Open Access Full Text Article

ORIGINAL RESEARCH

LncRNA FOXD3-AS1 Promotes Tumorigenesis of Glioma via Targeting miR-128-3p/SZRD1 Axis

Zhang Li Ming Li Pengcheng Xia Lili Wang Zhiming Lu (p

Department of Clinical Laboratory, Shandong Provincial Hospital, Cheeloo College of Medicine, Shandong University, Jinan, 250012, People's Republic of China



Correspondence: Zhiming Lu Department of Clinical Laboratory, Shandong Provincial Hospital, Cheeloo College of Medicine, Shandong University, Jinan, Shandong, 250012, People's Republic of China Tel +86-13658608997 Email luzhiming@sdu.edu.cn **Background:** The aim of the current study was to investigate the roles of L RNA *FOXD3-AS1* (*FOXD3-AS1*) in the glioma progression, and its inderly, mechanical of competing endogenous RNA (ceRNA) network of *FOXD3-AS1* miR-128-3p/s/RD

Materials and Methods: The FOXD3-AS1 resistor and its prognostic relation were detected by bioinformatics tool. Next, gliop cell (HS683, 1, 51, T98G, and SNB-19) were used to verify the FOXD3-AS1 excession. Furth more me roles of the FOXD3-AS1 /miR-128-3p/SZRD1 axis on the glice a devolpment in the o and in vivo were examined. **Results:** Bioinformatics analysis showed that *YD3-AS1* was upregulated in the glioma and linked with poor prognosis. Constantly, FOXD3-, V level was overexpressed in the glioma cell lines (HS683 and U21). Subsequently, we verified that silencing of FOXD3-AS1 (si-FOXD3-AS1) restrained the ell proliferation invasion, and tumor growth in vivo, and induced G0/G1 arrest, and promoted as tosis. Further study also stated that FOXD3-AS1 interacted with miR-128-3p and was the tangenee of miR-128-3p. Moreover, overexpression of miRration and metastasis of glioma, and reduced the SZRD1 level. 128-3p restrained e cell y nat miR-128-3p inhibitor could reverse the suppressive impact of si-Rescue accev illustr FOX 5-ASI n the genma progression. Similarly, SZRD1 overexpression could neutralize the hiR-128-, mimic on glioma progression. dences of

Conjugon: *FORE -AS1* promoted the tumorigenesis of glioma, and exerted its function to module *SZRD1* by targeting miR-128-3p.

Keyword, zlioma, LncRNA FOXD3-AS1, MiR-128-3p, SZRD1, proliferation, invasion

Introduction

As a dominating primary intracranial tumor, glioma is featured by high aggression, poor prognosis, and high lethality.¹ On the basis of the grading criteria, glioma is classified as low-grade glioma (LGG, I–II) and high-grade glioma (HGG, III–IV).² Of these, patients with LGG have longer overall survival in contrast to HGG.³ Glioblastomas (GBM), refers to the most aggressive type (grade IV), with a proportion of 65% in brain tumors.⁴ Despite advances in therapeutic treatment, most patients achieve unsatisfactory efficacy and have poor prognosis, especially with GBM.⁵ Like other tumors, the progression of glioma is a complex procedure involving changes of pathology, genes, and pathways.⁶ Currently, more attention has been paid to the molecular mechanism of glioma. Thus, elucidating the molecular mechanism is an urgent need for glioma therapy.

Accumulating evidence has stated that LncRNAs exert crucial roles on the progression of various cancers including glioma.^{8–10} Moreover, a previous study has reported

© 121 Li et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/terms.ph you hereby accept the freems. Non-commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial uses of this work, please see paragraphs 4.2 and 5 of our Terms (http://www.dovepress.com/terms.php). that LncRNAs as competing endogenous RNAs (ceRNAs) by sponging miRNAs to directly target the mRNAs is the key mechanism for regulation of tumor growth and metastasis.¹¹ In terms of glioma, recent studies have verified this point.^{12,13} Those findings suggested that the LncRNA as ceRNA may be the underlying mechanism to glioma development.

LncRNA FOXD3-AS1 (FOXD3-AS1) is highly expressed and may exert the crucial function on cancer progression through the ceRNA network, including lung cancer, maligadenocarcinoma.14-16 melanoma. and colon nant Consistently, FOXD3-AS1 is upregulated in glioma, which has been demonstrated in clinical specimen detection.¹⁷ However, no reports of FOXD3-AS1 as ceRNA regulating glioma development are available. Microarray studies have been conducted to screen the aberrantly expressed miRNAs in glioma and found that miR-128-3p is observably declined.^{18,19} Moreover, miR-128-3p is the target gene of FOXD3-AS1, which has been elucidated in cervical cancer.²⁰ Another study has also demonstrated that miR-128-3p suppresses the proliferation and metastasis of glioma cells via binding to 3'-UTR of SZRD1, thereby influencing the glioma tumorigenicity.²¹ Thus, we speculated that FOXD3-AS1 regulating miR-128-3p/SZRD1 axis may be an underlying mechanism in glioma progression. In the current study, first verified that functioned as ceRNA to regulate the gliom progression. In addition, miR-128-3p/SZRD1 axis a novel target of FOXD3-AS1 in regulation of gliomatic .evelo nent may supply theoretical foundation for further study.

