

Albumin-encapsulated Nanoparticles of Naproxen Platinum(IV) Complexes with Inflammation Inhibitory Competence Displaying Effective Antitumor Activities in vitro and in vivo

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Background: Platinum(IV) complexes with inflammation inhibitory properties are much favored in improving antitumor activities. Nanodrug-delivery system as a preferable measure for antitumor therapy are widely explored in platinum(IV) drug delivery.

Purpose: The aim for this study was to develop novel bovine serum albumin (BSA) nanoparticles (NPs) based on naproxen platinum(IV) complexes to display a synergistic antitumor mechanism targeting cyclooxygenase-2 (COX-2), metalloproteinase-9 (MMP-9) and inducible nitric oxide synthase (iNOS).

Methods: Herein, we reported the preparation of two BSA NPs of naproxen platinum(IV) complexes, and their antitumor activities were investigated in vitro and in vivo.

Results: Both NPs possessed relatively uniform size and good stability for 30 days in aqueous solution. They exhibited prominent antitumor activities in vitro, and showed great potential in reversing drug resistance. Furthermore, these two NPs played superior tumor growth suppression in vivo in contrast to the free compounds, which were comparable to that of cisplatin and oxaliplatin, but induced lower toxic influences than platinum(II) drugs especially to spleen and liver. Moreover, the naproxen platinum(IV) NPs could decrease tumor inflammation targeting COX-2, MMP-9 and iNOS, and decreasing NO production, which would be in favor of enhancing the antitumor competence, and reducing toxicity.

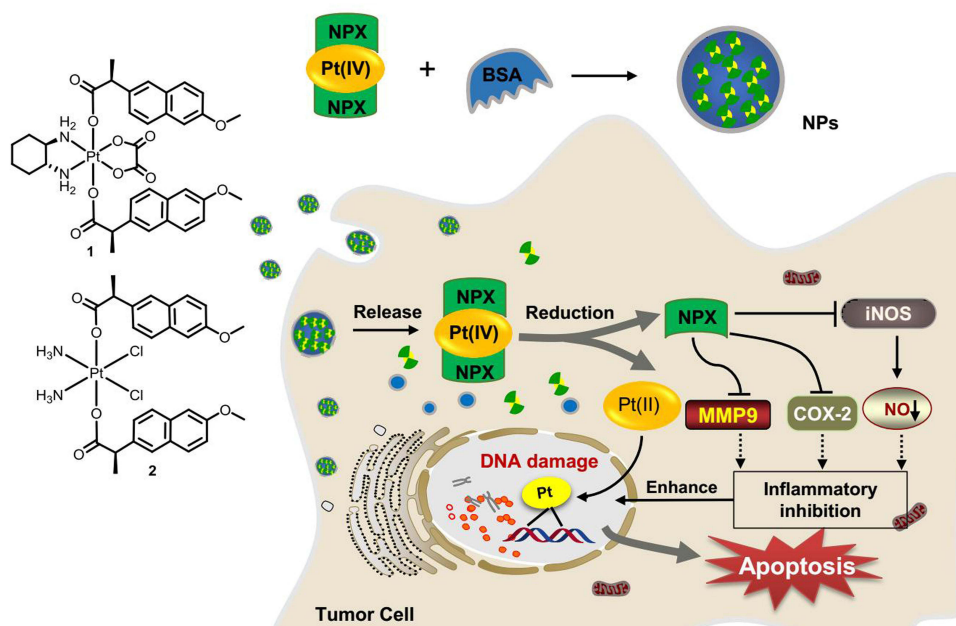
Conclusion: Taken together, BSA NPs of naproxen platinum(IV) complexes demonstrated a powerful antitumor efficacy in vitro and in vivo. The platinum(IV) NPs with inflammation inhibitory competence targeting multiple enzymes reported in this work afford a new strategy for the development of antitumor therapy to overcome drawbacks of clinical platinum(II) drugs.

Keywords: bovine serum albumin nanoparticles, cyclooxygenase-2, metalloproteinases-9, inducible nitric oxide synthase, synergistic antitumor efficacy

Introduction

Cancer is a major public problem worldwide and seriously threatens human health and life.^{1,2} Chemotherapy as a systemic treatment is widely applied in the treatment of various tumors for its superior effectiveness, especially to the tumors at advanced stages. Nanodrug-delivery system (nano-DDS) provides a new effective tool for chemotherapy in clinics.³ Nanodelivery of various chemotherapeutic drugs has received tremendous attention in the past decades owing to their superiority in

Graphical Abstract



improving drug solubility, increasing accumulation in tumors, reducing side effects and prolonging blood circulation time. Moreover, taking advantage of the enhanced permeability and retention (EPR) effect in tumor tissues, nanoparticles (NPs) facilitate drug targeting to tumor tissues, and further influence their activities and toxic properties.^{4,5} The investigation on nano-DDS of chemotherapeutics has been a hot topic in cancer treatments.

Platinum drugs play a pivotal role in cancer treatment in clinics. Especially cisplatin, carboplatin, and oxaliplatin are highly effective in cancer chemotherapy.^{6,7} The injection of platinum(II) drugs usually involves a high peak of drug level above the maximum tolerable concentration (MTC) in the plasma, rapid elimination and low bioavailability, which causes severe side effects, drug resistance, etc, and further hinders their scope of use. Nano-DDS exhibits unique advantages in drug controlled release and is extensively exploited for platinum drug delivery.⁸ Nevertheless, low lipophilicity and poor stability of traditional platinum(II) drugs limit the formation of the NPs especially when injected into the bloodstream.^{9,10} Platinum(IV) complexes as prodrugs of platinum(II) drugs with d^6 octahedral geometry show rather enhanced lipophilicity and stability.^{11,12} Large amounts of platinum(IV) based nano-DDS have been developed and exhibited significant superiority over free platinum drugs.

Serum albumin nanocarriers represent one of the most promising carriers recently generated because of their good biocompatibility, low immunogenicity, biodegradability, and favorable toxicological profiles.^{13,14} Moreover, albumin nanocarriers are potent in improving the stability of sensitive therapeutic cargos and possess much potential in targeting tumor cells.¹⁵ Especially, the market of serum albumin nanomedicine Abraxane[®] (paclitaxel) lightens the enthusiasm of researchers on serum albumin nanoparticles.¹⁶ Bovine serum albumin (BSA) with high sequence homology (76.52%) to human serum albumin (HSA), wide source and low cost has been widely investigated.^{17–20} Whereas, the serum albumin NPs loaded platinum(IV) drugs were seldom investigated. Moreover, it has been proven that the conjunction of platinum(IV) drugs with albumin could significantly decrease the reduction rate and enhance stability of drugs in whole blood.²¹ Thereby, it is of great interest for us to design a series of BSA nanoparticle-loaded platinum(IV) complexes and further investigate their antitumor properties.

Inflammation is an important hallmark in cancer progression associated with the occurrence, development and metastasis of malignancy.^{22,23} Inflammation inhibition during the cancer treatment plays essential roles in improving efficacy of chemotherapeutics.^{24,25} Several non-steroid anti-inflammatory drugs (NSAIDs) like aspirin, ibuprofen,

indomethacin, and flurbiprofen, which display dramatic anti-inflammatory properties mainly targeting cyclooxygenases (COXs), have been incorporated with the platinum(IV) system to form a series of promising complexes.^{26–38} The mechanism detection discloses that these complexes display prominent inflammation inhibitory properties besides causing serious DNA damage. Our previous work reported a series of mono-naproxen platinum(IV) complexes with prominent antitumor activities, which showed great potential in inhibiting tumor migration by synergistic inhibition of COX-2 and matrix metalloproteinases (MMPs).³⁹ More recently, Guo and Wang's group reported the efficient antitumor performance of dual naproxen platinum(IV) complex **2**.⁴⁰ It targeted COX-2 and PD-L1 via multiple pathways to significantly inhibit the immune escape of tumor cells and finally improve antitumor efficacy. Thereby, naproxen platinum(IV) complex represents a promising scaffold for further development of novel platinum drugs.

In view of the potent activities of NSAID derived platinum(IV) complexes and the advantages of nano-DDS, nanoparticles of NSAID platinum(IV) would be effective to facilitate cellular uptake and improve apoptosis-inducing ability.³² However, nano-DDS based on biocompatible BSA loading such platinum(IV) complexes has not been reported. Inspired by these observations and as a continuation of our interest in developing new platinum antitumor drugs,^{41–43} two dual naproxen platinum(IV) complexes **1** and **2** with oxaliplatin and cisplatin cores were prepared (Figure 1). Then, BSA nanoparticles **1-NPs** and **2-NPs** loaded with dual naproxen platinum(IV) complexes were designed, prepared, and characterized. Their antitumor activities and toxic properties were evaluated in vitro and in vivo, and their likely antitumor mechanisms were also detected.

Materials and Methods

Chemistry

General

All reactions were carried out under an atmosphere of nitrogen in flame-dried glassware with magnetic stirring unless otherwise indicated. Cisplatin and oxaliplatin were purchased from Boyuan Chemical Co. Ltd (Jinan, China). Other reagents were obtained from Sigma, Aladdin, Innochem, and J&K Scientific Ltd. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker (500 MHz and 125 MHz). All NMR chemical shifts were referenced

to residual solvent peaks or TMS as an internal standard. All coupling constants *J* were quoted in Hz. Mass spectra (MS) were obtained on a Shimadzu LC-MS/MS 8040 mass spectrometer with ESI ionization. The particle size (Z-average), polydispersity index and zeta potential were detected by dynamic light scattering (DLS, Zetasizer Nano ZSP, Malvern, UK). Transmission electron microscope (TEM) was recorded on JEM-2100 (JEOL, Japan). The BSA was purchased from J&K. Genomic DNA mini-preparation kit, was purchased from Beyotime, China. UV-vis spectra were measured on a Scinco S-3100 UV-vis spectrophotometer. The fluorescence spectra were recorded on a Hitachi F-7000. The cells were purchased from Procell Life Science & Technology Co., Ltd (China).

