

An Acceptor- π -Donor Structured Organic Chromophore for NIR Triggered Thermal Ablation of Tumor via DNA Damage-Mediated Apoptosis

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Introduction: It will be challenging to develop high-performance organic chromophores for light-triggered thermal ablation of the tumor. Besides, the mechanisms of organic chromophores for tumor therapy remain unclear. Herein, an acceptor- π -donor (A- π -D) structured organic chromophore based on 2-dicyanomethylenethiazole named PTM was developed for photothermal therapy (PTT) of tumors.

Methods and Results: Biocompatible PTM nanoparticles (PTM NPs) were fabricated by enclosing PTM with Pluronic F-127. The results of optical and photothermal properties of PTM NPs showed robust near-infrared (NIR) absorption, excellent photostability and high photothermal conversion efficiency (56.9%). The results of flow cytometry, fluorescence microscopy, apoptosis, CCK-8 assays and animal experiments showed that PTM NPs had a good killing effect on tumors under NIR laser irradiation. Furthermore, mechanistic studies, RNA-seq and biological analysis revealed that PTM NPs can cause tumor cell death via DNA damage-mediated apoptosis.

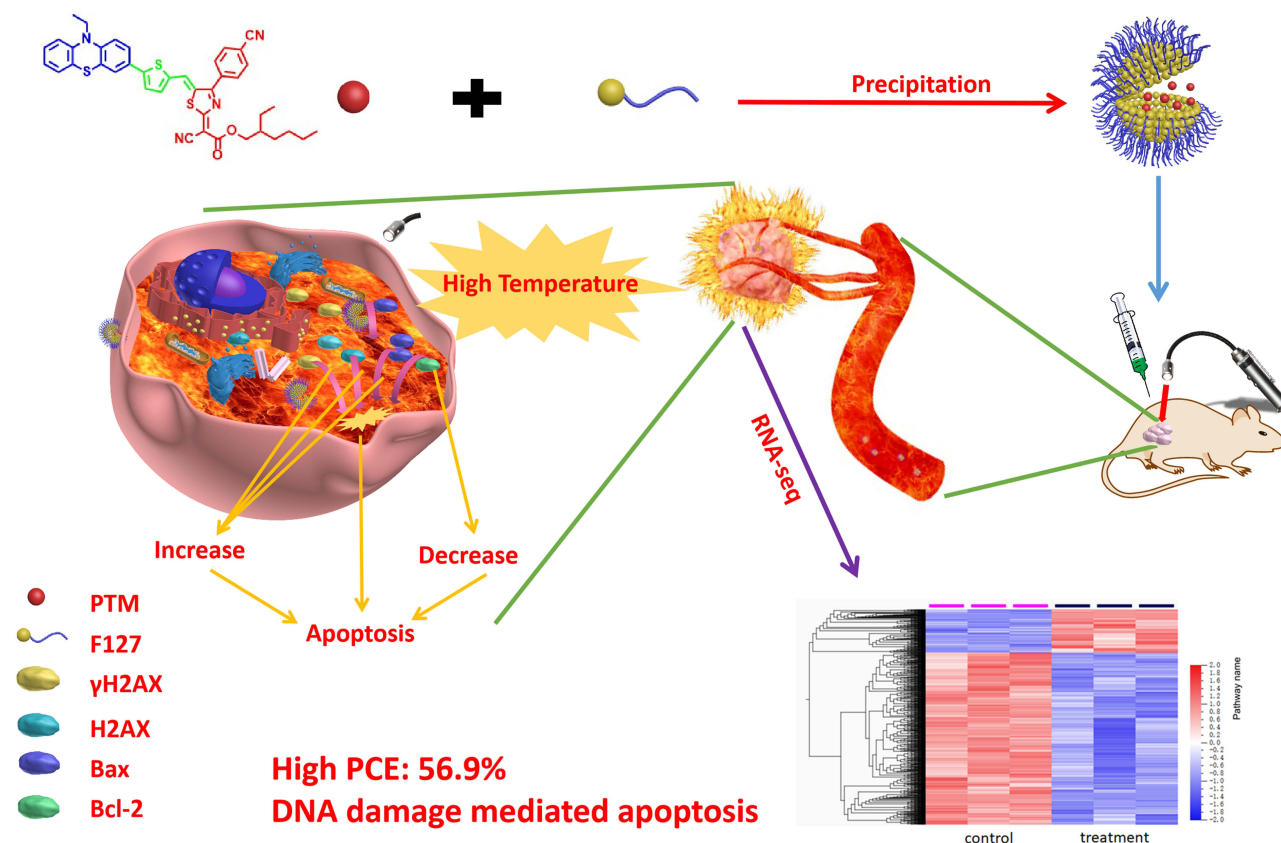
Conclusion: Light-induced thermal ablation effects of PTM NPs in vitro and vivo were surveyed. Collectively, our studies provided a new approach to developing a safe and effective photothermal agent for cancer treatment.

