

A study on the preparation and characterization of plasmid DNA and drug-containing magnetic nanoliposomes for the treatment of tumors

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Purpose: To explore the preparation and characterization of a novel nanosized magnetic liposome containing the PEI-As₂O₃/Mn_{0.5}Zn_{0.5}Fe₂O₄ complex.

Methods: Mn_{0.5}Zn_{0.5}Fe₂O₄ and As₂O₃/Mn_{0.5}Zn_{0.5}Fe₂O₄ nanoparticles were prepared by chemical coprecipitation and loaded with PEI. The PEI-As₂O₃/Mn_{0.5}Zn_{0.5}Fe₂O₄ complex was characterized using transmission electron and scanning electron microscopy, X-ray diffraction, energy dispersive spectrometry, and Fourier transform infrared spectroscopy. Cell transfection experiments were performed to evaluate the transfect efficiency. Magnetic nanoliposomes were prepared by rotatory evaporation and their shape, diameter, and thermodynamic characteristics were observed.

Results: Mn_{0.5}Zn_{0.5}Fe₂O₄ and PEI-As₂O₃/Mn_{0.5}Zn_{0.5}Fe₂O₄ nanoparticles were spherical, with an average diameter of 20–40 nm. PEI-As₂O₃/Mn_{0.5}Zn_{0.5}Fe₂O₄ was an appropriate carrier for the delivery of a foreign gene to HepG2 cells. Energy dispersive spectrometry results confirmed the presence of the elements nitrogen and arsenic. Nanoliposomes of approximately 100 nm were observed under a transmission electron microscope. Upon exposure to an alternating magnetic field, they also had good magnetic responsiveness, even though Mn_{0.5}Zn_{0.5}Fe₂O₄ was modified by PEI and encased in liposomes. Temperatures increased to 37°C–54°C depending on different concentrations of PEI-As₂O₃/Mn_{0.5}Zn_{0.5}Fe₂O₄ and remained stable thereafter.

Conclusion: Our results suggest that PEI-As₂O₃/Mn_{0.5}Zn_{0.5}Fe₂O₄ magnetic nanoliposomes are an excellent biomaterial, which has multiple benefits in tumor thermotherapy, gene therapy, and chemotherapy.

Keywords: nanoliposomes, magnetic fluid hyperthermia, As₂O₃, DNA

Introduction

Completion of the human genome project has greatly increased the discovery of functional and disease-related genes. Gene therapy has become one of the most active research areas in biomedicine in the 21st century. One of the main advantages of gene therapy over traditional therapy is the potential to target the expression of therapeutic genes in desired cells or tissues. To achieve targeted gene expression, we developed a novel nonviral gene delivery system.

Arsenic trioxide (As₂O₃) has been adopted from traditional Chinese medicine and used successfully to treat refractory or relapsed acute promyelocytic leukemia.¹ In recent years, some researchers have found that As₂O₃ was not only effective in leukemia treatment, but also effective in inhibiting several animal and human solid cancers, such as neuroblastoma, liver, and gastric cancers.^{2–4} However, there is a lack of drug specificity with the intravenous or oral administration of As₂O₃, resulting in the requirement

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of large doses to achieve high local concentrations. This can cause nonspecific toxicity and other adverse side effects. Moreover, As_2O_3 has been generally considered to be an extremely effective environmental cocarcinogen for some human malignancies, especially for renal, skin, and lung cancer.⁵⁻⁷ Therefore, we investigated an alternative means of administering As_2O_3 to enhance its curative effects and reduce toxicity.

Clinically, the use of thermotherapy, such as heating certain organs or tissues to temperatures between 41°C and 46°C, also known as hyperthermia, has been long-used in the treatment of tumors. With the development of nanotechnology, magnetic nanoparticles have been used for tumor hyperthermia, to absorb energy from an applied high frequency alternating magnetic field (AMF). This treatment is known as magnetic fluid hyperthermia (MFH).⁸ MFH is more effective in the uniform heating of deeply-situated tumors, with relatively good targeting. Generally, in this technique, magnetic fluids are dispersed into the target tissue, which are then heated through the application of an external AMF.⁹

In this paper, we aimed to develop a novel nanosized magnetic liposome containing the PEI- As_2O_3 / $\text{Mn}_{0.5}\text{Zn}_{0.5}\text{Fe}_2\text{O}_4$ (PEI- As_2O_3 /MZF) complex. Herein we also show multiple antitumor functions, including thermotherapy of magnetic MZF nanoparticles, gene therapy of PEI-modified complexes, and chemotherapy using As_2O_3 . With further research, it may develop into a new approach for the treatment of tumors.