Synthetic Procedures

The synthetic procedures for oxoplatin **O1–O2** were supplied in the supporting information according to procedures reported in the literature.^{42,44}

Preparation of Compound 1

Complex **1** was prepared as reported in our previous work.³⁹ A solution of naproxen (134 mg, 0.58 mmol) and *N,N,N',N'*-tetramethyl-O-(benzotriazol-1-yl)uranium tetrafluoroborate (TBTU) (187 mg, 0.58 mmol) in dry DMF 5 mL was stirred at room temperature for 10 min after injection of *N,N,N*-triethylamine (TEA) (80 µL, 0.58 mmol). Subsequently, oxoplatin **O1** (86 mg, 0.20 mmol) was added, and the reaction mixture was stirred vigorously at 50°C for 48 h in the dark. After the reaction completed, the solvent was evaporated and the residue was purified by silica gel column chromatography using dichloromethane-methanol (80: 1, v/v). Complex **1** was obtained as white solid (54.8 mg, 32%), with 97.9% purity as determined by HPLC (methanol-H₂O, 70%).

¹H NMR (500 MHz, DMSO-*d*₆) δ 7.80–7.65 (m, 6H), 7.39–7.34 (m, 2H), 7.27 (s, 2H), 7.15–7.10 (m, 2H), 3.92–3.82 (m, 8H), 2.26–2.03 (m, 2H), 1.98–1.78 (m, 2H), 1.46–1.36 (m, 6H), 1.26–1.20 (m, 2H), 1.16–1.07 (m, 2H), 0.77–0.49 (m, 2H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 182.2, 163.6, 157.5, 137.3, 133.6, 129.5, 128.8, 127.1, 127.0, 125.8, 119.0, 106.1, 61.3, 55.6, 46.9, 31.2, 23.5, 19.6. ESI-MS: calcd for [M + H]⁺: 857 (M = C₃₆H₄₀N₂O₁₀Pt), found: 857.³⁹

Preparation of Compound 2

A solution of naproxen (134 mg, 0.58 mmol) and TBTU (187 mg, 0.58 mmol) in dry DMF 5 mL was stirred at

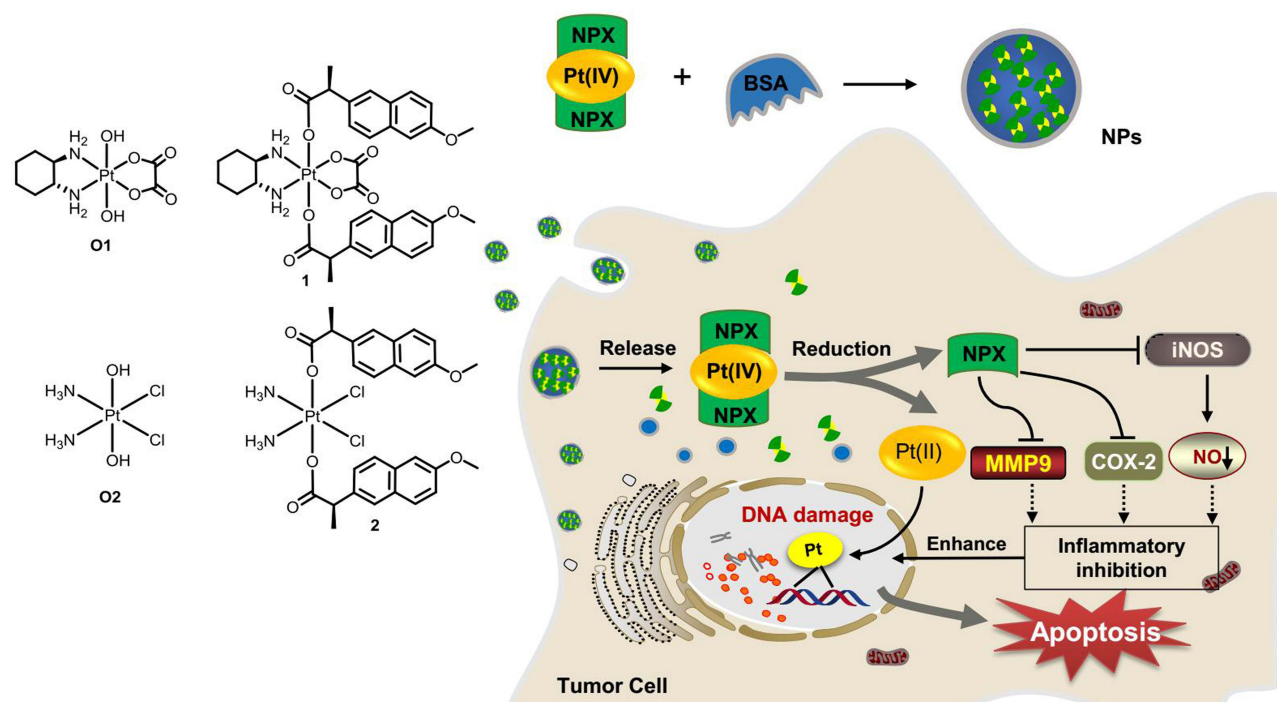


Figure 1 Structure of oxoplatins **O1** and **O2**, dual naproxen platinum(IV) complexes **1** and **2**, the preparation of nanoparticles **1-NPs** and **2-NPs**, and their likely antitumor mechanism.

Abbreviations: NPs, nanoparticles; BSA, bovine serum albumin; COX-2, cyclooxygenase-2; MMP-9, metalloproteinase-9; iNOS, inducible nitric oxide synthase.

room temperature for 10 min after injection of TEA (80 μ L, 0.58 mmol). Subsequently, oxoplatin **O2** (67 mg, 0.20 mmol) was added, and the reaction mixture was stirred vigorously at 50°C for 48 h in dark. After the reaction completed, the solvent was evaporated and the residue was purified by silica gel column chromatography using dichloromethane-methanol (70:1, v/v). Complex **2** was obtained as white solid (36.4 mg, 24%). 98.1% purity determined by HPLC (methanol-H₂O, 70%).

¹H NMR (500 MHz, DMSO-*d*₆) δ 7.82–7.68 (m, 6H), 7.48 (d, *J* = 8.4 Hz, 2H), 7.27 (s, 2H), 7.13 (dd, *J* = 8.9, 1.9 Hz, 2H), 6.55 (s, 6H), 3.86 (s, 6H), 3.07 (d, *J* = 7.0 Hz, 2H), 1.41 (d, *J* = 7.1 Hz, 6H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 182.47, 157.40, 137.84, 133.51, 129.55, 128.83, 127.47, 126.82, 125.97, 118.83, 106.14, 55.61, 46.88, 46.22, 20.29. ESI-MS: Calcd. for [M+H]⁺: 759 (M = C₂₈H₃₂Cl₂N₂O₆Pt), found: 759.

Preparation of Nanoparticles 1-NPs and 2-NPs

Briefly, free compound **1** or **2** (5 mg, 1.0 eq.) was dissolved in 1.5 mL ethanol containing 5% dichloromethane under continuous ultrasonication. The organic solution was added dropwise into 15 mL deionized water with BSA (complex **1** or **2**/BSA; 2.0 eq./1.0 eq.,

3.0 eq./1.0 eq., 5.0 eq./1.0 eq. and 7.5 eq./1.0 eq.), and the mixture was stirred for 20 min vigorously at room temperature. Then, the organic solvent was evaporated under vacuum to about 12 mL. The obtained NPs were then homogenized by a D-3L homogenizer (PhD, USA) at optimal homogenization conditions of 100 MPa for 15 min to reduce the particle size. Then, deionized water was supplied, and solution of nanoparticles 15 mL was obtained.

Characterization of NPs

The particle size (Z-average), polydispersity index and zeta potential of NPs were detected by dynamic light scattering (DLS, Zetasizer Nano ZSP, Malvern, UK) with an integrated 4 mV He-Ne laser (λ =633 nm) and the backscattering detection (scattering angle θ =173°) at set temperature. Experiments were performed in triplicate.

The morphology of NPs was analyzed by transmission electron microscope (TEM, JEM-2100, JEOL, Japan). One drop of NP (100 μ g/mL of free complex) was placed on a carbon-coated copper grid and stained with uranyl acetate (2%, w/v). The TEM observations were performed at an accelerating voltage of 80 kV.

Biological Evaluation

In vitro Cellular Cytotoxicity Assays

The naproxen platinum(IV) nanoparticles **1-NPs**, **2-NPs** and free compounds **1**, **2** were evaluated for antitumor activities against four tumor cell lines including human lung cancer (A549), human ovarian cancer (SKOV-3), murine colon cancer (CT-26) and a cisplatin-resistant cell line A549R, and a human normal liver cell line LO-2 using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) assay with cisplatin, oxaliplatin, and naproxen as reference drugs.⁴⁵ A BSA nanoparticle without drugs was prepared and tested as a vehicle. The cells were maintained in RPMI1640 medium containing 10% FBS in a humidified atmosphere containing 5% CO₂ at 37°C. The A549R was maintained with 2 µg/mL cisplatin. Phosphate buffered saline (PBS) contained 137 mM NaCl, 2.7 mM KCl, 10 mM Na₂HPO₄ and 2 mM KH₂PO₄ (pH 7.4). MTT solution at concentration of 5 mg·mL⁻¹ for MTT assays was prepared before use.