Keywords: NIR absorbing chromophore, 2-dicyanomethylenethiazole, photothermal therapy, DNA damage induced apoptosis, RNA-seq, biological analysis

Introduction

Thermal ablation of tumors is an application of hyperthermia, which will result in irreversible cell injury, the tumor apoptosis and coagulative necrosis ultimately.^{1,2} At present, several thermal ablation methods have been developed, including radio-frequency ablation (RFA),^{3,4} microwave ablation (MWA)^{5,6} and high-intensity focused ultrasound (HIFU).^{7,8} However, expensive instruments were used in these methods. Compared with methods mentioned above, non-invasive photothermal therapy (PTT) be more economical, more effective, more precise targeting at the tumor, and minimize the damage to surrounding healthy tissues. PTT can kill cancer cells and bacterial biofilm via heat energy generated by photothermal agents under NIR laser irradiation.⁹⁻¹⁵ In recent years, various inorganic nanomaterials have been actively employed for photothermal therapy of tumors, including gold nanoparticles,¹⁶ carbon nanomaterials,¹⁷ palladium nanosheets,¹⁸ and transitional

Graphical Abstract



metal dichalcogenide.^{19,20} Although superior efficiency in ablation tumors has been achieved by using inorganic photothermal agents, the potential long-term toxicity and poor biodegradation remain the major challenges. On the other hand, the NIR-absorbing (750~1700 nm) organic photothermal agents have been explored owing to their great biocompatibility and can be metabolized rapidly in biological tissue, including NIR absorbing conjugated polymers,²¹ cyanines²² and porphyrins.²³ However, the poor photostability, severe photodegradation, and the low efficiency of photothermal conversion hamper their further applications. Dibenzamide dyes exhibit excellent molar absorption and subtly tunable optical properties, but their biological applications are mainly focused on fluorescence imaging,²⁴ their maximum absorption and emission in the visible range are limited. Terrylenediimide-based nanomedicines have high photothermal conversion efficiency, but the preparation process with certain inaccuracies is complex and unstable.^{25,26} Hence, it is crucial to develop a new class of NIR absorbing organic chromophores for thermal ablation in tumor.

A- π -D structured organic chromophores have been demonstrated to be an excellent option for photothermal therapy due to their excellent photostability and strong molar absorption coefficient.^{27–30} Besides, chromophores with A- π -D structure have rigid planar structures and strong π - π interaction, which enhance the non-radiative decay and improve photothermal efficiency.³¹ To date, efforts have been made to improve the PTT efficacy of A- π -D structured organic chromophores in many researches, the strategies proposed including increasing acceptor strength, extending conjugation length to reduce band gap, and red-shift the absorption maxima.³² To this effect, numbers of electron-deficient groups have been exploited, including diketopyrrolopyrrole,^{33,34} thiadiazolobenzotriazole,^{35,36} benzo[1,2-c:4,5-c']bis([1,2,5]thiadiazole),^{37,38} and indan.^{39,40} Among them, 2-dicyanomethylenethiazole, an electron-withdrawing core, can be combined with the electron-donating group to build an A- π -D organic chromophore for broadening the absorption spectrum, reducing the optical gap, and decreasing the HOMO level.^{41–45} However, 2-dicyanomethylenethiazole

molecules exhibit some limitations, such as weak absorbance in the near-infrared (NIR) region and low photothermal conversion efficiency. Therefore, it is highly desirable to explore new 2-dicyanomethylenethiazole chromophores with high PTT performance.

As reported, the hyperthermia treatment can inhibit cancer cells tumorigenesis via the necroptosis pathway or DNA damage-mediation tumor apoptosis pathway.^{46–51} However, the underlying mechanisms are still elusive. The comprehensive understanding of photothermal agents might provide new options for efficient cancer therapy. As shown in Graphical Abstract, in this contribution, we developed an A- π -D structured organic chromophore (PTM) based on 2-dicyanomethylenethiazole for light-induced tumor thermal ablation. PTM was directly encapsulated into an amphiphilic copolymer Pluronic F-127 through the nanoprecipitation method to obtain nanoparticles, PTM NPs. These agents shown robust photostability, good biocompatibility, and high PCE (56.9%). Moreover, PTM NPs exhibited cancer cells killing capacity via DNA damage mediated apoptosis induced by heat generation in cells (Scheme 1). Both in vitro and in vivo experiments demonstrated that PTM NPs had effective anti-tumor potential. Our comprehensive studies of the mechanisms and underlying PTM NPs performance will improve the understanding of photothermal therapy and augment the future application of photothermal agents.

Materials and Methods

Chemicals and Materials

All reagents and solvents were commercially provided. CCK-8 kit was provided from Dojindo Laboratories (Kumamoto, Japan). Calcein AM/PI Detection Kit and Hematoxylin and Eosin Staining Kit were purchased from NanJingKeyGen Biotech Co., Ltd. (China). 5-(10-Ethyl-phenothiazin-3-yl)thiophene-2-carbaldehyde was prepared according to reference.⁵²

Design and Synthesis of PTM

5-(10-Ethyl-phenothiazin-3-yl)thiophene-2-carbaldehyde (3.36 g, 10 mmol) and 2-dicyanomethylenethiazole 1 (3.82 g, 10 mmol) and was added into 30 mL acetic anhydride, and stirred at 160 °C for 12 h under N₂ protection. The mixture was drop added into saturated Na₂CO₃ aq. The dark blue solid was filtrated and purified by column chromatography using a mixture of DCM/MeOH (19: 1). Yield: 4.8 g (67%). ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.86 (s, 1H),