Experimental details

Main reagents and cell line

As_2O_3 (Sigma Aldrich, St Louis, MO); PEI (Sigma Aldrich); phosphatidyl choline (Lipoid LLC, Newark, NY); cholesterol (Sigma Aldrich); human liver cancer HepG2 cell lines were provided by the Institute of Biochemistry and Cell Biology, Shanghai Institute of Biological Science, Chinese Academy of Sciences, China.

Methods

Preparation of MZF and PEI- As_2O_3 /MZF nanoparticles

MZF nanoparticles were prepared using chemical coprecipitation. They were examined by transmission electron microscopy (TEM) (H-600; Hitachi, Tokyo, Japan) and scanning electron microscopy (SEM) (JSM-6360LV; Jeol, Tokyo, Japan). The element composition and content of the nanoparticles were analyzed by X-ray diffraction (XRD) and energy dispersive spectrometry (EDS) (Vantage; Thermo

Noran, Middleton, WI). MZF was subsequently modified by As_2O_3 and adsorbed by PEI, according to a previously-described method. The adsorption efficiency of PEI was demonstrated by Fourier transform infrared spectroscopy (FTIR) (560; Thermo Nicolet, Madison, WI) and EDS.

Cell transfection

Prior to transfection, pGFP plasmid DNA and PEI- As_2O_3 /MZF nanoparticles were diluted separately in serum- and supplement-free medium. Subsequently, 6 μg plasmid DNA per well was complexed with various amounts of PEI- As_2O_3 /MZF-NPs (10 mg/mL), corresponding to a nitrogen/phosphorus (N/P) ratio of particles to DNA in a final volume of 400 μL serum- and supplement-free medium. The complexes were then incubated at room temperature for 30 minutes. As a reference,¹⁰ PEI (complexed with DNA at an N/P ratio of 7) or commercially-available transfection reagents, such as Lipofectamine™ (Invitrogen, Carlsbad, CA) were used in accordance with the manufacturer's instructions.

HepG-2 cells were seeded in six well plates, at an initial density of 2×10^5 cells/well in 2 mL growth medium. After incubation for 18 hours, when cells had reached 80% confluency, the medium was replaced with 1.6 mL serum-free media and 400 μL PEI- As_2O_3 /MZF-NPs/pGFP complexes, followed by a further incubation for 4 hours. The medium was then exchanged for fresh medium containing serum

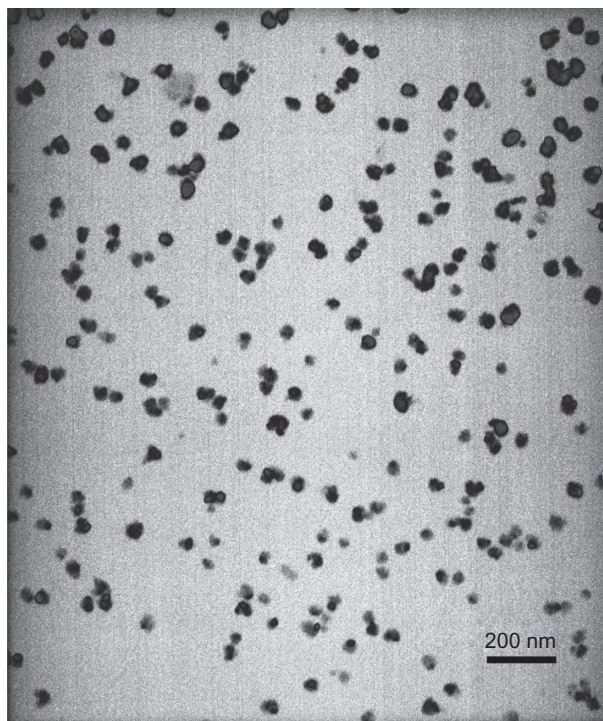


Figure 1 TEM image of $\text{Mn}_{0.5}\text{Zn}_{0.5}\text{Fe}_2\text{O}_4$ nanoparticles.