Cells were seeded in 96-well plates at 5000 cells per well in 100 µL of complete medium, and incubated for 24 h in a 5% CO₂ atmosphere at 37°C. Then freshly prepared culture medium 100 µL containing drugs at different concentrations were added. The cells were further incubated for another 48 h or 72 h. After that, freshly prepared MTT solution (5 mg·mL⁻¹) 20 µL was added. The resultant mixtures were incubated for 4 h to allow viable cells to reduce the yellow tetrazolium salt into dark blue formazan crystals. After removal of the medium, formazan was dissolved in DMSO (150 µL) and quantified by a microplate reader (570 nm). The IC₅₀ values were calculated using GraphPad Prism 6 based on three parallel experiments.

In vivo Antitumor Assay

Male BALB/c mice (18–20 g) were purchased from the Laboratory Animal Center, Shandong University (Shandong, China). This study was performed in strict accordance with the NIH guidelines for the care and use of laboratory animals (NIH Publications No. 8023, revised 1978), and was approved by the Institute Animal Ethics Committee of Liaocheng University (Liaocheng, China).

Murine CT-26 cells (5×10⁵) suspended in PBS were injected subcutaneously into the right flank of BALB/c mice and therapy was started when tumor nodules were palpable (day 6). The mice were divided into seven groups randomly, saline; cisplatin (2 mg Pt/kg, i.v.); oxaliplatin

(4 mg Pt/kg, i.v.); compound **1** (4 mg Pt/kg, i.p.), **1-NPs** (4 mg Pt/kg, i.v.); compound **2** (2 mg Pt/kg, i.p.), **2-NPs** (2 mg Pt/kg, i.v.). Freshly prepared solutions of drugs were administered four times, on days 6, 9, 12, and 14 post-tumor inoculation. Tumor volume (*V*) was determined as $V=L \times W^2/2$, where the length (*L*) and width (*W*) were measured. At day 15, 24 h after the last drug application, the mice were euthanized. The tumors were excised and weighed. The blood and tissues including liver, heart, spleen, lung, and kidney were collected. Then, tissues were formalin-fixed in 4% formaldehyde overnight, paraffin-embedded, and cut with a microtome (5 µm sections) for hematoxylin and eosin (H&E) staining and immunohistochemical detection.

Determination of NO Production

The cells RAW 264.7 (1×10⁶ cells/well) in 100 µL media were preincubated in 96-well plates for 24 h. Then the media was removed. Fresh media 100 µL containing lipopolysaccharide (LPS) (1 µg/mL) and drug at different concentrations were added and the cells were further incubated for 24 h. Then, the growth medium 75 µL was mixed with the same volume of Griess reagent for 5 min and absorbance was measured at 570 nm. The accumulation amount of nitrite was determined as an index of NO. Three replicates were performed for all the treatments.

AST, ALT, BUN and CRE Detection

The blood of mice was collected in Eppendorf tubes and centrifuged at 1500 g for 15 min to collect serum samples. Then the AST, ALT, BUN and CRE were detected using enzyme-linked immunosorbent assay kits according to the manufacturer's instructions: serum aspartate aminotransferase (AST ELISA kit, Nanjing Jiancheng Bioengineering Institute, China), alanine aminotransferase (ALT ELISA kit, Nanjing Jiancheng Bioengineering Institute, China), blood urea nitrogen (BUN ELISA kit, Nanjing Jiancheng Bioengineering Institute, China) and creatinine (CRE ELISA kit, Nanjing Jiancheng Bioengineering Institute, China). The OD values were recorded on an automatic microplate reader (Tecan Spark).

COX-2 Inhibition

The COX inhibitory experiment was carried out using a cyclooxygenase 2 inhibitor screening kit (Beyotime, China) according to the manufacturer's protocol. The rhCOX-2 was applied in the experiments. The inhibitions of compounds **1** and **2** at different concentrations (5, 20 µM) and naproxen at corresponding concentrations (10, 40

μM) to COX-2 were tested. Then the reduced systems of complexes **1** and **2** with ascorbic acid (AsA) (Compd.-AsA) were prepared by incubation of compound **1/2** solution under treatment of AsA (1 mM) for 48 h to allow the platinum(IV) to totally convert to platinum(II) complexes with release of two equivalent of naproxen. The compound **1**-AsA and compound **2**-AsA were also tested at concentrations of 5 and 20 μM . Then, cisplatin and oxaliplatin at 20 μM were also evaluated. The sample with no rhCOX-2 inhibitor was evaluated as blank, and celecoxib was used at the suggested concentration in the protocol (100 nM) as positive reference sample.

Immunohistochemical Detection

Tumor tissues of **1**-NPs, **1**, oxaliplatin and saline treated groups were stained using MMP-9 and iNOS staining assays according to the standard procedure. Immunohistochemistry was performed on formalin-fixed, paraffin-embedded organism sections. Following deparaffinization and hydration, sections were immersed into citrate buffer, boiled in a microwave for antigen repair and blocked in 10% normal goat serum (Sigma, USA) for 60 min. Then, sections were incubated at 4°C with primary antibody MMP-9 (Servicebio, China; DILution ratio, 1:100) or iNOS (Proteintech; DILution ratio, 1:800) overnight. Subsequently, sections were washed in PBS and incubated with goat anti-mouse IgE-HRP secondary antibody (Servicebio, China; DILution ratio, 1:200) for 50 min at room temperature. After washed with PBS, the sections were visualized with 3,3'-diaminobenzidine (DAB) (Servicebio, China), and counterstained using hematoxylin. The MMP-9 and iNOS index was calculated as the mean optical density (percent) by choosing five random fields with an optical microscope.

Accumulation of Platinum in Whole Cells and DNA

The CT-26 cells were seeded in 6-well cell culture plate and incubated for 12 h at 37 °C. Then the cells were treated with platinum compounds (100 μM) including nanoparticles **1**-NPs, **2**-NPs, complexes **1**, **2** and reference drugs cisplatin, oxaliplatin for 10 h in incubator. After that, all the cells were harvested, collected and washed for three times with PBS. Approximately one million cells were mineralized with 70% HNO_3 (LC), and the platinum in cells was determined with ICP-MS. Results were presented as the mean of three determinations for each data point. To test the DNA platination of the tumor cells, the DNA of approximately one million cells

was isolated with a Genomic DNA Mini-Preparation Kit (Beyotime, China). Then the DNA was mineralized with 70% HNO_3 and measured by ICP-MS.

Platinum in Tumor, Liver and Kidney

Tissues including tumor, liver and kidney from nanoparticles **1**-NPs, **2**-NPs, free compounds **1**, **2**, reference drugs cisplatin and oxaliplatin treated groups were collected and mineralized with 70% HNO_3 (LC). The platinum in tissues was determined with ICP-MS.

Fluorescence and UV-vis Spectra Assays for BSA Interaction

The fluorescence spectra were recorded on a Hitachi F-7000 spectrofluorometer using a quartz cell of 1.00 cm. The slits of both excitation and emission were 5.0 nm. The quenching spectra of BSA were recorded by the gradually addition of compound **1/2** (0.0, 8.0, 14.0, 20.0 μM) to BSA solution (4.0 μM) in buffer (10 mM Tris-HCl/10 mM NaCl, pH 7.4). The well-mixed solutions were holding for 15 min for equilibrium and tested under excitation of 298 nm at a scan rate of 300 nm·min⁻¹ under excitation of 280 nm. The fluorescence spectra of free complexes (20.0 μM) were also detected.

Then, the UV-vis spectra were also employed to testify the BSA interaction of compounds **1** and **2**. The solution of BSA (10 μM) was recorded as line a. The solution of BSA-Compound (1:1) (10 μM) was recorded as line b. The solution of compound (10 μM) was recorded as line c. Line d was obtained by the subtraction of line b and line c.

Results and Discussion

Chemistry

The Synthesis of Naproxen Platinum(IV) Complexes
Naproxen platinum(IV) complexes **1** and **2** were prepared by the condensation of naproxen with oxoplatins **O1** and **O2** in the presence of TBTU and TEA (Figure S1). Oxoplatins **O1** and **O2** were obtained by the oxidation of cisplatin, oxaliplatin with hydrogen peroxide as reported in our previous work.^{46,47}

The Particle Size, Zeta Potential and Morphology

The BSA nanoparticles of complexes **1** and **2** were prepared for four drug/BSA ratios (complex **1** or **2**/BSA; 2.0 eq./1.0 eq., 3.0 eq./1.0 eq., 5.0 eq./1.0 eq. and 7.5 eq./1.0 eq.) as shown in Tables S1 and S2. The results revealed that drug/BSA ratio of 3.5:1 is suitable for preparing the **1**-NPs due to

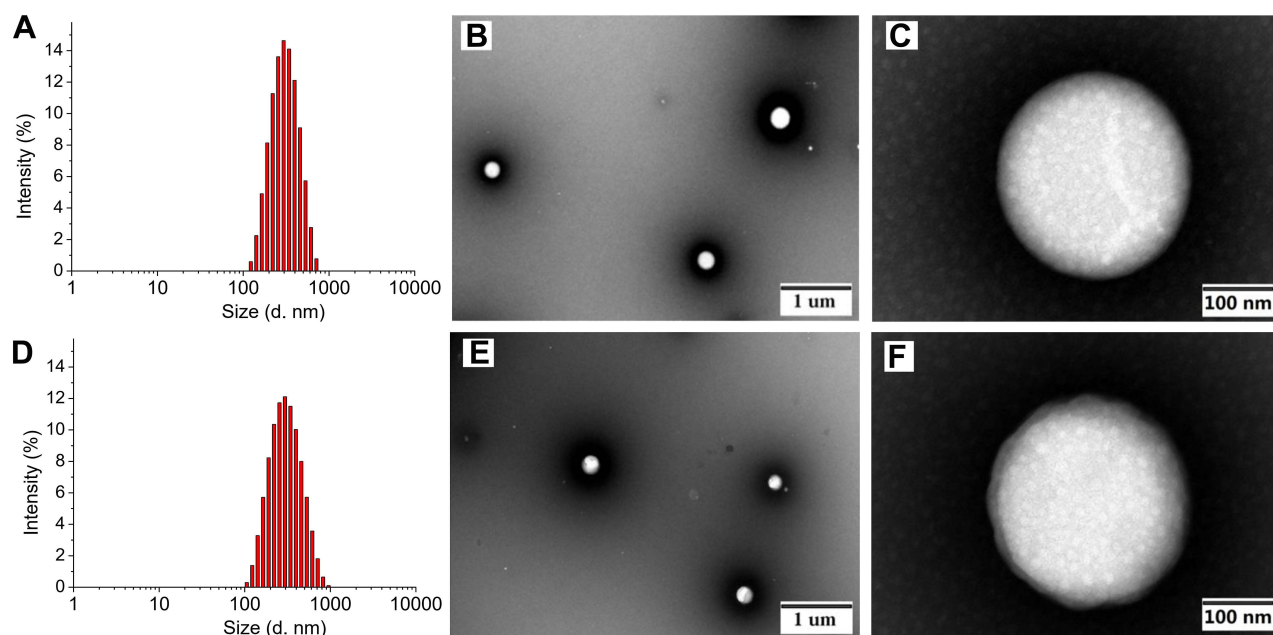


Figure 2 Mean hydrodynamic diameter determined by DLS, and TEM images of **1-NPs** (**A–C**) and **2-NPs** (**D–F**).

