models of wound repair. We also explored the relationship between VEGFA and the mechanisms whereby Exo-derived miR-20b-5p impacts wound healing.

Materials and Methods

Ethical Oversight

The institutional review board of Tongji Hospital, Tongji University School of Medicine approved all human research aspects of this study. The Laboratory Animal Management Committee of Tongji Hospital, Tongji University School of Medicine approved all animal studies.

Exosome Isolation and Assessment

Samples of blood were obtained from 10 control patients and 10 diabetes patients (all male) in citrate phosphate dextrose-containing tubes. After collection, blood was centrifuged for 15 min at 3000 \times g at 4°C. Serum was then isolated and spun again with the same settings.

Supernatants were then collected and spun for 30 min at 10,000 \times g, after which they were spun for 70 min at 100,000 \times g. Isolated Exos were washed thrice with PBS and spun under the same ultracentrifugation settings prior to resuspension in 15 mL of PBS and ultrafiltration with a 15-mL Amicon Ultra-15 Centrifugal Filter (Millipore, MA, USA) by spinning the solution at 4000 \times g to a final 200 μ L volume.

The morphological properties of isolated Exos were assessed by mixing them for 30 min with 4% osmium tetroxide and then applying them to a copper grid. Particles were then stained using 1% phosphotungstic acid and assessed via transmission electron microscopy (TEM). The expression of miR-20b-5p in Exo preparations was evaluated via qPCR. When performing experimental treatments using the Exos, Exo samples from each group were pooled to ensure consistency. All experiments were repeated in triplicate.

Animals and Treatments

C57BL/6 mice (male, 6 weeks-old, 18–20 g; 60 total) were anesthetized with pentobarbital sodium (50 mg/kg, i.p., Sigma-Aldrich, MO, USA). Mice were then shaved, and a full-thickness cutaneous wound (10 mm in diameter) was generated on the dorsal surface for each mouse. After wounding, animals were separated at random into two experimental cohorts, with three groups per cohort. Animals in the first cohort were treated with either PBS (100 μ L of PBS, applied to wounds), Con-Exos (100 μ L at 50 μ g/mL, applied to wounds), or Dia-Exos (100 μ L at 50 μ g/mL, applied to wounds). Mice in the second cohort were treated with either PBS (as above), agomiR-20b-5p (100 μ L at 20 μ mol/L, applied to wounds), or antagomiR-20b-5p (100 μ L at 20 μ mol/L, applied to wounds). For all treatments, wounds were subcutaneously injected at four different points (25 μ L each) on days 0, 3, 5, 7, 9, and 11 after wound generation. In addition, the ImageJ software was used to assess wound closure on days 0, 3, 5, 7, 10, and 14 based upon the following formula:

$$Cn = (A0 - An) / A0 \times 100\%$$

where Cn corresponds to the reduction in wound area (as a percentage) on the indicated days, A0 corresponds to the baseline wound area, and An corresponds to the wound area on the indicated day of analysis.

All mice were euthanized on day 14 after wound generation, and samples of skin were collected. For experimental models of diabetes, mice were fed a high-fat diet for 4 weeks and were then administered streptozotocin

(STZ; 40 mg/kg/day i.p.) once per day for seven days. When mice had a fasting blood glucose >11.1 mmol/L in two consecutive assay measurements, they were deemed diabetic.

Cell Culture

HSFs (FuHeng Biology, Shanghai, China) were grown in high-glucose DMEM (Gibco, NY, USA) containing 10% fetal bovine serum (FBS) at 37°C in a humidified 5% CO₂ incubator. Cells were transduced with agomiR/antagomiR constructs (200 mM; GenePharma, Shanghai, China) or siRNA-VEGFA (50 nM; RiboBio, Guangzhou, China) using Lipofectamine 3000 (Thermo Fisher Scientific, MA, USA) based on provided directions.

CCK-8 Assay

To evaluate proliferation and viability, HSFs were added to 96-well plates for 24, 48, or 72 h, after which the CCK-8 reagent was added to each well for 2 h in serum-free media. Absorbance was then evaluated at 450 nm.

Luciferase Reporter Assay

WT or mutant (MUT) versions of the VEGFA 3'-UTR region containing the predicted binding site for miR-20b-5p were cloned into the pMIR luciferase reporter plasmid. Lipofectamine 3000 was then used to co-transfect cells with these plasmids and agomiR-20b-5p. Following a 24 h incubation, the luciferase activity in these cells was assessed with a Dual-Luciferase Reporter Assay kit (Promega, Madison, WI, USA), and data were analyzed using the SpectraMax M5 program (Molecular Devices, CA, USA).

ELISA

Basic fibroblast growth factor (bFGF) levels in HSFs were measured following a 48 h growth period in serum-free media with a sandwich ELISA kit (R&D, USA). Cells in collected samples were counted, and ELISA assay results were normalized to cell counts, with bFGF levels being determined using an appropriate standard curve. This same approach was also used to measure VEGFA levels in individual samples.