Materials and Metho Cell Culture

Glioma cell lines (HS683, 6251, T98G, and NB-19) and human normal astronue (HF1) were purchased from BeNa Culture Collection Grajing, Coma), and cultured in DMEM (P1, 15, 21) superstant with 10% FBS (Solarbio, Dajing, Coma), and c00 U/mL penicillin and 100 mg/mL superstant collumbio, Dalian, China).

Cell Transfection

After reaching 80% confluence, HS683 and U251 cells were transfected with siRNA-*FOXD3-AS1* (si-*FOXD3-AS1*) and negative control (si-NC) (Genechem, Shanghai, China) respectively according to the Lipofectamine 2000 protocol. Similarly, pcDNA-*SZRD1* (OE-*SZRD1*) and empty vector was synthesized by Genechem (Shanghai, China). The miR-128-3p inhibitor, miR-128-3p mimic and its negative control were obtained from RiboBio (Guangzhou, China). Cells in

Dovepress

the control group were untreated with anything. After being cultured for 48 h, cells were harvested.

Quantitative Real-Time PCR Assay

Total RNA was extracted by TRIzol reagent (CWBio, Beijing, China). The first strand cDNA for FOXD3-AS1 and SZRD1 was synthesized by cDNA Synthesis kit (Accurate Biology, Changsha, China). Next, PCR reaction was performed on CFX connect detection system (Bio-Rad, Hercules, CA, USA). GAPDH was used as the internal control for quantification of FOXD3-AS1 and SZRD1 levels 6 for miR-128-3p level, respectively. Primers were disted as the following: FOXD3-AS1 forward, 5'- ACCAG. SGAAGGA CACGA-3', reverse, 5'-AGAAGCACC CTGTC ATCC-2 d reverse, 5'-3p forward, 5'-TCACAC GAACC GTC-CAGTGCGTGTCGTGC GT, SZRD1 forward, 5'-ATG AGGAGGTCGC AGAO and verse, 5'-GGAAGG CTATCGTCC . ATC-3'; GA D. forward, 5'-AGAAGG CTGGGGCTCATT1 3', reverse, 5'-AGGGGCCATCCA CA-3', and reverse, GC 5'- GAACGCTTCACGA GCGTG-3'. The relative expression was quantified by AT $2^{-\Delta\Delta}$ method.

Veston Blotting Assay

Protein was extracted from HS683 and U251 cells using IPA lysis buffer containing protease inhibitor (Bosterbio, Wuhan, China). Protein was separated and then transferred onto PVDF membranes. After blockage with 5% skimmed milk, the membranes were incubated with anti-SZRD1 (1:3000, ab95957, Abcam, MA, USA), and anti-GAPDH (1:5000, 60004-1-Ig, Proteintech), followed by HRPconjugated secondary antibody (1:5000, ZB-2301, ZSGB). The protein blots were visualized using the enhanced chemiluminescence (ECL) kit and captured using chemiluminescent (Tanon, imaging system Shanghai, China). GAPDH was used as internal control.

3-(4, 5-Dimethylthiazol-2-YI)-2,5-Diphenyltetrazolium Bromide (MTT) Assay

After transfection, HS683 and U251 cells were cultured $(2 \times 10^3 \text{ cells/well})$ for 24 h, 48 h, 72 h, and 96 h. Afterwards, MTT with concentration of 5 mg/mL was added for incubation of four hours. The absorbance at 490 nm was detected by spectrophotometer (Multiskan Sky, Thermo Fisher, Waltham, MA, USA).

Flow Cytometry Assay

For analysis of cell apoptosis, HS683 and U251 cells $(1 \times 10^6 \text{ cells/mL})$ were obtained after 48 h of transfection, and washed with cool phosphate buffered saline (PBS) three times. Cell apoptosis was detected by flow cytometer (NovoCyte, Agilent, Palo Alto, CA, USA) with annexin V-propidium iodide (PI) kit (Meilunbio, Dalian, China). To detect cell cycle, collected cells were fixed in the 75% ethanol. Having been washed, the cells were stained with PI, followed by analysis with flow cytometer (NovoCyte, Agilent, Palo Alto, CA, USA).

Transwell Assay

HS683 and U251 cells $(1 \times 10^5$ cells/mL) were placed in a transwell chamber (Corning, Lowell, MA, USA), and the culture medium was in the lower chamber. Subsequently, the invasion and migration cells were fixed with 4% paraformaldehyde and then dyed with 0.1% crystal violet. The number of cells was quantified by microscope (CKX53, Olympus, Tokyo, Japan).