Abbreviations: DLS, dynamic light scattering; TEM, transmission electron microscope; NPs, nanoparticles.

the minimum size (280.2 ± 6.5 nm), narrow distribution ($PDI = 0.32 \pm 0.03$) and high zeta potential (-29.5 ± 0.3 mV). Meanwhile, drug/BSA ratio of 5.0:1 is suitable for preparing the **2-NPs**, which affords particles with minimum size of 273.1 ± 1.5 nm, low PDI of 0.30 ± 0.02 , and high zeta potential of -22.8 ± 0.8 mV (Figure 2). It was proven that the NPs with negative surface charge would show preferential uptake at the tumor site with higher cationic charge density in comparison with normal organs.^{48,49} Thereby, the negative zeta potential of the NPs indicates that **1-NPs** and **2-NPs** keep stable in aqueous environment, and these facts might be a factor accounting for the high accumulation of nanoparticles in tumor cells and tissues as depicted by cellular uptake results (see, tumor target properties section). Subsequently, the morphology of **1-NPs** and **2-NPs** were observed by TEM. Both particles are mostly uniform spheres in shape without sign of agglomeration with size of 275 nm and 283 nm respectively. These results coincide with the size analyzed by DLS.

To illustrate the assembled mechanism of **1-NPs** and **2-NPs**, the interaction of complexes **1** and **2** with BSA were investigated by fluorescence and UV-vis absorption spectroscopies. The fluorescence spectroscopy is widely applied in the exploration of structural and conformational changes of proteins. The fluorescence spectra in Figure S2 demonstrate that the emission fluorescence of BSA at 340 nm is quenched with the addition of platinum(IV) complexes **1** or **2**. These

facts indicate that complexes **1** and **2** could combine with BSA noncovalently. Then, UV-vis spectra of BSA in the absence and presence of compounds **1** and **2** (Figure S3) were measured to determine the binding mechanism. The differential absorption spectra (curve d) between compound **1/2**-BSA complex (curve a) and platinum(IV) complex **1/2** (curve c) are apparently different from the spectra of BSA (curve b) at equivalent concentrations. Conclusively, the binding of platinum(IV) complexes **1** and **2** with BSA is a static quenching process due to the complex formation.

Stability Studies

The long-term storage stability studies of **1-NPs** and **2-NPs** were performed at 4°C with size measurement by DLS. The nanoparticles **1-NPs** and **2-NPs** exhibit no dramatic change of size, PDI and zeta potential during 30 days storage at 4°C , $\text{pH} = 7.4$. The results in Figure 3 (Table S3) manifested that the mean hydrodynamic diameter of **1-NPs** was 266.0 ± 16.7 nm with a narrow distribution ($PD = 0.37 \pm 0.01$) on day 30, which was similar to the original size of 280.2 ± 6.5 nm. Meanwhile, the size of **2-NPs** was approximately 297.1 ± 4.9 nm at 30 days, which had no significant difference compared to the size at day 0 (273.1 ± 1.5 nm). Moreover, the PDI and zeta potential of both nanoparticles also kept stable during the 30 days storage (Table S8). All these facts demonstrate that **1-NPs** and **2-NPs** could remain stable for at least 30 days in the aqueous phase under condition of

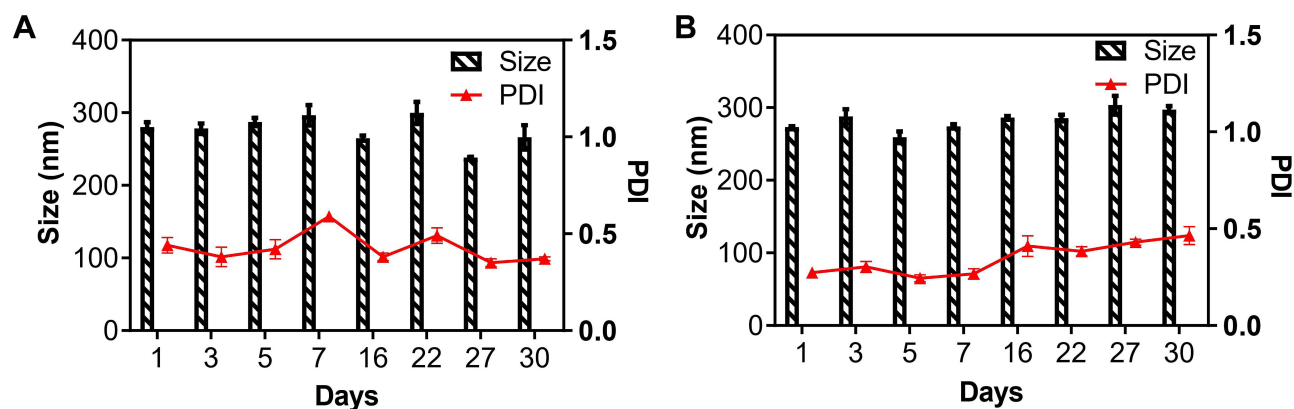


Figure 3 The particle size and PDI of (A) 1-NPs and (B) 2-NPs during storage of 30 days (pH=7.4, T=4°C).

Abbreviations: PDI, polydispersity index; NPs, nanoparticles.

T=4°C, pH=7.4. Then, to detect the influence of temperature on stability, 1-NPs and 2-NPs were also evaluated at 25°C and 37°C. Results in [Tables S4, S5, S9 and S10](#) revealed that the increased temperature remarkably decreased the stability of NPs. The size of both 1-NPs and 2-NPs increased significantly at day five both at 25°C and 37°C accompanied with the drop Zeta potential. To further detect the influence of pH, the change of size, PDI and zeta potential of NPs were also tested at pH=6.4 (the pH in TME) at 4°C and 37°C. Results in [Tables S6, S7, S11 and S12](#) manifested that low pH led to no significant change of 1-NPs and 2-NPs at 4°C in seven days. However, accelerated change of size and zeta potential of 1-NPs and 2-NPs were observed at 37°C (pH=6.4) in comparison with that in pH=7.4. Thereby, the optimized storage conditions of the BSA NPs in this work was T=4°C, pH=7.4.

Antitumor Activity in vitro

The in vitro antitumor activities of BSA-based naproxen platinum(IV) nanoparticles 1-NPs and 2-NPs and free complexes 1 and 2 on four tumor cell lines including human ovarian cancer (SKOV-3), murine colon cancer (CT-26), human lung cancer (A549), and a cisplatin-resistant cell line A549R were evaluated by MTT assay with cisplatin and oxaliplatin as reference drugs. Then, a human normal liver cell LO-2 was also evaluated and the results were given as IC₅₀ values in [Table 1](#). To evaluate the effects of incubation time on antitumor activities, the IC₅₀ values for 1, 2, 1-NPs and 2-NPs against SKOV-3 were tested after 48 h and 72 h incubation. It could be observed that the incubation time prolonging from 48 h to 72 h induces no dramatic change of the

IC₅₀ values. Thereby, the 48 h incubation time was selected and applied in the following evaluation.

Free naproxen platinum(IV) hybrids 1 and 2 display effective antitumor activities to all tumor cells with IC₅₀ values below 11.3 μM. Especially, complex 2 with cisplatin core exhibits promising activities (IC₅₀ ≤ 0.32 μM) in vitro, which are significantly more effective than cisplatin and oxaliplatin. However, the free compounds possess rather poor solubility in water with high *Alog P*-values for complexes 1 (6.263) and 2 (6.182) respectively. Thereby, it is of great importance to prepare nanoparticles of such naproxen platinum(IV) complexes, which would improve solubility of drugs and further influence their activities.