7.67~7.69 (d, 2H), 7.57(s, 1H), 7.45~7.47(d, 2H), 7.40~7.42 (d, 2H), 7.32~7.36(d, 2H), 7.28(s, 1H), 7.18~7.24(t, 2H), 6.94~7.05(m, 2H), 6.81(s, 1H), 3.81~3.84(d, 2H), 3.16~3.19 (t, 2H), 1.59(s, 1H), 1.31~1.49(m, 8H), 0.91~0.95(m, 9H). ¹³C NMR (100 MHz, CDCl₃), δ (ppm): 172.16, 166.78, 163.41, 148.70, 147.68, 146.71, 143.83, 141.58, 141.16, 137.38, 136.40, 134.27, 133.11, 132.03, 131.28, 130.60, 129.31, 128.16, 127.85, 126.54, 124.43, 121.11, 120.18, 119.30, 115.98, 115.12, 111.26, 95.94, 64.48, 49.56, 44.41, 31.76, 29.14, 26.55, 26.04, 15.83, 15.04, 14.33. IR (ν^{-1} , KBr): cm⁻¹ 3453, 2986, 2845, 2618, 1618, 1523, 1483, 1366, 1252, 1034, 863.

PTM NPs Preparation

PTM NPs were constructed by the coprecipitation method. Pluronic F-127 (100 mg) and PTM (10 mg) were mixed in THF (1 mL). THF solution was drop added into DI water (10 mL) under ultrasonic condition. The mixture was bubbling N₂ to remove THF completely. PTM was obtained after dialysis to remove the free F-127.

Animals

All the animal experiments were conducted in accordance with the institutional guidelines for the care and use of laboratory animals at Southern Medical University, Guangzhou, China, and Regulations for the Administration of Affairs Concerning Experimental Animals (1991.7, revised 2017). All the animal experiments have been approved by Southern Medical University.

Statistical Analysis

All data were expressed as means \pm standard deviation (SD) or means \pm standard errors. All figures shown in this article were obtained from at least three independent experiments. Analysis of variance was employed for multiple group comparisons, and results of $p < 0.05$ were considered statistically significant.

Result and Discussion

Synthesis, Optical and Photothermal Property of PTM NPs

NIR absorption for A- π -D structured organic chromophore (PTM) was built by using 2-dicyanomethylenethiazole as a strong acceptor, allowing for greater electron delocalization and thus lowering the band gap, phenothiazine as a donor, and planar thiophene ring as a π -conjugation unit, which

provided effective conjugation and large extinction coefficient. The synthetic routine of PTM is shown in [Scheme S1](#) and [S2](#). The synthesis process, NMR spectrum, and IR spectrum are shown in [Figure S1–S5](#) (Supporting information).

To further enhance the biocompatibility, hydrophobic PTM and Pluronic F-127 were dissolved in THF and allowed to self-assemble into PTM NPs. First of all, the absorption property of PTM NPs was evaluated, as shown in [Figure 1A](#), PTM NPs (200 $\mu\text{g/mL}$) have a broad UV-vis absorption arrange from 550 nm to 1000 nm with the maximum absorption located at ~ 680 nm, which permits deep tissue penetration and exhibits the potential to accomplish efficient photothermal therapy for deep tumor. Furthermore, the dynamic light scattering (DLS) experiment showed that PTM NPs exhibited hydrodynamic diameters around 110 nm ([Figure 1B](#)). Interestingly, the size of PTM NPs in PBS keeps stable within

28 days, which confirms the robust stability ([Figure S6](#)). In addition, TEM and SEM were measured to confirm the morphology and size of PTM NPs ([Figure S7–S8](#)).

The light-induced thermal activity of PTM NPs was also explored. The relation between laser power and temperature variation of PTM NPs in PBS was surveyed ([Figure 1C](#)). The temperature of PTM NPs solutions increased quickly with improving the laser power, confirming that the temperature elevation is proportional to the laser power density. As depicted in [Figure 1D](#), when the concentration of PTM NPs reached 160 $\mu\text{g/mL}$, the solution temperature rose rapidly to 72.4°C under NIR laser irradiation (808 nm, 1.5 W/cm^2 , 10 min). It is worth noting that PTM NPs also reached effective hyperthermia (45°C) with the concentration of 40 $\mu\text{g/mL}$ under the same conditions, demonstrated that PTM