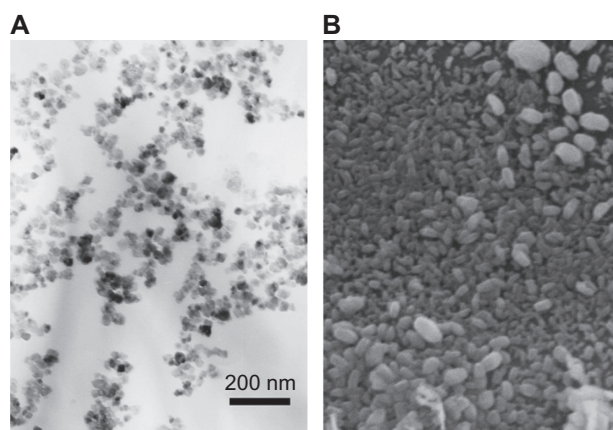


Figure 2 PEI-As₂O₃/MZF as seen by TEM (A) and SEM (B).

Abbreviations: PEI, polyethyleneimine; TEM, transmission electron microscopy; SEM, scanning electron microscopy.

and incubated further. Cells were observed using inverted fluorescence microscopy at 24 hours and 48 hours post-transfection, to evaluate transfection efficiency.

Preparation of nanoliposomes, containing PEI-As₂O₃/MZF and heating tests

Magnetic nanoliposomes were obtained by the combined methods of rotatory film and high-pressure homogenization. A heat test was performed to detect the thermodynamic

characteristics of the magnetic nanoliposomes. Various doses of nanoliposomes were decentralized in 0.9% sodium chloride, with concentrations of PEI-As₂O₃/MZF as follows: 5, 10, 15, 20, 30 g/L. The output electric current was 300A and the fluid was heated for 1 hour with temperature measurements every 5 minutes.

Results and discussion

Characterization of MZF and PEI-As₂O₃/MZF nanoparticles

MZF nanoparticles were evaluated by TEM, XRD, and EDS. Using TEM, MZF particles appeared round or elliptical in shape, with good dispersion and were 20–30 nm in diameter (Figure 1). These nanoparticles were identified as Mn_{0.5}Zn_{0.5}Fe₂O₄ by XRD and EDS.

Figures 2A and 2B show PEI-As₂O₃/MZF nanoparticles under TEM and SEM. The nanoparticles were brown spherical particles, with diameters ranging from 30 to 40 nm. Furthermore, they suspended stably in water, with good diffusibility. FTIR showed specific peaks at 1,471 cm⁻¹, 2,826 cm⁻¹, and 2,954 cm⁻¹, as compared to As₂O₃/MZF complexes (Figure 3). The EDS results verified the presence of the elements manganese, zinc, iron, arsenic, and