As disclosed in [Table 1](#), the BSA nanoparticles 1-NPs and 2-NPs loaded naproxen platinum(IV) complexes 1 and 2, respectively show effective antitumor activities, while the vehicle BSA exhibits no toxicity to tumor cells. Notably, all the IC₅₀ values for 2-NPs with cisplatin platinum(IV) complex are lower than reference drug cisplatin, meanwhile 1-NPs with oxaliplatin core also display comparable or even better activities than oxaliplatin. Furthermore, 1-NPs and 2-NPs show much potential in overcoming drug resistance of A549R with low RF values of 0.4 and 0.7, which are significantly lower than cisplatin (RF=2.6) and free complex 1 (RF=1.2). Then, a normal cell line LO2 was tested to judge the toxicities of tested NPs to normal cells. Tumor selectivity index (SI) is determined as a ratio of the IC₅₀ of LO-2 to the average IC₅₀ of tumor cells.^{39,43} Nanoparticles possess dramatically improved SI values in comparison with the free complexes that 1-NPs and 2-NPs afford high SI values of 3.9 and 3.0 which are over 2.0–5.6 times higher than compounds 1 (SI=1.5) and 2 (SI=0.7) as well as platinum(II) drugs cisplatin (SI=1.1) and oxaliplatin (SI=1.0). These facts

Table I Cytotoxicity Profiles of Naproxen Platinum(IV) Complexes Toward Four Carcinoma Cell Lines, and One Normal Human Cell Line Expressed as IC₅₀ (μM)

Compounds	Alog P ^a	SKOV-3 (48 h)	SKOV-3 (72 h)	CT-26 (48 h)	A549 (48 h)	A549R (48 h)	RF ^b	Average IC ₅₀ (48 h) ^c	LO-2 (48 h)	SI ^d
1 ³⁰	6.263	11.3±0.8	15.4±0.4	8.2±0.6	10.2±1.0	12.0±0.4	1.2	10.4	15.3±2.5	1.5
1-NPs	/ ^e	3.8±0.5	2.9±0.3	2.3±0.4	9.7±2.1	3.6±0.7	0.4	4.8	18.7±4.6	3.9
2	6.182	0.08±0.01	0.11±0.02	0.32±0.07	0.14±0.04	0.04±0.01	0.3	0.14	0.09±0.01	0.7
2-NPs	/	0.8±0.1	0.6±0.1	1.0±0.2	1.5±0.2	1.0±0.3	0.7	1.1	3.3±0.4	3.0
Npx	2.849	/		>100	>100	>100	/	/	>100	/
Vehicle	/	>100		>100	>100	>100	/	/	>100	/
CDDP	/	1.9±0.4	1.2±0.3	1.2±0.3	2.8±0.6	7.1±1.3	2.6	3.0	3.4±1.2	1.1
OLP	/	5.9±1.3	7.5±1.8	4.1±0.7	6.7±1.8	2.4±0.6	0.4	4.3	4.8±0.7	1.0

Notes: ^aThe values of Alog P were predicted by Discovery Studio; ^bRF=IC₅₀(A549R)/IC₅₀(A549); ^cAverage IC₅₀ values for four tumor cell lines; ^dSI = IC₅₀(LO-2)/Average IC₅₀; ^eNot tested or not calculated.

Abbreviations: NPs, nanoparticles; Npx, naproxen; RF, resistant factor; SI, selectivity index, CDDP, cisplatin; OLP, oxaliplatin.

indicate the low toxicities of the prepared BSA nanoparticles to normal cells.

Ultimately, BSA based naproxen platinum(IV) nanoparticles **1-NPs** and **2-NPs** display prominent antitumor activities to all tested tumor cells and exhibit much potential in overcoming drug resistance of cisplatin. Both nanoparticles show low toxicities to normal cells in contrast to cisplatin and oxaliplatin as well as the free platinum(IV) complexes **1** and **2**. The antitumor activities in vivo and action mechanism of the NPs were investigated in the following experiments.

Antitumor Efficacy in vivo

Colonic carcinoma as one of the most common cancers worldwide is a major cause of morbidity and mortality.⁵⁰ Platinum drugs occupy an important position in colonic carcinoma treatment. In addition, naproxen has been proven to be favorable for treating colonic carcinoma to combine with platinum drugs. Herein, BALB/c mice bearing CT-26 homograft tumors were selected as models to study the effectiveness of BSA nanoparticles **1-NPs** and **2-NPs** to compare with free naproxen platinum(IV) complexes **1** and **2**, using cisplatin and oxaliplatin as reference drugs. Complex **1** and **1-NPs** with oxaliplatin core and oxaliplatin were administrated at dosage of 4 mg Pt/kg. Then, cisplatin was given at a low dosage of 2 mg Pt/kg taking its highly toxic properties in consideration, and the cisplatin-derived compound **2** and **2-NPs** were also administrated at the same dosage. The nanoparticles **1-NPs**, **2-NPs** and platinum(II) drugs cisplatin and oxaliplatin were administrated via

intravenous injection (i.v.) with saline as blank, while free complexes **1** and **2** were administrated via intraperitoneal injection (i.p.) for their poor solubility in saline.

The results in Figure 4 showed that nanoparticles **1-NPs**, **2-NPs** show remarkable inhibition of the tumor growth in vivo in contrast to saline group ($P<0.001$), which are relatively more effective than the free compounds **1** and **2** ($P<0.05$), and this trend is contrary to the results in vitro. Tumor for the saline group rapidly grows to 1586 mm³ in 15 days (Figure 4A and C), while the volumes of reference drugs oxaliplatin and cisplatin groups are 477 mm³ and 399 mm³. Nanoparticles **1-NPs** and **2-NPs** suppress the tumor volumes to 654 mm³ and 562 mm³ effectively, which are smaller than the corresponding free complex treated groups **1** (1041 mm³, $P<0.001$) and **2** (836 mm³, $P<0.05$) respectively. Then, tumor weight was measured after the mice were sacrificed and given in Figure 4B. Tumor growth inhibition rate (TGI) as an important index to judge effectiveness of antitumor drugs was calculated as a ratio of tumor weight of the drug-treated group to that of the saline group, and was depicted above the columns. Nanoparticles **1-NPs** and **2-NPs** show higher TGI of 64.7% and 58.9% than free complexes **1** (TGI=45.8%, $P<0.05$) and **2** (TGI=41.2%, $P<0.05$). The hematoxylin and eosin (H&E) staining of tumor tissues in Figure 4D shows that **1-NPs**, **2-NPs** and free compound **1** and **2** induce obvious apoptosis and necrosis in tumor tissues in comparison with saline, and this evidence is similar to that in the cisplatin and oxaliplatin treated groups.

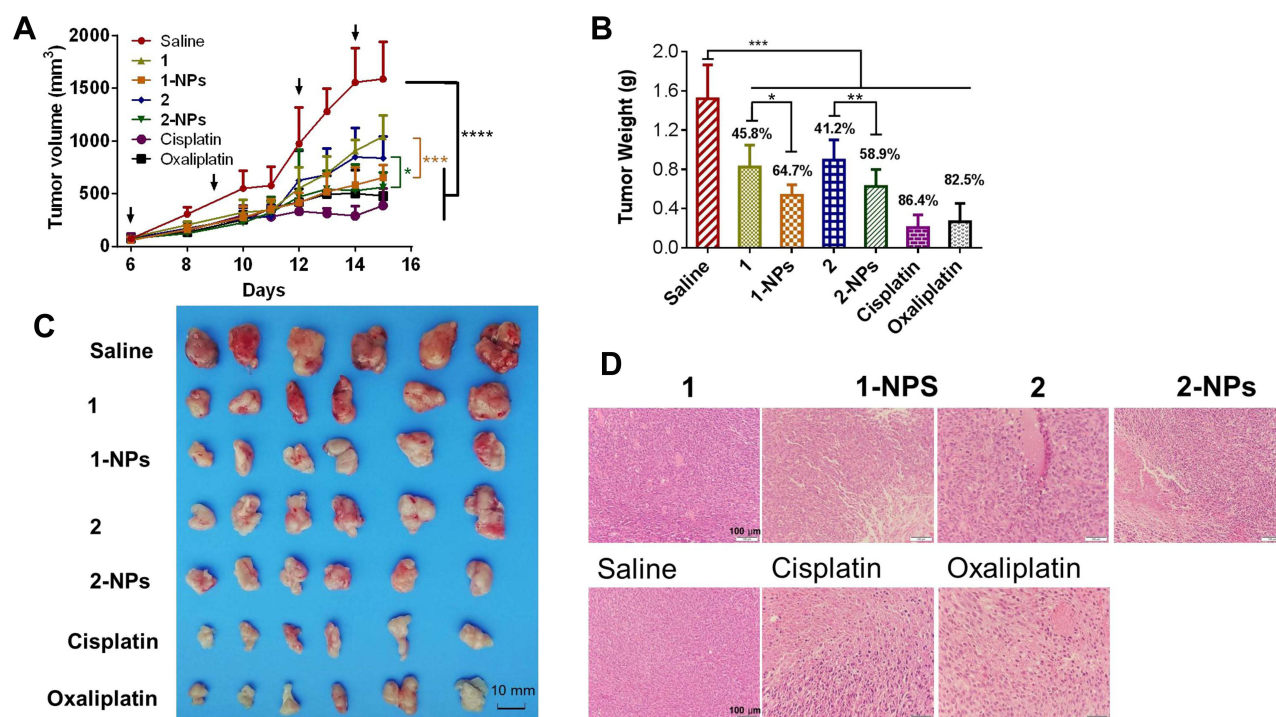


Figure 4 In vivo antitumor activities of nanoparticles and free naproxen platinum(IV) compounds to CT-26 homograft tumors in BALB/c mice. Dosage: 4 μ g Pt/kg for **1-NPs** (i.v.), **1** (i.p.) and oxaliplatin (i.v.); 2 μ g Pt/kg for **2-NPs** (i.v.), **2** (i.p.) and cisplatin (i.v.). Drugs were injected on days 6, 9, 12, 14 posttumor inoculation as indicated by the arrows in Figure 4A. **(A)** Tumor growth as a function of time. **(B)** Tumor weight in each group at the end of the experiment. The TGI of the tested drugs are depicted above the column (TGI=tumor weight of drug treated group/tumor weight of saline group). **(C)** The images of the tumors at the end of the experiment. **(D)** The H&E staining of slices from tumor tissues.

Notes: Results are presented as the mean \pm SD (n=6). * P <0.05, ** P <0.05, *** P <0.001, **** P <0.001.