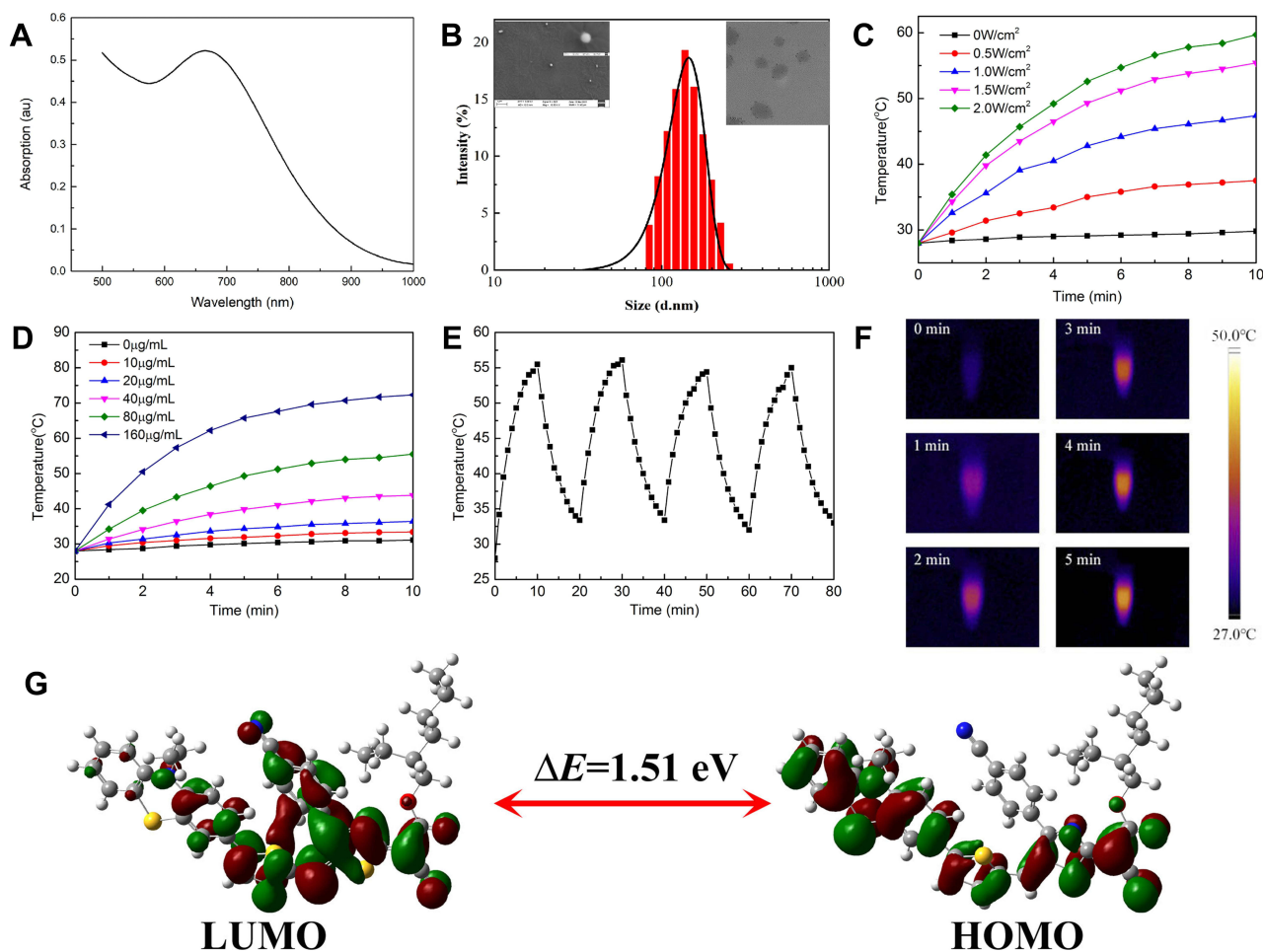


Figure 1 (A) The absorption spectrum of PTM NPs (200 $\mu\text{g/mL}$) in Water. (B) DLS of PTM NPs (10 $\mu\text{g/mL}$) insert SEM and TEM of PTM NPs. (C) Photothermal heating curves of PTM NPs (80 $\mu\text{g/mL}$) irradiated for 10 min using a 808 nm laser at varied power densities (0, 0.5, 1.0, 1.5, and 2.0 W/cm^2). (D) The temperature evolution of PTM NPs with different concentrations under 808 nm laser irradiation (1.5 W/cm^2 , 10 min). (E) Temperature elevation of PTM NPs (120 $\mu\text{g/mL}$) under five on/off cycles (1.0 W/cm^2). (F) Corresponding near infrared photographs of PTM NPs (80 $\mu\text{g/mL}$) under NIR laser irradiation (808nm, 1.5 W/cm^2 , 5 min). (G) HOMO and LUMO calculated by DFT at the B3LYP/6-31G (d,p) basis set.

NPs displayed excellent photothermal properties even under a low concentrations.

The photothermal conversion efficiency of PTM NPs was evaluated to be ~56.9% according to the reported method (Figure S9),⁵³ which is significantly superior to most previously reported photothermal reagents, including porphyrin (62.5%),⁵⁴ phthalocyanine-based nanodots (45.7%),⁵⁵ cyanine dyes (26.6%),⁵⁶ polypyrrole NPs (45%),⁵⁷ dopamine-melanin NPs (40%),⁵⁸ Au nanorods (21.0%),⁵⁹ MoS₂ nanosheets (24.4%).⁶⁰ All these results confirmed that PTM NPs can act as a promising candidate for photothermal therapy.

The photostability of PTM NPs was also investigated. The temperature changes of PTM NPs solution (120 µg/mL) were continuously monitored under NIR laser irradiation (808 nm, 1.0 W/cm², 10 min) and then cooling to r.t. naturally for five heating/cooling cycles. As shown in Figure 1E, the photothermal capacity changes slightly. The photobleaching stability of PTM NPs was further determined by continuous irradiation of PTM NPs in PBS (120 µg/mL) upon NIR laser decreased slightly, and no obvious color change was observed (Figure S10). These results confirmed that PTM NPs have excellent stability against photobleaching. The temperature of PTM NPs in aqueous dispersions was also investigated upon NIR laser irradiation (808 nm, 1.5 W/cm², 5 min). Infrared thermal images after irradiation were also present in Figure 1F.

After the UV-vis absorption and photothermal exploration of PTM NPs, density functional theory (DFT) calculation was performed (Figure 1G). The highest occupied molecular orbital (HOMO) of the phenothiazine group were occupied by electron clouds, whereas the 2-dicyanomethylenethiazole unit dominated the lowest unoccupied molecular orbital (LUMO) with the narrow band gap ($E_g = 1.51$ eV).