Dual-Luciferase Reporter Assay

The wild type (WT) and mutant (MU) fragments of *FOXD3-AS1* were cloned into the pmirGLO vector to establish the recombinant vectors (pmirGLO-*FOXD3-AS1*-WT and pmirGLO-*FOXD3-AS1*-MU). Classifiently, V and MU fragments of *SZRD1* were cloted into the above mentioned vector. Subsequently, 8 vg/m oppirol Co and 16 µg/mL miR-128-3p minut or mimit NC were co-transfected into U251 cells Are stransfection for 48 h, luciferase activities were detected.

RNA Immun precipitation (RIP) Assay

RIP assay was concreted as a reviously described.²² Briefly, trasfected HSo 2 and U251 cells were lysed and inclubated with magnetic beads conjugated with human as A gonaute2 (Ago2) antibody, negative control (anti-IgG) or input Control (Millipore) for six hours. Subsequently, in nunoprecipitated RNA was isolated and detected by qRT-PCR.

Tumorigenesis Assay in vivo

A total of 72 nude mice (five weeks old) were obtained from Hangzhou Ziyuan Laboratory nimal Science and Technology Co. Ltd (Hangzhou, China). The mice were divided into six groups, which were inoculated with HS683 and U251 cells $(1 \times 10^6 \text{ cells/mL})$ transfected with control, sh-*FOXD3-AS1*, sh-NC; control, miR-128-3p mimic, and mimics-NC, respectively (12 mice per group). The nude mice in the control group were injected with glioma cells untreated. The tumor volume was assessed from one week to six weeks after tumor inoculation (once every week). Subsequently, mice were sacrificed by cervical dislocation and the tumor tissues were collected for the further experiments.

Bioinformatics Analysis

The FOXD3-AS1 expression offles GSE147352) in glioma patients were rieved fro. the Gene Expression Omnibus <u>GEO</u> database. The dataset included 85 GBM amples, 18 GG amples and 15 normal samples. Sine excession was estimated using fragments per dloba f transport per million mapped reads (FPL) value and the Boxplot was visualized using the ggp package in R Language. The differentimpression by veen normal and tumor groups was halyzed by R package Deseq2 on basis of following riteria: fold change >2 and Padj <0.05. Samples were ected ac ording to inclusion criteria as following: (1) patients were diagnosed as glioma; (2) data were availas on expression of FOXD3-AS1; and (3) data of samples on FOXD3-AS1 expression had corresponding control group. Samples were excluded according to following exclusion criteria: (1) FOXD3-AS1 expression was not included; (2) control group was not included; and (3) cell samples were used.

The Cancer Genome Atlas (TCGA) data were applied to analyze the relationship between FOXD3-AS1 expression and patient's prognosis. 638 glioma patients with FOXD3-AS1 expression and survival information were included into current analysis. The survival analysis was performed using R language with "Survival" package and survival curves were plotted using "Survminer" package. Samples were collected according to inclusion criteria as following: (1) patients were diagnosed as glioma including LGG and GBM; (2) data were available on expression of FOXD3-AS1; and (3) data of samples on FOXD3-AS1 expression had corresponding survival information. Samples were excluded according to following exclusion criteria: (1) survival information was not included; and (2) patients were diagnosed as other types of cancer except glioma. The target bites between miRNA and lncRNA (miRNA and mRNA) were predicted by StarBase v3.0.

Statistical Analysis

Data were presented as the means \pm standard deviation (SD) and analyzed by SPSS 20.0 statistical software. The differences in the multiple groups were compared by oneway ANOVA followed by LSD test, respectively. P<0.05 was termed as statistically significant.

Results

FOXD3-AS1 Was Upregulated in Glioma and Correlated with Poor Prognosis

To investigate the function of FOXD3-AS1 in glioma, we downloaded the FOXD3-AS1 expression profiles (GSE147352) in glioma patients from GEO database. As displayed in Figure 1A and B, the FOXD3-AS1 expression in the GBM and LGG tissues (tumor group) was higher than that in normal tissues (normal group) (P<0.01) (Figure 1A and

GSE147352-GBM

B). Consistently, data from TCGA database showed that FOXD3-AS1 in the tumor group including GBM and LGG was higher than that in the normal group (P < 0.01) (Figure 1C). In addition, it also found that FOXD3-AS1 expression in glioma patients including GBM and LGG was negatively correlated with overall survival (P < 0.01) (Figure 1D). Similarly, FOXD3-AS1 expression in glioma cell lines (HS683 and U251) was higher than that of HEB (Figure 1E) (P < 0.01), indicating that HS683 and U251 cell lines can be used for further study.