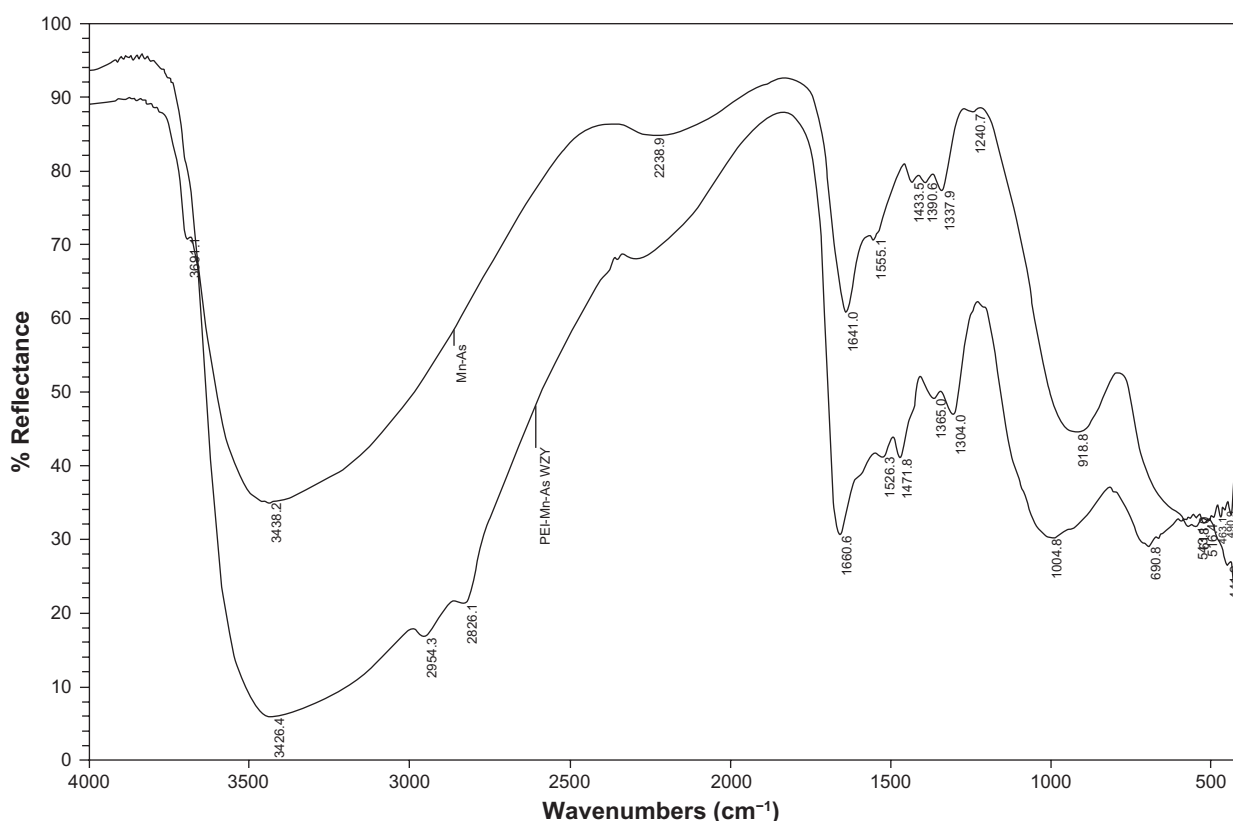


Figure 3 FTIR of PEI-As₂O₃/MZF: the top curve is As₂O₃/MZF and the bottom is PEI-As₂O₃/MZF.

Abbreviations: FTIR, Fourier transform infrared spectroscopy; PEI, polyethyleneimine.

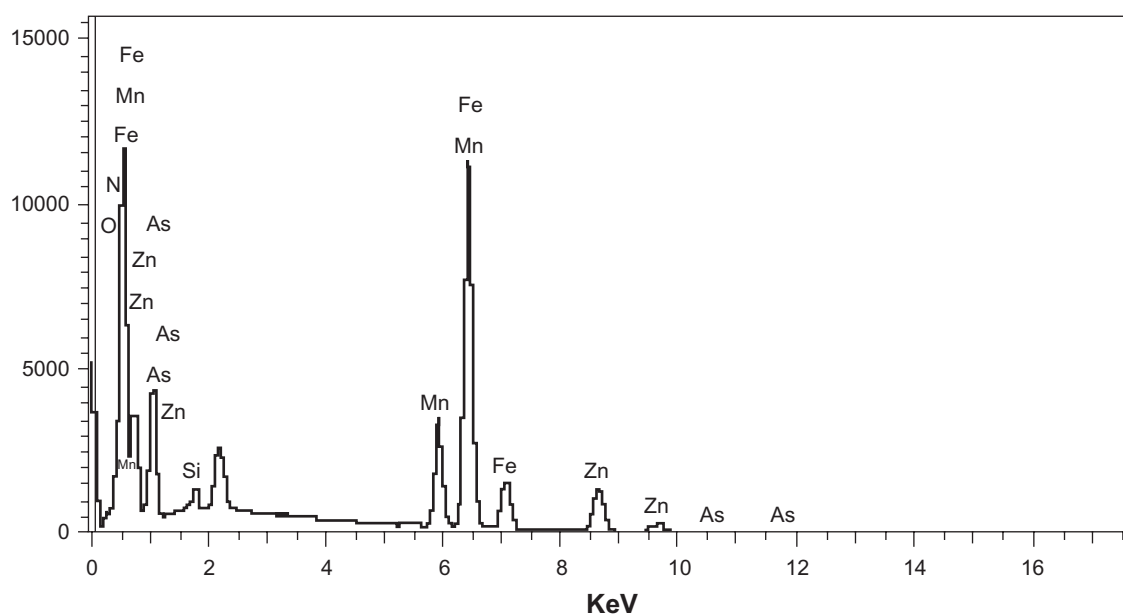


Figure 4 EDS results of PEI-As₂O₃/MZF.