Laser-Induced Thermal Ablation of PTM NPs in vitro

It is fundamental for phototherapy agents to exhibit low dark toxicity but high toxicity under laser radiation. Mouse breast cancer 4T1 cell line donated by the Department of Cell Biology, School of Basic Medical Sciences, Southern Medical University that were purchased from the National Infrastructure of Cell Line Resource (China) were employed in this study. ACCK-8 analysis for toxicity evaluation of PTM NPs in 4T1 cells without irradiation demonstrated that these agents had excellent biocompatibility (Figure S11). On the other hand, the cell viability decreased to 27% when incubation of PTM NPs (200 µg/mL) upon 808 nm irradiating (1.5 W/cm², 10 min), suggesting PTM NPs may be amenable to photoactive therapy.

Fluorescent images of 4T1 cells dyed with calcein AM and propidium iodide after PTM NPs with photothermal therapy treatment are illustrated in Figure 2A, strong red

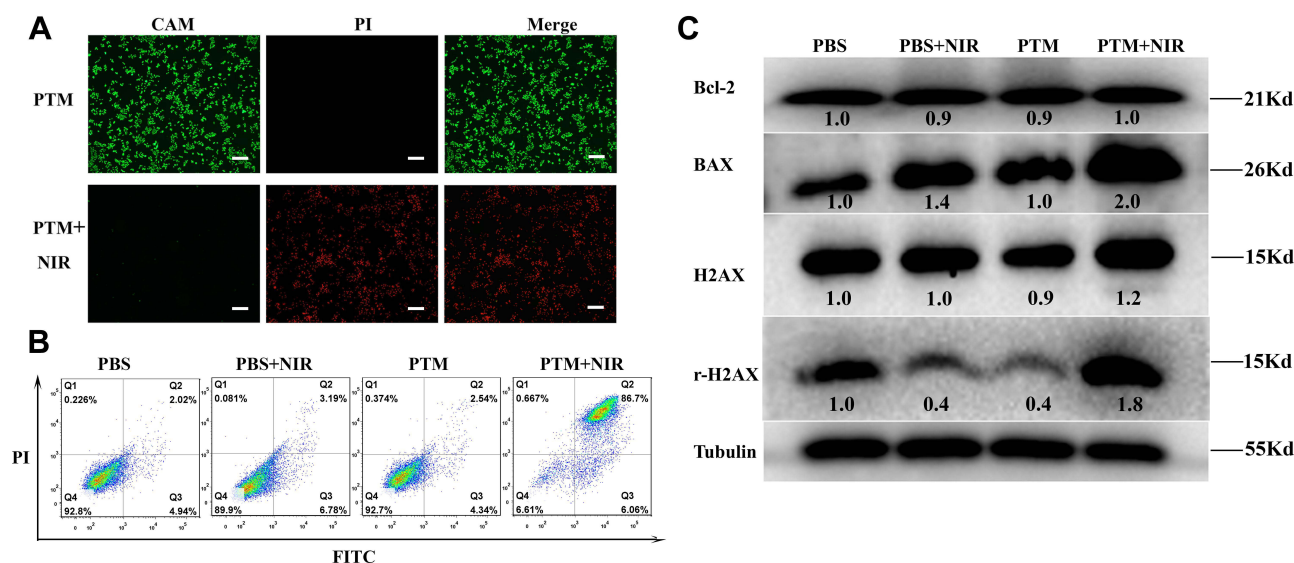


Figure 2 (A) AM/PI fluorescence imaging of 4T1 cells with PTM NPs (200 µg/mL) in the absence or presence of NIR laser irradiation (808nm, 1.5 W/cm², 10 min). The scale is 100µm. (B) Representative FCM profiles of 4T1 cells with PTM NPs (80 µg/mL) in the absence or presence of NIR laser irradiation (808 nm, 1.5 W/cm², 5 min). (C) PTT induced the Bcl-2, BAX, H2AX, and γH2AX expression in 4T1 cells after different treatments.

fluorescence in cells was observed. However, the group of PTM NPs treated or laser alone has almost no red fluorescence (Figure S12), confirming that the cells were killed after co-incubation with PTM NPs and 808 nm laser irradiation. Apoptosis assay was then employed to investigate the apoptosis and necrosis rate induced by activated PTT with PTM NPs. As displayed in Figures 2B, 4T1 cells treated with PTM NPs upon 808 nm laser irradiation displayed an apoptotic ratio to be ~92.76%, which is notably superior to that in other groups.

To clarify the cell death pathway in charge of the observed effects, we surveyed the effects of PTM NPs on Bax, Bcl-2, H2AX, and γ H2AX. It is well known that Bcl-2 has anti-apoptotic properties, which is usually opposite to Bax. The balance between Bcl-2 and Bax is crucial for cell apoptosis or survival.^{61–63} γ H2AX indicates the specific phosphorylation at serine 139 of the histone H2AX, the generation of double-strand breaks always

induces γ H2AX formation.^{64,65} Therefore, the expression of γ H2AX reflects the DNA damage. We analyzed their levels by Western blotting (WB) analysis with tubulin as the loading control. As exhibited in Figure 2C, the expression of Bax improved, while the Bcl-2 level diminished and the ratio between Bcl-2 and Bax declined when 4T1 cells were treated with PTM NPs and 808 nm laser radiation, which proved that apoptosis of cells increased. Moreover, PTM NPs + NIR laser irradiation significantly increased the expression of γ H2AX, confirming that this PTT treatment results in DNA damage. These datas verified the hypothesis that DNA damage might be the anti-cancer mechanism for PTT.