Figure I FOXD3-AS1 was upregulated in glioma and correlated with poor prognosis. (A and B) Difference in FOXD3-AS1 expression from GEO GSE147352 dataset between the tumor group (GBM/LGG) and normal group; (C) difference in FOXD3-AS1 expression from TCGA database between the tumor group (GBM and LGG) and normal group; (D) Kaplan-Meier method was utilized to analyze the overall survival of glioma patients based on FOXD3-AS1 expression from TCGA database; (E) the FOXD3-AS1 expression in the glioma cell lines (HS683, U251, T98G, and SNB-19); data were shown as the mean ±SD of three independent experiments. *vs HEB cell line, P<0.01. Abbreviations: FOXD3-AS1, LncRNA FOXD3-AS1; GBM, glioblastoma multiforme; LGG, low-grade gliomas; HEB, human normal astrocyte.

of *FOXD3-AS1* and miR-128-3p in the anti-Ago2 group compared with anti-IgG group (P<0.01) (Figure 2B and C). Luciferase reporter assay stated that overexpression of miR-128-3p significantly reduced the luciferase activity of *FOXD3-AS1*-WT (P<0.01) (Figure 2D). Thus, the aforementioned findings indicated that *FOXD3-AS1* could directly regulate the miR-128-3p.

Silencing of FOXD3-AS1 Suppressed Proliferation and Invasion of Glioma Cells via Modulating miR-128-3p

To examine the roles of FOXD3-AS1 on glioma, silencing of FOXD3-AS1 (si-FOXD3-AS1) was applied in HS683 and U251 cells. qRT-PCR results revealed FOXD3-AS1 was notably decreased, but miR-128-3p expression was prominently increased in the si-FOXD3-AS1 group, in comparison with si-NC and control groups (P < 0.01) (Figure 3A). Following, MTT results showed that cell viability in the si-FOXD3-AS1 group was notably reduced after 48 h, however, inhibition of miR-128-3p elevated the cell viability after 48 h compared with si-NC and control groups (P<0.05) (Figure 3B and C). Similarly, si-FOXD3-AS1 suppressed the invasion abilities, while inhibition of mi 3p rescued the roles of si-FOXD3-AS1 on invasion of H 683 and U251 cells (P<0.01) (Figure 3D–G). No-conspice difference was observed among the si-F XD3-SI+mi 128-3p inhibitor, si-NC, and Control groups (P> (05)

Knockdown of FOXD3-AS1 Influenced Cell Cycle and Promoted Apoptosis of Glioma Cells via Modulating miR-128-3p

As shown in Figure 4A–D, the cell proportion at G0/G1 phase was prominently increased in the si-FOXD3-AS1 group, however miR-128-3p inhibitor reduced the cell proportion at G0/G1 phase (P<0.01). The cell proportion at different phases in si-FOXD3-AS1+miR-128-3p inhibitor group exhibited no obvious difference in contrast to control and si-NC groups (P>0.05). In the aspect of cell apoptosis, we scover that apoptosis rate was notably increased the si- $FO\lambda$ 3-AS1 group, whereas it was decreated in miR-12-3p inhibitor group in contrast to control and si VC coups (P < 0.01). Moreover, miR-1, 3p is abitor can reverse the influences of si-OXD3 I on propriority of P < 0.01) (Figure 4 Collecti v ne aforementioned results concluded that FOXD3-AS1 influenced cell cycle and d apopto. of glioma cells via modulating iR-128-3p.

Suncing of FOXD3-AS1 Inhibited Tymorigenesis in vivo via Modulating mix-128-3p

To validate the effect of *FOXD3-AS1* on glioma in vitro, xenograft tumor model was conducted. First, the



Figure 2 *FOXD3-AS1* sponged miR-128-3p in glioma. (**A**) Complementary binding sites between *FOXD3-AS1* and miR-128-3p; (**B** and **C**) RIP assay was performed to validate the prediction in HS683 and U251 cells; *vs anti-IgG group, P<0.01; (**D**) relative luciferase activity in U251 cells among the groups; **vs control or mimics-NC group, P<0.01. Data were shown as the mean ±SD of three independent experiments.

Abbreviations: FOXD3-AS1, LncRNA FOXD3-AS1; RIP, RNA immunoprecipitation.



Figure 3 Silencing of FOXD3-AS1 suppressed proliferation at a payasion to the poly a modulating miR-128-3p. (**A**) Transfection efficacy in HS683 cells was detected by qRT-PCR; (**B** and **C**) cell viability in HS683 and U251 cells was explicitly by MTT assay, (**D**–**G**) the invasion ability in HS683 and U251 cells (Scale bar=20 μ m). *, ** vs control or si-NC group, *P*<0.05, *P*<0.01; data were shown the mean ±5, wf three independent experiments. **Abbreviation**: FOXD3-AS1, LncRNA FOXD3-AS

transfection efficacy was shown in Figure A, *FOXD3-AS1* expression was a creased while miR-128-3p level increased after transfected with sh-*FOXD3-AS1* (P<0.01). Subsequently we found that the transmission size and weight of the sh-*FOXD3-AS1* aroun was remarkably lessened in contrast to control and sh-NC groups (P<0.05) (Figure 5B–G).