Abbreviations: EDS, energy dispersive spectrometry.

nitrogen (Figure 4). Further cell transfection experiments suggested that PEI-As₂O₃/MZF nanoparticles were efficient carriers for gene transfection. The observed green cellular fluorescence was due to pGFP expression, which was confirmed by inverted fluorescence microscopy 24 hours post-transfection. The expression of GFP was much clearer at 48 hours post-transfection (Figure 5). This high transfer efficiency was due to the inherent proton absorption properties of PEI.¹¹ However, the use of PEI as a gene carrier is limited by cytotoxicity and nonspecific interactions with serum proteins. In our study, we used nanoliposomes to engulf the PEI-As₂O₃/MZF complexes and reduce their toxic side effects, which has not been previously reported.

Properties of nanoliposomes containing PEI-As₂O₃/MZF

We prepared magnetic nanoliposomes containing PEI-As₂O₃/MZF complexes, with an average diameter of

approximately 100 nm. The encapsulation ratio of As₂O₃ was analyzed by fluorescence spectrophotometry and determined to be 75.33%. An in vitro heating test was performed to detect the thermodynamic characteristics of the nanoliposomes. The temperature of the nanoliposomes exposed to AMF rose rapidly within the first 5 minutes and slowly continued to increase from 5–40 minutes, remaining stable thereafter. The temperature rose from 37°C to 54°C, depending on various doses of PEI-As₂O₃/MZF (Figure 6). The results showed that nanoliposomes had good absorption properties in high-frequency alternating electromagnetic fields, due to the presence of magnetic nanoparticles. The modified PEI, As₂O₃, and lipid membrane did not influence the magnetic responsiveness of MZF. Therefore, we can select a suitable temperature range (42°C–46°C) for tumor hyperthermia by adjusting the concentration of PEI-As₂O₃/MZF.

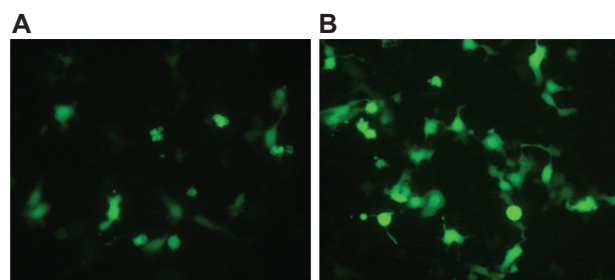


Figure 5 HepG2 cells observed under fluorescence microscopy (×400) 24 hours after transfection (A) and 48 hours after transfection (B).

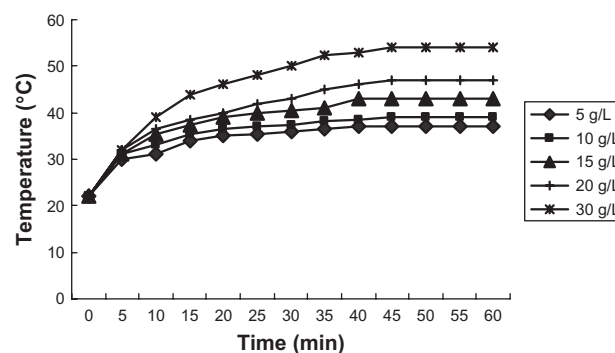


Figure 6 Heating test result of magnetic nanoliposomes.

Conclusion

Nanotechnology has been increasingly applied in the medical field, with potential treatments for cancer. In our study, we prepared a new kind of magnetic nanoliposome containing PEI-As₂O₃/MZF complexes. It has multiple antitumor applications, including thermotherapy using magnetic MZF nanoparticles, gene therapy with PEI-modified complexes, and chemotherapy using As₂O₃. In future studies, we will apply this novel material to tumor therapy.

Acknowledgments

This work was supported by the Chinese National 863 Plan (No: 2007 AA03Z356), and the Chinese National Natural Science Foundation (No: 30770584).

Disclosure

The authors declare no conflicts of interest with respect to the authorship and/or publication of this article.

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