Western Blotting

Lysates were prepared from cells or supernatants, were diluted 1:5 with loading buffer, and were boiled for 5 min at 95°C. Samples of protein were then separated via SDS-PAGE, transferred to PVDF membranes, and blots were blocked using 5% non-fat milk in TBST for 1 h at room temperature. Blots were then probed overnight with anti-

CD81 (1:500; ab59477), anti-VEGFA (1:1000; ab52917), TSG101 (1:1000, ab83) or anti-GAPDH (1:10,000, ab37168) (all from Abcam, Cambridge, UK). Blots were then probed using HRP-linked secondary antibodies at 37°C for 1 h, and protein bands were detected via LiDE110 scanner (Canon, Tokyo, Japan), with the AlphaEaseFC software (Alpha Innotech, Genetic Technologies, Inc, FL, USA) being used to measure band density.

qPCR

TRIzol (Invitrogen) was used to extract total from samples, after which cDNA was prepared from 1 µg total of RNA per sample with the Verso cDNA Synthesis Kit (Thermo Fisher Scientific). A SeraMir Exosome RNA Purification Kit (System Bioscience, CA, USA) was utilized to isolate miRNAs from Exo preparations, after which cDNA was prepared with the Taqman microRNA assay kit (Applied Biosystems, CA, USA). All qPCR reactions were conducted using a StepOne Real-Time PCR Instrument (Life Technologies, CA, USA). The 2^{-ΔΔCT} approach was used to evaluate relative gene expression, while GAPDH served as a normalization control. All primers are shown in Table 1.

Statistical Analysis

All assays were repeated in triplicate, and data are means ± SD. Student's t-tests and one-way ANOVAs were employed to compare data with GraphPad Prism 8.0 (GraphPad Software, CA, USA). P<0.05 was the significance threshold.

Results

Exosome Characterization

Con-Exos and Dia-Exos were assessed via TEM and Western blotting. In line with the results of prior studies, we found these particles to be cup- or sphere-like in shape (Figure 1A and C).¹⁶ Western blotting revealed these Exos to be enriched for exosomal surface marker proteins including CD81 and TSG101 (Figure 1B and D). As such, we concluded that these preparations were Exos, and they were utilized for subsequent experiments.

Dia-Exos Suppress the in vitro Functionality of HSFs

HSF morphology was assessed via fluorescent staining, revealing these cells to have ovoid nuclei and a flattened spindle-like or stellate shape. These cells were then treated with Con-Exos or Dia-Exos, and their proliferation was assessed via CCK-8

Table 1 microRNAs and mRNA Primer Sequences

microRNA or Gene Names	Primer Sequence (5' to 3')
hsa-miR-20b-5p-Forward	CGCCATCAAAGTGCTCATAGTGC
hsa-miR-20b-5p-Reverse	ATCCAGTGCAGGGTCCGAGG
hsa-U6-Forward	CTCGCTTCGGCAGCACACA
hsa-U6-Reverse	AACGCTTCACGAATTTGCGT
hsa-Cyclin D1-Forward	TTGCCCTCTGTGCCACAGAT
hsa-Cyclin D1-Reverse	TCAGGTTTCAGGCCTTGCACT
hsa-Cyclin D3-Forward	CTGGCCATGAACTACCTGGA
hsa-Cyclin D3-Reverse	CCAGCAAATCATGTGCAATC
hsa-VEGFA-Forward	TGCAGATTATGCGGATCAAACC
hsa-VEGFA-Reverse	TGCATTACATTTGTTGTGCTGTAG
hsa- α -SMA-Forward	TGACCCAGATTATGTTTGAGACC
hsa- α -SMA-Reverse	CCAGAGTCCAGCACAATACCA
hsa-Col-1-Forward	AGGCCACGCATGAGCCGAAG
hsa-Col-1-Reverse	GCCATGCGTCAGGAGGGCAG
hsa-Col-3-Forward	AGGATCTGAGGGCTCGCCAGG
hsa-Col-3-Reverse	AGCCACCAGACTTTTCACCTCCA
hsa-MMPI-Forward	AAAGGGAATAAGTACTGGGC
hsa-MMPI-Reverse	CAGTGTTTTCTCAGAAAGAG
hsa-GAPDH-Forward	CCGTTGAATTGCGCGTGA
hsa-GAPDH-Reverse	TGATGACCCTTTGGCTCCC

assay. This analysis revealed that HSF treatment with 50–100 μ g/mL of Dia-Exos markedly suppressed the proliferation of these cells in a dose-dependent fashion relative to treatment with equivalent doses of Con-Exos (Figure 2A). Levels of

Cyclin D1 and *Cyclin D3*, which are associated with proliferation, were also decreased following Dia-Exo treatment of HSFs (Figure 2B and D). Dia-Exos also suppressed *Bcl-2* levels and enhanced *Bax* expression in these cells (Figure 2C and E). Dia-Exos also significantly enhanced α -SMA production and suppressed collagen I (Col 1) and collagen III (Col 3) expression (Figure 2F). ELISAs were further conducted, revealing that Dia-Exo-treated HSFs secreted lower levels of VEGFA and bFGF relative to Con-Exo-treated cells (Figure 2G and H). Together, these findings suggested that Dia-Exos can impair the functionality of HSFs in

Dia-Exos Suppress Wound Healing in vivo

Next, we local administered Dia-Exos to mice bearing full-thickness cutaneous dorsal wounds, revealing that such treatment markedly compromised the rate of wound closure relative to control treatment (Figure 3A and B). On day 14 post-wounding, skin samples were collected and levels of collagen-related markers were assessed via qPCR and Western blotting. This analysis revealed significant increases in α -SMA and significant decreases in Collagen 1 and 3 expression in Dia-Exo-treated mice relative to other treatment groups (Figure 3C and D). As such, we concluded that Dia-Exos can suppress the wound healing process in vivo.