MiR-128-3p Targeted the 3'-UTR of SZRD1

As shown in Figure 6A, the 3'-UTR of *SZRD1* contains a targeting site for the region of miR-128-3p. Luciferase reporter assays showed that the relative luciferase activity of *SZRD1*-3'-UTR-WT was remarkably decreased in overexpressed miR-128-3p group (Figure 6B). Western blotting indicated that si-*FOXD3-AS1* suppressed the *SZRD1* expression, while miR-128-3p inhibitor elevated the *SZRD1* level in comparison with control and si-NC groups (*P*<0.01). Surprisingly, *SZRD1* level in the si-*FOXD3-AS1*+miR-128-3p inhibitor group did not differ from control and si-NC groups (*P*>0.05) (Figure 6C–F). Collectively, *SZRD1* is the target gene of miR-128-3p regulated by *FOXD3-AS1*.

Overexpressed miR-128-3p Suppressed Proliferation and Invasion of Glioma Cells via Modulating SZRD1

MiR-128-3p mimic and OE-*SZRD1* were co-transfected into HS683 and U251 cells, and it turned out that miR-128-3p expression was remarkably elevated in the miR-128-3p



Figure 4 Silencing of FOXD3-AS1 influences of cycle and provide apoptosis of glioma cells via modulating miR-128-3p. (A–D) Cell cycle of HS683 and U251 cells among the different groups was measured by flow cyclemetry. (E–H) The poptosis rate of HS683 and U251 cells was measured by flow cytometry. *vs control or si-NC group, P<0.01. Data were shown as the mean aSD of the hindependent experiments. Abbreviation: FOXD3-AS1, LncPue, FOXD3-AS1.

as decreased (P<0.01). Besides, RDI mimic group while influer of miR-128-3p in the OE-SZRD1 rsed aspect of ,ZRD1 evel (7A). MTT results showed that certiabilit wered after transfection with miR-128-3p mine whereas increased after transfection with OE-SZDR1 (P<0.6 There was no distinct difference in the OD₄₉₀ value among the miR-128-3p mimic+OE-SZRD1, mimics-NC and control groups (P>0.05) (Figure 7B and C). Consistently, the number of invasion cells was reduced in miR-128-3p mimic group compared with mimics-NC and control groups (P<0.01). Nonetheless, transfection with OE-SZRD1 enhanced the invasion ability of miR-128-3p mimic (P<0.01) (Figure 7D-G).

Overexpression of miR-128-3p Inhibited Tumorigenesis in vivo via Modulating SZRD1

As shown in Figure 8A, qRT-PCR results validated transfection of miR-128-3p mimic (P<0.01). In vivo, it indicated that tumor size and weight of miR-128-3p mimic group was conspicuously decreased compared with control and mimics-NC groups (P<0.01) (Figure 8B–G). Importantly, the schematic for the regulatory relationship among *FOXD3-AS1*, miR-128-3p, and *SZRD1* in glioma was shown in Figure 9. Those finding proved that overexpression of miR-128-3p restrained tumorigenesis in vivo via modulating *SZRD1*.



Figure 6 MiR-128 $(\mathbf{p}, \mathsf{targetec}, \mathsf{p}, \mathsf{s}', \mathsf{rargetec}, \mathsf{p}, \mathsf{s}', \mathsf{s}', \mathsf{rargetec}, \mathsf{s}', \mathsf{s}', \mathsf{s}', \mathsf{rargetec}, \mathsf{s}', \mathsf{$

Discussion

To our knowledge, dysregulation of LncRNAs frequently occurs in the various cancers, which is deemed as a master regulator for disease progression.²³ Here, our study showed that *FOXD3-AS1* was upregulated in glioma and correlated with poor prognosis using bioinformatic analysis. In addition, *FOXD3-AS1* level in the glioma cell lines was consistent with bioinformatic analysis. Furthermore, knockdown of *FOXD3-AS1* suppressed the tumorigenesis

of glioma via targeting miR-128-3p/*SZRD1* axis, as evidenced by cell viability, invasive ability, apoptosis rate, cell cycle, and tumorigenesis in vivo.

FOXD3-AS1, is a 963-bp lncRNA, which is located in the chromosome 1p31.3 upstream of *FOXD3* promoter,²⁴ indicating that its function is closely associated with adjacent protein-coding transcripts.²⁵ Beyond that, *FOXD3-AS1* acts as independent prognostic indicator for prediction of neuroblastoma progression, which has been proved.²⁴



Figure 7 Overexpressed mik-128-sp suppressed proliferation and it is not of grama cells via modulating SZRDT. (A) Transfection efficacy in HS683 cells was shown; (B and C) influence of miR-128-3p/SZRDT axis on cell viability as tested. MTT assa, (D–G) effect of miR-128-3p/SZRDT axis on invasion ability (Scale bar=20 μ m); *, **vs control or mimics-NC group, P<0.05, P<0.01. Data was shown as the test of three independent experiments.



Figure 8 Overexpression of miR-128-3p inhibited tumorigenesis in vivo via modulating SZRD1. (A) Transfection efficacy was shown; (B–G) the tumor size and weight were shown. *, **vs control or mimics-NC group, P<0.05, P<0.01. Data were shown as the mean ±SD of six mice.



Figure 9 Schematic for the regulatory relationship among FOXD3-AS1, miR-128-3p, and SZRD1 in glioma.