Abbreviations: NPs, nanoparticles; i.p., intraperitoneal; i.v., intravenous; TGI, growth inhibition rate; H&E, hematoxylin and eosin.

Toxicity in vivo

The potential toxicities of the nanoparticles were assessed systematically during the treatment in vivo to compare with free complexes, cisplatin and oxaliplatin. Body weight as a primary indicator measuring toxic properties of drugs was monitored and supplied in Figure 5A. It is disclosed that cisplatin (2 mg Pt/kg) and oxaliplatin (4 mg Pt/kg) possess the most serious toxicity to mice by inducing body weight loss of 19.8% and 16.1% in comparison with the saline group at the end of the experiment (P <0.01). Moreover, anorexia and dyskinesia, which associate with toxicity, were observed in the cisplatin and oxaliplatin treated groups for approximately eight hours after administration. In contrast, the tested groups **1-NPs** and **2-NPs** as well as free complexes **1** and **2** induce no apparent signs of muscle loss, dehydration, anorexia, locomotor impairment or other unusual symptoms during the treatment, and cause rather lower weight loss of mice (0.2%–10.7%) than cisplatin and oxaliplatin (P <0.01). These facts reflect the lower toxic effects of naproxen platinum(IV) nanoparticles and free compounds in

comparison with platinum(II) drugs. Furthermore, nanoparticles possess reduced toxic properties in comparison with free naproxen platinum(IV) complexes that the loss of body weight for nanoparticle **2-NPs** (5.6%) is rather lower than free compound **2** (10.6%) (P <0.001), meanwhile **1-NPs** (2 mg Pt/kg) also induces relatively lower body weight loss (0.2%) than free complex **1** (2.5%).

The toxic effects of drugs on the organs including heart, liver, spleen, lung and kidney were evaluated by H&E staining assay and organ weight indexes. The nanoparticles **1-NPs**, **2-NPs**, and free complexes **1**, **2** as well as the reference drugs cisplatin and oxaliplatin cause no appreciable histological differences of the organs to compare with the saline group (Figure 5C). However, the organ weight indexes in Figure S4 as ratios of organ weight to body weight reveal remarkable suppressions of spleen induced by cisplatin and oxaliplatin in comparison with the saline group (P <0.01), which indicates the spleen damage caused by the platinum(II) drugs. Meanwhile nanoparticles **1-NPs** and **2-NPs** improve spleen indexes to some extent, and free

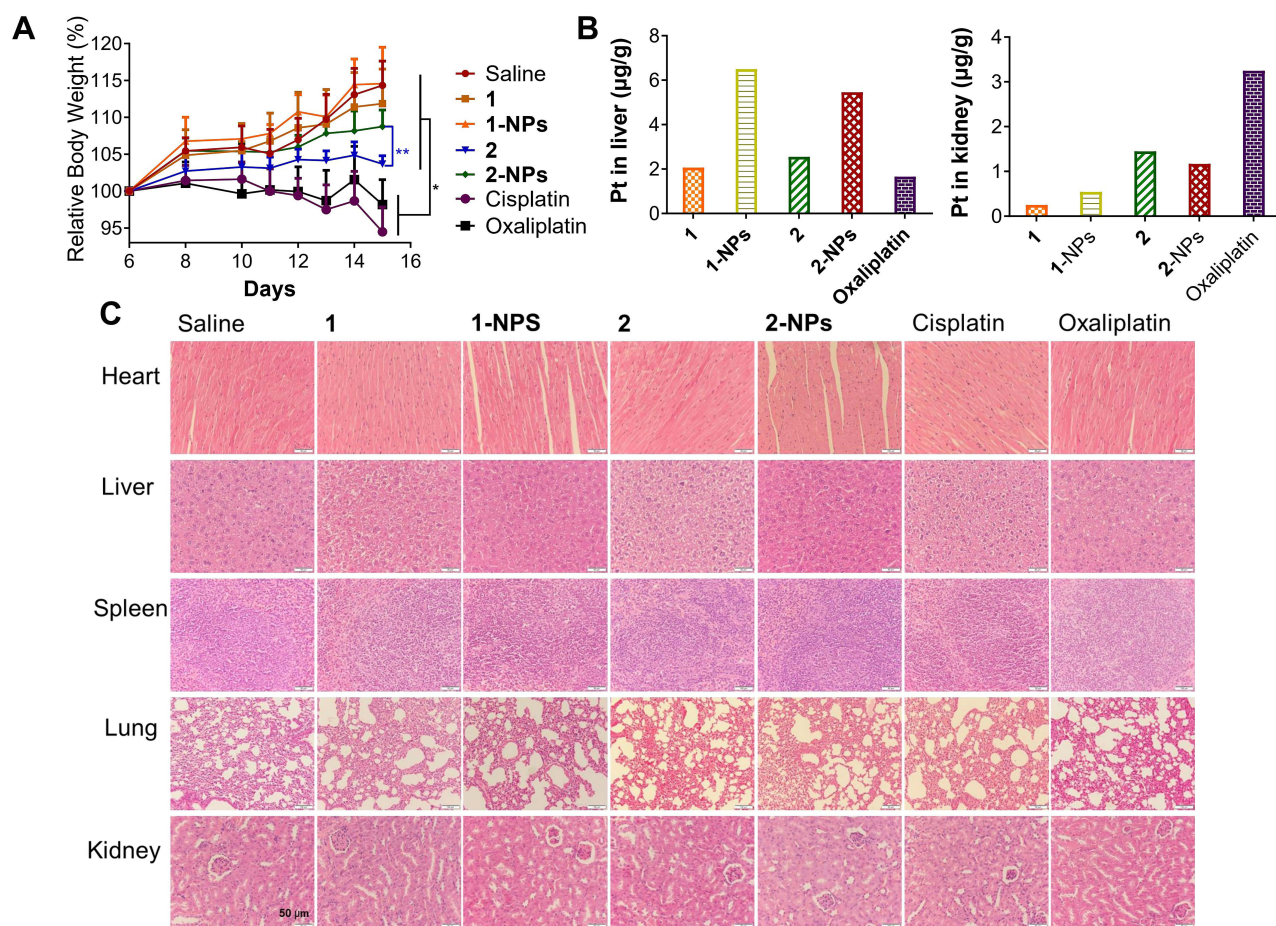


Figure 5 In vivo toxicities of tested compounds and nanoparticles to BALB/c mice. **(A)** The body weight of the mice during the treatment. **(B)** Drug accumulation in liver and kidney. **(C)** The H&E staining of slices from heart, liver, spleen, lung and kidney.

Notes: * $P < 0.01$, ** $P < 0.001$.

Abbreviations: NPs, nanoparticles; H&E, hematoxylin and eosin.

complexes **1** and **2** induce no significant influences. These results are probably attributed to the reduced toxic properties of the nanoparticles. Then, the variations of heart, liver, lung and kidney indexes for all tested drugs are negligible in contrast to saline.

It is widely proven that nephrotoxicity, hepatotoxicity and neurotoxicity are the key factors influencing the toxicities of platinum drugs. The drug accumulation in liver and kidney were also detected. Results in Figure 5B reveal that the BSA nanoparticles have rather different distribution in liver and kidney with the free naproxen platinum(IV) compounds. The platination levels of **1-NPs** and **2-NPs** in liver are higher than free compounds **1** and **2** as well as oxaliplatin. Meanwhile, oxaliplatin accumulates at a remarkably higher level in the kidney than naproxen platinum(IV) complexes and nanoparticles. The different distributions manifest the different

metabolic pathways of the BSA NPs with platinum(II) drugs in vivo, which would further influence toxicities. Subsequently, the nephrotoxicity and hepatotoxicity were detected by evaluating aspartate aminotransferase (AST), alanine aminotransferase (ALT), blood urea nitrogen (BUN) and creatinine (CRE) levels in serum. Noteworthy, oxaliplatin causes significant enhanced ALT and AST levels in comparison with saline indicating its impacts on liver function (Figure S5), although no obvious histological differences or liver index change are observed. In contrast, nanoparticles **1-NPS**, **2-NPS** display no significant effects on ALT and AST levels in serum. Then, free complexes **1** and **2** cause relatively high AST levels in comparison with **1-NPS** and **2-NPS** indicating that the BSA nanoparticles of naproxen platinum(IV) complexes exhibit lower liver toxicity than the free compounds as well as oxaliplatin. As for

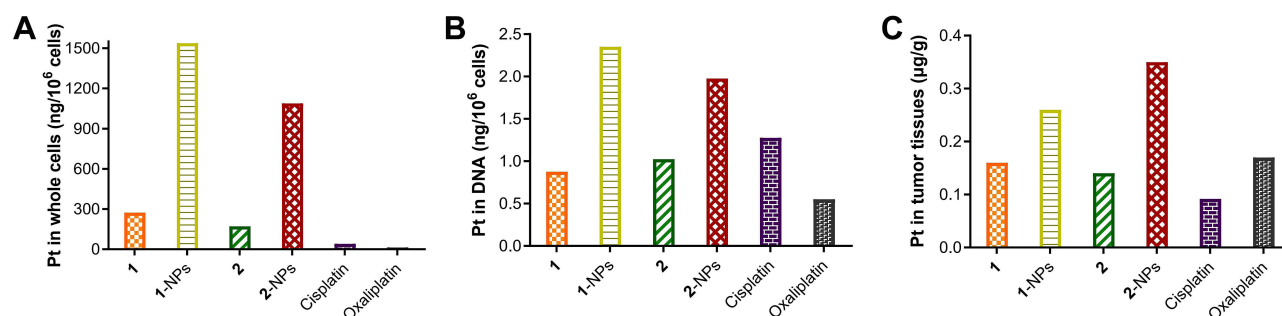


Figure 6 Accumulation of nanoparticles **1-NPs**, **2-NPs** and complexes **1**, **2** in (A) CT-26 cells, (B) DNA after treatment of 10 h at concentration of 100 μM in vitro and in (C) tumor tissues in vivo.