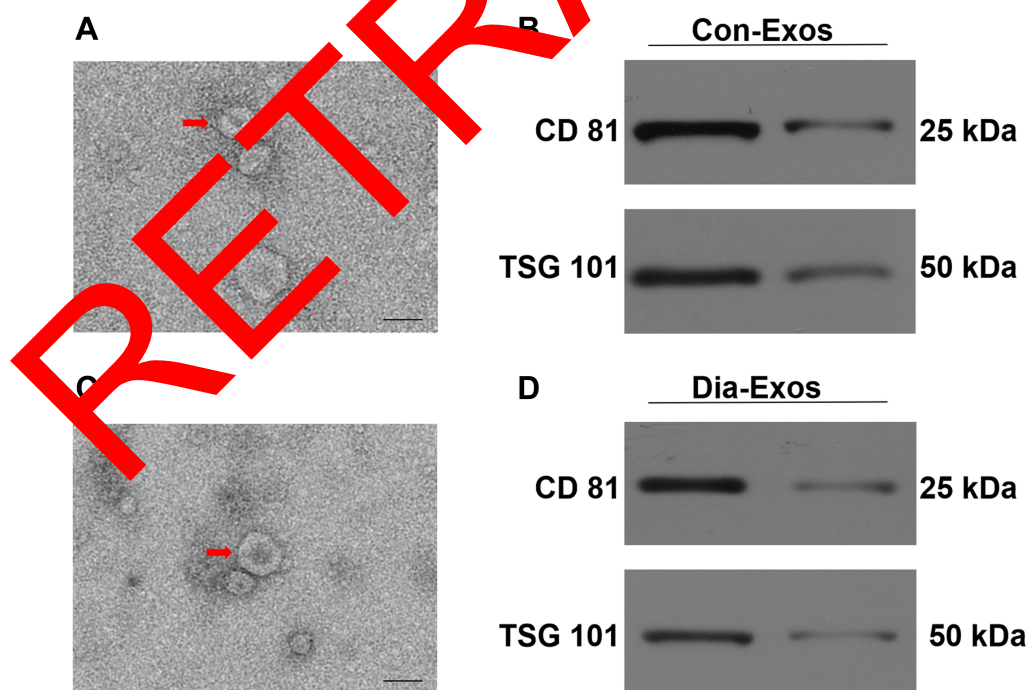


Figure 1 Exosomal characterization. (A) Con-Exos ultrastructural characteristics were assessed via TEM, with typical morphological characteristics being marked with red arrows; (B) CD81 and TSG101 protein levels in Con-Exos; (C) Dia-Exo ultrastructural characteristics were assessed via TEM; (D) CD81 and TSG101 protein levels in Dia-Exos. Scale: 100 nm.