Similarly, dysregulation of FOXD3-AS1 expression is observed in numerous cancers.14,26,27 In terms of glioma, Chen et al¹⁷ have clarified that si-FOXD3-AS1 inhibited the tumor development as demonstrated by cell proliferation, migration, and invasion. Consistently, our study found that FOXD3-AS1 was highly expressed in glion and si-FOXD3-AS1 restrained the tumor growth in viv and in vitro. In addition, glioma patients with overexpressed FOXD3-AS1 had lower overall surviver, indi ting FOXD3-AS1 was closely associated with por pro Furthermore, Wang et al²⁸ have demonstrued that FOXD3-AS1 may be functioning ceRNA to kert the pivotal role in the regulation can progression. To date, knowledge about *XD3-AS1* NA network involved in glioma prograssion remains vague.

MiR-128-3p was in cally accovered in GBM,²⁹ which attracts more attention at put ent because it acts as a tumor early liagnos dicator.^{30,31} A similar suppressor ar study has stified the miR-128-3p can increase the sensirapy of colorectal cancer.32 In the gliotivity of chem. mas, Bendahou al³³ have revealed that patients with high miR-128-3p expression have longer overall survival than low miR-128-3p using bioinformatic analysis. A similar study has demonstrated that miR-128 may be effective therapy for disruption of tumor-relevant phenotypes and tumor ingrowth.³⁴ Furthermore, a previous study has reported that miR-128-3p restrained the cell proliferation and accelerated apoptosis.18 Aforementioned findings were aligned with our results. Our study showed that overexpression of miR-128-3p restrained proliferation and tumorigeness in viv and accelerated apoptosis of glioma cells. contrary, hibition of miR-128-3p accelerated he glioma development. In regard to the interaction bet en miRNA and LncRNAs, previous studies have been ported. F example, a previous study reported by s uncovered that miR-128-3p recuses the Fu et a on of LncRNA PVT1 on glioma tumorigenesis. reg nother study also showed that inhibition of miR-128-3p liminates the influence of si-LINC00346 on metastasis of ioma.²¹ However, the relationship between miR-128-3p and FOXD3-AS1 in glioma has not been reported. Accordingly, our study firstly disclosed that miR-128-3p inhibitor reversed the effects of si-FOXD3-AS1 on glioma progression. Taken together, FOXD3-AS1 may be involved in the modulation of glioma development via sponging miR-128-3p.

In the current study, we also found *SZRD1* is the targeted gene of miR-128-3p. *SZRD1*, a highly conserved protein, is first found in cervical cancer, which functions as a tumor suppressor.³⁵ Conversely, *SZRD1* exerts the push roles on tumor development in the oligodendrogliomas, lymphoma, and glioma,^{21,36,37} which are in accordance with our results that overexpression of *SZRD1* facilitated the tumorigenesis. Moreover, our study displayed that miR-128-3p mimic reduced the *SZRD1* expression, whereas overexpression of *SZRD1* reversed the function of miR-128-3p mimic on inhibition of glioma development, indicating that miR-128-3p was bound with *SZRD1*. This results were in line with a previous study, which revealed the interaction between

miR-128-3p and *SZRD1*.²¹ The aforementioned findings manifested that miR-128-3p modulated the proliferation, and invasion of glioma cells via targeting *SZRD1*.

There were some limitations in the current study. First, the overall survival of mice among different groups was not examined in this study. In future, we will explore the effects of compounds on survival of mice including sh-*FOXD3-AS1* and miR-128-3p. In addition, whether over-expression of *FOXD3-AS1* influences on behaviors of glioma cell lines is unclear. Importantly, comparison of the growth rate and invasion among the glioma cell lines will be a potential research direction.

Conclusions

FOXD3-AS1 promoted proliferation, and invasion of glioma cells via regulation of the miR-128-3p/*SZRD1* axis. Those findings clarified that the *FOXD3-AS1*/miR-128-3p/*SZRD1* network may be an underlying mechanism for glioma development.

Ethics Approval and Informed Consent

The study was approved by the ethical committee of Shandong Provincial Hospital, Cheeloo College of Medicine, Shandong University (No. 2020 400) and of experiments conformed to Guide for the Care and Use of Laboratory Animals.

Author Contribution

All authors made a significant contraction to the work reported, whether the vis in the conception, study design, execution, acquisition of dua, analysis and interpretation, or in all these areas, nook part in drafting, revising or critically eviewed, the aciely gave final approval of the version to be prolished; have agreed on the journal to which the noice has been submitted; and agree to be accountable recall aspects of the work.

Funding

The study was funded by Major Science and Technology Innovation Project of Shandong Province (2020SFXGFY03-2).

Disclosure

The authors report no conflicts of interest in this work.