In vivo Photothermal Properties

Five-week-old female nude mouse (n=20) were purchased from the Animal Center of Southern Medical University. All animal procedures were performed in accordance with

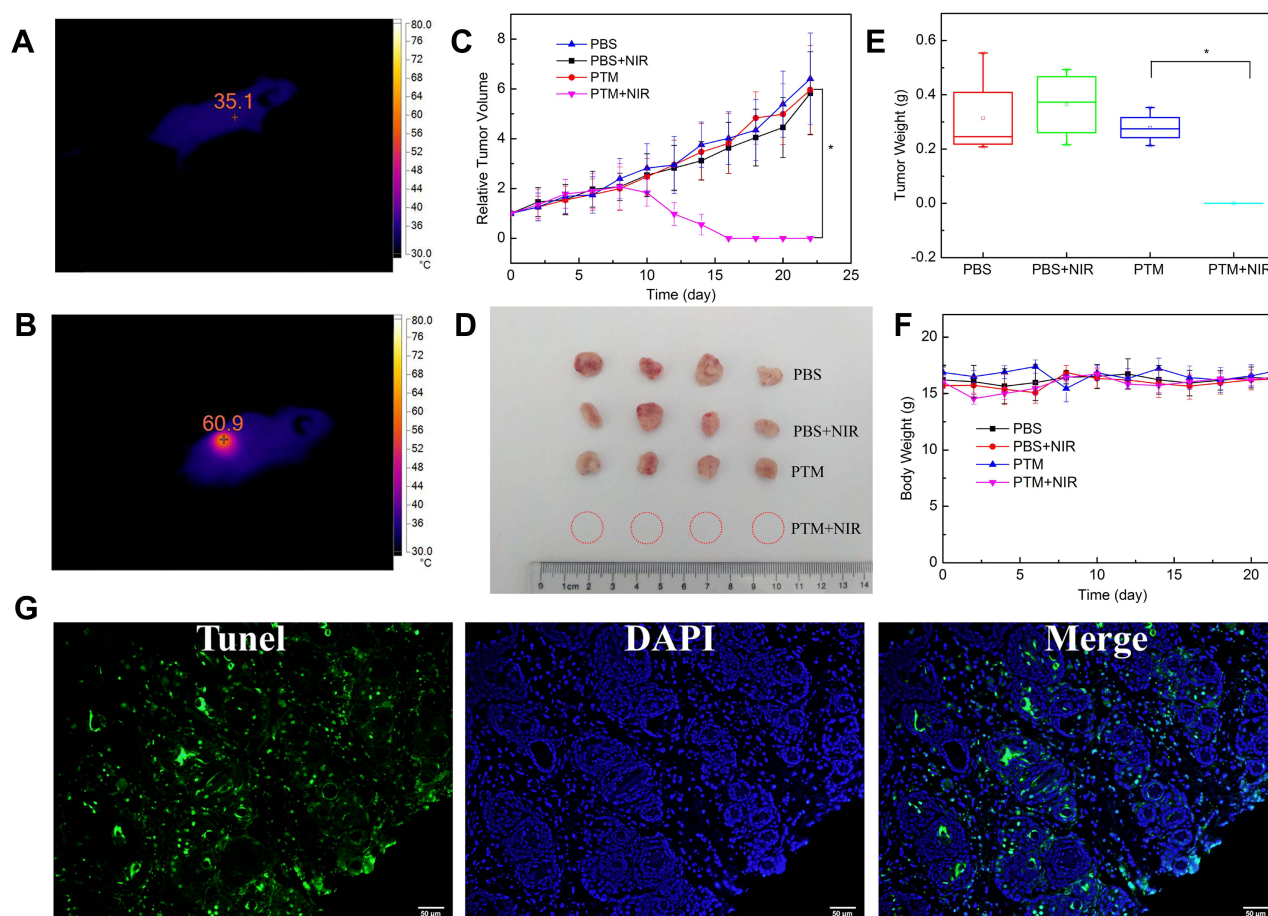


Figure 3 (A) and (B) Tumor temperature of PTM + NIR laser irradiation group before and after irradiation (808nm, 1.0 W/cm², 1 min). (C) Tumor growth curves of mice after different treatments. Results are expressed as mean \pm S.E. * $P < 0.05$ compared with the PTM+NIR laser irradiation group. (D) Photographs of the dissected tumors after different treatment. (E) Dissected tumors weight after different treatment. Results are expressed as mean \pm S.E. * $P < 0.05$ compared with the PTM+NIR laser irradiation group. (F) Body weight curve of mice after PTM+NIR laser irradiation group. (G) TUNEL assay of the tumor tissues after PTT.

the “Guidelines for the Care and Use of Laboratory Animals of Southern Medical University” and were approved by the Animal Ethics Committee of Southern Medical University.

Finally, *in vivo* photothermal efficacy was conducted by post-intratumoral injection. Firstly, temperature changes in tumor was monitored by infrared thermography during irradiation (Figure 3A and B). The tumor surface temperature of the PTM group rose to 60.9°C upon 808 nm laser irradiation for 1 min which provided adequate hyperthermia to kill cancer cells. What's more, the tumor surface temperature of the PBS injection group was roughly unchanged upon 808 nm laser irradiation for 1 min (Figure S13).