Abbreviation: NPs, nanoparticles.

CRE and BUN, there are no obvious differences observed in the tested groups with the saline group. Summarily, BSA NPs of naproxen platinum(IV) hybrids display reduced toxic effects in comparison with platinum(II) drugs in vivo, especially to the spleen and liver.

Tumor Target Properties

Tumor target properties of chemotherapeutic drugs influence their antitumor activities and toxicities dramatically. To evaluate the tumor target properties of nanoparticles, the accumulation of **1-NPs** and **2-NPs** in CT-26 cells in vitro and in tumor tissues in vivo were tested by ICP-MS to compare with compounds **1**, **2**, cisplatin, oxaliplatin. Moreover, platinum drugs exert antitumor activities mainly through coordination to DNA via the platinum fragment. Therefore, the platinum accumulation in DNA of CT-26 cells was also evaluated. Results in Figure 6 manifest that nanoparticles **1-NPs** and **2-NPs** significantly improve the drug accumulation both in CT-26 cells in vitro and in tumor tissues in vivo in comparison with free compounds **1** and **2** as well as the platinum(II) drugs cisplatin and oxaliplatin. Nanoparticles **1-NPs** and **2-NPs** lead to 5.6 and 6.4 folds enhanced uptake in whole cells in contrast to **1** and **2**, which are over 37.4 and 26.5 times higher than cisplatin and oxaliplatin (Figure 6A). Subsequently, **1-NPs** and **2-NPs** also lead to relatively higher platination level in DNA than free complexes **1**, **2** and platinum(II) drugs (Figure 6B). These results also supply direct evidence of the DNA damage caused by the platinum(II) complex released from the nanoparticles. Notably, the accumulation of **1-NPs** and **2-NPs** in tumor tissues are 1.6 and 2.5 folds higher than **1** and **2**, respectively (Figure 6C), which are also higher than cisplatin and oxaliplatin.

COX Inhibition

COX as the most important inflammatory factor has appeared as a promising target for reducing tumor-associated inflammation during the cancer chemotherapy treatment. Thereby, the COX inhibitory properties of naproxen platinum(IV) complexes **1** and **2**, which could be released by **1-NPs** and **2-NPs**, were tested with naproxen and celecoxib as reference drugs. The reduced systems of complexes **1** and **2** (compound-AsA) were also evaluated, which were prepared by the incubation of compound solutions in the presence of AsA (1 mM) for 48 h to allow the platinum(IV) totally converting to divalent form. Results in Figure 7 manifest that celecoxib could effectively inhibit

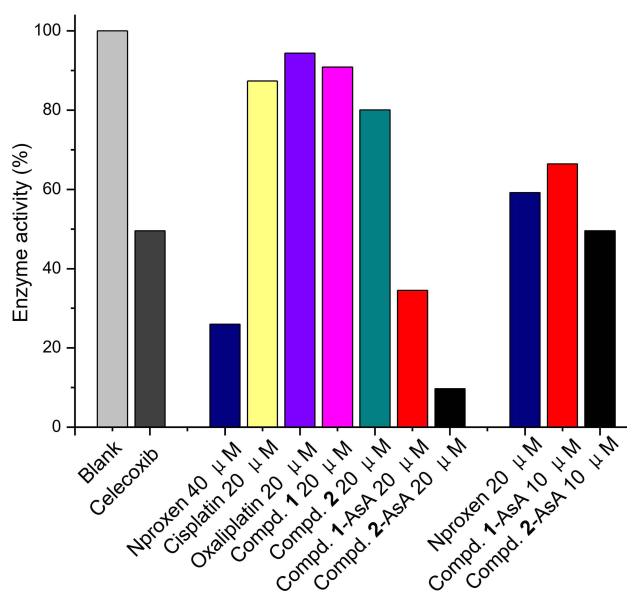


Figure 7 The inhibition of compounds **1**, **2**, naproxen and the reduced systems of complexes **1**, **2** (Compd-AsA) at different concentrations to COX-2 with celecoxib as reference drug.

Abbreviation: COX-2, cyclooxygenase-2.

the activity of COX-2 to 49.5% at concentration of 100 nM. Naproxen platinum(IV) complexes **1** and **2** exhibit negligible inhibitions of COX-2 in tetravalent form even at high concentration of 20 μ M, meanwhile their reduced systems compound **1**-AsA and compound **2**-AsA display prominent COX-2 inhibition of 65.5% and 90.3%, which are similar to naproxen (74.0%). Furthermore, the COX-2 inhibitory competence of the reduced naproxen platinum(IV) complexes is in a dose-dependent manner that low concentration of 5 μ M leads to significantly less effective COX-2 inhibition than that of high dosage of 20 μ M. Thereby, naproxen platinum(IV) complexes **1** and **2** as well as the nanoparticles **1**-NPs and **2**-NPs exert effective COX-2 inhibitory properties after reduction and release naproxen, which would decrease tumor-associated inflammation in tumor tissues and further increase antitumor activities.

iNOS Suppression and NO Production Inhibition

Nitric oxide (NO) was an important pro-inflammatory mediator in the regulation of inflammation progresses. Accumulated evidence has indicated that excessive production of NO in the tumor tissues could promote the development of inflammation and eventually lead to inflammatory tissue damage and further stimulate tumor growth and migration by inducing cell proliferation, reducing immune surveillance and promoting angiogenesis.^{51–55} Herein, LPS induced RAW 264.7 macrophages, which

were widely used as a model for studying the inflammatory responses,⁵⁶ were applied to evaluate the effects of naproxen platinum(IV) nanoparticles on the NO production and detect their potential effects on anti-inflammation. Results in Figure 8 indicate that naproxen platinum(IV) nanoparticles **1**-NPs and **2**-NPs could inhibit production of NO in RAW 264.7 macrophages, which are significantly more effective than naproxen, cisplatin and oxaliplatin. The conversion of complex **1** to nanoparticles **1**-NPs improve the activities effectively, while both complex **2** and **2**-NPs display satisfactory inhibition of NO production.

The inducible nitric oxide synthase (iNOS) as a calcium-independent isoform of nitric oxide synthase highly expresses in many tumors, and is the crucial enzyme responsible for the production of NO, which could lead to high levels of NO in the micromolar range. Moreover, there is emerging evidence showing that iNOS plays a central role in the process of tumor initiation and/or development.^{57,58} Thus, the expressions of iNOS in tumor tissues treated by **1**-NPs and free compound **1** were detected by immunohistochemistry. Results in Figure 9 reflect that oxaliplatin significantly upregulates iNOS in tumor tissues during its antitumor progress. Whereas, the expressions of iNOS in **1**-NPs treated group is significantly lower than that of oxaliplatin ($P<0.001$). Furthermore, **1**-NPs is also more effective than the free drug **1** ($P<0.001$) in iNOS inhibition, which is in line with the NO production inhibition in vitro.

Accordingly, the title naproxen platinum(IV) nanoparticle drugs show much potential in inhibiting NO production and iNOS expression, which would further influence tumor-associated inflammation.

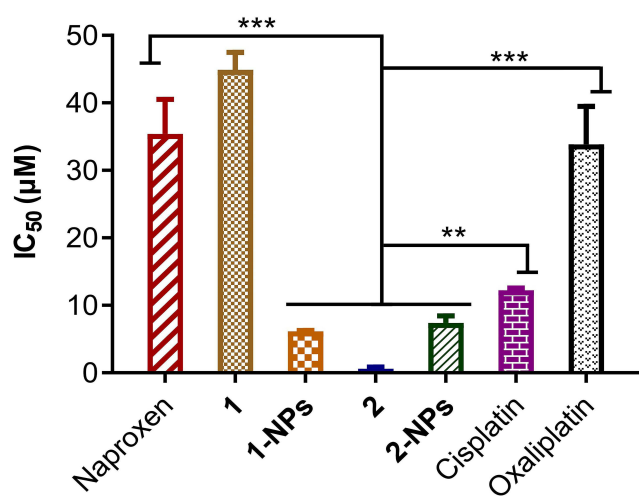


Figure 8 NO production inhibition of drugs **1**, **2**, **1**-NPs and **2**-NPs and reference drugs naproxen, cisplatin and oxaliplatin in LPS induced RAW 264.7.

Notes: Results are presented as the mean \pm SD. ** $P<0.01$, *** $P<0.001$.

Abbreviations: NPs, nanoparticles; LPS, lipopolysaccharide.