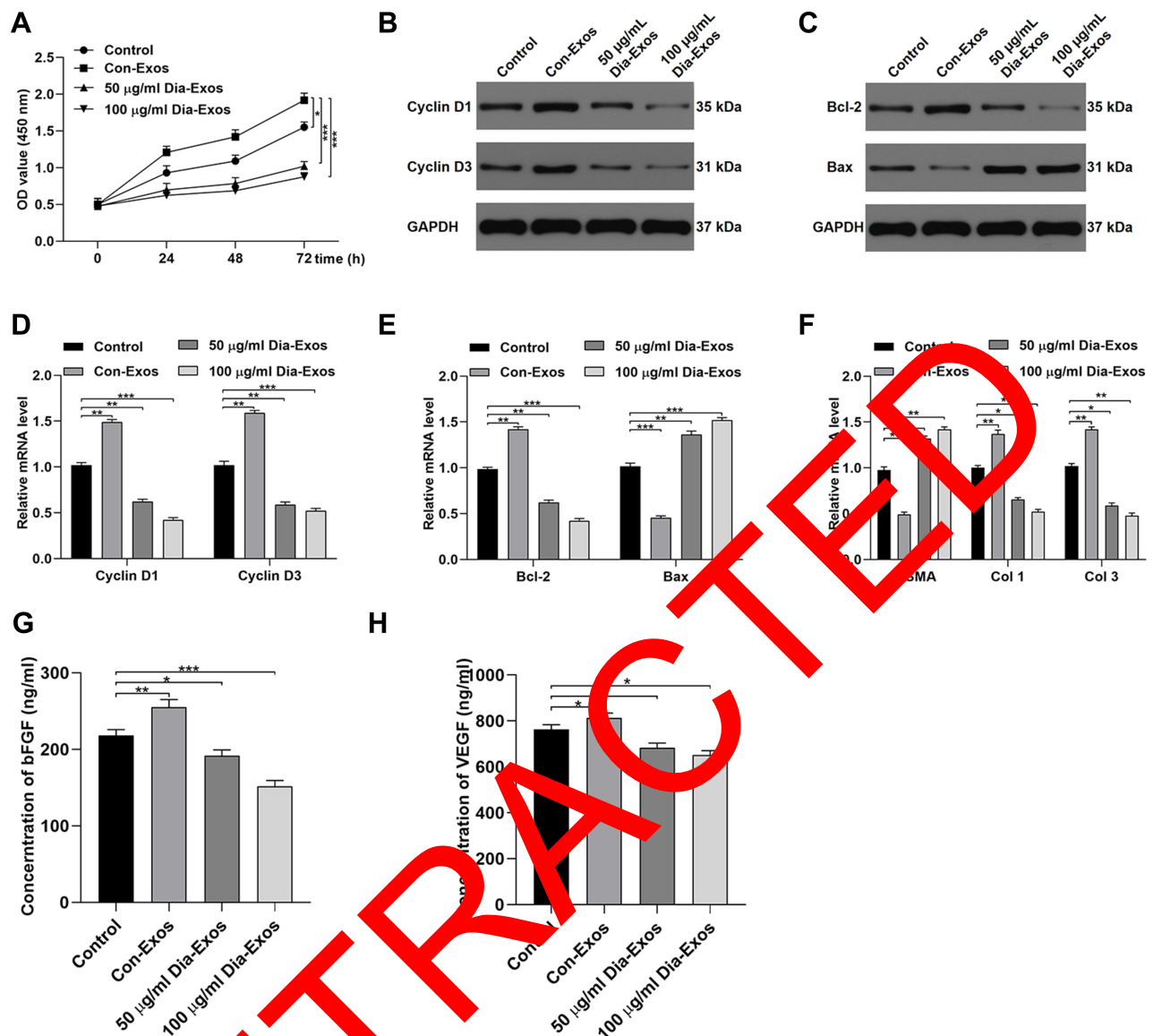


Figure 2 Dia-Exos suppress the survival and functionality of HSFs in vitro. (A) CCK-8 assays were conducted to measure HSF proliferation following exosome treatment; (B) Western blotting was used to quantify Cyclin D1 and Cyclin D3 protein levels following Dia-Exo treatment; (C) Western blotting was used to quantify Bcl-2 and Bax protein levels following Dia-Exo treatment; following Dia-Exo treatment, qPCR was used to evaluate the expression of (D) Cyclin D1 and Cyclin D3 and (E) Bcl-2 and Bax; (F) The impact of Con-Exo or Dia-Exo treatment on collagen-related marker expression in HSFs was measured to evaluate collagen synthesis; (G–H) ELISAs were used to measure bFGF and VEGF secreted by HSFs. Data are means \pm SD from triplicate experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

Suppression of Exosome-Derived miR-20b-5p Enhances the in vitro Functionality of HSFs

In line with prior study results, we determined that Dia-Exos exhibited significant increases in miR-20b-5p expression relative to Con-Exos (Figure 4A).¹⁴ When samples of serum were compared between patients with DFUs and patients with foot injuries that were unrelated to diabetes, we similarly observed significantly higher miR-20b-5p levels in samples from individuals with DFUs (Figure 4B). We also compared the

expression of this miRNA in mice with and without diabetes (Figure S1). Next, we examined miR-20b-5p as a mediator of the impact of Dia-Exos on HSF functionality by directly transfecting these cells with mimics or inhibitors of this miRNA (agomiR-20b-5p and antagomiR-20b-5p, respectively). As expected, antagomiR-20b-5p transfection suppressed the expression of this miRNA (Figure S2 and Figure 4C). Subsequent CCK-8 assays showed that cells treated with antagomiR-20b-5p were more proliferative than control cells (Figure 4D), and these cells expressed higher levels of *Cyclin*