After that, 4T1 tumor-bearing mice were randomly batched into four groups: PBS control group (group A), 808 nm laser irradiation only (group B), PTM NPs (2.5 mg/kg) treated only (group C), PTM NPs (2.5 mg/kg) and NIR laser irradiation (808 nm, 2.0 W/cm², 5 min) (group D). Changes in tumor size and body weight were monitored in all the groups. Notably, no apparent abnormal body weight changes and other signs of toxic side effects were observed in all treated groups. As shown in

Figure 3C–E, groups A, B, and C all displayed minimal tumor inhibition, implying that laser exposure or PTM NPs themselves have little influence on the tumor growth. However, Group D (PTM NPs upon 808 nm laser irradiation) displayed tumor ablation, confirming the apoptosis-induced potential, which is well in consonance with the flow cytometry analysis. All the groups demonstrated similar body weight variation curves (Figure 3F), indicating that PTM NPs exhibited slightly side effects. Additionally, apparent apoptosis and necrosis were observed in PTT treated group (Figure 3G), indicated that PTM NPs plus 808 nm laser radiation triggers cell apoptosis and necrosis.

Light Induces DNA Damage in Cancer Cells by the PTM NPs

Owing to the excellent photothermal performance of PTM NPs observed *in vitro* and *in vivo* experiments, we wonder the action anti-tumor mechanisms of PTM NPs. To further understand the action mechanisms of PTM NPs, RNA-seq analysis was carried out to identify differentially regulated genes after PTM NPs treatment (Figure 4A and B). Enriched in metabolism pathways,

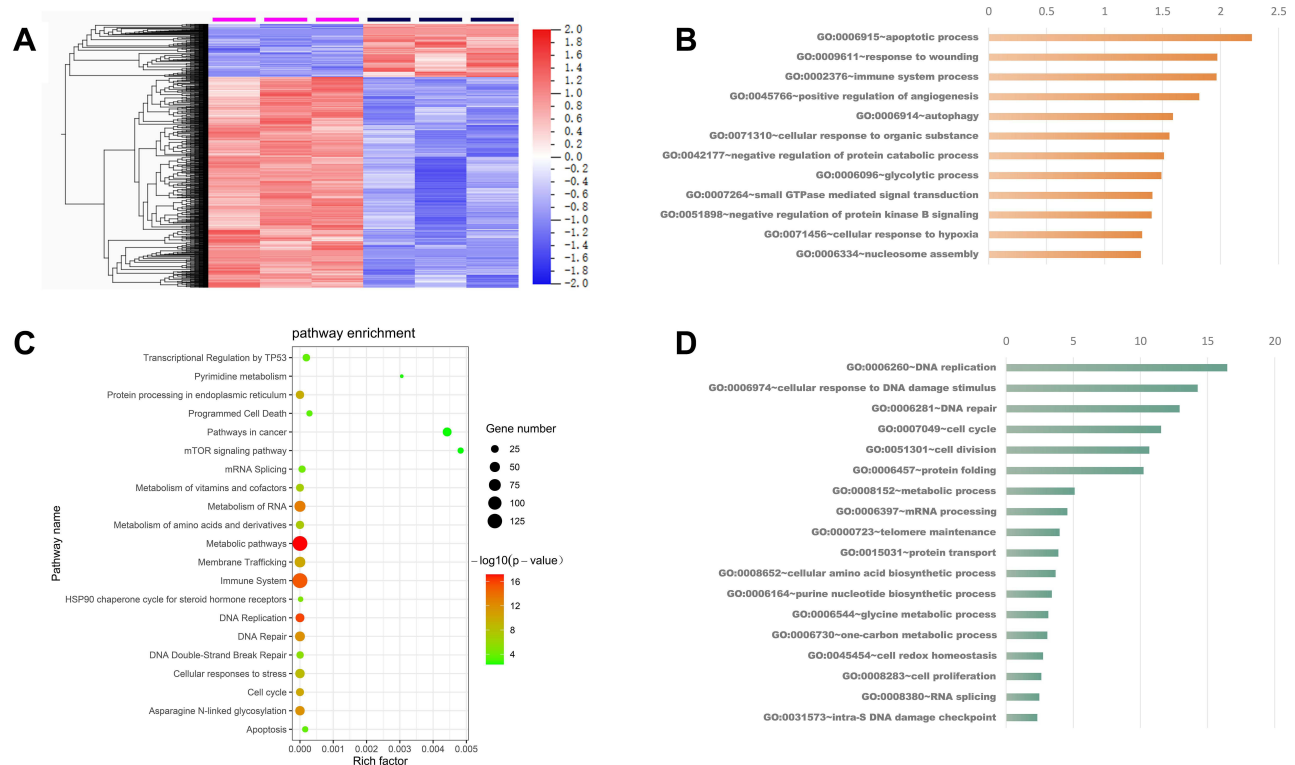


Figure 4 (A) Heatmap showed the differential expressed genes after PTM NPs treated. (B) Enriched KEGG pathway analysis of differentially expressed genes after PTM NPs treated. (C) and (D) Enriched GO biological process of up-regulated genes (orange) and down-regulated genes (blue) in PTM NPs treated group. The X-axis of the histogram is $-\log_{10}$ (P-value) of individual terms calculated by right-sided hypergeometric test and corrected with Bonferroni. GO categories are indicated on Y-axis.

immune system, DNA repair, cell cycle, and mRNA splicing was observed (Figure 4C and D). The expression of genes involved in the apoptotic process, glycolytic process, and immune system process were up-regulated. In contrast, the genes involved in DNA repair, cell cycle, metabolic process, protein folding, and cellular amino acid biosynthetic process were down-regulated. The results confirmed that the enriched expressed genes are involved in the cell apoptosis pathways, including the focal adhesion, adherend junction, and Wnt signaling pathway after PTM NPs + NIR laser irradiation treatment.