Detection of MMP-9

MMPs are widely found in inflammation and various tumor tissues, and display pivotal roles in influencing tumor growth, invasion and metastasis. Moreover, MMPs play a synergic function with COXs in promotion of inflammation in tumors. MMPs have been considered as potential diagnostic and prognostic biomarkers in many types and stages of cancer.^{59,60} Herein, the expressions of MMP-9 in tumor tissues of **1**-NPs, **1**, oxaliplatin and saline groups were immunohistochemically stained and given as Figure 10. MMP-9 overexpresses in saline group with a mean optical density of $16.3\pm1.9\%$. Meanwhile, **1** and **1**-NPs remarkably decrease the expression of MMP-9 to $1.9\pm0.5\%$ and $2.0\pm0.8\%$ respectively,

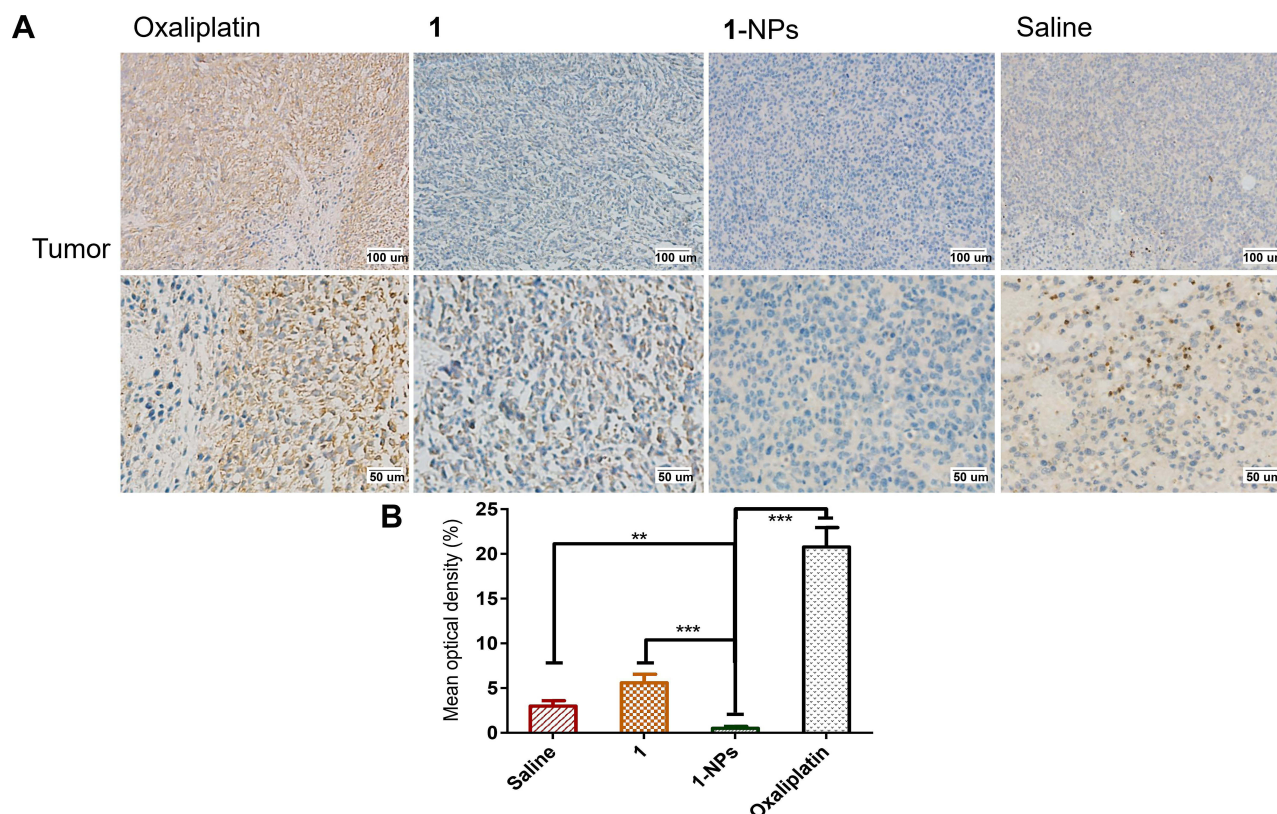


Figure 9 Photography of immunohistochemical staining of iNOS in tumor tissues from mice treated by **1**, **1-NPs**, oxaliplatin and saline. **(A)** Representative micrographs. **(B)** Quantified data of immunohistochemistry analysis.

Notes: Results are presented as the mean \pm SD (n=5). **P<0.01, ***P<0.001.

Abbreviations: iNOS, inducible nitric oxide synthase; NPs, nanoparticles.

which are significantly lower than oxaliplatin ($6.5 \pm 0.5\%$). The promising MMP-9 inhibitory potential of naproxen platinum(IV) nanoparticles would put much influence on reducing inflammation in tumor tissues and improving antitumor activities.

Conclusion

In conclusion, two BSA-encapsulated nanoparticles **1-NPs** and **2-NPs** of naproxen platinum(IV) complexes **1** and **2** were prepared and evaluated for antitumor activities in vitro and in vivo. The nanoparticles possessed relatively uniform size and good stability for 30 days in aqueous solution ($T=4^{\circ}\text{C}$, $\text{pH}=7.4$). Both of them display effective antitumor activities in vitro, and exhibit great potential in overcoming drug resistance of cisplatin with resistant factors ($\text{RF} < 0.68$) significantly lower than cisplatin ($\text{RF}=2.57$). Moreover, **1-NPs** and **2-NPs** accumulate at higher levels in whole tumor cells, DNA and tumor tissues than cisplatin and oxaliplatin as well as free platinum(IV) complexes **1** and **2**, and show low

toxicity with high tumor selective indexes of 3.0 and 3.9, which are significantly more promising than cisplatin ($\text{SI}=1.1$) and oxaliplatin ($\text{SI}=1.0$). Furthermore, these two nanoparticles play superior tumor growth suppression in vivo with high TGI of 64.7% and 58.9% in contrast to free compounds **1** and **2** (45.8%, 41.7%), which are comparable to that of cisplatin and oxaliplatin, and induce lower toxic influences than platinum(II) drugs especially to the spleen and liver. The naproxen platinum(IV) nanoparticles display antitumor activities by a symmetric mechanism (Figure 1) that nanoparticles target tumor cells and would release naproxen platinum-(IV) complexes **1** and **2** in cytoplasm. Then the platinum-(II) core would be released after reduction, and further conjunct with DNA to cause serious DNA damage. The further mechanism detections in vitro and in vivo disclose that the inflammation in tumor tissues could be reduced through the suppression of COX-2, iNOS, MMP-9 and the decrease of NO production, which would increase the sensitivity of tumor cells to platinum

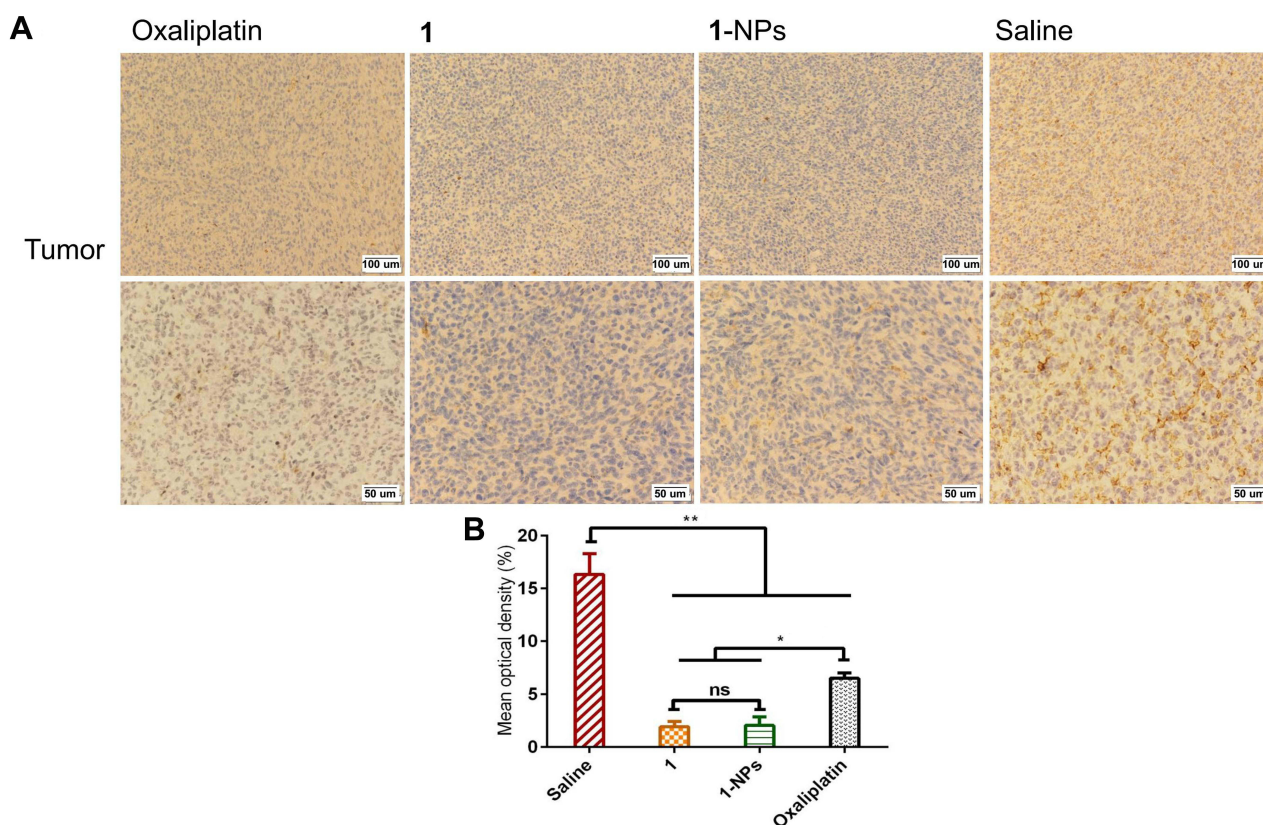


Figure 10 Photography of immunohistochemical staining of MMP-9 in tumor tissues from mice treated by **1**, **1-NPs** oxaliplatin and saline. **(A)** Representative micrographs. **(B)** Quantified data of immunohistochemistry analysis.

Notes: Results are presented as the mean \pm SD (n=5). * P <0.001, ** P <0.001, ns, no significant difference.

Abbreviations: MMP-9, metalloproteinase-9; NPs, nanoparticles.

drugs, prevent the development of cancer and enhance the antitumor activities of the nanoparticles. These encouraging results highlight the value of naproxen platinum(IV) nanoparticles **1-NPs** and **2-NPs** as antitumor agents, and further investigations based on such drugs are underway in our laboratory.

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Disclosure

Zhengping Wang is an employee of Liaocheng High-Tech Biotechnology Co., Ltd, Liaocheng, China. The authors report no other potential conflicts of interest in this work.

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