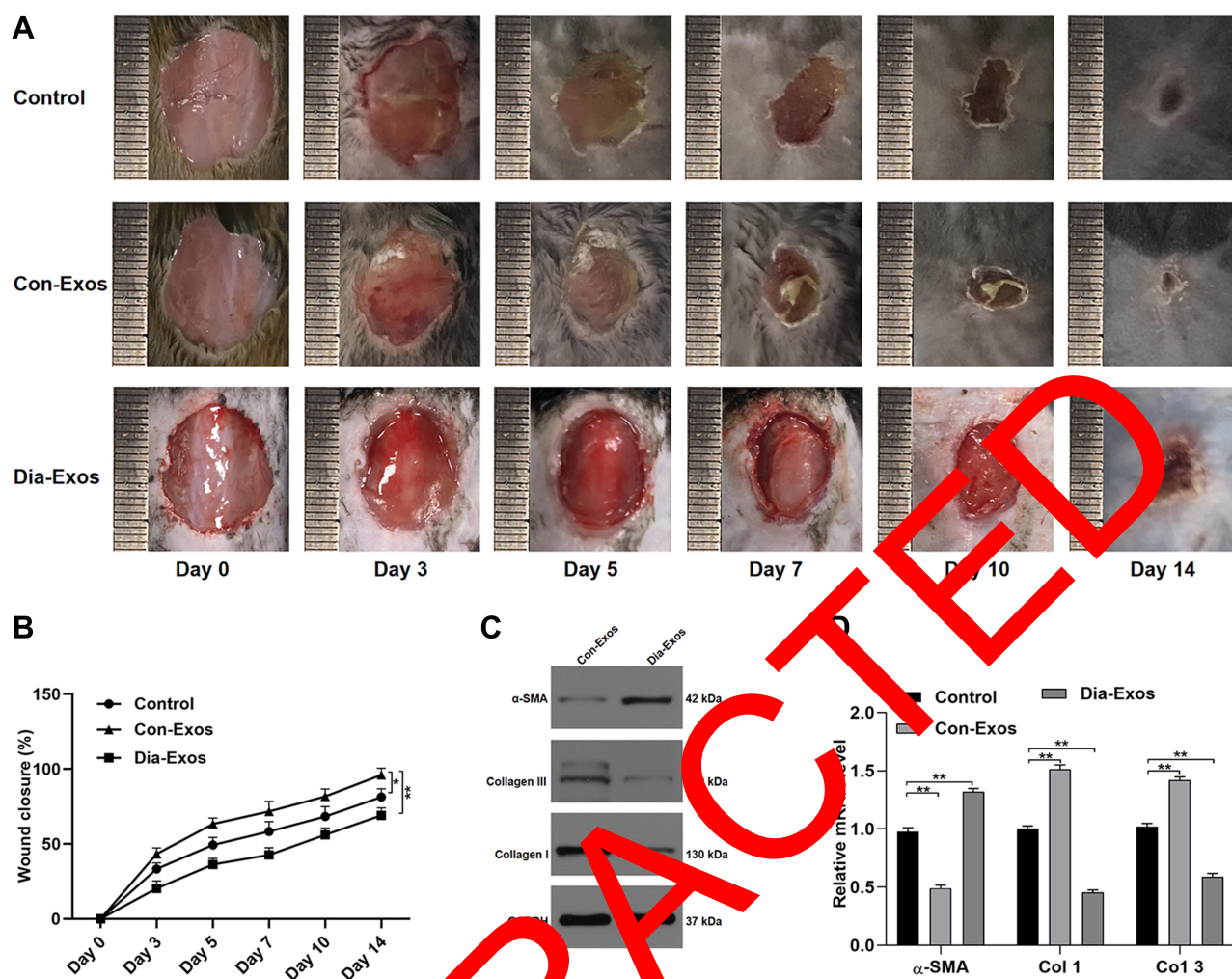


Figure 3 Dia-Exos suppress in vivo wound healing. (A) Wound images on days 0, 3, 5, 7, 10, and 14 after wound generation; (B) wound closure rates were measured with the ImageJ program (n=10 per group); (C and D) markers of collagen synthesis in skin tissue were assessed via qPCR and Western blotting (n=10/group). Data are means \pm SD from triplicate experiments. * $p < 0.05$, ** $p < 0.01$.

Dl and *Cyclin D3* (Figure 4F), as well as lower levels of *Bax* and increased levels of *Bcl-2* (Figure 4F). When we examined the impact of antagonist miR-20b-5p on the secretion of soluble collagen by HSFs, we found that antagonist miR-20b-5p significantly suppressed α -SMA levels and enhanced collagen 1 and 3 expression (Figure 5C). IHCAs further confirmed that inhibiting this miRNA markedly enhanced HSF secretion of VEGFA and bFGF (Figure 4H and I). As such, these data revealed that inhibiting miR-20b-5p was sufficient to enhance the functionality of HSFs in vitro.

miR-20b-5 Suppresses VEGFA Expression and Thereby Inhibits the in vitro Functionality of HSFs

We next examined the mechanistic basis for miR-20b-5p-mediated inhibition of HSF functionality. To that

end, online predictive tools such as TargetScan (http://www.targetscan.org/vert_70/), miRDB (<http://mirdb.org/>), and miRWalk (<http://zmf.umm.uni-heidelberg.de/apps/zmf/mirwalk2/>) were used to identify predicted targets of this miRNA. In total, 24 such predicted targets were identified (Figure 5A). As VEGFA was identified as one of these target genes and has previously been linked to wound healing,¹⁷ we next explored its functional role in the context of miR-20b-5p-mediated wound repair. We began by conducting a luciferase reporter assay, which confirmed that this miRNA bound to the VEGFA 3'-UTR in a WT but not mutated form (Figure 5B). We then utilized an siRNA construct to knock down VEGFA in HSFs in order to demonstrate the importance of this growth factor in the wound healing process (Figure 5C and D). Such VEGFA knockdown