References

- Morgan LL. The epidemiology of glioma in adults: a "state of the science" review. *Neuro Oncol.* 2015;17(4):623–624. doi:10.1093/ neuonc/nou358
- Louis DN, Perry A, Reifenberger G, et al. The 2016 World Health Organization classification of tumors of the central nervous system: a summary. *Acta Neuropathol*. 2016;131(6):803–820. doi:10.1007/ s00401-016-1545-1
- Rich JN, Bigner DD. Development of novel targeted therapies in the treatment of malignant glioma. *Nat Rev Drug Discov.* 2004;3 (5):430–446. doi:10.1038/nrd1380
- Hanif F, Muzaffar K, Perveen K, Malhi SM, Simjee SU. Glioblastoma multiforme: a review of its epidemiology and pathogenesis through clinical presentation and reatment. *Asian Pac J Cancer Prev.* 2017;18(1):3–9. doi:10.22034/Art/CP.2017.18.1.3
- Mohtashami E, Shafaei-Bajesta, J, Mollazadeh Leet al. The current state of potential therapeuti modal as for gliobla oma multiforme: a clinical review. *Cur Drug Vetab.* 2120;21(8):564–578. doi:10.2174/13892002.46662007141016
- Pan W, Li G, Yang, Miao Jacevealing by potential pathogenesis of glioma by utilizing reglinera associated protein-protein interaction network. *Phylol Onco. Res.* 201 21(2):455–462. doi:10.1007/ s12253-01-0848-9
- 7. Rajeskar, Pan Banik P, et Insights into molecular therapy of glioma: current hallenges and next generation blueprint. *Acta acol Sin.* 20, 138(5):591–613. doi:10.1038/aps.2016.167
- Arun G, Diermeier SL, Spector DL. Therapeutic targeting of long non-coding DNAs in cancer. *Trends Mol Med.* 2018;24(3):257–277. doi:10.1016 nolmed.2018.01.001
 - Chi Y, Wang D, Wang J, Yu W, Yang J. Long non-coding RNA in the hogener of cancers. *Cells*. 2019;8(9):1015. doi:10.3390/cells8091015
- 10. Peng 2, Liu C, Wu M. New insights into long noncoding RNAs and their roles in glioma. *Mol Cancer*. 2018;17(1):61. doi:10.1186/ 512943-018-0812-2
- Qi X, Zhang DH, Wu N, Xiao JH, Wang X, Ma W. ceRNA in cancer: possible functions and clinical implications. *J Med Genet*. 2015;52 (10):710–718. doi:10.1136/jmedgenet-2015-103334
- Xin J, Zhao YH, Zhang XY, Tian LQ. LncRNA NFIA-AS2 promotes glioma progression through modulating the miR-655-3p/ZFX axis. *Hum Cell*. 2020;33(4):1273–1280. doi:10.1007/s13577-020-00408-9
- 13. Zhou XY, Liu H, Ding ZB, Xi HP, Wang GW. IncRNA SNHG16 promotes glioma tumorigenicity through miR-373/EGFR axis by activating PI3K/AKT pathway. *Genomics*. 2020;112(1):1021–1029. doi:10.1016/j.ygeno.2019.06.017
- Zeng ZL, Zhu HK, He LF, et al. Highly expressed lncRNA FOXD3-AS1 promotes non-small cell lung cancer progression via regulating miR-127-3p/mediator complex subunit 28 axis. *Eur Rev Med Pharmacol Sci.* 2020;24(5):2525–2538. doi:10.26355/eurrev_202003_20520
- Chen X, Gao J, Yu Y, Zhao Z, Pan Y. LncRNA FOXD3-AS1 promotes proliferation, invasion and migration of cutaneous malignant melanoma via regulating miR-325/MAP3K2. *Biomed Pharmacother*. 2019;120:109438. doi:10.1016/j.biopha.2019.109438
- 16. Wu Q, Shi M, Meng W, Wang Y, Hui P, Ma J. Long noncoding RNA FOXD3-AS1 promotes colon adenocarcinoma progression and functions as a competing endogenous RNA to regulate SIRT1 by sponging miR-135a-5p. *J Cell Physiol.* 2019;234(12):21889–21902. doi:10.1002/jcp.28752
- Chen ZH, Hu HK, Zhang CR, et al. Down-regulation of long non-coding RNA FOXD3 antisense RNA 1 (FOXD3-AS1) inhibits cell proliferation, migration, and invasion in malignant glioma cells. *Am J Transl Res.* 2016;8(10):4106–4119.
- Qu C, Yan C, Cao W, et al. miR-128-3p contributes to mitochondrial dysfunction and induces apoptosis in glioma cells via targeting pyruvate dehydrogenase kinase 1. *IUBMB Life*. 2020;72(3):465–475. doi:10.1002/iub.2212

