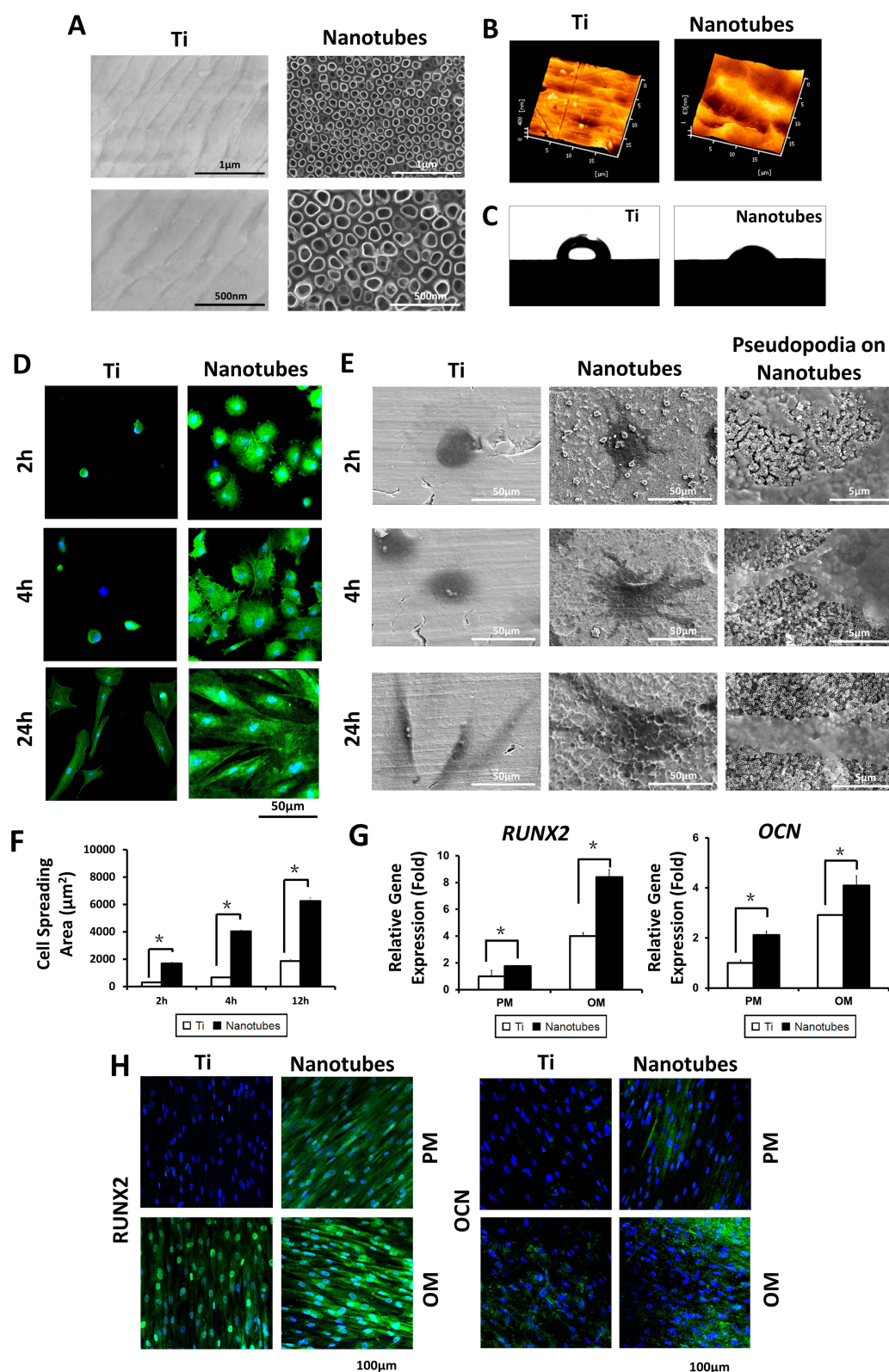


## The Epigenetic Mechanisms of Nanotopography-Guided Osteogenic Differentiation of Mesenchymal Stem Cells via High-Throughput Transcriptome Sequencing [Corrigendum]

Lv L, Liu Y, Zhang P, et al. *Int J Nanomedicine*. 2018;13:5605–5623.

It was brought to the authors' attention that FITC and OCN images had been misused with images from their previous *Biomaterials* article (2015; 39: 193–205). The authors reviewed their original data and found a mistake had occurred during the assembly of the FITC and OCN images in Figure 1D and H on pages 5609 and 5610. The immunofluorescence experiments, from which the FITC and OCN images were obtained, were performed together with the study published in their 2015 *Biomaterials* article. The authors intended to further describe the underlying mechanism of why nanotubes were much better in osteogenesis than smooth Ti surface by RNA sequencing and high-throughput analyses based on the results of their earlier study. The purpose of Figure 1 was to provide an overview of how 70nm nanotubes were better than smooth Ti surface in cell adhesion and osteogenic ability but images from the wrong treatment groups were selected in error. The authors had retained all original data and were able to make corrections without impacting the results and conclusions of the article. The authors apologize for any inconvenience caused.

The correct Figure 1 is as follows.



**Figure 1** Surface characterization, cell morphology, and osteoinductive ability of TiO<sub>2</sub> nanotubes.

**Notes:** (A) Scanning electron microscopic (SEM) observation of TiO<sub>2</sub> nanotubes and smooth Ti surfaces. (B) Atomic force microscopy (AFM) images of TiO<sub>2</sub> nanotubes and Ti surfaces. (C) Photographs of contact-angle measurement of water. (D) Confocal micrographs of human adipose-tissue-derived stem cells (hASCs) on TiO<sub>2</sub> nanotubes and Ti surfaces after 2, 4, and 24 hours of culture. (E) SEM observation of cell morphology on TiO<sub>2</sub> nanotubes and Ti surfaces after 2, 4, and 24 hours of culture. (F) Cell spreading area on TiO<sub>2</sub> nanotubes and Ti surfaces after 2, 4, and 24 hours of culture. (G) Gene expression of osteogenic-related genes quantified by quantitative real-time PCR. \*P < 0.05. (H) Immunofluorescent staining of RUNX2 and osteocalcin (OCN) in hASCs cultured on TiO<sub>2</sub> nanotubes and Ti surfaces after 7 days of osteoinduction. OCN and RUNX2 are colored green, while nuclei are blue.

**Abbreviations:** OM, osteoinduction medium; PM, proliferation medium.

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