The Biocompatibility of PTM NPS

The *in vivo* toxicology of PTM NPs was examined. Hematoxylin and eosin (H&E) staining was carried out to measure the histological changes of main organs (Figure 5A). No obvious pathological variation of major organs was observed in all the groups, confirmed that the excellent biocompatibility of PTM NPs. Moreover, blood biochemical analyses and hematology

markers were investigated to examine the *in vivo* toxicity of PTM NPs (Figure 5B). No apparent distinction was detected in PTM NPs treated groups. All these results highlight the good biological compatibility and safety of PTM NPs.

Conclusion

In conclusion, a new NIR-absorbing A- π -D type organic chromophore (PTM) based on 2-dicyanomethylenethiazole was prepared for laser-induced thermal ablation tumors. The encapsulation of PTM into an amphiphilic copolymer F-127 yields PTM NPs with good biocompatibility, excellent photostability, and high photothermal conversion efficiency (56.9%). Furthermore, the therapeutic mechanism and RNA-seq and biological analysis revealed that hyperthermia induces cell apoptosis via DNA-damage. *In vivo* investigation, PTM NPs demonstrated excellent tumor ablation under irradiation. All these results confirmed that the A- π -D structured small molecule had great potential for NIR-triggered phototherapy of the tumor.

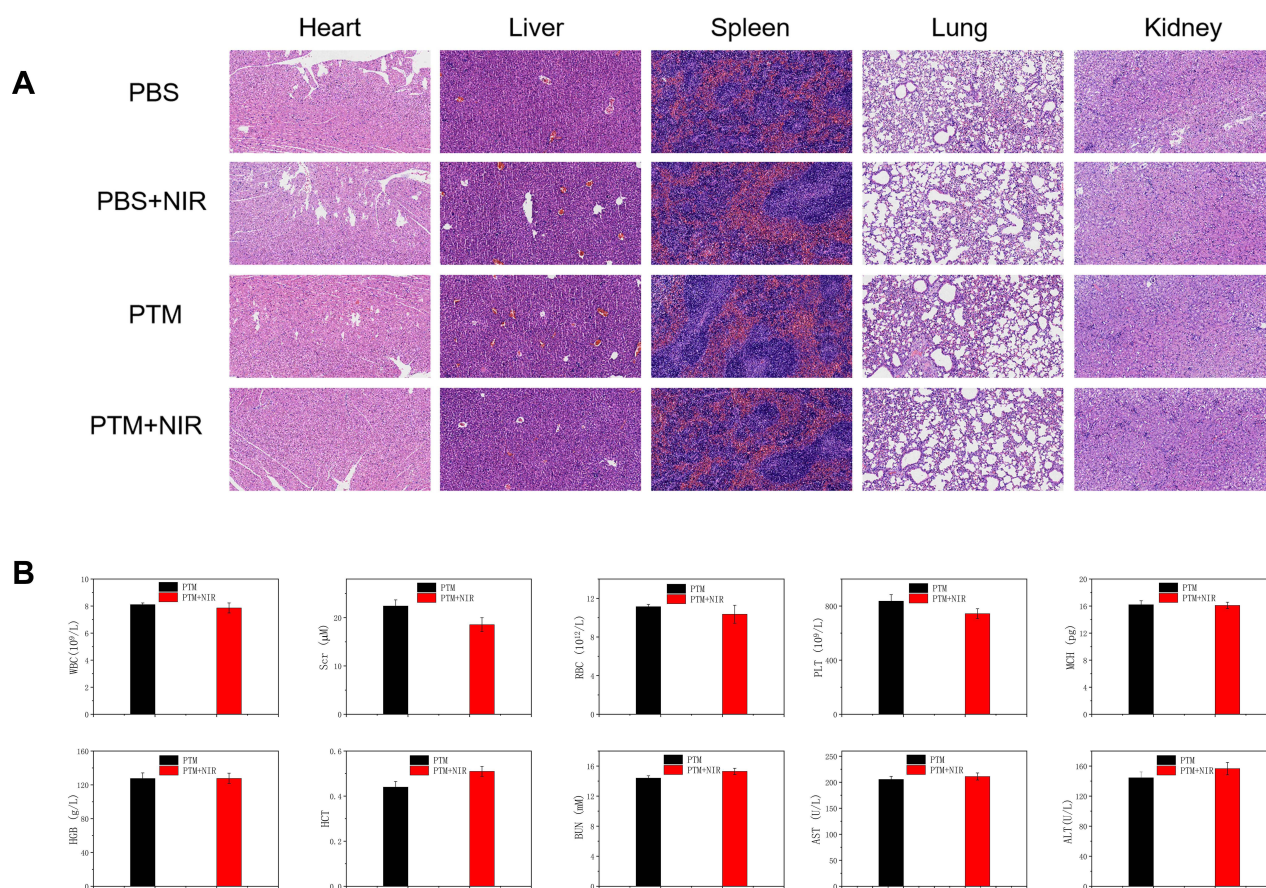


Figure 5 (A) H&E staining images of major organs in the mice after the intravenous injection. (B) The influence of PTM NPs on the hematopoietic system of mice.

Acknowledgments

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Disclosure

The authors report no conflicts of interest in this work.

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