- Wang BC, Ma J. Role of microRNAs in malignant glioma. *Chin Med J (Engl)*. 2015;128(9):1238–1244. doi:10.4103/0366-6999.156141
- 20. Yang X, Du H, Bian W, Li Q, Sun H. FOXD3-AS1/miR-128-3p/ LIMK1 axis regulates cervical cancer progression. *Oncol Rep.* 2021;45(5). doi:10.3892/or.2021.8013
- 21. Geng YB, Pan CC, Xu C, et al. Long non-coding RNA LINC00346 regulates proliferation and apoptosis by targeting miR-128-3p/ SZRD1 axis in glioma. *Eur Rev Med Pharmacol Sci.* 2020;24 (18):9581–9590. doi:10.26355/eurrev_202009_23046
- 22. Fu C, Li D, Zhang X, Liu N, Chi G, Jin X. LncRNA PVT1 facilitates tumorigenesis and progression of glioma via regulation of MiR-128-3p/GREM1 axis and BMP signaling pathway. *Neurotherapeutics*. 2018;15(4):1139–1157. doi:10.1007/s13311-018-0649-9
- Peng WX, Koirala P, Mo YY. LncRNA-mediated regulation of cell signaling in cancer. *Oncogene*. 2017;36(41):5661–5667. doi:10.1038/ onc.2017.184
- 24. Zhao X, Li D, Huang D, et al. Risk-associated long noncoding RNA FOXD3-AS1 inhibits neuroblastoma progression by repressing PARP1-mediated activation of CTCF. *Mol Ther.* 2018;26 (3):755–773. doi:10.1016/j.ymthe.2017.12.017
- 25. Preker P, Almvig K, Christensen MS, et al. PROMoter uPstream Transcripts share characteristics with mRNAs and are produced upstream of all three major types of mammalian promoters. *Nucleic Acids Res.* 2011;39(16):7179–7193. doi:10.1093/nar/gkr370
- 26. Guan Y, Bhandari A, Xia E, Yang F, Xiang J, Wang O. IncRNA FOXD3-AS1 is associated with clinical progression and regulates cell migration and invasion in breast cancer. *Cell Biochem Funct*. 2019;37 (4):239–244. doi:10.1002/cbf.3393
- 27. Ma WG, Shi SM, Chen L, Lou G, Feng XL. SP1-induced lncRNA FOXD3-AS1 contributes to tumorigenesis of cervical cancer by modulating the miR-296-5p/HMGA1 pathway. J Cell Biochem. 2021;122(2):235–248. doi:10.1002/jcb.29846
- Wang X, Zhou J, Xu M, et al. A 15-IncRNA signature pred survival and functions as a ceRNA in patients with colorec cancer. *Cancer Manag Res.* 2018;10:5799–5806. doi:10.2147 CMAR.S178732

- 29. Ciafrè SA, Galardi S, Mangiola A, et al. Extensive modulation of a set of microRNAs in primary glioblastoma. *Biochem Biophys Res Commun.* 2005;334(4):1351–1358. doi:10.1016/j.bbrc.2005.07.030
- 30. Zhao L, Li R, Xu S, et al. Tumor suppressor miR-128-3p inhibits metastasis and epithelial-mesenchymal transition by targeting ZEB1 in esophageal squamous-cell cancer. *Acta Biochim Biophys Sin* (Shanghai). 2018;50(2):171–180. doi:10.1093/abbs/gmx132
- 31. Pan J, Zhou C, Zhao X, et al. A two-miRNA signature (miR-33a-5p and miR-128-3p) in whole blood as potential biomarker for early diagnosis of lung cancer. *Sci Rep.* 2018;8(1):16699. doi:10.1038/s41598-018-35139-3
- 32. Liu T, Zhang X, Du L, et al. Exosome-transmitted miR-128-3p increase chemosensitivity of oxaliplatin-resistant colorectal cancer. *Mol Cancer*. 2019;18(1):43. doi:10.1186/s12943-019-0981-7
- Bendahou MA, Ibrahimi A, Boutarbouch March formatics analysis of differentially expressed genes and min AAs in low grade gliomas. *Cancer Inform.* 2020;19:1176 120969692. doi:10.1177/ 1176935120969692
- 34. Kosti A, Barreiro R, Guardiar DA, et a Synergispe of proneurogenic miRNAs provides an ore effective strongy target glioma stem cells. *Cancers (Basel)*, 2021;13(2), 39. doi:10.3390/ cancers13020289
- 35. Zhao N, Zhang J, He M, Sal. SZRD is a novel protein that functions as a potential tunin supressor in cervical cancer. *J Cancer*. 27,7;8(1):2132–2141. doi:10.7150/jca.18806
- 36. Gladitz J, Klink B, Schrt M. Network-based analysis of oligodendroeling predicts november gene candidates within the region of the 1p/19q co-deletion. Acad Neuropathol Commun. 2018;6(1):49. h:10.1186/s40473-018-0544-y
- 37. Gereras J, Handelli R, Nakamura N. Artificial intelligence analysis of the expression data predicted the prognosis of patients with diffus parager cell lymphoma. *Tokai J Exp Clin Med.* 2020;45 (1):37–48.

Cancer Management and Research

Dovepress

Publish your work in this journal

Cancer Management and Research is an international, peer-reviewed open access journal focusing on cancer research and the optimal use of preventative and integrated treatment interventions to achieve improved outcomes, enhanced survival and quality of life for the cancer patient. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit http://www.dovepress.com/testimonials.php to read real quotes from published authors.

Submit your manuscript here: https://www.dovepress.com/cancer-management-and-research-journal