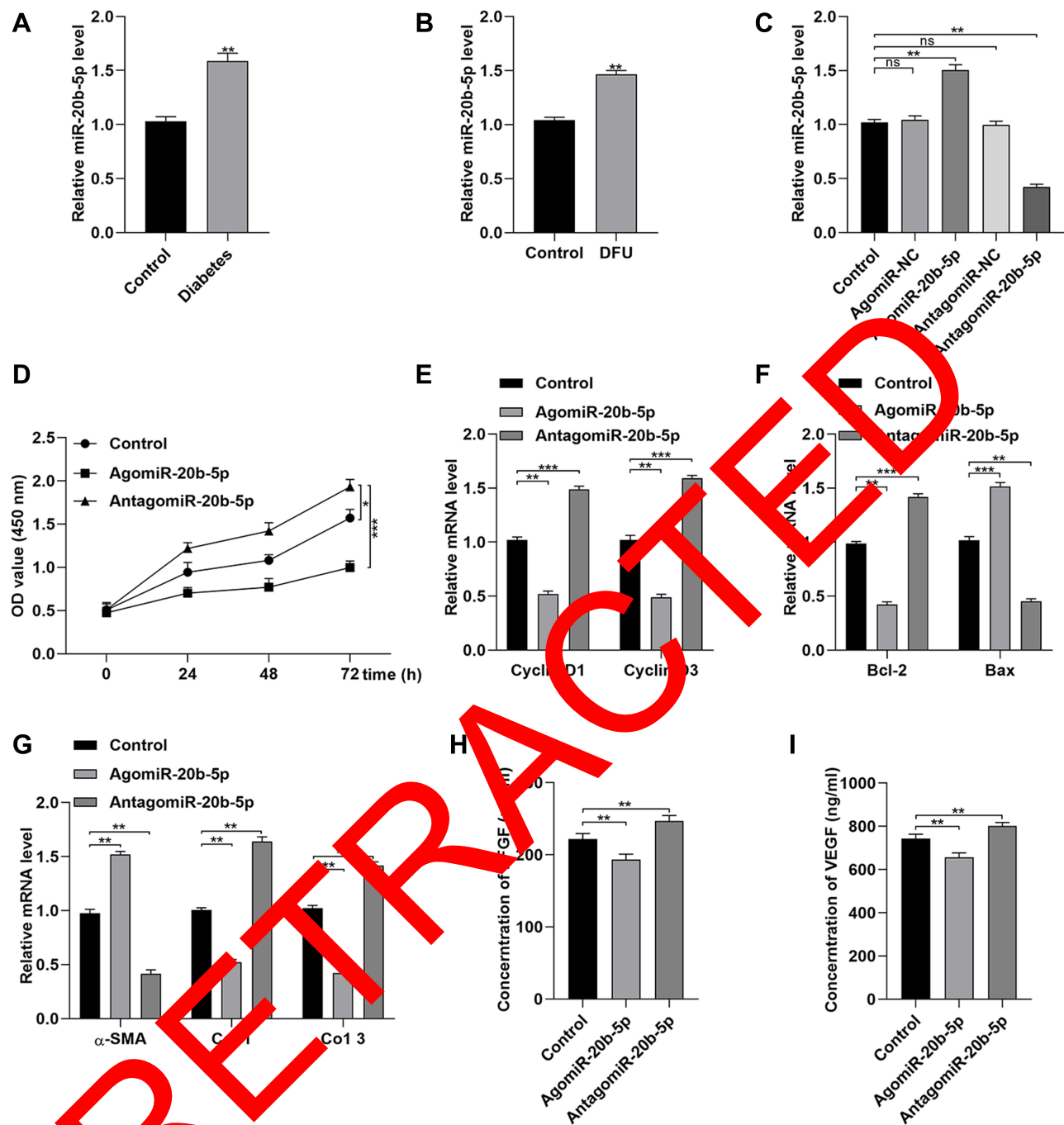


Figure 4 Diabetes exhibit high levels of miR-20b-5p expression. (A) miR-20b-5p expression in diabetes patients and controls (n=10/group); (B) miR-20b-5p levels in patients with non-healing foot wounds and DFUs (n=10/group); (C) agomiR-20b-5p treatment enhanced miR-20b-5p expression by HSFs, as shown via qPCR; (D) the impact of agomiR-20b-5p on HSF proliferation was quantified via CCK-8 assay; (E) treatment with agomiR-20b-5p suppressed Cyclin D1 and Cyclin D3 expression; (F) qPCR was used to examine the impact of agomiR-20b-5p on Bcl-2 and Bax expression; (G) collagen-related markers were evaluated to gauge the impact of miR-20b-5p on collagen synthesis; (H and I) ELISAs were used to measure bFGF and VEGFA secretion by HSFs treated as indicated. Data are means \pm SD from triplicate experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

suppressed HSF proliferation and promoted apoptotic death. Inhibiting miR-20b-5p was able to partially reverse siRNA-VEGFA-mediated impairment of HSFs (Figure 5E and F). Knocking down VEGFA also promoted α -SMA expression while suppressing that of

collagen 1 and 3 (Figure 5G). ELISAs also revealed that VEGF knockdown compromised HSF-derived VEGFA and bFGF secretion (Figure 5H and I). Overall, this indicated that miR-20b-5p disrupts HSF functionality by inhibiting VEGFA expression.

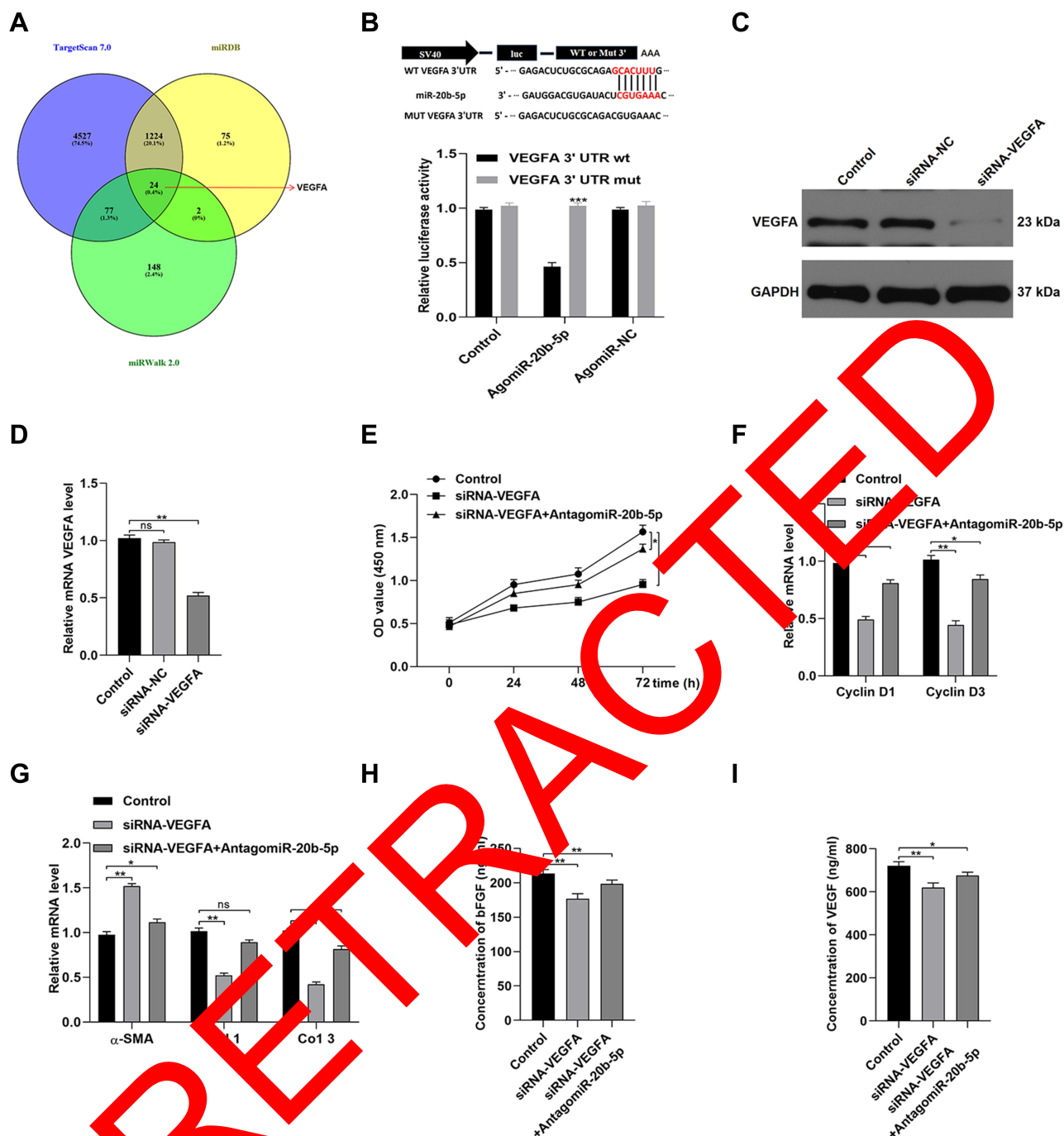


Figure 5 MiR-20b-5p suppresses VEGFA expression and thereby controls HSF functionality. **(A)** miRDB, TargetScan, and miRWalk were utilized to identify possible targets of miR-20b-5p; **(B)** a luciferase reporter assay was used to examine the interactions between miR-20b-5p and VEGFA, with data being analyzed via the SpectraMax M5 software; **(C and D)** the impact of miR-20b-5p on VEGFA expression was quantified via qPCR and Western blotting; **(E)** a CCK-8 assay revealed that miR-20b-5p inhibition was sufficient to partially suppress the anti-proliferative impact of siRNA-mediated VEGFA knockdown in HSFs; **(F)** qPCR was used to measure the expression of Cyclin D1 and Cyclin D3; **(G)** the impact of VEGFA expression on collagen synthesis in HSFs was evaluated based upon collagen-related marker expression; **(H and I)** ELISAs were used to measure bFGF and VEGFA secretion by HSFs treated as indicated. Data are means \pm SD from triplicate experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

Knockdown of miR-20b-5p Improves Wound Healing

Lastly, we explored miR-20b-5p-mediated inhibition of wound healing in vivo by injecting mice bearing full-thickness dorsal

wounds with PBS, agomiR-20b-5p, antagomiR-20b-5p, siRNA-NC, siRNA-VEGFA, or siRNA-VEGFA + AntagomiR-20b-5p on days 0, 3, 5, 7, 9, and 11 post-wounding. A significantly quicker wound closure rate was

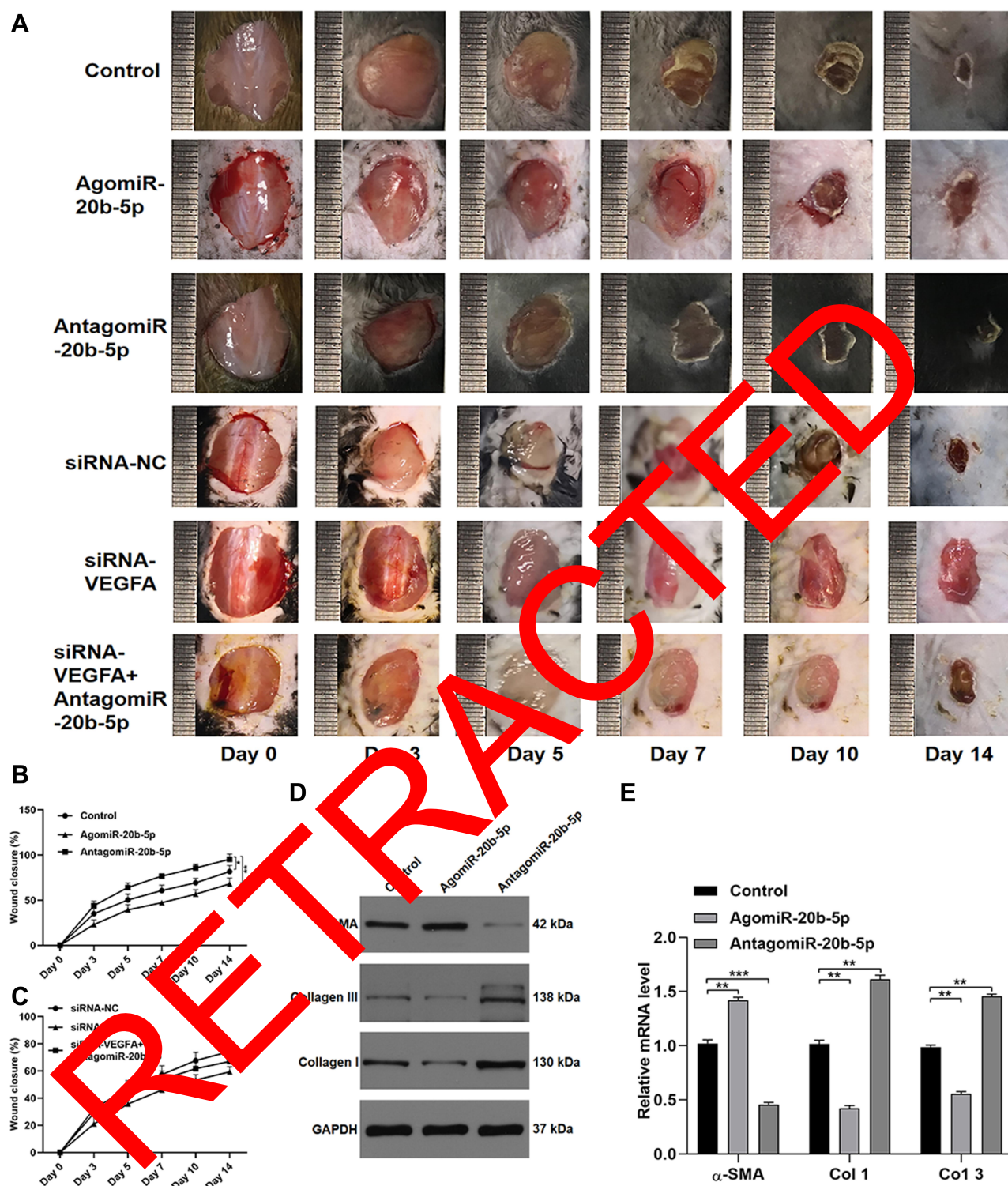


Figure 6 miR-20b-5p suppresses in vivo wound healing. (A) Wound closure on days 0, 3, 5, 7, 10, and 14 after wound generation; (B–C) wound closure rates were measured with the ImageJ program (n=10 per group); (D and E) markers of collagen synthesis in skin tissue were assessed via qPCR and Western blotting (n=10/group). Data are means \pm SD from triplicate experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

seen in the antagomiR-20b-5p group and a better wound closure in siRNA-VEGFA+AntagomiR-20b-5p group compared with the siRNA-VEGFA group, indicating that

antagomiR-20b-5p can partially reverse the impaired effect of siRNA-VEGFA on wound repair (Figure 6A–C). When samples of wounded skin were collected on day 14 after

wound generation, we found that antagomiR-20b-5p-treated mice exhibited significantly lower α -SMA levels and significantly higher collagen 1 and 3 levels as compared to other experimental groups (Figure 6D and E). This suggested that miR-20b-5p inhibition is sufficient to expedite wound healing, making it an attractive therapeutic strategy in those with diabetes.

Discussion

Plasma has been increasingly studied as a tool for tissue engineering applications in recent years.¹⁸ Plasma-derived exosomes are released by a wide range of different cells and that can be internalized by nearby or distant cells, thereby altering their functionality,¹⁹ delivering contents such as cholesterol, proteins, and nucleic acids.²⁰ As they can protect the encapsulated molecules from external factors, Exos are ideal nanovesicles for use when delivering miRNAs and other bioactive materials to target cells in an effort to modulate gene expression.²¹ Fibroblasts are able to readily internalize serum-derived Exos²² and are essential mediators of wound healing owing to their ability to promote collagen synthesis and localized remodeling of diverse tissue types.⁶ As such, we explored the ability of diabetes patient-derived Exos to alter fibroblast functionality in vitro. This experimental approach revealed that Dia-Exos suppressed fibroblast proliferation and induced their apoptotic death, thereby compromising wound healing. In prior studies, Exos were shown to be retained in wound sites for up to three days.²³ We therefore injected Exos into wound sites in model mice every two days and monitored the impact of such treatment on overall wound healing, confirming the ability of Dia-Exos to compromise the wound repair process.

As miRNAs are present at high levels in circulation, fibroblasts likely interact with and are regulated by these non-coding RNAs. Herein, we determined that miR-20b-5p was overexpressed in Dia-Exos and was able to negatively regulate the functionality of fibroblasts. Following miR-20b-5p agonist transfection, fibroblast collagen synthesis and proliferation were markedly compromised, whereas they underwent apoptotic death at higher rates. Similar findings were also observed in vivo. MiR-20b-5p is a miR-17 family member that is thus related to miR-106a-363, miR-17-192, and the miR-106b-25 cluster.²⁴ Dysfunction of the pancreatic islets is tightly linked to the onset of diabetes, and exosomal miRNAs have recently been shown to be differentially expressed under stress

upon release from the islets.²⁵ It is thus possible that exosomal miR-20b-5p in Dia-Exos is derived from islet cells.

VEGFA is a critical regulator of wound healing, tissue remodeling, and collagen production.²⁶ Impaired VEGFA signaling activity is believed to be a key cause of disrupted wound healing in DFU, and several studies have shown VEGFA to be associated with fibroblasts.²⁷ Herein, we identified VEGFA as a miR-20b-5p target gene such that when its expression was suppressed, fibroblast function and wound repair were adversely impacted.

Herein, we also explored the adverse impact of Dia-Exos on HSFs, and our results suggested that the transfer of miR-20b-5p into target cells may be the primary mechanism whereby these nanovesicles suppress normal wound healing. Therapeutic approaches that target these Dia-Exos and/or miR-20b-5p may therefore improve healing in diabetes patients.

Conclusion

In conclusion, we found that Dia-Exos suppress cutaneous wound repair by suppressing fibroblast functionality in vivo and in vitro. Overexpressing miR-20b-5p in Dia-Exos suppresses wound repair by inhibiting the expression of VEGFA. As such, Exos or similar nanoparticles loaded with miR-20b-5p inhibitors may offer therapeutic value as a means of enhancing wound repair in those with diabetes.

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Disclosure

The authors report no conflicts of interest in this work.

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